



HHS Public Access

Author manuscript

Immunol Cell Biol. Author manuscript; available in PMC 2010 April 01.

Published in final edited form as:

Immunol Cell Biol. 2009 October ; 87(7): 546–553. doi:10.1038/icb.2009.38.

Extraction and characterization of the rhesus macaque T cell receptor β -chain genes

Hui Yee Greenaway¹, Monica Kurniawan¹, David A Price^{2,3}, Daniel C Douek³, Miles P Davenport¹, and Vanessa Venturi^{1,*}

¹Complex Systems in Biology Group, Centre for Vascular Research, University of New South Wales, Kensington NSW 2052, Australia

²Department of Medical Biochemistry and Immunology, Cardiff University School of Medicine, Cardiff CF14 4XN, UK

³Human Immunology Section, Vaccine Research Center, NIAID/NIH, Bethesda MD 20892, USA

Abstract

Rhesus macaque models have been instrumental for the development and testing of vaccines prior to human studies and have provided fundamental insights into the determinants of immune efficacy in a variety of infectious diseases. However, the characterization of antigen-specific T cell receptor (TCR) repertoires during adaptive immune responses in these models has previously relied on human TCR gene assignments. Here, we extracted and characterized TCR β -chain (TRB) genes from the recently sequenced rhesus macaque genome that are homologous to the human TRB genes. Comparison of the rhesus macaque TRB genes with the human TRB genes revealed an average best-match similarity of 92.9%. Furthermore, we confirmed the usage of most rhesus macaque TRB genes by expressed TCR β sequences within epitope-specific TCR repertoires. This primary description of the rhesus macaque TRB genes will provide a standardized nomenclature and enable better characterization of TCR usage in studies that utilize this species.

Keywords

T cell; T cell receptor; T cell receptor repertoire

INTRODUCTION

The rhesus macaque is widely used as a non-human primate model to study infection and immunity due to the close genetic relationship with humans (~93% average human-macaque sequence identity¹) and the homology between human and rhesus pathogen genomes^{2, 3}. Indeed, rhesus macaques have been used to study fundamental aspects of immunology, including the development and maintenance of T cell memory⁴, immunodominance⁵ and the aging immune system⁶. There have also been many studies of

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:http://www.nature.com/authors/editorial_policies/license.html#terms

*Address correspondence and reprint requests to Dr Vanessa Venturi, Complex Systems in Biology Group, Centre for Vascular Research, University of New South Wales, Kensington NSW 2052, Australia. E-mail address: v.venturi@unsw.edu.au.

immune responses in rhesus macaque models of human infections such as human immunodeficiency virus (HIV)⁷, influenza virus^{8, 9}, tuberculosis¹⁰, Epstein-Barr virus (EBV)^{11, 12}, cytomegalovirus (CMV)^{4, 13–15}, smallpox¹⁶, measles¹⁷ and severe acute respiratory syndrome (SARS)¹⁸. Furthermore, rhesus macaques have been instrumental in the design and testing of vaccines against infections such as HIV¹⁹ and smallpox ¹⁶.

The various roles of T lymphocytes in adaptive immune responses to infection, which include the provision of helper functions to other immune cells and cytolytic control of infected cells, require that T cell populations recognize a large variety of foreign peptides bound to major histocompatibility complex (MHC) molecules. This recognition is facilitated by a diverse repertoire of T cell receptors (TCRs). The TCR repertoires that respond to different peptide-MHC epitopes can vary greatly. Indeed, diversity estimates range from ~10 to >1000 different TCRs responding to a specific epitope^{20–23}. Moreover, some epitope-specific TCR repertoires can feature biased usage of TCR V β (TRBV) or J β (TRBJ) genes, or distinct patterns of amino acid usage within the third complementarity-determining region (CDR3)²⁴. Studies of the TCR repertoire can provide valuable information about the molecular evolution of an immune response and the factors that shape clonotype selection *in vivo*²⁵. Furthermore, it is becoming increasingly apparent that the clonotypic structure of an epitope-specific T cell response can have important implications for the immune control of some viral infections. For example, one issue of current debate that has important consequences for the rational design of immunotherapeutic and vaccination strategies^{24, 26} is whether a restricted TCR repertoire responding to a highly variable pathogen could be associated with the emergence of viral mutants that escape T cell recognition at this epitope^{27–31}.

Many studies of T cell immunity in rhesus macaque models of infection have utilized TCR repertoire data to gain additional insights^{5, 14, 30, 32–43}. In particular, a large number of studies have characterized the TCR repertoires of target CD4⁺ T cell populations or CD8⁺ T cell populations involved in the control of simian immunodeficiency virus (SIV) in rhesus macaques^{5, 30, 32–39, 41–43}. Most of these studies have relied on human TCR gene homology to identify V and J gene usage. Although the rhesus macaque TCR D β (TRBD) and TRBJ genes have previously been sequenced⁴⁴, the TRBV genes were not previously available. Here, we present the TRBV, TRBD and TRBJ genes extracted from the rhesus macaque genome¹ on the basis of their homology with the human TRB genes. In addition, we demonstrate extracted TRB gene usage in expressed TCR β sequences by using an existing database of 7218 TCR β sequences involved in CD8⁺ T cell responses specific for the immunodominant Mamu-A*01-restricted SL8/TL8 (S/TTPESANL; Tat, residues 28–35) and CM9 (CTPYDINQM; Gag, residues 181–189) epitopes derived from SIV^{30, 45}. The TRB genes extracted from the rhesus macaque genome will enable more accurate characterization of rhesus macaque TCR β repertoires.

