

## RESEARCH PAPER

## Urocortin promotes the development of vasculitis in a rat model of thromboangiitis obliterans via corticotrophin-releasing factor type 1 receptors

Youhua Xu<sup>1\*</sup>, Rongjian Zhang<sup>1\*</sup>, Jie Chen<sup>1</sup>, Qichun Zhang<sup>1</sup>, Juejin Wang<sup>1</sup>, Jue Hu<sup>1</sup>, Xiaowei Guan<sup>1</sup>, Lai Jin<sup>1</sup>, Hong Fu<sup>1</sup>, Bo Gui<sup>2</sup>, Yuanyuan Guo<sup>1</sup> and Shengnan Li<sup>1</sup>

<sup>1</sup>Department of Pharmacology, Nanjing Medical University, Nanjing, China, and <sup>2</sup>Department of Anesthesiology, First Affiliated Hospital of Nanjing Medical University, Nanjing, China

**Background and purpose:** Urocortin is a locally expressed pro-inflammatory peptide. Here we have examined the effects of urocortin on sodium laurate-induced peripheral arterial vasculitis in rats, modelling the mechanisms of thromboangiitis obliterans (TAO).

**Experimental approach:** Peripheral vasculitis in rats was induced by sodium laurate and graded by gross appearance on the 12th day after injection. Histological changes in rat femoral arteries were assessed by histopathology and transmission electron microscopy. Blood cell counts, blood rheology, blood coagulation and plasma urocortin, thromboxane B<sub>2</sub>, prostaglandin E<sub>2</sub> and soluble intercellular adhesion molecule-1 levels were measured. Expression of urocortin, corticotrophin-releasing factor (CRF<sub>1/2</sub>) receptors, cyclooxygenase (COX)-2 and intercellular adhesion molecule-1 (ICAM-1) at both mRNA and protein levels were determined by RT-PCR and Western blot.

**Key results:** Rats showed grossly visible signs and symptoms of TAO on the 12th day after sodium laurate injection. In these rats, blood was in a hypercoagulable state; plasma urocortin, prostaglandin E<sub>2</sub> and soluble intercellular adhesion molecule-1 levels were elevated; and the expression of urocortin, CRF<sub>1</sub> and CRF<sub>1α</sub>-receptors, COX-2 and ICAM-1 in rat femoral arteries were markedly increased. Exogenous urocortin, given for 12 days after sodium laurate, exacerbated the hypercoagulable state and augmented expression of CRF<sub>1α</sub>-receptors, COX-2 and ICAM-1. These effects were abolished by a CRF<sub>1</sub>-receptor antagonist, NBI-27914, or a non-selective CRF-receptor antagonist, astressin, but not by the CRF<sub>2</sub>-receptor antagonist, antisauvagine-30, given with exogenous urocortin.

**Conclusion and implications:** Urocortin exacerbated the hypercoagulable state and vasculitis in a model of TAO induced by sodium laurate in rats, via CRF<sub>1</sub>-receptors. COX-2 and ICAM-1 might also have contributed to this exacerbation.

*British Journal of Pharmacology* (2009) **157**, 1368–1379; doi:10.1111/j.1476-5381.2009.00210.x; published online 30 June 2009

**Keywords:** Urocortin; thromboangiitis obliterans; vasculitis; CRF<sub>1</sub>-receptor; CRF<sub>2</sub>-receptor

**Abbreviations:** CRF, corticotrophin-releasing factor; CRF<sub>1</sub> receptor, corticotrophin-releasing factor type 1 receptor; CRF<sub>2</sub> receptor, corticotrophin-releasing factor type 2 receptor; ICAM-1, intercellular adhesion molecule-1; PG, prostaglandin; TAO, thromboangiitis obliterans; TXA<sub>2</sub>, thromboxane A<sub>2</sub>; TXB<sub>2</sub>, thromboxane B<sub>2</sub>

## Introduction

Vasculitis is a pathological process characterized by inflammatory damage to blood vessels, which may be induced by genetic abnormalities, autoimmune or physical damage.

Thromboangiitis obliterans (TAO) is a non-atherosclerotic, segmental, inflammatory disease that most commonly affects the small- and medium-sized arteries, veins and nerves of the extremities (Olin, 2000). In the characteristic acute-phase lesion, in association with occlusive cellular thrombosis, the acute inflammation that involves all layers of the vessel wall leads TAO to be classified as a vasculitis (Puéchal and Fiessinger, 2007). However, the exact aetiology for TAO is not yet well defined (Olin, 2000). In sodium laurate-induced arterial occlusive disease of rats, the injected sodium laurate is supposed to cause endothelial cell damage that may lead to the aggregation of platelets in peripheral vascular beds

Correspondence: Dr Shengnan Li, Department of Pharmacology, Nanjing Medical University, 140 Hanzhong Road, Nanjing 210029, China. Email: snli@njmu.edu.cn

\*These authors contributed equally to this work.

Received 27 November 2008; revised 12 January 2009; accepted 15 January 2009

(Ashida *et al.*, 1980). The progression of the disease in this model of vasculitis resembles that reported in the patients with TAO (Nakata *et al.*, 1976; Nielubowicz *et al.*, 1980).

Urocortin, a peptide of the corticotrophin-releasing factor (CRF) family (Vaughan *et al.*, 1995), is expressed in both central and peripheral tissues (Fekete and Zorrilla, 2007). Besides its cardiovascular protective property (Okosi *et al.*, 1998; Oki and Sasano, 2004), urocortin is now considered to be a potent immunomodulatory factor that participates in immune responses (Fekete and Zorrilla, 2007). In contrast to its systemic indirect immunosuppressive effects on the hypothalamic–pituitary adrenal axis, urocortin acts as a locally expressed, autocrine or paracrine, pro-inflammatory factor in a series of inflammatory diseases, such as rheumatoid arthritis and osteoarthritis (Kohno *et al.*, 2001) and ulcerative colitis (Saruta *et al.*, 2004). Moreover, it can stimulate the release of pro-inflammatory mediators under inflammatory conditions (Saruta *et al.*, 2004; Theoharides *et al.*, 2004). Our previous study demonstrated the overexpression of urocortin in lung tissues of rats with allergic asthma and inhalation of aerosolized urocortin increased pulmonary vascular permeability via mast cell infiltration and activation (Wu *et al.*, 2006). This process was mediated by CRF receptors (CRF<sub>1</sub> and CRF<sub>2</sub>) (Singh *et al.*, 1999; nomenclature follows Alexander *et al.*, 2008). We demonstrated for the first time that mast cell degranulation induced by urocortin was mediated by increasing intracellular calcium concentration via CRF<sub>1</sub>-receptors (Wu *et al.*, 2008). Moreover, our data showed that CRF played a significant role in promoting the development of atherosclerosis (Wu *et al.*, 2009), which also exhibits features of vasculitis (Cipollone and Fazio, 2006). All these results suggested that endogenous urocortin might participate in the pathophysiology of many inflammatory conditions, as a pro-inflammatory mediator.

We therefore hypothesized that urocortin, which is synthesized and secreted by the systemic vasculature, may be a potent, locally expressed, pro-inflammatory factor in TAO, which is classified as a vasculitis with endothelial dysfunction (Joras *et al.*, 2006). In this study, we assessed the role of urocortin in a rat model of TAO induced by the intravascular injection of sodium laurate. We also investigated the type of CRF receptors involved in these effects of urocortin.

## Methods

### Animals

All animal care and this investigation conform to the *Guide for the Care and Use of Laboratory Animals* published by the US National Institutes of Health (NIH Publication No. 85–23, revised 1996), and were approved by the Ethical Review Board of Nanjing Medical University. Male Wistar rats (200–250 g) were purchased from Shanghai laboratory animal centre. Food and water were given *ad libitum* unless otherwise noted.

### Experimental protocol

Male Wistar rats were randomly divided into seven groups (eight animals per group) as follows: (i) normal group; (ii) sham operated group; (iii) vasculitis (TAO model) group; (iv)

urocortin group; (v) urocortin + NBI-27914 group; (vi) urocortin + antisauvagine-30 group; and (vii) urocortin + astressin group. Rats with sodium laurate-induced vasculitis were prepared according to the method described previously (Ashida *et al.*, 1980). Briefly, rats were anaesthetized with 10% (w/v) chloral hydrate (3.5 mL·kg<sup>-1</sup>, i.p.). The left hind leg was shaved and the femoral artery was exposed by surgical incision and retraction of muscles. Sodium laurate solution (10 mg·mL<sup>-1</sup> in normal saline, adjusted to pH 8.0, 0.1 mL per animal) was injected into the left femoral artery while the sham operated group were treated with normal saline as vehicle. All groups were subjected to the surgical procedure except for the normal group. Urocortin (3 nmol·kg<sup>-1</sup>) and CRF<sub>1/2</sub> receptor antagonists (15 nmol·kg<sup>-1</sup>) (Li *et al.*, 2008) were given s.c., 2 h after sodium laurate injection and once a day for the following 11 days.

The gross appearance of the rat hind legs was checked daily after the operation. The degree of the disease on the 12th day was graded according to the system developed by Murakami *et al.* (1995), as follows: 0, normal appearance; 1, change in nail colour; 2, change in digit colour; 3, gangrene of digit; 4, loss or mummification of digit.

At the end of the experiment, rats were anaesthetized with 10% chloral hydrate. A blood sample was drawn from the common carotid artery and anti-coagulated with heparin, sodium citrate or EDTA, depending on the variable measured. Plasma was frozen at –80°C for biochemical analysis. Left femoral arteries were collected and divided into two: one was frozen at –80°C until used to provide a homogenate; the other was taken for histological examination.

