

NIH Public Access

Author Manuscript

J Am Chem Soc. Author manuscript; available in PMC 2010 September 23.

Published in final edited form as:

J Am Chem Soc. 2009 September 23; 131(37): 13232–13233. doi:10.1021/ja905125b.

Intelligent medical diagnostics via molecular logic

Tania Konry and **David R. Walt**

Department of Chemistry, Tufts University, 62 Talbot Avenue, Medford, MA 02155, USA

Abstract

In this communication, we describe the integration of microarray sensor technology with logic capability for screening combinations of proteins and DNA in a biological sample. In this system, we have demonstrated the use of a single platform amenable to both protein detection and protein-DNA detection using molecular logic gates. The pattern of protein and DNA inputs results in fluorescence outputs according to a truth table for AND and INHIBIT gates, thereby demonstrating the feasibility of performing medical diagnostics using a logic gate design. One possible application of this technique would be for the direct screening of various medical conditions that are dependent on combinations of diagnostic markers.

> Molecular logic gates and molecular computational systems have used a variety of recognition mechanisms including proteins, ¹⁻⁵ biochemical pathways in living cells, ⁶⁻¹⁵ and DNA¹⁶⁻³¹. Molecular logic and computation may also be applied to medical diagnostics. For example, if abnormal results were detected during medical examination, they could be interpreted using Boolean logic resulting in intelligent diagnostics. In this communication, we report the integration of microarray sensor technology with logic capability for screening combinations of proteins and DNA in a biological sample. In this system, the reporter and receptor molecules perform simple logic operations by coupling multiple molecular recognition inputs to a fluorescence signal output.

> Previously developed molecular AND gates by the de Silva group allowed the creation of a "lab on a molecule" prototype, which responded to inputs of the electrolytes $Na⁺$, $H⁺$, and Zn^{2+} in water leading to an enhanced fluorescence signal as output.²⁸ In addition, the NOTIF logic function was previously demonstrated in designed synthetic peptide networks that mimic some basic logic functions of more complex biological networks.³² In this communication, we demonstrate both AND and INHIBIT (NOTIF) logic gates that respond to the presence of both protein and DNA in a sample. Such a system potentially could be used for performing smart diagnostics. For instance, increased airway obstruction in patients with chronic obstructive pulmonary disease (COPD) or bronchial asthma is frequently associated with bacterial respiratory infections, especially Haemophilus influenzae (H. influenzae), and is accompanied by the secretion of proinflammatory cytokines, such as IL-8 protein.³²⁻³⁷ It is possible to describe the resulting logic network of bacterial DNA and IL-8 protein using a Boolean operator truth table (Figure 1A).

> Our system is designed to determine if both a protein and a nucleic acid sequence are present in a sample, or if only protein is present. A schematic illustration of the protein and protein-DNA sensors is shown in Figures 2A and B, respectively. The fiberoptic microarray was prepared by loading monoclonal antibody (MAb)-functionalized microspheres into microwells created by selectively etching the distal end of optical fiber bundles. ³⁸ Every microsphere on the array is encoded with a unique optical barcode comprised of fluorescent dye incorporated

david.walt@tufts.edu.

into each microsphere. The identity of each microsphere in the random array is then determined using image-processing techniques to allow positional registration of the entire array (Figure 3 A, B (1)). MAb-functionalized microspheres were used as receptors for capturing protein on the surface followed by binding of a secondary antibody labeled with DNA (capture probe) via a biotin-avidin bridge (described in detail in the Supporting Information).

To use this platform for both protein detection and protein-DNA detection, two fluorescent probes were designed for the detection step. If both DNA and protein are present in the sample, the Cy3-labeled signal probes hybridize to the complementary target DNA sequence (Figure 2A) while the remaining secondary antibodies free of target DNA hybridize to Cy5-signal probes (Figure 2B). No Cy5 fluorescence is obtained when the capture probe is saturated with a high concentration of target DNA (50 nM) (Figure 2D $(1,1)$). The detection of the specific DNA sequence in the designed system is possible only if the protein is also present. Therefore, Cy3 signal indicates the detection of both target DNA and protein. Conversely, if target DNA is not present in the sample solution, only the Cy5-labeled signal probes hybridize to the secondary antibody labeled with complementary sequence (Figure 2B).

