

Review

Binding and Docking Interactions of NO, CO and O₂ in Heme Proteins as Probed by Density Functional Theory

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Abstract: Dynamics and reactivity in heme proteins include direct and indirect interactions of the ligands/substrates like CO, NO and O₂ with the environment. Direct electrostatic interactions result from amino acid side chains in the inner cavities and/or metal coordination in the active site, whereas indirect interactions result by ligands in the same coordination sphere. Interactions play a crucial role in stabilizing transition states in catalysis or altering ligation chemistry. We have probed, by Density Functional Theory (DFT), the perturbation degree in the stretching vibrational frequencies of CO, NO and O₂ molecules in the presence of electrostatic interactions or hydrogen bonds, under conditions simulating the inner cavities. Moreover, we have studied the vibrational characteristics of the heme bound form of the CO and NO ligands by altering the chemistry of the proximal to the heme ligand. CO, NO and O₂ molecules are highly polarizable exerting vibrational shifts up to 80, 200 and 120 cm⁻¹, respectively, compared to the non-interacting ligand. The importance of Density Functional Theory (DFT) methodology in the investigation of the heme-ligand-protein interactions is also addressed.

Keywords: density functional theory; ligand molecules; protein cavities; proximal effect

1. Introduction

We can divide NO, CO and O₂ ligand molecules interactions inside a protein into two different groups. The first one, called the inner cavity interactions, results from interactions of the ligand molecules with amino acid residues inside the protein cavities and represents those ligand molecules in the unbound-to-the-metal states. The second one, called the bound interactions, deals with the bound forms of the ligand molecules and either the related changes in the coordination sphere of the associated metal or other interactions that influence their binding energies and chemistry. As for the latter we will focus on the proximal interactions in the heme bound forms of the ligand molecules and in the deprotonation/protonation events in the coordination sphere of the Cu_B metal of Cytochrome *c* Oxidase (CcO). In this paper we report previous theoretical studies from our lab, as well as new calculations on model systems dealing with both kinds of interactions, which taken together, we use the term “protein effect”. Several theoretical studies [1-11] primarily based on Density Functional Theory (DFT), have provided insight into the geometries, electronic structures, and binding energies for the NO, CO and O-O complexes.

1.1. Inner Cavity Interactions

Inner protein cavities play a crucial role in controlling the dynamics, as well as their reactivity in reactions with small ligands like O₂, CO and NO by accommodation and/or docking [12-14]. These cavities act like local storage sites for small molecules near the active site, resulting in an increased effective concentration of the ligand. Neutral, positively or negatively charged residual side chains form such structural characteristics in the proteins. Thus, ligands interact with charged side chains or hydrogen bonding networks. In addition, they exert active role in the ligand binding process. Electrostatic interactions exert significant contribution in stabilizing the three dimensional architecture of either individual proteins or even between different biological systems. In addition, such interactions in or near the active site determine the reactivity and functionality of enzymatic systems. Thus, it is crucial for such events to be probed by theoretical calculations and compared to experimental data.

Myoglobin (Mb) is found in large concentrations in muscles storing and delivering molecular oxygen to mitochondria of red muscle cells. Theoretical characterization of CO/NO and O₂ ligand binding to ferrous (Fe²⁺) Mb becomes a significant step towards a better understanding of more complex biological systems, like CcO or nitric oxide reductases (NOR). CO dissociates from the heme-iron of the active site, during photolysis of complexes like Mb-CO [15-17] or CcO *ba*₃-CO [18] and transiently moves to a cavity nearby. Dynamics of such migration has been studied experimentally by ps and fs time-resolved (TR) [15,16], as well as infrared (FTIR) spectroscopy in conjunction with thermal desorption spectroscopy (TDS) [17]. Theoretical Simulations (like molecular dynamics) have probed the cavities near the active site to study ligand migration after photodissociation, and possible ligand pathways to and from the active site cavity [12,19-25]. Three different CO bound states to heme-iron (A₀ 1,967 cm⁻¹, A₁ 1,945 cm⁻¹, A₂ 1,932 cm⁻¹) have been identified, as well as a number of unbound/free ligand states (B₁ 2,130 cm⁻¹/B₂ 2,120 cm⁻¹, C and D) [15,17,26-29]. These differences in $\nu(\text{CO})$ vibrational frequencies correspond to diverse interactions with the surrounding residues. $\nu(\text{CO})$

in CcO ba_3 exerts vibrational frequencies at 2,131 and 2,146 cm^{-1} attributed to B_1 and B_0 respectively, [18] after *photodissociation* of the ba_3 -CO complex.