### Assessment of blood cell counts, blood coagulation and blood rheology

Blood cell counts, blood coagulation and blood rheology were measured by the clinical laboratory in the First Affiliated Hospital of Nanjing Medical University.

### Plasma urocortin, thromboxane B<sub>2</sub>, prostaglandin E<sub>2</sub> and soluble intercellular adhesion molecule-1 assays

Plasma urocortin, prostaglandin (PG)E<sub>2</sub> and soluble intercellular adhesion molecule (sICAM)-1 were determined by ELISA Kits (purchased from Phoenix Pharmaceuticals, USA; R&D Systems, USA; Boster Biological Technology, China respectively). Plasma thromboxane (TX)B<sub>2</sub> was measured with an enzyme immunoassay kit (Cayman Chemical, USA) according to the manufacturer's protocol.

### Assessment by light and transmission electron microscopy

For light microscopy, samples of rat femoral artery were fixed in 10% neutral formalin for at least 24 h. After being embedded in paraffin, the tissues were cut into 5-µm-thick sections and stained with haematoxylin and eosin before examination.

For electron microscopy, samples of arteries were immediately placed in sodium acetate-buffered 2.5% glutaraldehyde solution and left overnight. The subsequent procedures were kindly performed by the Key Laboratory of Antibody

**Table 1** Summary of the RT-PCR primer sequences used to amplify GAPDH, urocortin, CRF<sub>1</sub>-receptors, CRF<sub>2</sub>-receptors, COX-2 and ICAM-1 from rat tissues

		Sequences	Product size (bp)	Annealing T(C)
GAPDH	Sense	TCCCAGAGCTGAACGGGAGCTCACTG	339	68.1
	Antisense	TGGAGGCCATGTAGGCCATGAGGTCCA		
Urocortin	Sense	GCTACGCTCCTGGTAGCGTTGCTGCTTCTG	356	68.1
	Antisense	GCCGATCACTTGCCACCGAGTGAATATG		
CRF <sub>1</sub>	Sense	AAGGCGGATCCAGGCAGTAGAGA	508	60.8
	Antisense	TCCCGGTAGCCATTGTTTGTCTGTG		
CRF <sub>1α</sub>	Sense	TCCTACGCAACGCCAC	147	58.8
	Antisense	AGCAGCCCTCACCGAAC		
CRF <sub>2</sub>	Sense	CTGGTGGCTGCTTTCCTGCTTTTC	425	58.1
	Antisense	ATGGGGCCCTGGTAGATGTAGTCC		
COX-2	Sense	TTCACCAGACAGATTGCTGGC	530	63.5
	Antisense	AGTCTGGAGTGGGAGGCACTTG		
ICAM-1	Sense	AGAAGGACTGCTTGGGGAA	332	58.1
	Antisense	CCTCTGGCGGTAATAGGTG		

CRF, corticotrophin-releasing factor; COX, cyclooxygenase; ICAM-1, intercellular adhesion molecule-1.

Technique, Ministry of Health, Nanjing Medical University. Artery specimens were then cut into 0.5- $\mu$ m-thick sections, the sections stained with uranyl acetate and lead citrate and examined in a JEOL 2000FX transmission electron microscope operated at 80 kV.

#### Semi-quantitative RT-PCR analysis

Total RNAs were extracted from femoral arteries, using TRIzol (Invitrogen) according to the manufacturer's protocol. For cDNA synthesis, Moloney murine leukemia virus (Invitrogen) was applied as the reverse transcriptase. For PCR reaction, Taq DNA polymerase (Promega) was used in the reaction system. Primers applied in the experiment were synthesized from published sequences (Baigent and Lowry, 2000; Ohnaka *et al.*, 2000; Taal *et al.*, 2000; Brar *et al.*, 2004) (Table 1). The products were analysed by electrophoresis in 2.0% agarose gel containing 0.5  $\mu$ g·mL<sup>-1</sup> ethidium bromide. Specific genes were verified by their predicted sizes. GAPDH was set as the internal control.

#### Western blot analysis

The protein samples were separated on a 10% SDS-polyacrylamide gel and electrophoretically transferred to PVDF membranes in Tris-glycine transfer buffer. Then membranes were blocked in 5% (wt/vol) instant non-fat dried milk for 2 h at room temperature, and incubated with primary antibody for CRF<sub>1</sub>-receptor (1:400), CRF<sub>2</sub>-receptor (1:400), COX-2 (1:1000), ICAM-1 (1:500) or  $\beta$ -actin (1:200, Boster Biological Technology) at 4°C overnight. After three washes with TBST (50 mmol·L<sup>-1</sup> Tris-HCl, pH 7.5, 150 mmol·L<sup>-1</sup> NaCl, 0.05% Tween 20), the membranes were incubated with secondary horseradish peroxidase-conjugated IgG for 2 h at room temperature. Immunoreactive proteins were visualized by LumiGLO chemiluminescent reagent and peroxide (Cell Signaling Technology). The light-emitting bands were detected with X-ray films (Kodak, Rochester, NY).

#### Statistical analysis

Data were expressed as means  $\pm$  SEM. The significance for the difference among groups was analysed with SPSS 11.0 by one-

way ANOVA with Student–Newman–Keuls multiple comparison methods. The Kruskal–Wallis *H* test was used to compare the gross appearance of the vasculitis among groups. *P* < 0.05 was considered statistically significant.

#### Materials

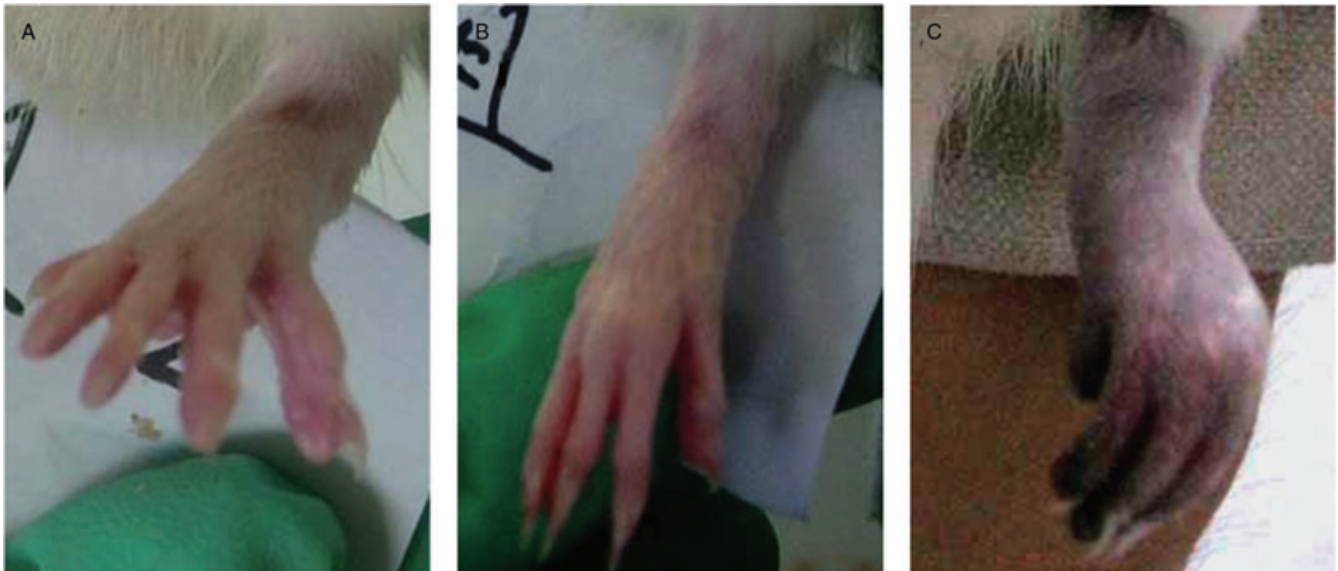
Rat urocortin, CRF<sub>1</sub>-receptor antagonist NBI-27914, CRF<sub>2</sub>-receptor antagonist, antisauvagine-30 and the non-selective CRF-receptor antagonist, astressin were purchased from Sigma (Missouri, USA). Sodium laurate was provided by International Laboratory (California, USA). Polyclonal urocortin and antibodies to CRF<sub>1</sub>- and CRF<sub>2</sub>-receptors were obtained from Santa Cruz Biotechnology (California, USA). Polyclonal COX-2 and monoclonal ICAM-1 antibodies were from Abcam (Cambridge, UK). All other reagents used were derived from commercial sources.

## Results

#### Rats showed gross signs and symptoms of TAO and histological changes after sodium laurate injection

The rat model of TAO, induced by sodium laurate, has been widely accepted. In our study, as reported previously (Nakata *et al.*, 1976; Nielubowicz *et al.*, 1980), the whole left hind paw went pale 2 or 3 min after injection of sodium laurate. On the 12th day, in sham operated and normal rats (Figure 1A and B), no ischaemic signs were found; while rats in the vasculitis (TAO model) group showed typical signs and symptoms of TAO (Figure 1C), with four rats displaying obvious signs of gangrene, and two with loss or mummification of paws.

Histological changes were also observed in sections of the femoral artery examined by light microscopy (Figure 2A and B). The sections from the TAO model rats (Figure 2B), stained with haematoxylin and eosin, showed obvious thrombi, which resulted in the narrowing or complete occlusion of the vessel lumen, and some showed recanalization of the artery. To investigate ultrastructural changes in the arterial tissue, sections were examined by transmission electron microscopy.