RESULTS

Rhesus macaque TRBV genes

A total of 72 TRBV genes were extracted from chromosome 3 of the rhesus macaque genome (Table 1 and Rhesus_macaque_TRBV.fsa in Supporting Information). The human

TRBV gene corresponding most precisely to each rhesus macaque TRBV gene was identified on the basis of the highest percentage match between the nucleotide sequences for the TRBV genes (i.e. V-GENE in the IMGT standardized labels). The percent similarity between the nucleotide sequences for the rhesus macaque and the best-match human TRBV genes ranged between 78.3% and 96.5%, with an average similarity of 92.2%. We could not identify a one-to-one correspondence between all rhesus macaque and human TRBV genes (Figure 1). In many cases, one human TRBV gene was found to be the best match to more than one of the TRBV genes extracted from the rhesus macaque genome. For example, the human TRBV6-5 gene had the highest percent similarity of all human TRBV genes to five of the rhesus macaque TRBV genes; in contrast, the human TRBV6-6 gene was not the best match to any of the rhesus macaque TRBV genes. For five of the 72 TRBV genes, only partial sequences were available from the rhesus macaque genome (Table S1 in Supporting Information) and only two of these partial TRBV genes were incomplete at the 3' end, which would influence their use in analysis of the CDR3. The human TRBV17 subgroup, consisting of just one gene, was the only one for which no corresponding TRB gene was found in the rhesus macaque genome (using a cutoff of 75% similarity).

We also compared the TRBV exons (i.e. L-PART1+V-EXON in the IMGT standardized labels) between the rhesus macaque and best-match human TRBV genes (Table 1). The percent identities between the nucleotide sequences for the rhesus macaque and human TRBV exons ranged between 72.7% and 96.5%, with an average of 92.9%. The similarities between the rhesus macaque and human TRBV exons at the amino acid sequence level ranged between 19.5% and 94.7%, with an average of 85.3%.

Rhesus macaque TRBD genes

The two TRBD genes extracted from the rhesus macaque genome were found to have 95.0% and 92.8% agreements at the nucleotide level with the corresponding human TRBD genes (Table 2 and Rhesus_macaque_TRBD.fsa in Supporting Information). The percent similarities between the rhesus macaque and human TRBD exon (i.e. D-REGION in the IMGT standardized labels) nucleotide sequences were 84.6% and 75.0%.

The rhesus macaque TRBD genes have been sequenced in a previous study⁴⁴. The TRBD1 gene extracted from the rhesus macaque genome does not differ from that reported in this previous study. A 1.2% difference was found between the TRBD2 gene reported here and that reported previously, with a single nucleotide difference occurring in the 5' spacer. Thus, there are no differences in the TRBD2 D-REGION extracted from the rhesus macaque genome compared with that reported previously⁴⁴.

Rhesus macaque TRBJ genes

For each of the 14 human TRBJ genes, there was one corresponding TRBJ gene found on chromosome 3 of the rhesus macaque genome (Table 3 and Rhesus_macaque_TRBJ.fsa in Supporting Information). The percent similarities between the rhesus macaque TRBJ genes and the corresponding human TRBJ genes are shown in Table 3 (range: 92.1% and 98.7%; average: 96.1%).

A comparison of the rhesus macaque and human TRBJ exons (i.e. J-REGION in the IMGT standardized labels) revealed percent similarities of nucleotide sequences ranging between 90.2% and 100%, with an average similarity of 95.4% (Table 3). The similarities between the translated TRBJ exons of the rhesus macaque and human genes ranged between 81.3% and 100%, with an average similarity of 92.3% (Table 3).

We compared the TRBJ genes extracted from the rhesus macaque genome with those reported in a previous study⁴⁴. The only differences found were in the TRBJ1-6 and TRBJ2-1 genes, which differed by 1.9% and 2%, respectively. A single nucleotide difference in the 20th nucleotide position of the TRBJ1-6 exon resulted in a difference of a single amino acid (i.e. the TRBJ1-6 exon from the rhesus macaque genome contained H in the 7th amino acid position instead of Y). In the TRBJ2-1 gene, a single nucleotide difference in the 31st nucleotide position of the exon did not result in any amino acid differences between the TRBJ2-1 exon extracted from the rhesus macaque genome and that reported by Cheynier *et al.*⁴⁴

Use of the rhesus macaque TRB genes by expressed TCR β sequences

To demonstrate the use of the TRB genes extracted from the rhesus macaque genome by expressed TCR β sequences, we used an existing database of 7218 TCR β sequences involved in CD8⁺ T cell responses specific for the immunodominant Mamu-A*01-restricted SIV-SL8/TL8 and SIV-CM9 epitopes in 20 rhesus macaques^{30, 45}. Each of these TCR β sequences was aligned with the TRB gene exons to determine the most likely TRBV, TRBJ and TRBD gene usage. In Table 4 and Table 5 we show the rhesus macaque TRB genes that were found to be most likely used by at least one of the TCR β sequences. The genes used by the TCR β sequences included 54 of the 72 TRBV genes, both TRBD genes, and 13 of the 14 TRBJ genes. The highest percent homology and longest match between each TRB gene and a TCR β sequence is also shown. Of the 18 rhesus macaque TRBV genes not used by the TCR β sequences, 12 either didn't begin with a start codon or contained stop codons when translated (Table 1). The rhesus macaque TRBJ2-2P gene, which is homologous to the human TRBJ2-2P gene (qualified by IMGT as having an "Open Reading Frame" functionality), was the only TRBJ gene not used by the TCR β sequences. Deviations between the rhesus macaque TRB genes and TCR β sequences were mostly attributed to the full-length genes not being used by the TCR β sequences, owing to nucleotides being cleaved during TCR gene recombination. However, allelic differences could also exist between the single rhesus macaque sequenced in the genome project and the 20 SIV-infected macaques from which the TCR β sequences were obtained.

Possible allelic variants of the TRB genes used by the TCR β sequences were not identified due to the level of uncertainty associated with distinguishing allelic variants from sequencing errors, in either the rhesus macaque genome or TCR β sequences, when there were often small numbers of TCR β sequences per rhesus macaque using a particular TRB gene. However, we investigated whether the nucleotide sequence variants of the TRBJ1-6 and TRBJ2-1 genes reported by Cheynier *et al.*⁴⁴ were used in our collection of epitope-specific TCR β sequences. The previously reported variant of the TRBJ1-6 gene was found to be used by some TCR β sequences, suggesting that this is an allelic variant of the TRBJ1-6

gene extracted from the rhesus macaque genome. The TRBJ2-1 gene variant was not used by any of the TCR β sequences. This TRBJ2-1 gene variant may be an allelic variant that was not present in any of the 20 rhesus macaques in which the Mamu-A*01-restricted SIV-SL8/TL8- and SIV-CM9-specific TCR β repertoires were studied but it is also possible that the single nucleotide difference in the TRBJ2-1 gene reported Cheynier *et al.*⁴⁴ is due to sequencing error.