Heme-protein HemeAT [30] functions as a signal transducer and molecular O_2 sensor. Three states of CO have been spectroscopically identified. Two of them are bound states, at 1,967 (no interactions of the CO with the environment) and 1,928 cm^{-1} (hydrogen bonding interactions with the bound CO). A third $\nu(\text{CO})$ vibration at 2,065 cm^{-1} does not shift under photolytic conditions and has been attributed to free CO in a cavity [31].

By changing the electrostatic environment near the free NO, CO and O_2 on small models and in b3lyp/6-31g(d, p) level of theory we calculate the stretching vibrational frequencies $\nu(\text{NO})$, $\nu(\text{CO})$, $\nu(^3\text{O}_2)$ to construct diagrams of frequencies vs. interactions. It is interesting to note that especially for the free, unbound CO molecule, $\nu(\text{CO})$ can become as low as that of the bound CO, depending on the electrostatic interactions with amino acid side chains [31].

Aside from the importance of CO as a molecular probe, several biological processes involve NO, as a cellular molecular messenger that controls a number of physiological events. It is known to inhibit O_2 reduction by CcO or to be involved in additional oxidation and nitrification degrading proteins [32,33] and DNA [34].

1.2. The Bound Interactions

NO, CO and O_2 forms bound to heme groups are subject to distal or proximal interactions. Blomberg *et al.* [1] have shown that Cu_B in CcO has a similar effect on the binding energies of NO, CO and O_2 as the distal histidine in Mb. The heme iron is coordinated to a proximal histidine (His93) making the iron five coordinated with a free binding site for dioxygen. Above the free binding site resides the so called distal histidine (His64), which can form a hydrogen bond to the sixth iron ligand. One of the key features of the distal histidine is the discrimination between NO, CO and O_2 , favoring O_2 by mainly electrostatic interactions [28].

Hydrogen bonding networks that affect the basicity of an axial to the heme iron ligand to support oxidation states greater than Fe(III) have been reported [35,36]. CcO, sulfite reductase and CooA exhibit such proposed hydrogen bonding networks [35-38]. In the case of CcO, hydrogen bonding to the $\text{N}_\delta\text{-H}$ hydrogen of proximal histidine is the crucial step in understanding how reactivity of the heme-iron is controlled by the proximal environment in heme proteins [39].

A number of studies have addressed various aspects of hydrogen bonding effects to the proximal side. Franzen [40] has studied the effect of hydrogen bonding to the proximal histidine by donors such as H_2O , CH_3COO^- and $\text{CH}_3\text{CONHCH}_3$ on the $\nu(\text{Fe-C})$ and $\nu(\text{C-O})$ frequencies of CO bound models of peroxidases active site containing the catalytic Asp-His-Fe triad. Based on the DFT calculations, it was proposed that increased hydrogen bonding to the $\text{N}_\delta\text{-H}$ proton causes a rise in negative charge density on the imidazole ring and alters significantly the donor character of the imidazole π -system leading, in consequence, to an increased σ -bonding by the N_ϵ lone pair. The charge density on the iron metal is moved into the π^* orbitals of bound CO. Moreover, in another case, Hu *et al.* [41] studied several five-coordinate $[\text{Fe}(\text{II})(\text{Por})(2\text{-MeHIm})]$ derivatives by Mössbauer spectroscopy, showing that imidazole deprotonation leads to distinctly different axial and equatorial bond distances, also observed were significant deviations in the displacement of iron atom from the heme plane compared to neutral

imidazole species. It was thus proposed that such a switch altering heme chemistry is important for the biological function of heme proteins in cases of reversible binding of O₂.

Turning to CcO, the $\nu(\text{CO})$ frequency of the transient Cu_B-CO complex determines in great amount the conformational changes (*e.g.*, ligand dissociation) of the copper metal site in CcO, whereas both $\nu(\text{CO})$ and $\nu(\text{Cu-C})$ are reliable criteria for protonation/deprotonation phenomena. Therefore, we can derive structural information for the Cu_B His ligands from the $\nu(\text{CO})$ frequency of the Cu_B¹⁺-CO complex. The negative slope in the diagram of the DFT-optimized distances, $d(\text{C-O})$ versus $d(\text{Cu-C})$, implies the presence of a π -back-bonding in the Cu_B-CO complexes [42].