**Figure 1** Gross appearance of rat paws in normal, sham operated and thromboangiitis obliterans (TAO) model rats on the 12th day after sodium laurate injection. Paws in normal (A), sham operated (B) and TAO model (C) rats.

The endothelium in sections from normal rats looked intact with regular arrangement of the endothelial cells [Figure 2C (a)]; while in arteries from the TAO model rats, the artery intima was modified with gaps in the endothelial sheet and desquamation in many places, and proliferation and hypertrophy of endothelium were also extensively found [Figure 2C (b)]. Under the intima, smooth muscle cells were in a regular order in normal arteries [Figure 2C (c)]; while in arteries from the TAO group, muscle cells were disordered and most showed multiformity and degeneration [Figure 2C (d)]. These findings were comparable with those found in TAO patients (Cui Pan Cui *et al.*, 2000).

#### *Urocortin and CRF<sub>1</sub>-receptors were increased in TAO model rats*

As shown in Figure 3, plasma urocortin was increased markedly in the TAO model group, compared with the levels in the sham operated or normal groups ( $P < 0.05$ ). There was no difference between normal and sham operated group.

RT-PCR analysis (Figure 4A and B) showed that the mRNA for urocortin and CRF<sub>1</sub>-receptors were increased in femoral arteries from the TAO group relative to the levels found in the normal and sham operated rat femoral arteries. We also showed that mRNA for the CRF<sub>1α</sub>-receptor was increased in the model rats (Figure 4A and B). The levels of mRNA for the CRF<sub>2</sub>-receptor were not significantly altered in samples from the TAO group.

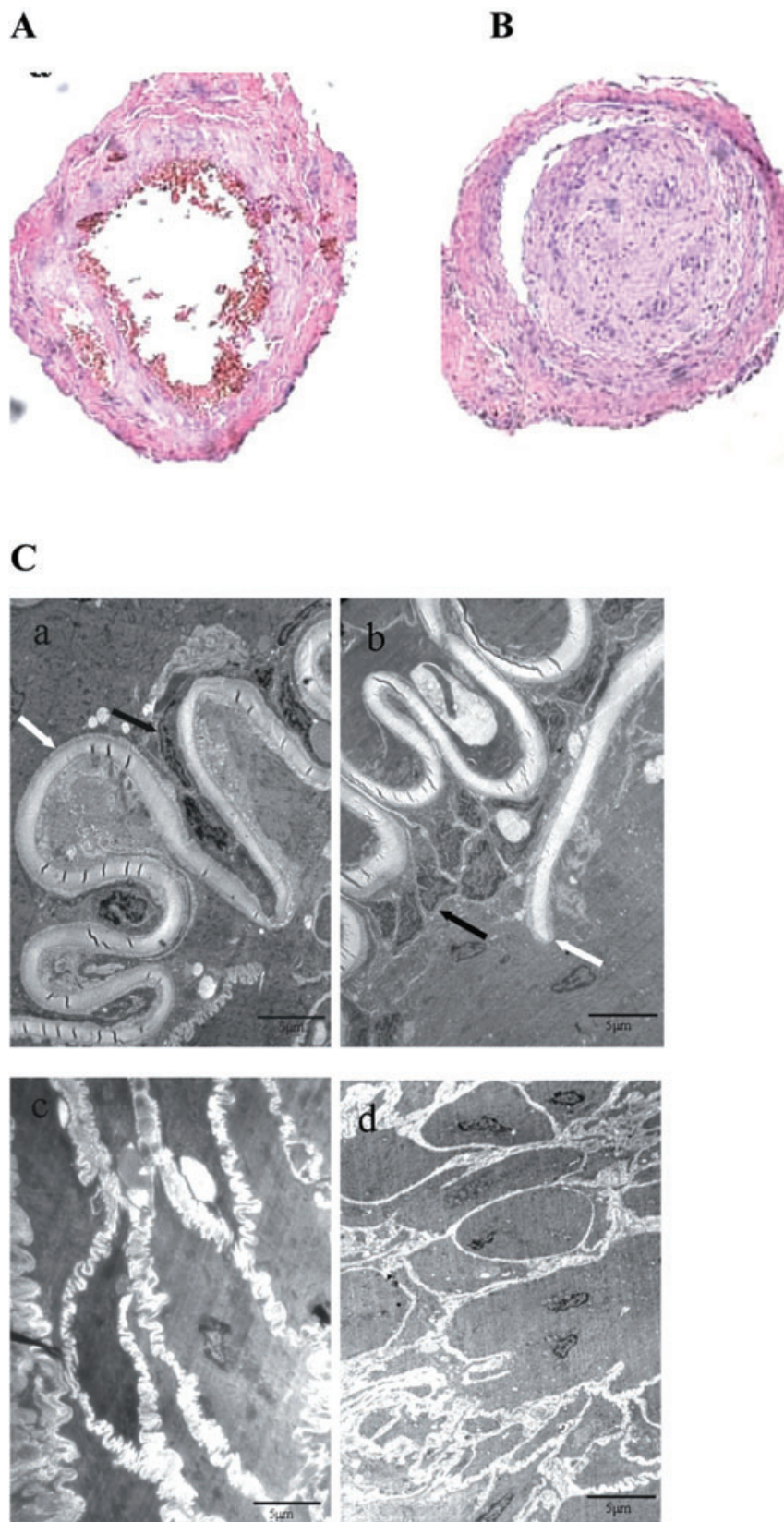
Subsequently, we also measured protein levels of CRF<sub>1</sub>- and CRF<sub>2</sub>-receptors in rat femoral arteries by Western blot analysis. As shown in Figure 4C and D, compared with sham operated group, the expression of CRF<sub>1</sub>-receptor protein in the TAO model group was significantly increased ( $P < 0.05$ ), while the expression of CRF<sub>2</sub>-receptor protein remained constant.

#### *Effects of exogenous urocortin and CRF<sub>1/2</sub> receptor antagonists on TAO model rats*

In order to ascertain whether urocortin contributed to the TAO model, exogenous urocortin and CRF<sub>1/2</sub> receptor antagonists were added to rats treated with sodium laurate. As shown in Figure 5A, adding exogenous urocortin for 12 days to the TAO model rats exacerbated the gross effects of the ischaemia, with three rats displaying obvious signs of gangrene and three with loss or mummification of paws. These signs of intensified ischaemia were markedly decreased in the groups given the CRF<sub>1</sub>-receptor antagonist, NBI-27914 (Figure 5B) or the non-selective CRF-receptor antagonist, astressin (Figure 5D) along with the exogenous urocortin. However, the CRF<sub>2</sub>-receptor antagonist, antisauvagine-30 (Figure 5C), did not modify the gross effects of exogenous urocortin.

To quantify the gross effects of ischaemia in each group, these effects were graded into five levels, as described in the *Methods* and shown in Table 2. Rats from each group were graded on the 12th day and the group grades compared, using the Kruskal–Wallis  $H$  test. Compared with the values from the sham operated group, those from the TAO model and TAO + urocortin groups were significantly higher, that is, more intense signs of ischaemia. In agreement with the gross observations, the grades in the groups given NBI-27914 or astressin were lower and not different from those in the sham operated group. No such attenuation of ischaemic signs was present in the group given antisauvagine-30 and no statistical significance was found between normal and sham operated groups (Table 2). These data indicate that antagonism of CRF<sub>1</sub>-receptors, but not of CRF<sub>2</sub>-receptors, could alleviate the gross signs of the vasculitis.

In further analysis of the effects of exogenous urocortin and CRF<sub>1/2</sub> receptor antagonists in the TAO model rats, blood cell counts, blood coagulation and blood rheology of each group were investigated. Blood platelet count was markedly elevated

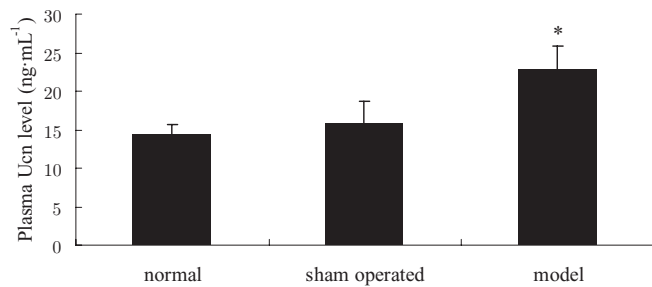


**Figure 2** Histological examination of femoral arteries in normal (A) and thromboangiitis obliterans (TAO) model (B) rats (haematoxylin and eosin staining, magnification: 100 $\times$ ). In (C), ultrastructure of femoral arteries in normal (a and c) and TAO model (b and d) rats. a and b, artery intima; c and d, medial membrane. Black arrows show endothelial cells and white arrows show inner elastic layer. In picture b, black arrow indicates endothelial cell proliferation, and the white arrow indicates discontinuity of the internal elastic lamina (transmission electron microscopy; magnification: 4000 $\times$ ).

( $P < 0.05$ ) in TAO model group (Table 3A). As expected, blood coagulation parameters (Table 3B) in the TAO model rats were very different from those in the sham operated group: prothrombin time, thrombin time and activated partial thromboplastin time were all significantly shortened while fibrinogen and D-dimer values were increased. Compared with the TAO model group, adding exogenous urocortin further shortened prothrombin time, thrombin time, activated partial thromboplastin time and increased blood platelet count, fibrinogen and D-dimer levels. Treatment with NBI-27914 and astressin reversed those parameters, to almost

normal levels. However, it seems that urocortin had no influence on other parameters of rat blood cells and on blood rheology (Table 3C). Compared with the TAO model or urocortin groups, treatment with antisauvagine-30 had no significant effect on any of the parameters measured.