This biochemical system can be described as a simple functional model of AND and INHIBIT (INH) logic gates (Figure 1B). The INHIBIT (INH) function can be interpreted as a particular integration of AND and NOT logic functions, where the output signal is inhibited by one of the active inputs. Therefore, the INH logic gate presented here queries protein presence or absence in a sample. If only protein is present in the sample (input $(0,1)$), the gate switches on and a Cy5 fluorescence emission is produced (output 1) (Figure 3A (2)). If protein is not present in the sample (inputs $(0,0)$; $(1,0)$), then the fluorescent output is "inhibited"; consequently, no Cy5 fluorescence or Cy3 is produced (output 0). AND logic is represented by the situation where the output of an AND gate occurs only if both inputs are present. The AND logic gate presented here queries two biological species (protein and DNA) in a sample to determine in a single test if they are both present. Experiments were performed by testing the AND gate sensor with all four possible input combinations of protein and DNA (Figure 2C). In the presence of both protein and DNA inputs in a sample (1,1), the AND gate switches on and a Cy3 fluorescence emission is produced (output 1) (Figure 3B (2)). No Cy5 fluorescence is obtained for (1,1) since the capture probe is saturated with a high concentration of target DNA (50 nM) (Figure 2D (1,1)). If one $(0,1; 1,0)$ or neither $(0,0)$ of the analytes is present in the sample, no Cy3 fluorescence is produced (output 0). In our system, the absence of input protein has the power to disable the entire system regardless of the presence of the other input—DNA.

In summary, we have demonstrated the use of a single platform amenable to both protein detection and protein-DNA detection using molecular logic gates. The pattern of protein and DNA inputs to fluorescence outputs executed according to the truth table for AND and INHIBIT gates demonstrates the feasibility of performing medical diagnostics using a logic gate design. One possible application of this technique would be for the direct screening of various medical conditions that are dependent on combinations of diagnostic markers.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

ACKNOWLEDGMENT

We thank the National Institute of Dental and Craniofacial Research (NIDCR) for their support of this work (Grant U01 DE017788). We further thank Dr. Timothy M. Blicharz, Dr. Christopher LaFratta, and Ryan B. Hayman from the Department of Chemistry at Tufts for their technical assistance.

J Am Chem Soc. Author manuscript; available in PMC 2010 September 23.

REFERENCES

- (1). Hjelmfelt A, Weinberger ED, Ross. J. Proc. Natl. Acad. Sci. U.S.A 1991;88:10983–10987.
- (2). Hjelmfelt A, Weinberger ED, Ross. J. Proc. Natl. Acad. Sci. U.S.A 1992;89:383–387.
- (3). Dueber JE, Yeh BJ, Chak K, Lim WA. Science 2003;301:1904–1908. [PubMed: 14512628]
- (4). Sivan S, Tuchman S, Lotan N. BioSystems 2003;70:21–33. [PubMed: 12753934]
- (5). Muramatsu S, Kinbara K, Taguchi H, Ishii N, Aida T. J. Am. Chem. Soc 2006;128(11):3764–3769. [PubMed: 16536551]
- (6). Margulies, David; Melman, Galina; Shanzer, Abraham. J. Am. Chem. Soc 2006;128(14):4865–4871. [PubMed: 16594723]
- (7). Gardner T, Cantor C, Collins J. Nature 2000;403:339–342. [PubMed: 10659857]
- (8). Guet C, Elowitz M, Hsing W, Leibler S. Science 2002;296:1466–1470. [PubMed: 12029133]
- (9). Hasty J, McMillen D, Collins JJ. Nature 2002;420:224–230. [PubMed: 12432407]
- (10). Atkinson Mariette R, Savageau Michael A, Myers Jesse T, Ninfa A. J. Cell 2003;113:597–607.
- (11). Raymo FM. Adv. Mater 2002;14:401–414.
- (12). De Silva AP, McClenaghan ND. Chem.Eur. J 2004;10:574–586.
- (13). Conrad, M.; Zauner, KP.; Sienko, T.; Adamatzky, A.; Rambidi, N. Molecular Computing. MIT Press; 2003. p. 1-31.chapter 1
- (14). Strack G, Ornatska M, Pita M, Katz E. J. Am. Chem. Soc 2008;130(13):4234–4235. [PubMed: 18321113]
- (15). Nurse P. Nature 2008;454:424–426. [PubMed: 18650911]
- (16). Adleman LM. Science 1994;266:1021–1024. [PubMed: 7973651]
- (17). Lipton RJ. Science 1995;268:542–545. [PubMed: 7725098]
- (18). Guarnieri F, Fliss M, Bancroft C. Science 1996;273:220–223. [PubMed: 8662501]
- (19). Ouyang Q, Kaplan PD, Liu S, Libchaber A. Science 1997;278:446–449. [PubMed: 9334300]
- (20). Faulhammer D, Cukras AR, Lipton RJ, Landweber LF. Proc. *Natl. Acad. Sci. U.S.A.* 2000;97:1385– 1389. [PubMed: 10677471]
- (21). Liu Q, Wang L, Frutos AG, Condon AE, Corn RM, Smith LM. Nature 2000;403:175–179. [PubMed: 10646598]
- (22). Mao C, LaBean TH, Reif JH, Seeman NC. Nature 2000;407:493–496. [PubMed: 11028996]
- (23). Benenson Y, Paz-Elizur T, Adar R, Keinan E, Livneh Z, Shapiro E. Nature 2001;414:430–434. [PubMed: 11719800]
- (24). Carbone A, Seeman NC. Proc. Natl. Acad. Sci. U.S.A 2002;99:12577–12582. [PubMed: 12232051]
- (25). Stojanovic MN, Stefanovic D. J. Am. Chem. Soc 2003;125:6673–6676. [PubMed: 12769576]
- (26). Saghatelian A, Voelcker NH, Guckian KM, Lin VSY, Ghadiri MR. J. Am. Chem. Soc 2003;125:346–347. [PubMed: 12517142]
- (27). Stojanovic MN, Stefanovic D. Nat. Biotechnol 2003;21:1069–1074. [PubMed: 12923549]
- (28). Yan H, Feng L, LaBean TH, Reif JH. J. Am. Chem. Soc 2003;125:14246–14247. [PubMed: 14624551]
- (29). Voelcker NH, Guckian KM, Saghatelian A, Ghadiri RM. Small 2008;28(4):427–431. [PubMed: 18350554]
- (30). Cockroft SL, Chu J, Amorin M, Ghadiri MRJ. Am. Chem. Soc 2008;130(3):818–820.
- (31). Winfree, Erik. ACM 2008;42(2):1–1.
- (32). Ashkenasy G, Ghadiri RM. J. Am. Chem. Soc 2004;126:11140–11141. [PubMed: 15355081]
- (33). Rutmam A, Dowling R, Wills P, Feldman C, Cole PJ, Wilson R. Antimicrobial agents and chemotherapy 1998:772–778. [PubMed: 9559781]
- (34). Khair OA, Devalia JL, Abdelaziz MM, Sapsford RJ, Davies RJ. Eur Respir J 1995;8:1451–1457. [PubMed: 8575568]
- (35). Tong HH, Chen Y, James M, Van Deusen J, Welling DB, Demaria TF. Infection and Immunity 2001:3678–3684. [PubMed: 11349030]
- (36). Chung KF. Eur. Respir. J 2001;18(34):50–59.