In the rest of the paper, we will focus on new theoretical models probing such as the above mentioned interactions and discussion in general to include previous theoretical works and conclusions. The accuracy of the B3LYP functional in DFT methodology has been tested in the extended G3 benchmark set [43], which consists of enthalpies of formation, ionization potentials, electron affinities and proton affinities for molecules containing first- and second-row atoms. The B3LYP functional gives an average error of 4.3 kcal/mol [43] for 376 different molecules.

2. Results and Discussion

Results will be focused, initially, on the unbound forms of the CO, NO and O₂ ligands (Section 2.1) and then theoretical calculations will be reported for metal-ligand complexes, such as Cu_B-CO from CcO, heme-Fe-NO and heme-Fe-CO (Section 2.2).

2.1. The Unbound Interactions inside Cavities

Recently [31], we have applied density functional theory to investigate the effect of charged residues on $\nu(\text{CO})$ of a docked CO molecule in a protein cavity. Because of the nature of these cavities, theoretical calculations have to be performed on simplified models with a restricted number of atoms. In these models, the charged residues in the protein cavity are represented by NH₄⁺ (*e.g.*, the arginine side chain) and HCOO⁻ (*e.g.*, aspartic acid). Figure 1 shows the models in which the free CO interacts with (i) HCOO⁻ (model A, HCOO⁻...CO), (ii) a simple molecule that provides both carboxyl and amino groups, such as Gly (model B), and (iii) NH₄⁺ (model C: CO...NH₄⁺, model E: NH₄⁺...CO). In the case of two NH₄⁺ molecules (model not shown), two unconstrained NH₄⁺ molecules strongly repel each other, whereas a constrained model (with fixed NH₄⁺...CO...NH₄⁺ distances) exhibits a large positive binding energy. Models A1–A8 show the effect of the C–C distance (negative charge near CO) on $\nu(\text{CO})$. In model A, the C–C distance is optimized at 2.98 Å without using constraints, whereas models A1–A8 have constrained C–C distances at 3.08, 3.18, 3.58, 3.78, 3.98, 4.18, 4.38, and 4.58 Å, respectively. All models except for A1–A8 are optimized without constraints. In model B, in which both the amino and carboxyl groups interact with the CO, the carboxyl group is deprotonated, whereas the amino group is neutral. Model C contains hydrogen-bonding interaction in addition to the positive charge effect on CO.

Figure 1. Empirical diagram depicting the polarizability extend, expressed in wavenumbers, of CO molecule inside protein cavities (figure taken from *Proc. Natl. Acad. Sci. USA* 2006, 103, 14796-14801 [31]).