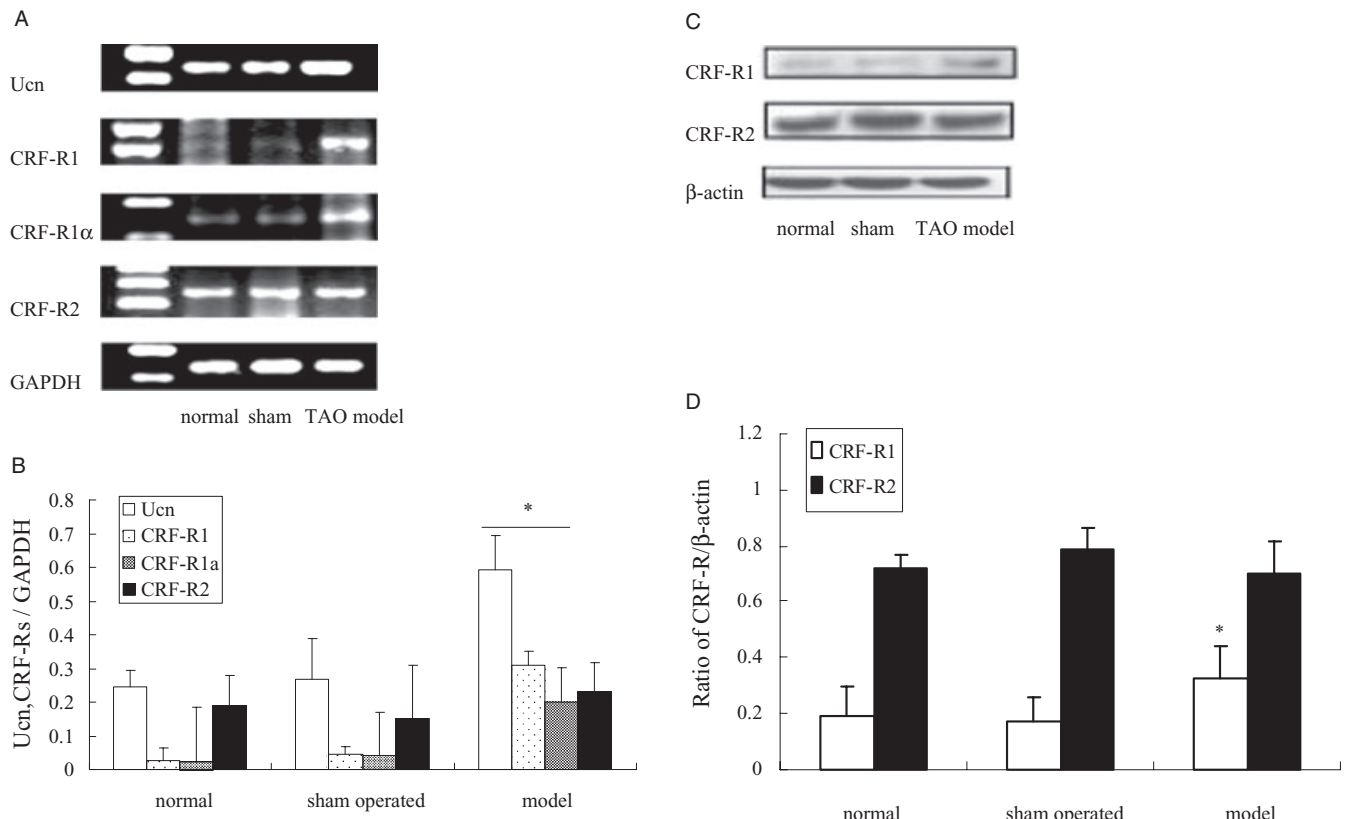
TXA<sub>2</sub>, a potent inducer of platelet aggregation and microvascular contraction (Silver *et al.*, 1973; 1974), was measured as its metabolite, TXB<sub>2</sub>. As shown in Figure 6, plasma TXB<sub>2</sub> was markedly raised in the TAO model group, over the sham or normal values ( $P < 0.05$ ), and was further elevated on addition of exogenous urocortin. Treatment with the CRF receptor antagonists, NBI-27914 or astressin but not antisauvagine-30, dramatically reduced plasma TXB<sub>2</sub> levels, relative to the TAO model group ( $P < 0.05$ ).



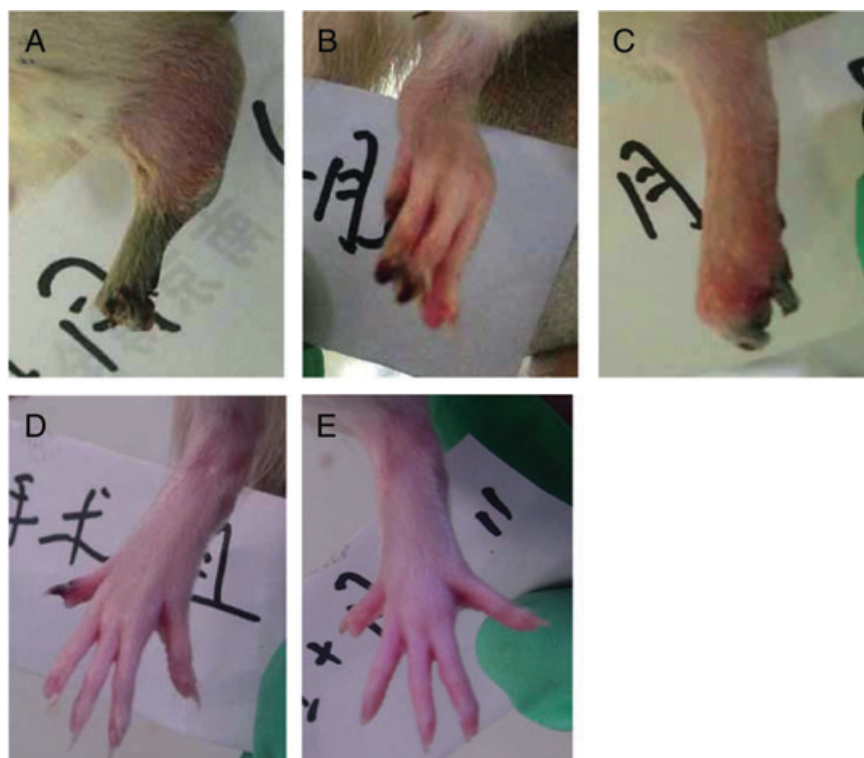
**Figure 3** Plasma urocortin (Ucn) levels in normal, sham operated and thromboangiitis obliterans model groups. Data are presented as means  $\pm$  SEM ( $n = 8$ ). \* $P < 0.05$ , versus sham operated group.

*Effects of urocortin and CRF<sub>1/2</sub> receptor antagonists on expression of COX-2 and ICAM-1*

COX-2 and ICAM-1 are two important components of many inflammatory diseases (Dubois *et al.*, 1998; Vane *et al.*, 1998; Witkowska, 2005). Clinical reports indicate that ICAM-1 expression is elevated in TAO patients (Halacheva *et al.*, 2002; Joras *et al.*, 2006) and treatment with COX-2 inhibitors can relieve the disease condition (Nizankowski *et al.*, 1985; Fiessinger and Schäfer, 1990). In the present study, RT-PCR



**Figure 4** Expression of mRNA and Western blot analysis for urocortin and corticotrophin-releasing factor (CRF)<sub>1/2</sub> receptors in femoral arteries from normal, sham operated and thromboangiitis obliterans (TAO) model rats. In (A), a representative record of the expression of mRNA is shown (Ucn, urocortin; CRF-R1, CRF<sub>1</sub>-receptor; CRF-R1 $\alpha$ , CRF<sub>1 $\alpha$</sub> -receptor; CRF-R2, CRF<sub>2</sub>-receptor) with GAPDH as an internal control. Summary data are shown in (B); data are presented as means  $\pm$  SEM ( $n = 8$ ). \* $P < 0.05$ , versus sham operated group. In (C), a representative Western blot for CRF<sub>1</sub>-receptors and CRF<sub>2</sub>-receptors is shown ( $\beta$ -actin as control) with summary data in (D); data presented as means  $\pm$  SEM ( $n = 8$ ). \* $P < 0.05$ , versus sham operated group.



**Figure 5** Gross appearance of rat paws after urocortin and corticotrophin-releasing factor type 1 and 2 receptor antagonists (12th day). A, urocortin group; B, urocortin + NBI-27914 group; C, urocortin + anti-sauvagine-30 group; D, urocortin + astressin group; E, sham operated group.

**Table 2** Grades of disease on the 12th day after sodium laurate injection

	Grade of disease					Significance
	0	1	2	3	4	
Normal	8	0	0	0	0	
Sham operated	8	0	0	0	0	
TAO model	2	0	0	4	2	**
Ucn	1	1	0	3	3	**
Ucn + NBI-27914	5	0	0	2	1	†
Ucn + antisauvagine-30	2	0	2	1	3	**
Ucn + astressin	5	1	1	1	0	†#

On the 12th day after sodium laurate injection, rats from each group were graded according to the gross appearance of the affected paw (see *Methods*). Kruskal–Wallis *H* test was carried out to compare the disease condition among groups.

\*\* $P < 0.01$ , versus sham operated group; # $P < 0.05$ , versus TAO model group; † $P < 0.05$ , versus Ucn group.

