



Published in final edited form as:

Fertil Steril. 2008 February ; 89(2 Suppl): e101–e102. doi:10.1016/j.fertnstert.2007.12.039.

Neuroendocrine systems as targets for environmental endocrine-disrupting chemicals

Andrea C. Gore

The University of Texas at Austin, Division of Pharmacology & Toxicology, College of Pharmacy, Austin, TX 78712, USA

Keywords

endocrine disruption; neuroendocrine; hypothalamus; GnRH; polychlorinated biphenyl; PCB; reproduction

Incontrovertible scientific evidence demonstrates that environmental endocrine-disrupting chemicals (EDCs) affect reproductive function (1). Nevertheless, the mechanisms for these effects remain elusive, in part because the control of reproductive physiology is so complex. The reproductive axis of all vertebrates comprises three interdependent levels of organization: the brain (specifically the hypothalamus), the anterior pituitary gland, and the gonad (ovary or testis). Each of these levels produces unique hormones, and each level also responds to the hormones produced by the other levels. Recent studies demonstrate that the three reproductive levels are also responsive to environmental EDCs. Here, I will discuss the compelling evidence that the neuroendocrine level of the reproductive axis, namely the hypothalamus, is a target for endocrine disruption.

Reproductive neuroendocrine function is driven by about 1000 neurons in the brain that synthesize and secrete a ten-amino acid peptide, gonadotropin-releasing hormone (GnRH) (2). GnRH release drives reproduction throughout the life cycle and this is the primary stimulus to the rest of the reproductive axis. Using a GnRH cell line, the GT1-7 cells (3), we found that polychlorinated biphenyls (PCBs) (4) and organochlorine pesticides such as methoxychlor and chlorpyrifos (5) altered GnRH gene expression, GnRH peptide release, and the morphology of the GT1-7 cells. Animal models also identify GnRH neurons as targets for both natural environmental estrogens such as coumestrol (6) and for industrial contaminants and pesticides (7). Interestingly, some of these studies reveal non-linear dose-response curves that are typical of hormonally active substances (8,9).

The hypothalamus is abundant in neurotransmitter and sex steroid hormone receptors that, together with GnRH neurons, form a neural network controlling reproductive physiology and behavior (10–12). Direct effects of EDCs have been shown on hypothalamic neurotransmitter systems, on the size of specific hypothalamic regions, as well as on numbers of cells expressing the estrogen receptor beta (13–15). These actions may underlie observations that prenatal exposure to low-doses of PCBs (16), or postnatal treatment with soy (17), significantly affected

Address correspondence to: Andrea C. Gore, PhD, The University of Texas at Austin, 1 University Station, A1915, Austin, TX 78712, Phone: 512-471-3669, Fax: 512-471-5002, andrea.gore@mail.utexas.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

aspects of mating behavior in female rats. These studies demonstrate a functional outcome for effects of EDCs in the neuroendocrine hypothalamus.

In conclusion, GnRH cells, and their regulatory neural network, are targets of EDCs. As a consequence, exposure to EDCs, particularly during critical developmental windows, results in impaired reproductive physiology and behavior. Importantly, the reproductive axis is only one of five neuroendocrine systems, the others being thyroid/metabolism, growth, stress, and lactation. These five neuroendocrine systems all play regulatory and homeostatic roles in the control of vertebrate physiology. In addition, there is cross-talk among these systems because the function of one neuroendocrine system impacts the functions of all the others. While this may increase the complexity of studying effects of EDCs it is also an important consideration for how an EDC may exert complex and diverse effects on the exposed individual. There is considerable need for additional research on effects of EDCs in neuroendocrine systems in order to better understand how the environment regulates basic physiologic processes.

References

1. Gore, AC. Endocrine-disrupting chemicals: From basic research to clinical practice. Totowa, NJ: Humana Press; 2007.
2. Gore, AC. GnRH: The Master Molecule of Reproduction. Norwell, MA: Kluwer Academic Publishers; 2002.
3. Mellon PL, Windle JJ, Goldsmith PC, Padula CA, Roberts JL, Weiner RI. Immortalization of hypothalamic GnRH neurons by genetically targeted tumorigenesis. *Neuron* 1990;5:1–10. [PubMed: 2196069]
4. Gore AC, Wu TJ, Oung T, Lee JB, Woller MJ. A novel mechanism for endocrine-disrupting effects of polychlorinated biphenyls: Direct effects on gonadotropin-releasing hormone (GnRH) neurons. *J Neuroendocrinol* 2002;14:814–823. [PubMed: 12372006]
5. Gore AC. Organochlorine pesticides directly regulate gonadotropin-releasing hormone gene expression and biosynthesis in the GT1-7 hypothalamic cell line. *Mol Cell Endocrinol* 2002;192:157–170. [PubMed: 12088877]
6. McGarvey C, Cates PS, Brooks NA, Swanson IA, Milligan SR, Coen CW, et al. Phytoestrogens and gonadotropin-releasing hormone pulse generator activity and pituitary luteinizing hormone release in the rat. *Endocrinology* 2001;142:1202–1208. [PubMed: 11181536]
7. Gore AC. Environmental toxicant effects on neuroendocrine function. *Endocrine* 2001;14:235–246. [PubMed: 11394642]
8. Cook R, Calabrese EJ. The importance of hormesis to public health. *Environ Health Perspect* 2006;114:1631–1635. [PubMed: 17107845]
9. Gore AC, Heindel JJ, Zoeller RT. Endocrine disruption for endocrinologists (and others). *Endocrinology* 2006;147:S1–3. [PubMed: 16690798]
10. Simerly RB, Chang C, Muramatsu M, Swanson LW. Distribution of androgen and estrogen receptor mRNA-containing cells in the rat brain: An in situ hybridization study. *J Comp Neurol* 1990;294:76–95. [PubMed: 2324335]
11. Quadros PS, Pfau JL, Goldstein AY, DeVries GJ, Wagner CK. Sex differences in progesterone receptor expression: a potential mechanism for estradiol-mediated sexual differentiation. *Endocrinology* 2002;143:3727–3739. [PubMed: 12239082]
12. Chakraborty TR, Gore AC. Aging-related changes in ovarian hormones, their receptors, and neuroendocrine function. *Exp Biol Med* 2004;229:977–987.
13. Shibutani M, Masutomi N, Uneyama C, Abe N, Takagi H, Lee KY, et al. Down-regulation of GAT-1 mRNA expression in the microdissected hypothalamic medial preoptic area of rat offspring exposed maternally to ethinylestradiol. *Toxicology* 2005;208:35–48. [PubMed: 15664431]
14. Patisaul HB, Fortino AE, Polston EK. Differential disruption of nuclear volume and neuronal phenotype in the preoptic area by neonatal exposure to genistein and bisphenol-A. *Neurotoxicology* 2007;28:1–12. [PubMed: 17109964]

15. Salama J, Chakraborty TR, Ng L, Gore AC. Effects of polychlorinated biphenyls (PCBs) on female reproductive development and estrogen receptor b expression. *Environ Health Perspect* 2003;111:1278–1282. [PubMed: 12896846]
16. Steinberg RM, Juenger TE, Gore AC. The effects of prenatal PCBs on adult female paced mating reproductive behaviors in rats. *Horm Behav*. 2007Epub ahead of print
17. Patisaul HB, Luskin JR, Wilson ME. A soy supplement and tamoxifen inhibit sexual behavior in female rats. *Horm Behav* 2004;45:270–277. [PubMed: 15053943]