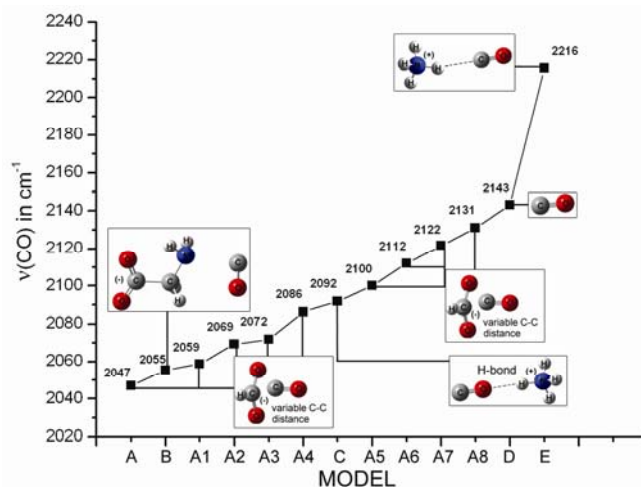


Figure 1 shows the calculated $\nu(\text{CO})$ trend versus the model. Positive charges around CO increase $\nu(\text{CO})$, whereas negative charges decrease it. Straub and Karplus [44] have calculated the $\nu(\text{CO})$ shifts in different CO–Imidazole dimer complexes at the 4–21G basis set level to vary between -17 cm^{-1} (H bonding C–O...H–N) to $+36 \text{ cm}^{-1}$ (N–H...C–O). Moreover, Nutt and Meuwly [45] have implemented molecular dynamics simulations on the photodissociated state of carboxymyoglobin based on a three-site charge model for CO to calculate the IR spectra of the free CO molecule in the heme pocket [45]. The IR spectrum obtained exhibits peaks between 2170 and 2200 cm^{-1} , corresponding to the signal from a single CO molecule docked in Mb. These signals are sensitive to the precise position and orientation of the CO molecule, as well as to the effect of the environment of the protein matrix (Mb) on the CO molecule. In general, the stretching frequencies $\nu(\text{CO})$ of heme-bound and photodissociated CO serve as powerful tools to probe the electrostatic fields and accessible space in the vicinity of the CO molecule inside the protein matrix. The docked, photodissociated CO ligands in the protein matrix display IR peaks, providing strong evidence for the existence of structurally well defined docking site(s). In analogy to the bound CO forms, the $\nu(\text{CO})$ of the docked CO is affected by the local environment through Stark effects of the local electric field acting on the CO dipole [12]. As our model tends to be more general and represent a wide variety of protein cavities, we have constructed an empirical diagram of $\nu(\text{CO})$ to investigate the effect of positive or negative charges and that of H-bonding interactions to different orientations of CO. The empirical diagram shows a significant variability in $\nu(\text{CO})$ ranging from 2047 cm^{-1} (interaction with only a negatively charged COO⁻ group) to 2216 cm^{-1} (interaction with only a positive charge). The calculations also show that $\nu(\text{CO})$ appears in the range of 2047 – 2131 cm^{-1} at 10 cm^{-1} steps when the CO distance to a negatively charged carboxyl group is varied. The combined interaction of carboxyl and amino groups to CO has a strong effect on $\nu(\text{CO})$, as shown by the 88 cm^{-1} down-shift from 2143 cm^{-1} of gas CO, even though CO is not interacting directly with the deprotonated carboxyl.

All above calculated vibrational frequencies are consistent with the Badger's rule [46]: $r_e = c_{ij}(1/\nu_e^{2/3}) + d_{ij}$, where r_e is the equilibrium C–O distance and ν_e refers to $\nu(\text{CO})$ (Figure 2). The

empirical parameters c_{ij} and d_{ij} were calculated to be 61.535 and 0.767, respectively. We observe a huge diversity in $\nu(\text{CO})$, reaching values in the region of CO bound metal complexes, like $\text{Cu}_B\text{-CO}$ in CcO [18]. In Figure 3 we draw three diagrams concerning trends for $\nu(\text{CO})/\text{CO}$: (A) binding energies for all models, (B) C–O distances in *angstroms* and (C) $\nu(\text{C–O})$ frequencies in relation to $\Delta q = q_C - q_O$ charge separation (charge difference between calculated carbon q_C and oxygen q_O charges) in the different A, A1-8, B, C, D and E cases.

Figure 2. Parameter fitting for Badger's rule on CO molecule based on the DFT models (figure taken from *Proc. Natl. Acad. Sci. USA* 2006, 103, 14796-14801, S.I., [31]).

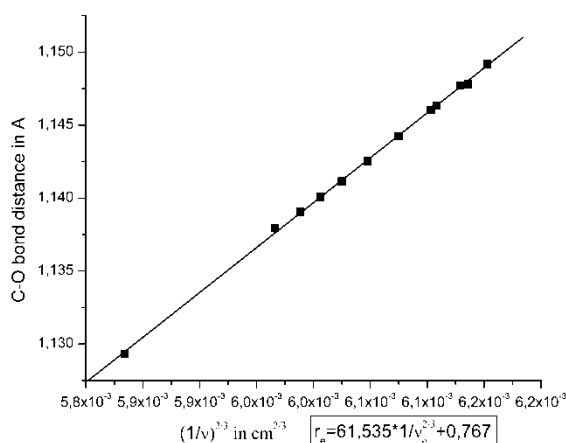
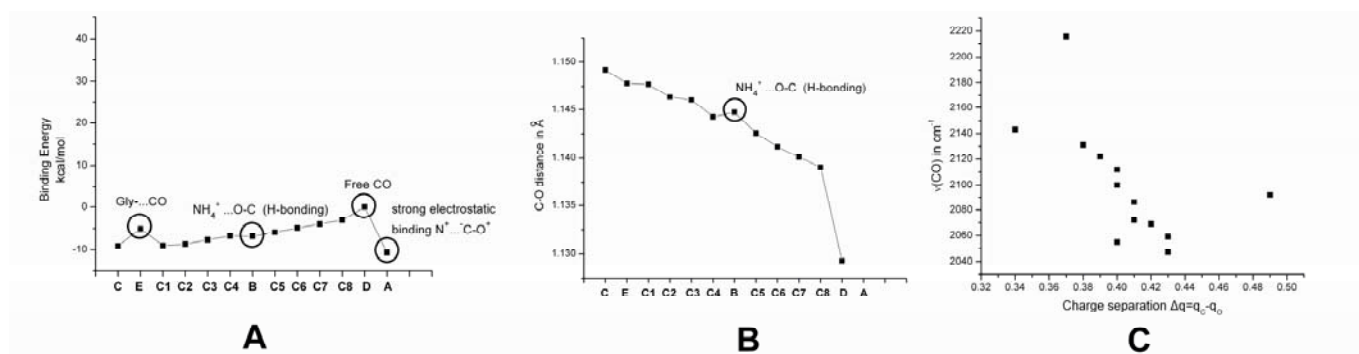
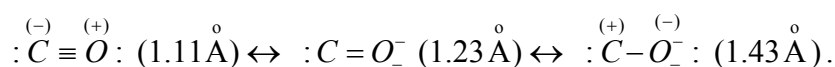