DISCUSSION

The assembly of reference TCR gene data sets for many species has often relied on the ad hoc sourcing of different TCR genes from various studies over time. Here, we report a reference set of TRB genes extracted from the rhesus macaque genome, most of which were expressed by TCR β sequences in our extensive database of TCR β repertoires involved in CD8⁺ T cell responses to the immunodominant Mamu-A*01-restricted SL8/TL8 and CM9 epitopes derived from SIV. Although there is a high degree of similarity (93.0%) between the exons of the rhesus macaque and human TRB genes, important interspecies differences exist. These interspecies differences are emphasized by the lack of a one-to-one correspondence between the rhesus macaque and human TRBV genes, and could potentially limit the accuracy of studies that rely on human TCR genes to characterize rhesus macaque TCR repertoires.

The rhesus macaque TRB genes described herein will not only aid in the identification of the TRBV and TRBJ genes used by TCR β sequences, they will also improve the accuracy of studies that aim to characterize the V(D)J recombination mechanisms that produce TCR β repertoires. Indeed, several of the extracted rhesus macaque TRB genes have already been used in a study of TCR β sequence sharing between macaques in the SIV-SL8/TL8-specific and SIV-CM9-specific CD8⁺ T cell responses³⁹. This study required predictions of the potential V(D)J recombination mechanisms involved in producing the observed epitope-specific TCR β repertoires, which were more reliable using the rhesus macaque TRB genes instead of the human TRB genes.

Rhesus macaques are frequently used to study fundamental aspects of immunology and investigate vaccine efficacy in a variety of infectious diseases. Increasing evidence, much of which has come from studies conducted with this non-human primate model, indicates that the clonotypic architecture of antigen-specific T cell populations is a fundamental determinant of immune control and disease outcome^{26, 45}. Thus, the rhesus macaque TRB genes presented here provide a valuable tool for dissecting the molecular features of TCR β repertoires that underlie such associations in this model.

METHODS

Extraction of TRB gene sequences from the rhesus macaque genome

The published rhesus macaque (*Macaca mulatta*) genome¹ is available from the National Center for Biotechnology Information (NCBI) Rhesus Macaque Genome Resources website (http://www.ncbi.nlm.nih.gov/projects/genome/guide/rhesus_macaque/). The TRB gene locus is located on chromosome 3 (Accession number: NC_007860.1). The rhesus macaque

chromosome 3 sequence was queried against all human TRB reference genes (obtained from the NCBI Human Resources website <http://www.ncbi.nlm.nih.gov/projects/genome/guide/human/>) using BLAST (Basic Local Alignment Search Tool)⁴⁶ to identify regions in the rhesus macaque sequence that resembled human TRB genes. Results were filtered to those with e-value 0.001, total alignment length 35% of the human reference gene, and total percent identity 75% with the human reference gene. These parameters were chosen to minimize false positive search results. Overlapping regions were merged and all regions were extended in both the 5' and 3' directions for regions missed in BLAST's local alignment search. Sequence alignments using ClustalW⁴⁷ were then performed to compare each region of the rhesus macaque genome with each human TRB gene from the NCBI human reference set. The best human match to each macaque region was identified and then used as a guide to determine the exact length and terminal ends of the rhesus macaque TRB gene sequences, as well as intron and exon positions.

Comparison of rhesus macaque and human TRB gene sequences

We assessed the similarity between the rhesus macaque and the NCBI human TRB reference gene sequences (or the IMGT human TRB reference gene if the NCBI reference gene sequence was partial) by identifying the human TRB gene that had the highest overall percentage identity with each rhesus macaque TRB gene using a ClustalW alignment. We encountered the following scenarios: (i) a clearly identifiable one-to-one correspondence between a rhesus macaque and a human TRB gene; (ii) a rhesus macaque TRB gene with reasonable similarity to a group of human TRB genes; and, (iii) a human TRB gene with no reasonable correspondence to a rhesus macaque TRB gene. We therefore adopted the following approach to labelling the rhesus macaque TRB genes. For each rhesus macaque TRB gene, we first identified the group of human TRB genes to which it was most similar (e.g. TRBV1). We then numbered all rhesus macaque TRB genes which were most similar to this same group of human TRB genes according to the order in which the TRB sequences were found in the rhesus macaque genome (e.g. TRBV1-1, TRBV1-2, etc.). The ImMunoGeneTics (IMGT)⁴⁸ nomenclature for TCR genes was used throughout.

Analysis of expressed epitope-specific TCR β sequences using the rhesus macaque TRB genes

For all Mamu-A*01-restricted SIV-SL8/TL8-specific and SIV-CM9-specific TCR β sequences, we performed a complete alignment analysis using the identified rhesus macaque TRB genes. This analysis determined for each epitope-specific TCR β sequence the best-percentage-match TRBV, TRBD and TRBJ genes over the longest alignment length by initially aligning the TRBV gene at the 5' end of the TCR β sequence and then aligning the TRBJ gene at the 3' end of the TCR β sequence. A minimum percentage match of 77% over an alignment length of at least 50 nucleotides was required for alignment of the TRBV genes. For alignment of the TRBJ genes, a minimum percentage match of 70% was required over the length of the TRBJ exon. The TRBD genes were then aligned to the sequence interval between the identified TRBV and TRBJ regions. A match to a string of two or more nucleotides was considered to originate from the TRBD gene.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

ACKNOWLEDGEMENTS

We thank Dr Mark Tanaka for assistance with the phylogenetic analysis and Associate Professor Andrew Collins for helpful discussions. This work was supported by the James S. McDonnell Foundation 21st Century Research Award/Studying Complex Systems, the Australian Research Council (ARC), and the National Institutes of Health (NIH). MPD is a Sylvia and Charles Viertel Senior Medical Research Fellow and DAP is a Medical Research Council (UK) Senior Clinical Fellow.