TAO, thromboangiitis obliterans; TAO model, single injection of sodium laurate only; Ucn, TAO model + daily urocortin for 12 days; Ucn + NBI 27914 or + antisauvagine-3 or + astressin, Ucn + daily injections of each antagonist for 12 days.

analysis (Figure 7) showed that mRNA for COX-2 and ICAM-1 extracted from femoral arteries were increased in the TAO model and urocortin groups, compared with the sham operated group ( $P < 0.05$ ) and that treatment with NBI-27914 or astressin but not antisauvagine-30 attenuated the increase due to urocortin group ( $P < 0.05$ ). Western blot assays to determine protein levels of COX-2 and ICAM-1 (Figure 8)

gave results largely in accordance with the RT-PCR data. Again, both NBI-27914 and astressin but not antisauvagine-30 blocked the effect of exogenous urocortin ( $P < 0.05$ ) on expression of COX-2 and ICAM-1.

We also measured plasma PGE<sub>2</sub>, the main metabolite of COX-2 and sICAM-1 in the experimental groups of rats. As shown in Figure 9, in accordance with the expression of COX-2 and ICAM-1 in rat femoral arteries, plasma PGE<sub>2</sub> and sICAM-1 were enhanced in the TAO model group, compared with the sham operated group ( $P < 0.05$ ) and were further increased on addition of exogenous urocortin ( $P < 0.05$  vs. TAO model group). As observed earlier, the CRF-receptor antagonists NBI-27914 and astressin but not antisauvagine-30 abolished these effects of urocortin ( $P < 0.05$ ).

## Discussion

Urocortin is a peptide of the CRF family. In our model of TAO in rats, urocortin potentiated the effects of the peripheral vasculitis and ischaemia, which could be attributed to exacerbation of the hypercoagulable state of the blood. This exacerbation was mediated by CRF<sub>1</sub>-receptors, probably the CRF<sub>1α</sub>-receptor subtype.

Urocortin was originally identified in the CNS in 1995 (Vaughan *et al.*, 1995) and more recently, it was found to be widely expressed in the periphery (Fekete and Zorrilla, 2007). Except for its protective roles in some diseases (Oki and Sasano, 2004), urocortin has been found to play a pro-inflammatory role in some inflammatory diseases, such as

**Table 3A** Effects of urocortin and CRF<sub>1/2</sub> receptor antagonists on rat blood cell counts

	Blood platelet count (10 <sup>8</sup> mL <sup>-1</sup> )	Red blood cell count (10 <sup>9</sup> mL <sup>-1</sup> )	Haemoglobin (g·L <sup>-1</sup> )	Leucocyte count (10 <sup>6</sup> mL <sup>-1</sup> )	Neutrophil (10 <sup>6</sup> mL <sup>-1</sup> )
Normal	7.12 ± 0.31	6.51 ± 0.31	131.25 ± 6.17	4.93 ± 0.21	1.08 ± 0.20
Sham operated	7.35 ± 0.51	6.21 ± 0.49	128.22 ± 4.73	5.01 ± 0.40	1.16 ± 0.11
TAO model	9.30 ± 0.49*	6.11 ± 0.63	129.98 ± 7.44	5.11 ± 0.61	1.19 ± 0.28
Ucn	11.71 ± 0.72*#	6.23 ± 0.14	130.05 ± 5.10	5.08 ± 0.58	1.09 ± 0.09
Ucn + NBI-27914	7.87 ± 0.62#†	6.83 ± 0.72	135.12 ± 9.39	4.95 ± 0.13	1.13 ± 0.19
Ucn + antisauvagine-30	11.07 ± 0.82	6.61 ± 0.42	132.26 ± 8.17	5.00 ± 0.46	1.14 ± 0.24
Ucn + astressin	7.61 ± 0.71#†	6.47 ± 0.55	128.78 ± 6.85	4.97 ± 0.71	1.15 ± 0.17

Values are presented as means ± SEM (*n* = 8). \**P* < 0.05, versus sham operated group; #*P* < 0.05, versus model group; †*P* < 0.05, versus urocortin group. CRF<sub>1/2</sub>, corticotrophin-releasing factor type 1 and 2 receptors; TAO, thromboangiitis obliterans; TAO model, single injection of sodium laurate only; Ucn, TAO model + daily urocortin for 12 days; Ucn + NBI 27914 or + antisauvagine-3 or + astressin, Ucn + daily injections of each antagonist for 12 days.

**Table 3B** Effects of urocortin and CRF<sub>1/2</sub> receptor antagonists on rat blood coagulation

	Prothrombin time (s)	Thrombin time (s)	Activated partial thromboplastin time(s)	Fibrinogen level (g·L <sup>-1</sup> )	D-dimer (ng·mL <sup>-1</sup> )
Normal	37.93 ± 1.61	25.27 ± 2.12	38.91 ± 3.10	2.98 ± 0.11	0.60 ± 0.02
Sham operated	38.20 ± 0.98	24.00 ± 1.91	35.90 ± 2.74	3.12 ± 0.20	0.57 ± 0.06
TAO model	26.50 ± 1.23*	10.33 ± 1.69*	20.03 ± 2.18*	5.94 ± 0.47*	1.34 ± 0.10*
Ucn	20.20 ± 1.50*#	8.07 ± 1.78*#	15.11 ± 3.53*#	6.90 ± 0.35*#	1.66 ± 0.19*#
Ucn + NBI-27914	35.38 ± 0.72#†	22.65 ± 3.12#†	36.80 ± 4.61#†	3.53 ± 0.29#†	0.78 ± 0.09#†
Ucn + antisauvagine-30	21.01 ± 1.11	11.20 ± 2.19	18.31 ± 3.50	6.28 ± 0.61	1.58 ± 0.21
Ucn + astressin	36.83 ± 1.31#†	23.79 ± 1.70#†	38.77 ± 5.07#†	3.30 ± 0.42#†	0.59 ± 0.13#†

Values are presented as means ± SEM (*n* = 8). \**P* < 0.05, versus sham operated group; #*P* < 0.05, versus model group; †*P* < 0.05, versus urocortin group. CRF<sub>1/2</sub>, corticotrophin-releasing factor type 1 and 2 receptors; TAO, thromboangiitis obliterans; TAO model, single injection of sodium laurate only; Ucn, TAO model + daily urocortin for 12 days; Ucn + NBI 27914 or + antisauvagine-3 or + astressin, Ucn + daily injections of each antagonist for 12 days

**Table 3C** Effects of urocortin and CRF<sub>1/2</sub> receptor antagonists on rat blood rheology

	Erythrocyte sedimentation rate (mm·h <sup>-1</sup> )	Haematocrit (%)	Blood viscosity		Plasma viscosity (mPa s)
			200 S <sup>-1</sup> (mPa s)	5 S <sup>-1</sup> (mPa s)	
Normal	1.08 ± 0.23	40.88 ± 1.34	5.68 ± 0.03	11.34 ± 1.39	1.39 ± 0.03
Sham operated	1.15 ± 0.09	40.03 ± 0.31	5.77 ± 0.28	12.01 ± 2.80	1.34 ± 0.05
TAO model	1.11 ± 0.05	39.60 ± 0.18	5.81 ± 0.41	11.73 ± 0.47	1.36 ± 0.02
Ucn	1.09 ± 0.17	39.21 ± 0.99	5.69 ± 0.18	11.58 ± 1.19	1.39 ± 0.04
Ucn + NBI-27914	1.13 ± 0.20	39.67 ± 2.12	5.71 ± 0.33	11.21 ± 1.30	1.34 ± 0.04
Ucn + antisauvagine-30	1.16 ± 0.09	40.30 ± 2.08	5.61 ± 0.01	11.94 ± 1.71	1.31 ± 0.01
Ucn + astressin	1.01 ± 0.17	39.63 ± 1.62	5.73 ± 0.17	11.84 ± 1.55	1.33 ± 0.02

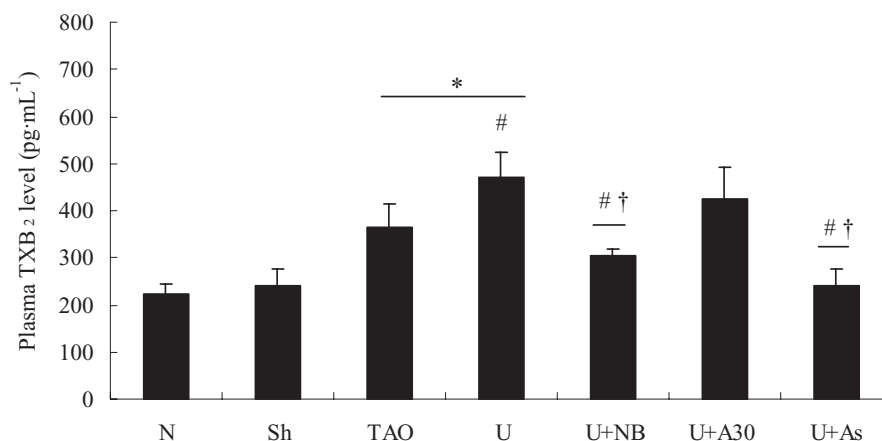
CRF<sub>1/2</sub>, corticotrophin-releasing factor type 1 and 2 receptors; TAO, thromboangiitis obliterans; TAO model, single injection of sodium laurate only; Ucn, TAO model + daily urocortin for 12 days; Ucn + NBI 27914 or + antisauvagine-3 or + astressin, Ucn + daily injections of each antagonist for 12 days Values are presented as means ± SEM (*n* = 8).

ulcerative colitis (Saruta *et al.*, 2004) and rheumatoid arthritis (Kohno *et al.*, 2001; Uzuki *et al.*, 2001). Our previous study demonstrated that CRF accelerated progression of atherosclerosis in LDLr<sup>-/-</sup> mice via CRF<sub>1</sub>-receptors (Wu *et al.*, 2009). However, its function in peripheral vasculitis and a potential role in modulating TAO progression are not well analysed.