Figure 3. (A) Binding energies of CO models. (B) C–O distances in Å of CO models. (C) Charge separation $\Delta q = q(\text{C}) - q(\text{O})$ observed in the CO models.



The CO molecule can exist in three different resonance forms, depending on the environment or metal ligation. The different resonance structures of CO are linked through the below equilibrium (experimental interatomic distances are also shown for clarity):

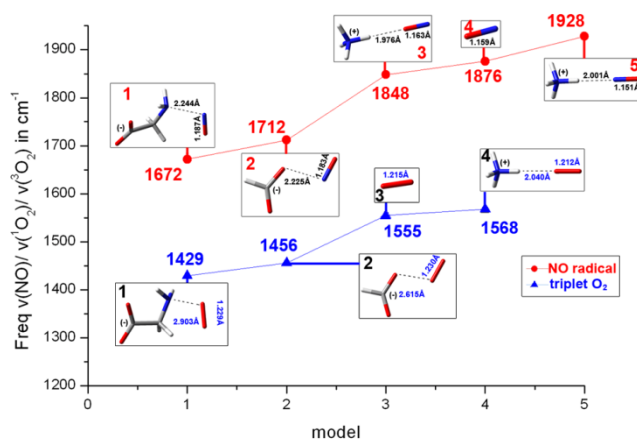


When a positively charged atom is near carbon atom of CO, the first form is the dominant one, as in that case, electrostatics (attraction) are the main cause of stabilizing such interaction. Further polarization of CO molecule towards this direction would lead to a further strengthening of the C–O bond (model E). In the opposite trend, if a negatively charged molecule approaches carbon atom of

CO, the last structure would prove to be the dominant one, as the C-O bond length is increased, while its strength is in turn decreased (models **A**, **A1-8**). A positive charge on the site of oxygen of CO would also lead to the third resonance structure, while an hydrogen bond would strengthen the C-O bond. In fact, these opposite effects collaborate in model **C** which includes positive charge and the hydrogen bonding interaction ($\text{CO}\dots\text{NH}_4^+$), as $\nu(\text{CO})$ appears at $2,092\text{ cm}^{-1}$ taking into account both contributions. In the case of Gly-CO interaction, as in model **B**, the C-O bond weakens and the third resonance structure dominates (C-O distance at 1.43 \AA). This could be due to a more complicated interaction between Gly and CO including a combination of electrostatic interactions from NH_2 , as well as COO^- . Binding energies in Figure 3 do not exert a smooth trend versus the $d(\text{C-O})$ distances. This is due to a) hydrogen bonding involved in CO interactions (model **C**) and b) in the intense electrostatic interaction in model **E**. Such trends are also depicted in the $\nu(\text{CO})$ versus $\Delta q = q_{\text{C}} - q_{\text{O}}$ charge separation diagram in the same figure. The negative slope indicates that as the charge separation between C and O increases (due to polarization), the C-O bond energy is decreased. Thus, calculations are fully consistent to the above mentioned equilibrium of CO resonance states.