REFERENCES

1. Gibbs RA, Rogers J, Katze MG, et al. Evolutionary and biomedical insights from the rhesus macaque genome. *Science*. 2007; 316:222–234. [PubMed: 17431167]
2. Rivaller P, Jiang H, Cho YG, Quink C, Wang F. Complete nucleotide sequence of the rhesus lymphocryptovirus: genetic validation for an Epstein-Barr virus animal model. *J Virol*. 2002; 76:421–426. [PubMed: 11739708]
3. Hansen SG, Strelow LI, Franchi DC, Anders DG, Wong SW. Complete sequence and genomic analysis of rhesus cytomegalovirus. *J Virol*. 2003; 77:6620–6636. [PubMed: 12767982]
4. Pitcher CJ, Hagen SI, Walker JM, et al. Development and homeostasis of T cell memory in rhesus macaque. *J Immunol*. 2002; 168:29–43. [PubMed: 11751943]
5. Hasegawa A, Moriya C, Liu H, et al. Analysis of TCRalpha combinations used by simian immunodeficiency virus-specific CD8+ T cells in rhesus monkeys: implications for CTL immunodominance. *J Immunol*. 2007; 178:3409–3417. [PubMed: 17339435]
6. Cicin-Sain L, Messaoudi I, Park B, et al. Dramatic increase in naive T cell turnover is linked to loss of naive T cells from old primates. *Proc Natl Acad Sci U S A*. 2007; 104:19960–19965. [PubMed: 18056811]
7. Lackner AA, Veazey RS. Current concepts in AIDS pathogenesis: insights from the SIV/macaque model. *Annu Rev Med*. 2007; 58:461–476. [PubMed: 17217334]
8. Fan J, Liang X, Horton MS, et al. Preclinical study of influenza virus A M2 peptide conjugate vaccines in mice, ferrets, and rhesus monkeys. *Vaccine*. 2004; 22:2993–3003. [PubMed: 15297047]
9. Kobasa D, Jones SM, Shinya K, et al. Aberrant innate immune response in lethal infection of macaques with the 1918 influenza virus. *Nature*. 2007; 445:319–323. [PubMed: 17230189]
10. Lewinsohn DM, Tydeman IS, Frieder M, et al. High resolution radiographic and fine immunologic definition of TB disease progression in the rhesus macaque. *Microbes Infect*. 2006; 8:2587–2598. [PubMed: 16952476]
11. Rivaller P, Carville A, Kaur A, et al. Experimental rhesus lymphocryptovirus infection in immunosuppressed macaques: an animal model for Epstein-Barr virus pathogenesis in the immunosuppressed host. *Blood*. 2004; 104:1482–1489. [PubMed: 15150077]
12. Fogg MH, Garry D, Awad A, Wang F, Kaur A. The BZLF1 homolog of an Epstein-Barr-related gamma-herpesvirus is a frequent target of the CTL response in persistently infected rhesus macaques. *J Immunol*. 2006; 176:3391–3401. [PubMed: 16517707]
13. Sequer G, Britt WJ, Lakeman FD, et al. Experimental coinfection of rhesus macaques with rhesus cytomegalovirus and simian immunodeficiency virus: pathogenesis. *J Virol*. 2002; 76:7661–7671. [PubMed: 12097580]
14. Price DA, Bitmansour AD, Edgar JB, et al. Induction and Evolution of Cytomegalovirus-Specific CD4+ T Cell Clonotypes in Rhesus Macaques. *J Immunol*. 2008; 180:269–280. [PubMed: 18097028]
15. Powers C, Fruh K. Rhesus CMV: an emerging animal model for human CMV. *Med Microbiol Immunol*. 2008; 197:109–115. [PubMed: 18193454]