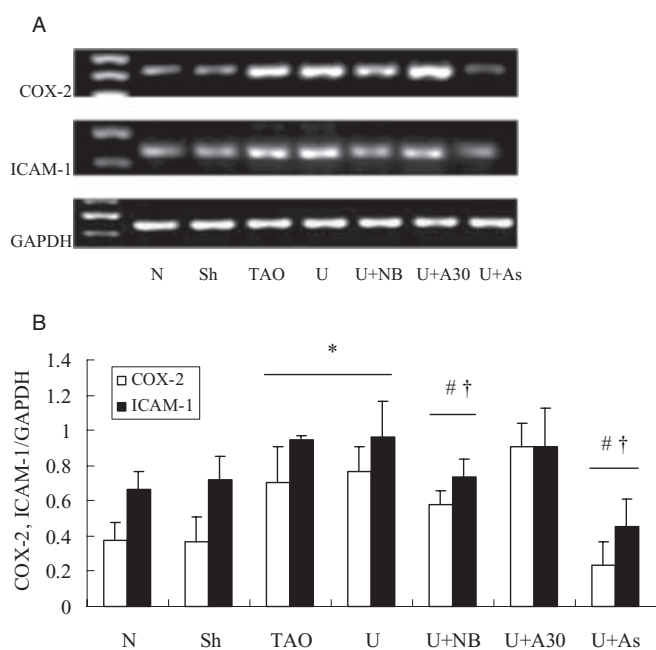
In the present study, rats with a sodium laurate-induced vasculitis were used to model TAO, which is also defined as a vasculitis. This model has previously been used to study TAO (Ashida *et al.*, 1980) and other groups have found that injection of sodium laurate into the rat femoral artery could mimic the signs and symptoms displayed in patients suffering from TAO (Nakata *et al.*, 1976; Nielubowicz *et al.*, 1980). Here, we observed that most of the model rats showed symptoms

typical of ischaemia and vasculitis in the hind limb, and that the femoral artery was structurally changed with thrombi or proliferation of endothelial cells. Our results were consistent with previous reports (Nakata *et al.*, 1976; Nielubowicz *et al.*, 1980; Cui Pan Cui *et al.*, 2000). We also found that expression of mRNA and protein for urocortin, CRF<sub>1</sub>- and CRF<sub>1α</sub>-receptors (but not for CRF<sub>2</sub>-receptors) were elevated in the femoral arteries from our TAO model rats, as was plasma urocortin. The CRF<sub>1α</sub>-receptor, widely expressed throughout the body, is the main mediator of the actions of CRF and urocortin (Teli *et al.*, 2008). Moreover, the CRF<sub>1α</sub>-receptor is the most efficient receptor in terms of stimulation of cAMP production, while the CRF<sub>1c</sub>- and CRF<sub>1β</sub>-receptors have a decreased CRF binding capacity (Pisarchik and Slominski, 2004). Taken together,





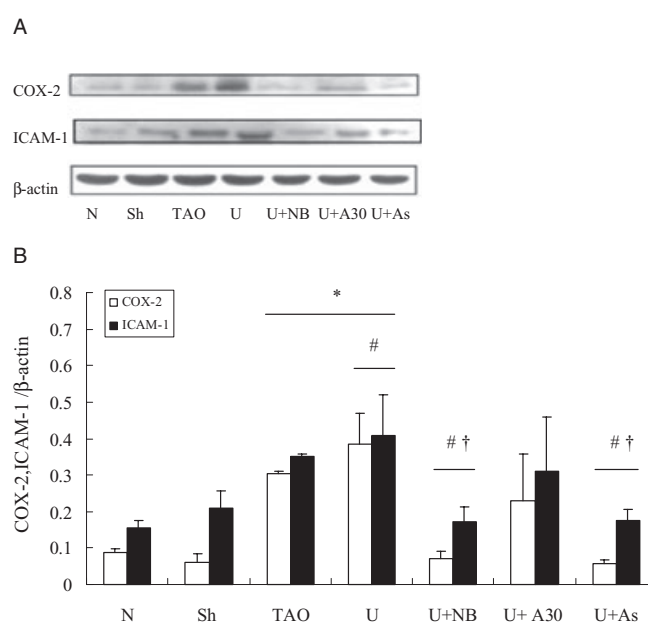
**Figure 6** Plasma thromboxane B<sub>2</sub> (TXB<sub>2</sub>) levels in experimental groups. TXB<sub>2</sub> levels were higher (than sham) in the thromboangiitis obliterans (TAO) model rats and further increased on addition of exogenous urocortin (Ucn; see *Methods*). These effects of urocortin were prevented by treatment with NBI-27914 or astressin, but not by antisauvagine-30. Values are presented as means  $\pm$  SEM ( $n = 8$ ). \* $P < 0.05$ , versus sham operated group; # $P < 0.05$ , versus TAO model group; † $P < 0.05$ , versus urocortin group. N, normal; Sh, sham; TAO, TAO model; U, urocortin; U + NB, U + NBI-27914; U + A30, urocortin + antisauvagine-30; U + As, urocortin + astressin.



**Figure 7** Expression of mRNA for cyclooxygenase (COX)-2 and intercellular adhesion molecule-1 (ICAM-1) in rat femoral arteries from experimental groups. In (A), a representative record of the expression of mRNA is shown, with GAPDH as an internal control. Summary data are shown in (B); data are presented as means  $\pm$  SEM ( $n = 8$ ). \* $P < 0.05$ , versus sham operated group; # $P < 0.05$ , versus TAO model group; † $P < 0.05$ , versus urocortin group. N, normal; Sh, sham; TAO, TAO model; U, urocortin; U + NB, U + NBI-27914; U + A30, urocortin + antisauvagine-30; U + As, urocortin + astressin.

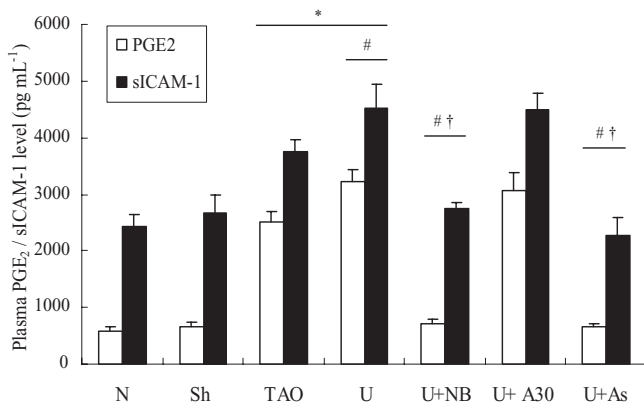
these observations suggest that urocortin does contribute to the signs and symptoms of TAO.

Previous reports have observed a hypercoagulable state of blood in TAO model rats (Shirakura *et al.*, 1994). In the present study, such a hypercoagulable state was observed in our rat model of vasculitis. Plasma TXA<sub>2</sub>, a potent inducer of platelet aggregation and microvascular contraction (Silver *et al.*, 1973; 1974), was measured as its stable product, TXB<sub>2</sub>,



**Figure 8** Western blot analysis of cyclooxygenase (COX)-2 and intercellular adhesion molecule-1 (ICAM-1) expression in rat femoral arteries from experimental groups. In (A), a representative blot is shown with  $\beta$ -actin as an internal control. Summary data are shown in (B); data are presented as means  $\pm$  SEM ( $n = 8$ ). \* $P < 0.05$ , versus sham operated group; # $P < 0.05$ , versus TAO model group; † $P < 0.05$ , versus urocortin group. N, normal; Sh, sham; TAO, TAO model; U, urocortin; U + NB, U + NBI-27914; U + A30, urocortin + antisauvagine-30; U + As, urocortin + astressin.

and was greatly elevated in the TAO model rats. This result could be mainly attributed to the augmentation of platelet aggregation, which is the main producer of TXA<sub>2</sub>. Previous studies indicated that serum levels of a soluble form of ICAM-1 (sICAM-1) were correlated with possibility of disseminated intravascular coagulation or maximum disseminated intravascular coagulation scores (Gando *et al.*, 2002; Chen *et al.*, 2005). Our data showed that plasma sICAM-1 was



**Figure 9** Plasma prostaglandin (PG)E<sub>2</sub> and soluble intercellular adhesion molecule (sICAM)-1 levels from experimental groups. Values are presented as means ± SEM (n = 8). \*P < 0.05, versus sham operated group; #P < 0.05, versus TAO model group; †P < 0.05, versus urocortin group. N, normal; Sh, sham; TAO, TAO model; U, urocortin; U + NB, U + NBI-27914; U + A30, urocortin + antisauvagine-30; U + As, urocortin + astressin.

increased in the TAO model rats, consistent with clinical findings of increased sICAM-1 in TAO patients (Joras *et al.*, 2006).

To investigate the role of urocortin in TAO, the TAO model rats were treated with exogenous urocortin and CRF<sub>1/2</sub> receptor antagonists. Adding urocortin significantly enhanced the grossly visible signs of ischaemia and vasculitis and further raised plasma TXB<sub>2</sub> and sICAM-1 levels. Treatment of these rats with the CRF<sub>1</sub>-receptor antagonist, NBI-27914 or the non-selective CRF-receptor antagonist, astressin, the gross signs of vasculitis were attenuated; plasma TXB<sub>2</sub> and sICAM-1 were returned to near normal and signs of the hypercoagulable state were decreased. When the rats were treated with the CRF<sub>2</sub>-receptor antagonist, antisauvagine-30, no such changes were observed.