In the models with NH_4^+ interacting with either O_2 or NO we introduce, in addition to the positive charge effect, a hydrogen bond, not always present in the CO models with NH_4^+ . Electrostatic fields, present in the protein cavities (or DFT models presented in this study) are responsible for a Stark Effect. An electric field induces a shift in $\nu(\text{CO})$ in addition to a change in dipole moment, μ , of CO due to excitation by absorption of a vibrational mode. DFT studies by Brewer and Franzen [47] indicate that for carbonyls and NO moieties, transitions in molecular geometry, due to an electric field, are the main contributors to a Stark Effect.

Figure 4. Empirical diagram depicting the extent of polarizability, expressed in wavenumbers, of NO and triplet O_2 molecules inside protein cavities.



Moreover, we simulate intercavity interactions inside a protein in cases where the ligand molecule is different from CO, by replacing it by either NO, or $^3\text{O}_2$ in the above presented models. Again, free ligands interact with charged or neutral moieties acting as amino acid side chains. Results are shown on Figure 4. All NO, CO and O_2 ligand-gases seem to behave in a similar way exhibiting a pronounced diversity in vibrational stretching frequencies due to polarizability. The environment plays a crucial role in each and every case. Again, no unusually high or questionable binding energies are observed

for any NO/O₂ models, as well as in CO. The general trend followed is that positively charged atoms near NO, CO or O₂ shift the respective stretching vibrational frequencies to higher energies, while negative charges to lower energies. All three gases exhibit a pronounced effect by the environment on their vibrational stretching frequencies, without the need to be bound on a metal (*e.g.*, heme-iron or Cu_B site in CcO).

Summarizing, we can accept that the variation of $\nu(\text{CO})$ of free CO ligand inside a protein cavity can reach up to 170 cm⁻¹ (2,216-2,047 cm⁻¹ based on theoretical models presented herein), without excluding more pronounced shifts. Such interactions are of course more complicated and combine effects from hydrogen bonds, positively and negatively charged side chains, but in any case the present study enforces us to reconsider the experimental results on $\nu(\text{CO})$ or $\nu(\text{NO})$, where bound and unbound states may exert vibrational frequencies in the same region of the spectrum.

2.2. The Bound Interactions

Table 1. Theoretical vibrational frequencies in cm⁻¹ of the various DFT models in groups A, B and C (table taken from *J. Phys. Chem. B* 2007, 111, 10502-10509, [42]).

Cu _B -CO models	Theoretical vibrations in cm ⁻¹ (dmol3-BLYP/DND)		
	$\nu(\text{C-O})$	$\nu(\text{Cu-C})$	$\delta(\text{Cu-C-O})$
A1	2031	414	326
A2	2020	425	
A3	2020	420	
A4	2023	421	
A5	2072	421	
B1	2049	419	
B2	2011	442	
B3	2040	421	
B4	2083	422	
B5	2147	444	
C1	2035	411	324
C2	1992	431	
C3	2078	411	
C4	2026	468	

Cu_B-CO complexes of Cytochrome *c* Oxidase [42]. Protonation/deprotonation events of groups near the catalytic center of CcO have been suggested to be involved or not in the delivery of protons for the dioxygen reduction chemistry and proton pumping [48-51]. Thus, it is important to probe such functional properties in the first coordination sphere of Cu_B. Protonation/deprotonation events at the binuclear center coupled to the dynamics and chemistry occurring at the heme-Fe is crucial in elucidating the functional properties of the enzyme itself. On this line, we have designed four groups

of models, referring to them as **A**, **B**, **C** (Figure 5) and **D** (Figure 6). In these models, Cu_B (I) is coordinated to three (**A1-A4**), and two (**A5**) imidazoles and the cross-linked His-Tyr is designed as a cross-linked imidazole-phenol (protonated) unit. In **A2-A4** the phenol is deprotonated. In **A3** and **A4** the phenolic O is H-bonded to H₃O⁺ and H₂O, respectively. In group **B**, the imidazoles coordinated to Cu_B in group **A** are substituted by NH₃ ligands, while the cross-link Im-phenol unit remains protonated in **B1** and deprotonated in **B2-B3**; in **B3** the phenolic O is H-bonded to H₃O⁺. **B4** is similar to **B1** but with one instead of two coordinated NH₃ ligands. **B5** lacks both NH₃ ligands of **B1**. In **C1-C2**, Cu_B is coordinated to three Imidazoles without the cross linked phenol unit. In **C1**, all imidazoles are protonated and in **C2** one of the three imidazoles is deprotonated. In **C3**, Cu is coordinated to two protonated imidazoles. In **C4**, Cu is coordinated to a deprotonated cross-linked Im-phenol unit. In group **D**, we have optimized structures of the binuclear Cu_B(I)-Fe(II) center in which a CH₃CH(OH)-group represents the hydroxyethylgeranylgeranyl or hydroxyethylfarnesyl side chain of the hemes (**D1-D2**) or is absent (**D3**). The distance between Cu(I) and Fe(II) is constrained to ~4,4 Å. In Figure 5 the HOMO of **A1** and **A2** are also presented.