16. Edghill-Smith Y, Venzon D, Karpova T, et al. Modeling a safer smallpox vaccination regimen, for human immunodeficiency virus type 1-infected patients, in immunocompromised macaques. *J Infect Dis.* 2003; 188:1181–1191. [PubMed: 14551889]
17. El Mubarak HS, Yuksel S, van Amerongen G, et al. Infection of cynomolgus macaques (*Macaca fascicularis*) and rhesus macaques (*Macaca mulatta*) with different wild-type measles viruses. *J Gen Virol.* 2007; 88:2028–2034. [PubMed: 17554037]
18. Rowe T, Gao G, Hogan RJ, et al. Macaque model for severe acute respiratory syndrome. *J Virol.* 2004; 78:11401–11404. [PubMed: 15452262]
19. McMichael AJ. HIV vaccines. *Annu Rev Immunol.* 2006; 24:227–255. [PubMed: 16551249]
20. Maryanski JL, Jongeneel CV, Bucher P, Casanova JL, Walker PR. Single-cell PCR analysis of TCR repertoires selected by antigen in vivo: a high magnitude CD8 response is comprised of very few clones. *Immunity.* 1996; 4:47–55. [PubMed: 8574851]
21. Pewe L, Heard SB, Bergmann C, Dailey MO, Perlman S. Selection of CTL escape mutants in mice infected with a neurotropic coronavirus: quantitative estimate of TCR diversity in the infected central nervous system. *J Immunol.* 1999; 163:6106–6113. [PubMed: 10570300]
22. Kedzierska K, Day EB, Pi J, et al. Quantification of repertoire diversity of influenza-specific epitopes with predominant public or private TCR usage. *J Immunol.* 2006; 177:6705–6712. [PubMed: 17082583]
23. Pewe LL, Netland JM, Heard SB, Perlman S. Very diverse CD8 T cell clonotypic responses after virus infections. *J Immunol.* 2004; 172:3151–3156. [PubMed: 14978121]
24. Turner SJ, Doherty PC, McCluskey J, Rossjohn J. Structural determinants of T-cell receptor bias in immunity. *Nat Rev Immunol.* 2006; 6:883–894. [PubMed: 17110956]
25. Nikolich-Zugich J, Slifka MK, Messaoudi I. The many important facets of T-cell repertoire diversity. *Nat Rev Immunol.* 2004; 4:123–132. [PubMed: 15040585]
26. Davenport MP, Price DA, McMichael AJ. The T cell repertoire in infection and vaccination: implications for control of persistent viruses. *Curr Opin Immunol.* 2007; 19:294–300. [PubMed: 17433874]
27. Cornberg M, Chen AT, Wilkinson LA, et al. Narrowed TCR repertoire and viral escape as a consequence of heterologous immunity. *J Clin Invest.* 2006; 116:1443–1456. [PubMed: 16614754]
28. Gillespie GM, Stewart-Jones G, Rengasamy J, et al. Strong TCR conservation and altered T cell cross-reactivity characterize a B*57-restricted immune response in HIV-1 infection. *J Immunol.* 2006; 177:3893–3902. [PubMed: 16951352]
29. Meyer-Olson D, Shoukry NH, Brady KW, et al. Limited T cell receptor diversity of HCV-specific T cell responses is associated with CTL escape. *J Exp Med.* 2004; 200:307–319. [PubMed: 15289502]
30. Price DA, West SM, Betts MR, et al. T cell receptor recognition motifs govern immune escape patterns in acute SIV infection. *Immunity.* 2004; 21:793–803. [PubMed: 15589168]
31. Yu XG, Lichterfeld M, Chetty S, et al. Mutually exclusive T-cell receptor induction and differential susceptibility to human immunodeficiency virus type 1 mutational escape associated with a two-amino-acid difference between HLA class I subtypes. *J Virol.* 2007; 81:1619–1631. [PubMed: 17121793]
32. Chen ZW, Kou ZC, Shen L, Reimann KA, Letvin NL. Conserved T-cell receptor repertoire in simian immunodeficiency virus-infected rhesus monkeys. *J Immunol.* 1993; 151:2177–2187. [PubMed: 8393899]
33. Chen ZW, Li Y, Zeng X, et al. The TCR repertoire of an immunodominant CD8+ T lymphocyte population. *J Immunol.* 2001; 166:4525–4533. [PubMed: 11254709]
34. Chen ZW, Kou ZC, Lekutis C, et al. T cell receptor V beta repertoire in an acute infection of rhesus monkeys with simian immunodeficiency viruses and a chimeric simian-human immunodeficiency virus. *J Exp Med.* 1995; 182:21–31. [PubMed: 7540651]
35. Chen ZW, Shen L, Regan JD, Kou Z, Ghim SH, Letvin NL. The T cell receptor gene usage by simian immunodeficiency virus gag-specific cytotoxic T lymphocytes in rhesus monkeys. *J Immunol.* 1996; 156:1469–1475. [PubMed: 8568249]

36. Chen ZW, Shen Y, Kou Z, et al. Prolonged dominance of clonally restricted CD4(+) T cells in macaques infected with simian immunodeficiency viruses. *J Virol.* 2000; 74:7442–7450. [PubMed: 10906197]
37. Shen L, Chen ZW, Letvin NL. The repertoire of cytotoxic T lymphocytes in the recognition of mutant simian immunodeficiency virus variants. *J Immunol.* 1994; 153:5849–5854. [PubMed: 7989780]
38. Sen P, Charini WA, Subbramanian RA, et al. Clonal focusing of epitope-specific CD8+ T lymphocytes in rhesus monkeys following vaccination and simian-human immunodeficiency virus challenge. *J Virol.* 2008; 82:805–816. [PubMed: 17977967]
39. Venturi V, Chin HY, Price DA, Douek DC, Davenport MP. The role of production frequency in the sharing of simian immunodeficiency virus-specific CD8+ TCRs between macaques. *J Immunol.* 2008; 181:2597–2609. [PubMed: 18684950]
40. Currier JR, Stevenson KS, Kehn PJ, Zheng K, Hirsch VM, Robinson MA. Contributions of CD4+, CD8+, and CD4+CD8+ T cells to skewing within the peripheral T cell receptor beta chain repertoire of healthy macaques. *Hum Immunol.* 1999; 60:209–222. [PubMed: 10321957]
41. Zhou D, Kou Z, Ibegbu C, et al. The disruption of macaque CD4+ T-cell repertoires during the early simian immunodeficiency virus infection. *J Med Primatol.* 1999; 28:174–180. [PubMed: 10593483]
42. Bostik P, Stephenson ST, Lynch RM, Cardona A, Ansari AA. Maintenance of CD4+ T cell TCR Vbeta repertoire heterogeneity is characteristic of apathogenic SIV infection in non-human primate model of AIDS. *Virology.* 2007; 369:324–328. [PubMed: 17889219]
43. Manuel ER, Charini WA, Sen P, et al. Contribution of T-cell receptor repertoire breadth to the dominance of epitope-specific CD8+ T-lymphocyte responses. *J Virol.* 2006; 80:12032–12040. [PubMed: 17035327]
44. Cheynier R, Henrichwark S, Wain-Hobson S. Sequence of the rhesus monkey T-cell receptor beta chain diversity and joining loci. *Immunogenetics.* 1996; 43:83–87. [PubMed: 8537129]
45. Price DA, Asher TE, Wilson NA, et al. Public clonotype usage identifies protective Gag-specific CD8+ T cell responses in SIV infection. *J Exp Med.* 2009; 206:923–936. [PubMed: 19349463]
46. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. *J Mol Biol.* 1990; 215:403–410. [PubMed: 2231712]
47. Thompson JD, Higgins DG, Gibson TJ. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 1994; 22:4673–4680. [PubMed: 7984417]
48. Lefranc MP, Giudicelli V, Ginestoux C, et al. IMGT, the international ImMunoGeneTics database. *Nucleic Acids Res.* 1999; 27:209–212. [PubMed: 9847182]
49. Saitou N, Nei M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol.* 1987; 4:406–425. [PubMed: 3447015]
50. Letunic I, Bork P. Interactive Tree Of Life (iTOL): an online tool for phylogenetic tree display and annotation. *Bioinformatics.* 2007; 23:127–128. [PubMed: 17050570]