From the gross, macroscopic findings (Figure 5 and Table 2), the non-selective CRF-receptor antagonist, astressin, could completely reverse whereas the selective CRF<sub>1</sub>-receptor antagonist, NBI-27914, could only partially reverse, the effects of urocortin and this might suggest a contribution from CRF<sub>2</sub>-receptors. However, selective blockade of CRF<sub>2</sub>-receptors with antisauvagine-30 had no significant effect. Overall, it is likely that urocortin promotes a hypercoagulable state, at least partially, through increasing plasma TXB<sub>2</sub> and sICAM-1 and that this promoting effect was modulated via CRF<sub>1</sub>-receptors, rather than CRF<sub>2</sub>-receptors.

COX-2, the inducible isoform of the rate-limiting enzyme in the biosynthesis of TXA<sub>2</sub> and PGs is well established as an important component of many inflammatory diseases (Dubois *et al.*, 1998; Vane *et al.* and., 1998). It plays a critical role in atherosclerosis and modulates atherosclerotic plaque stability through PG output (Cipollone and Fazio, 2006). In case-control studies, prostacyclin derivatives have shown to be more effective than placebo for the therapy of TAO patients (Nizankowski *et al.*, 1985; Fiessinger and Schäfer, 1990). Another key component of peripheral inflammatory disease, ICAM-1, is regarded as a marker for vasculitis (Witkowska, 2005), in conditions such as atherosclerosis (Wit-

kowska, 2005) and TAO (Halacheva *et al.*, 2002). In patients with TAO, inflammatory cells infiltrate local lesion sites, and ICAM-1 expression in endothelial cells and inflammatory sites was increased greatly (Halacheva *et al.*, 2002; Joras *et al.*, 2006). Our present study found that urocortin increased COX-2 and ICAM-1 expression in femoral arteries via CRF<sub>1</sub>-receptors, consistent with previous reports that in rat aortic smooth muscle cells, urocortin induced the expression of COX-2 in a time- and dose-dependent manner (Kageyama *et al.*, 2006). Also, CRF-related peptides (CRF and urocortin) up-regulated COX-2 expression and PG output in cultured human placental trophoblasts via CRF<sub>1</sub>-receptors (Gao *et al.*, 2008). Furthermore, CRF was biologically active in cultured keratinocytes and enhanced expression of hCAM and ICAM-1, stimulated by interferon-γ (Quevedo *et al.*, 2001).

Our present study showed that both endogenous and exogenous urocortin were involved in our model of TAO in rats. On the one hand, endogenous urocortin was increased in these TAO model rats. On the other hand, exogenous urocortin application exacerbated the condition of the TAO rats. Furthermore, treatment with NBI-27914 or with astressin, CRF-receptor antagonists, reduced the urocortin-enhanced TXB<sub>2</sub>, COX-2 and ICAM-1 expression to a level lower than those in the TAO model group, indicating antagonism of both endogenous and exogenous urocortin.

The pro-inflammatory role of CRF<sub>1</sub>-receptors in peripheral inflammatory sites has been demonstrated (Kohno *et al.*, 2001; Saruta *et al.*, 2004). Yokotani *et al.* (2001) found that activation of brain CRF<sub>1</sub>-receptors raised brain TXA<sub>2</sub>, which is involved in the central adrenomedullary outflow. In the synovium of rheumatoid arthritis patients, urocortin and CRF-receptors were overexpressed and were significantly correlated with the degree of inflammation (Kohno *et al.*, 2001). Furthermore, peripheral administration of CRF-receptor antibody (Karalis *et al.*, 1991) or the specific CRF<sub>1</sub>-receptor non-peptide antagonist, antalarmin (Webster *et al.*, 1996), prior to subcutaneous carrageenan injection, significantly suppressed inflammatory exudate volume and cell infiltration. Although there are reports indicating a protective role of urocortin in the cardiovascular system, this effect is mainly mediated via activating CRF<sub>2</sub>-receptors (Oki and Sasano, 2004). Recent studies have demonstrated that the CRF<sub>2</sub>-receptor is a tonic suppressor of vascularization (Bale *et al.*, 2002). Genetic deletion of CRF<sub>2</sub>-receptors resulted in profound postnatal hypervascularization in mice, which was characterized by both an increase in total vessel number and a dramatic increase in vessel diameter. Furthermore, CRF<sub>2</sub>-receptor activation can inhibit tumour growth via effects on vascularization and cell proliferation (Hao *et al.*, 2008; Wang *et al.*, 2008). However, in the present study, expression of mRNA or protein of CRF<sub>2</sub>-receptors were the same in normal, sham operated and TAO model groups. In addition, the selective CRF<sub>2</sub>-receptor antagonist, antisauvagine-30 had no obvious effect on the vasculitis or the hypercoagulable state in TAO rats.

Urocortin induced degranulation of rat lung mast cells by increasing intracellular calcium via CRF<sub>1</sub>-receptors (Wu *et al.*, 2008). In the present study, levels of CRF<sub>1</sub>-receptors were elevated in the TAO model group. This might represent both the infiltration of immune cells bearing CRF<sub>1</sub>-receptors and an effect of inflammatory mediators on CRF<sub>1</sub>-receptor

expression. On the one hand, the pro-inflammatory action of CRF peptides is partially the result of their effects on immune cells (such as mast cell, macrophages and lymphocytes) (Bamberger Wald Bamberger *et al.*, 1998; Theoharides *et al.*, 1998; Agelaki *et al.*, 2002), which can synthesize CRF peptides and express their receptors, CRF<sub>1</sub> and CRF<sub>2</sub> (Theoharides *et al.*, 1998; Baigent, 2001). On the other hand, inflammatory mediators could induce expression of CRF<sub>1</sub>-receptors (Inada *et al.*, 2009). Thus any increase of CRF<sub>1</sub>-receptors could be due to increased immune cells or to increased inflammatory mediators.

In conclusion, we have shown, in a model of TAO induced by sodium laurate in rats, that urocortin could exacerbate the pathological condition by potentiating the hypercoagulable state and the vasculitis. Blockade of CRF<sub>1</sub>-receptors attenuated the signs and symptoms of TAO by normalizing the hypercoagulable state. Up-regulation of COX-2 and ICAM-1 might also contribute to the exacerbation induced by urocortin.

## Acknowledgements

This work was supported by Key Project from Natural Scientific Foundation of Jiangsu Province (No. BK2006727), Project from New Century Excellent Talents (No. NCET-06-0507) and Project funded by the Summit of the 'Six Great Talents' of Jiangsu Province (06-C-017).

## Conflict of interest

The authors have no conflict of interest.

## References

Agelaki S, Tsatsanis C, Gravanis A, Margioris AN (2002). Corticotropin-releasing hormone augments proinflammatory cytokine production from macrophages in vitro and in lipopolysaccharide-induced endotoxin shock in mice. *Infect Immun* **70**: 6068–6074.

Alexander SPH, Mathie A, Peters JA (2008). Guide to receptors and channels (GRAC), 3rd edn. *Br J Pharmacol* **153** (Suppl. 2): S1–S209.

Ashida S, Ishihara M, Ogawa H, Abiko Y (1980). Protective effect of ticlopidine on experimentally induced peripheral arterial occlusive disease in rats. *Thromb Res* **18**: 55–67.

Baigent SM (2001). Peripheral corticotropin-releasing hormone and urocortin in the control of the immune response. *Peptides* **22**: 809–820.

Baigent SM, Lowry PJ (2000). mRNA expression profiles for corticotropin-releasing factor (CRF), urocortin, CRF receptors and CRF-binding protein in peripheral rat tissues. *J Mol Endocrinol* **25**: 43–52.

Bale TL, Giordano FJ, Hickey RP, Huang Y, Nath AK, Peterson KL *et al.* (2002). Corticotropin-releasing factor receptor 2 is a tonic suppressor of vascularization. *Proc Natl Acad Sci U S A* **99**: 7734–7739.

Bamberger CM, Wald M, Bamberger AM, Ergün S, Beil FU, Schulte HM (1998). Human lymphocytes produce urocortin, but not corticotropin-releasing hormone. *J Clin Endocrinol Metab* **83**: 708–711.

Brar BK, Chen A, Perrin MH, Vale W (2004). Specificity and regulation of extracellularly regulated Kinase1/2 phosphorylation through

Corticotropin-Releasing Factor (CRF) receptors 1 and 2 $\beta$  by the CRF/Urocortin Family of peptides. *Endocrinology* **145**: 1718–1729.

Chen DY, Lan JL, Lin FJ, Hsieh TY (2005). Association of intercellular adhesion molecule-1 with clinical manifestations and interleukin-18 in patients with active, untreated adult-onset Still's disease. *Arthritis Rheum* **53**: 320–327.

Cipollone F, Fazia ML (2006). COX-2 and Atherosclerosis. *J Cardiovasc Pharmacol* **47**: S26–S36.

Cui XM, Pan L, Cui L, Qi GC, Li ZB (2000). Ultrastructural diagnosis of thromboangiitis obliterans. *J Chin Electron Microsc Soc* **19**: 811–814.

Dubois RN, Abramson SB, Crofford L, Gupta RA, Simon LS, Van De Putte LB *et al.* (1998). Cyclooxygenase in biology and disease. *FASEB J* **12**: 1063–1073.

Fekete EM, Zorrilla EP (2007). Physiology, pharmacology, and therapeutic relevance of urocortins in mammals: ancient CRF paralogs. *Front Neuroendocrinol* **28**: 1–27.