Table 1 summarizes the calculated $\nu(\text{Cu-C})$, $\nu(\text{C-O})$ and selected $\delta(\text{Cu-C-O})$ frequencies for the **A-C** models. A close inspection of the data presented in Table 1 shows large frequency shifts in $\nu(\text{CO})$ when Cu_B loses one of its imidazole (or NH₃ in simpler models) ligands. When a Cu_B-Im bond scission occurs, $\nu(\text{CO})$ shifts to higher frequency by 43 cm⁻¹ (**A5** compared to **A1**), as the C-O bond strengthens due to geometrical and electronic structural changes in Cu. A relatively smaller effect (7 cm⁻¹) is calculated for $\nu(\text{Cu-C})$ in the same set of models. A change in the protonation state of one of the Cu_B imidazole ligands (comparison of **C1** to **C2**) has significant change in the back donation of electron density and therefore the vibrational frequencies with a calculated shift of 20-43 cm⁻¹, while deprotonation of the phenol unit (comparison **A1** to **A2**) strengthens the Cu-C and weakens the C-O bond leading to a frequency shift of 11 cm⁻¹. In the case where Cu lacks two of the three imidazoles (**C4**) or the two imidazoles are replaced by NH₃ groups, the deprotonation of the cross-linked phenol unit (**B5**→**C4** or **B1**→**B2**) has a pronounced effect of 120 cm⁻¹ and 38 cm⁻¹, respectively. The calculated frequencies indicate that electron density due to phenol deprotonation moves mainly to the Cu-C-O entity, altering bond lengths and vibrational frequencies. On the other hand, if Cu is coordinated to two imidazoles (structures **A1**, **A2**), electron density due to phenol deprotonation does not localize to the Cu-C-O moiety, but rather is delocalized on the whole Cu_B complex, including the histidine ligands, leading to a reduced $\Delta\nu(\text{CO})$ effect of 11 cm⁻¹.

Figure 6 shows geometry optimized binuclear models containing both Cu_B and heme Fe. Models **A**, **B**, **C** exhibit an almost linear Cu-C-O geometry compared to **D1** and **D2**, where the Cu-C-O tilt and bending is different when the heme Fe is present, but remains unaltered in deprotonation events.

Models **D1** and **D2** were altered in such way that CO is bound to heme-iron rather than to copper (structures not shown). Geometry optimization was performed on the same blyp/dnd (dmol3) level of theory. CO geometry (bending or tilting) is affected as the distance between the metal sites changes. The iron-copper distance was kept constant at 6.0, 5.0 and 4.5 Å for the above models and the rest of the active site was left free to be optimized. The tilt/bend of Fe-C-O moiety was calculated to be 87.9°/178.0° (6.0 Å), 84.7°/173.9° (5.0 Å) and 80.5°/169.1° (4.5 Å). The nature of this distortion either in favor of electrostatic interactions or steric hindrance can lead to discrimination between CO and O₂ by systems as CcO, Myoglobin (Mb) and Hemoglobin (Hb).

While in dioxygen a favorable overlap between its π^* -orbital and Fe- d_{z^2} orbital minimizes the binding energy by bending [52], this is not the case for CO, where these orbitals are not close in energy. Thus, the observed M-C-O distortion in CcO is induced. Metal bound CO has a character of $M-C^{(-)}-O^{(+)}$. Electrostatic repulsion occurs between the partially positively charged oxygen of CO and the positively charged metal not coordinating CO. Increasing the Fe- Cu_B distance ($4.5 \text{ \AA} \rightarrow 5.0 \text{ \AA}$ - step: $0.5/4.5 \text{ \AA} \rightarrow 6.0 \text{ \AA}$ - step 3×0.5) the relative changes in CO distortion (see above) would be proportional to a 1:3 scheme in the case of a steric hindrance-only assumption, as M-C-O would relax to the linear geometry proportionally to the step. In fact, these relative changes in tilt and bend appear to be consistent with a $\sim 1:1.8$ scheme. This could be justified by a more complex mechanism including electrostatic interactions in the active site inducing the observed distortions.