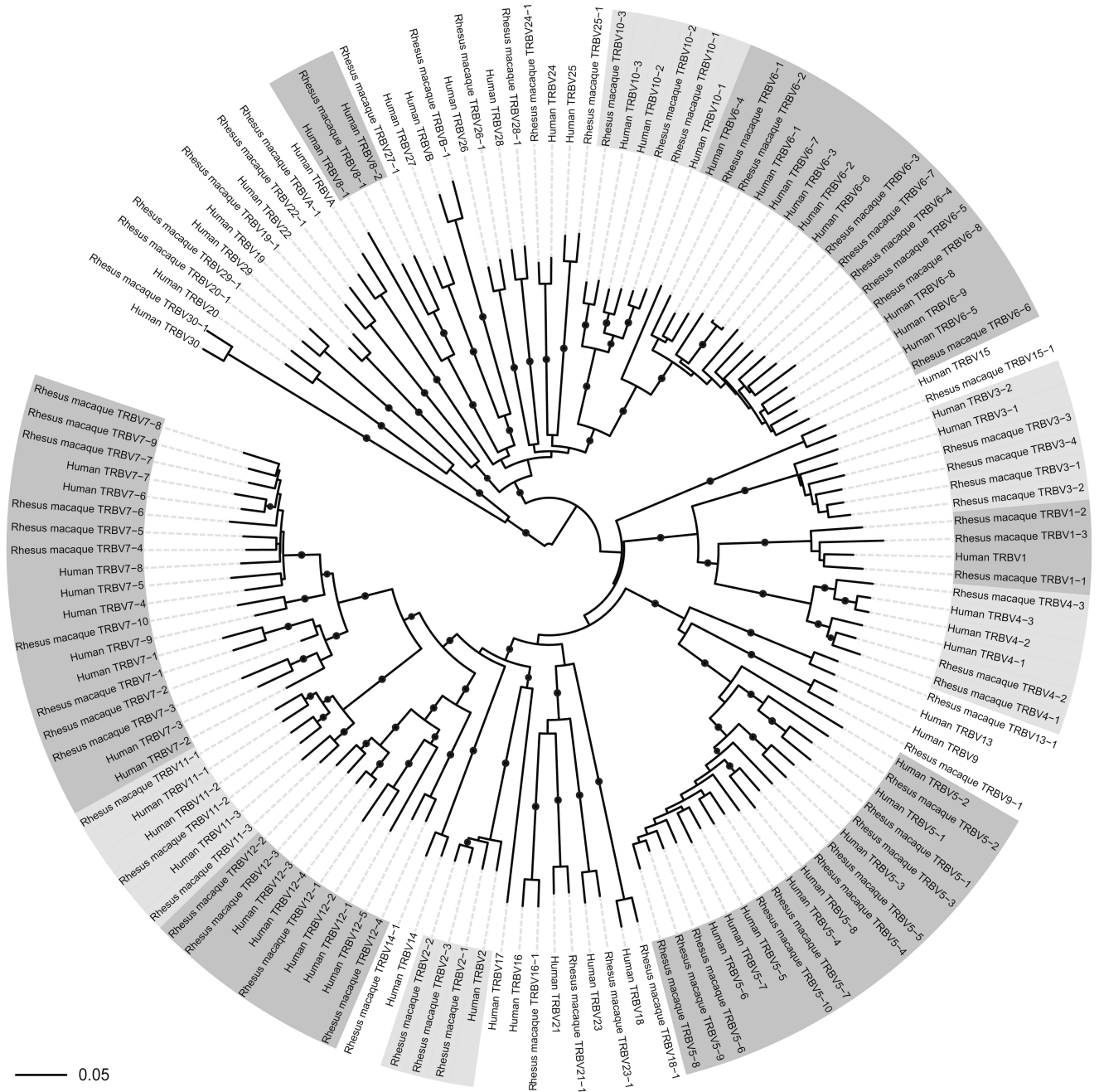


Figure 1. Unrooted circular phylogram showing the clustering relationships between all rhesus macaque and human TRBV gene sequences at the nucleotide level

Exons, introns and recombination signal sequences have been included and gene families consisting of multiple genes are highlighted. All TRBV gene sequences were aligned using ClustalW and the tree was constructed in ClustalW using the neighbour-joining method⁴⁹ and bootstrapped 1000 times. Branches with bootstrap values >80% are indicated with a black dot and branch lengths are those assigned by ClustalW. The tree was visualized using

the Interactive Tree of Life⁵⁰ (available at <http://itol.embl.de/>). Note that the tree has been rotated about the mid-point of the most distant nodes to assist visualization.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Table 1

Comparison of the rhesus macaque TRBV genes and their best human homologues.

Rhesus macaque gene	Best human homologue ¹	Gene nucleotide sequence			Exon nucleotide sequence			Exon amino acid sequence ³			Note
		Percent identity	Alignment length ²	Alignment length	Percent identity	Alignment length	Percent identity	Alignment length	Percent identity	Alignment length	
TRBV1-1	TRBV1	93.3	489	333	93.1	333	86.5	111	8		
TRBV1-2	TRBV1	91.0	490	334	90.1	334	64.0	111	8		
TRBV1-3	TRBV1	85.5	496	337	82.8	337			8		
TRBV2-1	TRBV2	93.5	475	347	94.8	347	89.6	115			
TRBV2-2	TRBV2	93.7	475	347	93.7	347	89.6	115			
TRBV2-3	TRBV2	94.3	475	347	93.9	347	87.8	115			
TRBV3-1	TRBV3-1	92.6	499	344	93.6	344	86.8	114			
TRBV3-2	TRBV3-1	92.8	499	344	92.4	344	84.2	114			
TRBV3-3	TRBV3-1	90.8	499	344	93.9	344	87.7	114			
TRBV3-4	TRBV3-1	92.6	499	344	93.6	344	86.0	114			
TRBV4-1	TRBV4-1	92.7	493	344	93.6	344	89.5	114			
TRBV4-2	TRBV4-1	94.7	493	344	94.8	344	93.0	114			
TRBV4-3	TRBV4-3	91.2	502	344	93.0	344	91.2	114			
TRBV5-1	TRBV5-1	93.5	509	343	93.9	343			7		
TRBV5-2	TRBV5-1	84.8	512	343	83.7	343			7, 8		
TRBV5-3	TRBV5-3	93.1	509	346	93.4	346	86.1	115			
TRBV5-4	TRBV5-8	93.1	506	343	93.6	343	88.6	114			
TRBV5-5	TRBV5-6	90.9	507	344	91.6	344	81.6	114			
TRBV5-6	TRBV5-6	94.7	505	343	95.0	343	91.2	114			
TRBV5-7	TRBV5-5	93.7	506	343	95.0	343	91.2	114			
TRBV5-8	TRBV5-6	93.1	506	343	93.9	343	90.4	114			
TRBV5-9	TRBV5-6	94.5	505	343	95.3	343	92.1	114			
TRBV5-10	TRBV5-5	92.1	506	343	92.1	343	87.7	114			
TRBV6-1	TRBV6-1	91.9	472	344	93.6	344	87.7	114			
TRBV6-2	TRBV6-1	94.3	351	295	93.9	295	87.6	97	4		
TRBV6-3	TRBV6-5	93.9	475	344	94.2	344	88.6	114			