Fiessinger JN, Schäfer M (1990). Trial of iloprost versus aspirin treatment for critical limb ischaemia of thromboangiitis obliterans: the TAO study. *Lancet* **335**: 555–557.

Gando S, Kameue T, Matsuda N, Hayakawa M, Ishitani T, Morimoto Y *et al.* (2002). Combined activation of coagulation and inflammation has an important role in multiple organ dysfunction and poor outcome after severe trauma. *Thromb Haemost* **88**: 943–949.

Gao L, Lu C, Xu C, Tao Y, Cong B, Ni X (2008). Differential regulation of prostaglandin production mediated by CRH receptor type 1 and type 2 in cultured human placental trophoblasts. *Endocrinology* **149**: 2866–2876.

Halacheva K, Gulubova MV, Manolova I, Petkov D (2002). Expression of ICAM-1, VCAM-1, E-selectin and TNF-alpha on the endothelium of femoral and iliac arteries in thromboangiitis obliterans. *Acta Histochem* **104**: 177–184.

Hao Z, Huang Y, Cleman J, Jovin IS, Vale WW, Bale TL *et al.* (2008). Urocortin2 inhibits tumor growth via effects on vascularization and cell proliferation. *Proc Natl Acad Sci USA* **105**: 3939–3944.

Inada Y, Ikeda K, Tojo K, Sakamoto M, Takada Y, Tajima N (2009). Possible involvement of corticotropin-releasing factor receptor signaling on vascular inflammation. *Peptides* **30**: 365–372.

Joras M, Poredos P, Fras Z (2006). Endothelial dysfunction in Buerger's disease and its relation to markers of inflammation. *Eur J Clin Invest* **36**: 376–382.

Kageyama K, Hanada K, Nigawara T, Moriyama T, Terui K, Sakihara S *et al.* (2006). Urocortin induces Interleukin-6 gene expression via Cyclooxygenase-2 activity in Aortic Smooth Muscle Cells. *Endocrinology* **147**: 4454–4462.

Karalis K, Sano H, Redwine J, Listwak S, Wilder RL, Chrousos GP (1991). Autocrine or paracrine inflammatory actions of corticotropin-releasing hormone in vivo. *Science* **254**: 421–423.

Kohn M, Kawahito Y, Tsubouchi Y, Hashiramoto A, Yamada R, Inoue KI *et al.* (2001). Urocortin expression in synovium of patients with rheumatoid arthritis and osteoarthritis: relation to inflammatory activity. *J Clin Endocrinol Metab* **86**: 4344–4352.

Li X, Hu J, Zhang R, Sun X, Zhang Q, Guan X *et al.* (2008). Urocortin ameliorates diabetic nephropathy in obese db/db mice. *Br J Pharmacol* **154**: 1025–1034.

Murakami T, Sawada K, Taneda K, Hayashi M, Katsuura Y, Tanabe H *et al.* (1995). Effect of isocarboxycyclin methyl ester incorporated in lipid microspheres on experimental models of peripheral obstructive disease. *Arzneimittelforschung* **45**: 991–994.

Nakata Y, Ban I, Hirai M, Shionoya S (1976). Onset and clinicopathological course in Burger's disease. *Angiology* **27**: 509–517.

Nielubowicz J, Rosnowski A, Pruszyński B, Przetakiewicz Z, Potemkowski A (1980). Natural history of Buerger's disease. *J Cardiovasc Surg* **21**: 529–540.

Nizankowski R, Królikowski W, Bielatowicz J, Szczeklik A (1985). Prostacyclin for ischemic ulcers in peripheral arterial disease. A random assignment, placebo controlled study. *Thromb Res* **37**: 21–28.

Ohnaka K, Numaguchi K, Yamakawa T, Inagami T (2000). Induction

- of Cyclooxygenase-2 by Angiotensin II in cultured rat vascular smooth muscle cells. *Hypertension* **35**: 68–75.
- Oki Y, Sasano H (2004). Localization and physiological roles of urocortin. *Peptides* **25**: 1745–1749.
- Okosi A, Brar BK, Chan M, D'Souza L, Smith E, Stephanou A *et al.* (1998). Expression and protective effects of urocortin in cardiac myocytes. *Neuropeptides* **32**: 167–171.
- Olin JW (2000). Thromboangiitis obliterans (Buerger's disease). *N Engl J Med* **343**: 864–869.
- Pisarchik A, Slominski A (2004). Molecular and functional characterization of novel CRFR1 isoforms from the skin. *Eur J Biochem* **271**: 2821–2830.
- Pu  chal X, Fiessinger JN (2007). Thromboangiitis obliterans or Buerger's disease: challenges for the rheumatologist. *Rheumatology* **46**: 192–199.
- Quevedo ME, Slominski A, Pinto W, Wei E, Wortsman J (2001). Pleiotropic effects of corticotropin releasing hormone on normal human skin keratinocytes. *In Vitro Cell Dev Biol Anim* **37**: 50–54.
- Saruta M, Takahashi K, Suzuki T, Torii A, Kawakami M, Sasano H (2004). Urocortin 1 in colonic mucosa in patients with ulcerative colitis. *J Clin Endocrinol Metab* **89**: 5352–5361.
- Shirakura S, Higo K, Takeda M, Karasawa A (1994). Antithrombotic effects of KW-3635, a thromboxane A<sub>2</sub>-receptor antagonist, in guinea pigs. *Jpn J Pharmacol* **65**: 93–98.
- Silver MJ, Smith JB, Ingerman C, Kocsis JJ (1973). Arachidonic acid induced human platelet aggregation and prostaglandin formation. *Prostaglandins* **4**: 863–875.
- Silver MJ, Hoch W, Kocsis JJ, Ingerman CM, Smith JB (1974). Arachidonic acid causes sudden death in rabbits. *Science* **183**: 1085–1087.
- Singh LK, Boucher W, Pang X, Letourneau R, Seretakis D, Green M *et al.* (1999). Potent mast cell degranulation and vascular permeability triggered by urocortin through activation of corticotropin-releasing hormone receptors. *J Pharmacol Exp Ther* **288**: 1349–1356.
- Taal MW, Zandi-Nejad K, Weening B, Shahsafaei A, Kato S, Lee KW *et al.* (2000). Proinflammatory gene expression and macrophage recruitment in the rat remnant kidney. *Kidney Int* **58**: 1664–1676.
- Teli T, Markovic D, Hewitt ME, Levine MA, Hillhouse EW, Grammatopoulos DK (2008). Structural domains determining signalling characteristics of the CRH-receptor type 1 variant R1beta and response to PKC phosphorylation. *Cell Signal* **20**: 40–49.
- Theoharides TC, Singh LK, Boucher W, Pang X, Letourneau R, Webster E *et al.* (1998). Corticotropin-releasing hormone induces skin mast cell degranulation and increased vascular permeability, a possible explanation for its proinflammatory effects. *Endocrinology* **139**: 403–413.
- Theoharides TC, Donelan JM, Papadopoulou N, Cao J, Kempuraj D, Conti P (2004). Mast cells as targets of corticotropin-releasing factor and related peptides. *Trends Pharmacol Sci* **25**: 563–568.
- Uzuki M, Sasano H, Muramatsu Y, Totsune K, Takahashi K, Oki Y *et al.* (2001). Urocortin in the synovial tissue of patients with rheumatoid arthritis. *Clin Sci* **100**: 577–589.
- Vane JR, Bakhle YS, Botting RM (1998). Cyclooxygenases 1 and 2. *Annu Rev Pharmacol Toxicol* **38**: 97–120.
- Vaughan J, Donaldson C, Bittencourt J, Perrin MH, Lewis K, Sutton S *et al.* (1995). Urocortin, a mammalian neuropeptide related to fish urotensin I and to corticotropin-releasing factor. *Nature* **378**: 287–292.
- Wang J, Xu Y, Xu Y, Zhu H, Zhang R, Zhang G *et al.* (2008). Urocortin's inhibition of tumor growth and angiogenesis in hepatocellular carcinoma via corticotrophin-releasing factor receptor 2. *Cancer Invest* **26**: 359–368.
- Webster EL, Lewis DB, Torpy DJ, Zachman EK, Rice KC, Chrousos GP (1996). In vivo and in vitro characterization of antalarmin a non-peptide corticotropin-releasing hormone (CRH) receptor antagonist: suppression of pituitary ACTH release and peripheral inflammation. *Endocrinology* **137**: 5747–5750.
- Witkowska AM (2005). Soluble ICAM-1: a marker of vascular inflammation and lifestyle. *Cytokine* **31**: 127–134.
- Wu Y, Xu Y, Zhou H, Tao J, Li S (2006). Expression of urocortin in rat lung and its effect on pulmonary vascular permeability. *J Endocrinol* **189**: 167–178.
- Wu Y, Hu J, Zhang R, Zhou C, Xu Y, Guan X *et al.* (2008). Enhanced intracellular calcium induced by urocortin is involved in degranulation of rat lung mast cells. *Cell Physiol Biochem* **21**: 173–182.
- Wu Y, Zhang R, Zhou C, Xu Y, Guan X, Hu J *et al.* (2009). Enhanced expression of vascular cell adhesion molecule-1 by corticotrophin-releasing hormone contributes to progression of atherosclerosis in LDL receptor-deficient mice. *Atherosclerosis* **203**: 360–370.
- Yokotani K, Murakami Y, Okada S, Hirata M (2001). Role of brain arachidonic acid cascade on central CRF1 receptor-mediated activation of sympatho-adrenomedullary outflow in rats. *Eur J Pharmacol* **419**: 183–189.