J Am Chem Soc. Author manuscript; available in PMC 2010 September 23.

- (37). Pauwels R, Verschraegen G, Van Der Straeten M. Allergy 1980;35(8):665–669. [PubMed: 6971585]
- (38). LaFratta CN, Walt DR. Chem. Rev 2008;108(2):614–637. [PubMed: 18229955]

J Am Chem Soc. Author manuscript; available in PMC 2010 September 23.

Konry and Walt Page 5

А. Input 1 Input 2 Outlet Outlet Outlet OR gate (Target DNA) (Protein) **INH** gate AND gate $\overline{0}$ $\overline{1}$ $\mathbf{1}$ $\mathbf{1}$ $\mathbf{1}$ $\overline{\mathbf{0}}$ $\overline{\mathbf{0}}$ $\overline{\mathbf{0}}$ $\overline{\mathbf{0}}$ $\overline{1}$ $\overline{\mathbf{0}}$ $\overline{1}$ $\overline{\mathbf{0}}$ $\overline{1}$ $\overline{1}$ \mathbf{B} . $\mathbb{N} \mathbb{H}$ Target DNA IL-8 protein $\mbox{\rm AND}$

(A) Truth table for logic gates. (B) AND and INHIBIT logic gates.

Figure 2.

Schematic illustration for (A) protein-DNA detection and (B) protein detection. Output averaged fluorescence intensities in the absence (0) or in the presence (1) (protein (1) = 10 nM, Target DNA (1) = 50 nM) of different inputs; (C) Cy3 signal (D) Cy5 signal.

Figure 3.

Fluorescence micrographs of a small section of a fiber-optic microarray (A, B) (1)) Positional registration of the entire Europium (Eu)-dye encoded microsphere array. (A (2)) Cy5 microarray signal image of IL8-protein (10 nM) and Target DNA (0 nM). (B (2)) Cy3 microarray signal image of IL8-protein (10 nM) and Target DNA (50 nM). Note the correspondence between the encoded microspheres with the Eu-dye and the microspheres showing fluorescence in the signal images.