Rhesus macaque gene	Best human homologue ¹	Gene nucleotide sequence		Exon nucleotide sequence		Exon amino acid sequence ³		Note
		Percent identity	Alignment length ²	Percent identity	Alignment length	Percent identity	Alignment length	
TRBV6-4	TRBV6-5	92.0	475	92.4	344	84.2	114	
TRBV6-5	TRBV6-9	91.5	424	90.8	355	84.7	111	5
TRBV6-6	TRBV6-5	92.2	475	94.5	344	87.7	114	
TRBV6-7	TRBV6-5	92.4	475	93.9	344	86.8	114	
TRBV6-8	TRBV6-5	91.0	390	91.3	298	83.8	99	5
TRBV7-1	TRBV7-1	83.6	329	93.1	290	83.3	96	4, 8
TRBV7-2	TRBV7-3	87.7	261	88.3	222	83.6	73	4
TRBV7-3	TRBV7-3	90.6	500	91.9	347	86.1	115	
TRBV7-4	TRBV7-7	89.8	549	90.8	347	84.3	115	
TRBV7-5	TRBV7-8	91.8	525	92.8	347	87.0	115	
TRBV7-6	TRBV7-6	89.7	532	92.8	347	87.8	115	
TRBV7-7	TRBV7-4	89.6	509	90.8	347	82.6	115	
TRBV7-8	TRBV7-6	87.4	546	89.2	361	37.2	121	8
TRBV7-9	TRBV7-6	92.8	538	94.5	347	89.6	115	
TRBV7-10	TRBV7-9	90.7	516	92.2	347	82.6	115	
TRBV8-1	TRBV8-2	86.7	511	85.1	336	30.4	115	8
TRBV9-1	TRBV9	94.7	514	96.2	343	93.9	114	
TRBV10-1	TRBV10-1	96.5	489	96.2	344	93.9	114	
TRBV10-2	TRBV10-2	95.1	489	95.3	344	93.9	114	
TRBV10-3	TRBV10-3	95.5	489	96.5	344	93.0	114	
TRBV11-1	TRBV11-1	93.3	490	93.9	347	87.0	115	
TRBV11-2	TRBV11-2	94.4	480	94.5	347	91.3	115	
TRBV11-3	TRBV11-3	93.9	477	94.2	347	88.7	115	
TRBV12-1	TRBV12-2	92.7	482	93.0	344	85.2	115	
TRBV12-2	TRBV12-3	91.8	486	93.1	347	87.8	115	
TRBV12-3	TRBV12-3	91.8	486	92.5	347	87.8	115	
TRBV12-4	TRBV12-5	94.9	486	94.5	347	92.2	115	
TRBV13-1	TRBV13	94.5	524	95.7	374	89.5	124	

Rhesus macaque gene	Best human homologue ¹	Gene nucleotide sequence			Exon nucleotide sequence			Exon amino acid sequence ³			Note
		Percent identity	Alignment length ²	Percent identity	Alignment length	Percent identity	Alignment length	Percent identity	Alignment length		
TRBV14-1	TRBV14	93.6	482	95.1	347	89.6	115				
TRBV15-1	TRBV15	94.3	508	94.8	344	89.5	114				
TRBV16-1	TRBV16	94.7	493	94.8	347	88.7	115				
TRBV18-1	TRBV18	95.0	658	94.8	347	90.4	115				
TRBV19-1	TRBV19	95.5	516	95.6	344	94.7	114				
TRBV20-1	TRBV20-1	91.9	713	92.5	335	83.8	111				
TRBV21-1	TRBV21-1	94.0	500	94.0	348	19.5	118				
TRBV22-1	TRBV22-1	78.3	493	72.7	348						6, 8
TRBV23-1	TRBV23-1	92.0	536	93.9	347	91.3	115				
TRBV24-1	TRBV24-1	95.2	517	95.9	345	90.4	115				
TRBV25-1	TRBV25-1	94.7	507	95.1	344	90.4	114				
TRBV26-1	TRBV26	94.5	524	95.1	344	73.9	115				8
TRBV27-1	TRBV27	94.9	512	95.6	344	93.9	114				
TRBV28-1	TRBV28	93.3	523	96.2	344	93.9	114				
TRBV29-1	TRBV29-1	94.1	661	96.4	335	93.7	111				
TRBV30-1	TRBV30	94.3	743	96.1	335	93.7	111				6, 8
TRBVA-1	TRBVA	92.7	493	91.1	316						8
TRBVB-1	TRBVB	88.3	563	87.4	422						
Average percent identity		92.2		92.9		85.3					

¹The best human homologue had the highest percent identity with the rhesus macaque gene nucleotide sequence.

²The alignment length is the total length across both the aligned rhesus macaque and human gene/exon sequences.

³The exon amino acid sequence was translated in the frame that yielded a start codon at the 5' end of the exon. Comparisons of the exon amino acid sequences were omitted for TRBV genes in which no start codon was found. For partial rhesus macaque exons missing a portion of sequence at the 5' end, the sequences were translated in the frame in which the start codon was found in the human homologues.

⁴The rhesus macaque gene is a partial sequence, with a missing portion of sequence at the 5' end of the gene. The percent identities between rhesus macaque and human genes and exons are calculated with the missing portion of rhesus macaque gene excluded.

⁵The rhesus macaque gene is a partial sequence, with a missing portion of sequence at the 3' end of the gene. The percent identities between rhesus macaque and human genes and exons are calculated with the missing portion of rhesus macaque gene excluded.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

⁹The human NCBI reference gene was a partial sequence so the human IMGT reference gene sequence was used for these comparisons.

⁷There was some ambiguity in the identification of the exons for the TRBV5-1 and TRBV5-2 genes (see Table 1 in Supporting Information).

⁸The rhesus macaque exon amino sequence contained stop codons or, for those genes in which no start codon was found, the amino acid sequences in all three translation frames contained stop codons.

Table 2

Comparison of the rhesus macaque TRBD genes and their human homologues.

Rhesus macaque gene	Human gene homologue	Gene nucleotide sequence		Exon nucleotide sequence	
		Percent identity	Alignment length ¹	Percent identity	Alignment length
TRBD1	TRBD1	95.0	80	84.6	13
TRBD2	TRBD2	92.8	83	75.0	16
Average percent identity		93.9		79.8	

¹The alignment length is the total length across both the aligned rhesus macaque and human gene/exon sequences.

Table 3

Comparison of the rhesus macaque TRBJ genes and their human homologues.

Rhesus macaque gene	Gene nucleotide sequence			Exon nucleotide sequence			Exon amino acid sequence ²		
	Human gene homologue	Percent identity	Alignment length ¹	Percent identity	Alignment length	Percent identity	Alignment length	Percent identity	Alignment length
TRBJ1-1	TRBJ1-1	98.7	76	97.9	48	93.3	15		
TRBJ1-2	TRBJ1-2	92.1	76	91.7	48	86.7	15		
TRBJ1-3	TRBJ1-3	94.9	78	92.0	50	87.5	16		
TRBJ1-4	TRBJ1-4	98.7	79	100.0	51	100.0	16		
TRBJ1-5	TRBJ1-5	94.9	78	92.0	50	87.5	16		
TRBJ1-6	TRBJ1-6	97.5	81	96.2	53	100.0	17		
TRBJ2-1	TRBJ2-1	97.4	78	96.0	50	100.0	16		
TRBJ2-2	TRBJ2-2	93.7	79	90.2	51	87.5	16		
TRBJ2-2P	TRBJ2-2P	96.2	53	95.7	46	86.7	15		
TRBJ2-3	TRBJ2-3	98.7	77	98.0	49	93.8	16		
TRBJ2-4	TRBJ2-4	94.9	78	96.0	50	81.3	16		
TRBJ2-5	TRBJ2-5	97.4	76	97.9	48	100.0	15		
TRBJ2-6	TRBJ2-6	95.1	81	96.2	53	94.1	17		
TRBJ2-7	TRBJ2-7	94.7	75	95.7	47	93.3	15		
Average percent identity		96.1		95.4		92.3			

¹The alignment length is the total length across both the aligned rhesus macaque and human gene/exon sequences.

²The TRBJ exons were translated in the frame that yielded the characteristic FGXG or LGXG motif.

Table 4Usage of the rhesus macaque TRBV genes by expressed TCR β sequences.

Rhesus macaque gene	Highest % usage by a TCR β sequence	Alignment length ¹
TRBV2-1	100	182
TRBV2-2	99.1	116
TRBV2-3	100	185
TRBV3-1	100	274
TRBV3-2	100	278
TRBV3-3	100	179
TRBV3-4	100	278
TRBV4-1	100	167
TRBV4-2	100	282
TRBV4-3	100	273
TRBV5-1	98.7	232
TRBV5-3	92.1	76
TRBV5-4	98.9	281
TRBV5-6	98.9	272
TRBV5-7	98.1	159
TRBV5-8	97.9	280
TRBV5-9	98.7	307
TRBV5-10	100	189
TRBV6-1	100	277
TRBV6-2	100	219
TRBV6-3	100	274
TRBV7-2	98.9	182
TRBV7-3	98.7	297
TRBV7-4	99.3	281
TRBV7-5	98.6	289
TRBV7-6	99.7	287
TRBV7-7	97.1	68
TRBV7-9	97.9	290
TRBV7-10	98.6	284
TRBV9-1	99.4	180
TRBV10-1	96.7	276
TRBV10-2	100	279
TRBV10-3	99.2	238
TRBV11-1	99.6	276
TRBV11-2	98.6	282
TRBV11-3	98.9	282
TRBV12-2	99.3	277
TRBV12-3	98.8	168

Rhesus macaque gene	Highest % usage by a TCR β sequence	Alignment length ¹
TRBV12-4	95.9	74
TRBV13-1	98.9	182
TRBV14-1	99.7	294
TRBV15-1	99.2	262
TRBV16-1	94.6	74
TRBV18-1	97.6	167
TRBV19-1	100	279
TRBV20-1	99.5	197
TRBV21-1	98.2	274
TRBV23-1	100	298
TRBV24-1	97.7	214
TRBV25-1	98.5	268
TRBV27-1	100	281
TRBV28-1	99.2	119
TRBV29-1	100	282
TRBV30-1	98.2	170

¹ The alignment length between the TRBV gene and the TCR β sequence is mostly determined by the length of the TCR β sequence to the 5' end of the CDR3.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Table 5Usage of the rhesus macaque TRBD and TRBJ genes by expressed TCR β sequences.

Rhesus macaque gene	Highest % usage by a TCR β sequence	Alignment length ¹
TRBD1	100	13
TRBD2	100	14
TRBJ1-1	100	48
TRBJ1-2	100	48
TRBJ1-3	100	50
TRBJ1-4	100	51
TRBJ1-5	100	50
TRBJ1-6	96.2	53
TRBJ2-1	100	50
TRBJ2-2	100	51
TRBJ2-3	100	49
TRBJ2-4	100	49
TRBJ2-5	100	48
TRBJ2-6	100	53
TRBJ2-7	100	47

¹The alignments were performed over the total length of the TRBD or TRBJ exon.