Figure 5. DFT models of diverse Cu-CO complexes in groups A, B and C (figure taken from *J. Phys. Chem. B* 2007, 111, 10502-10509, [42]).

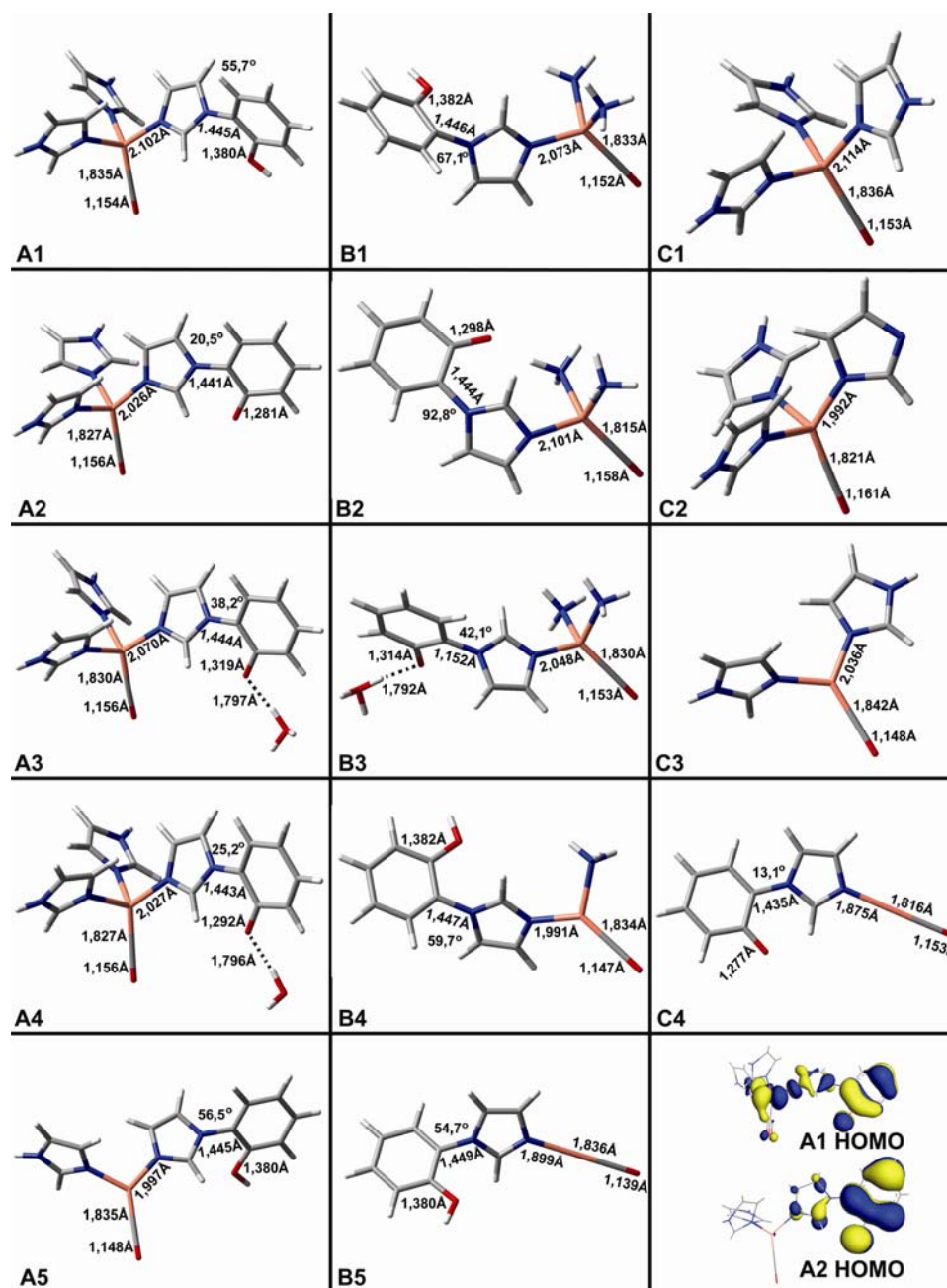
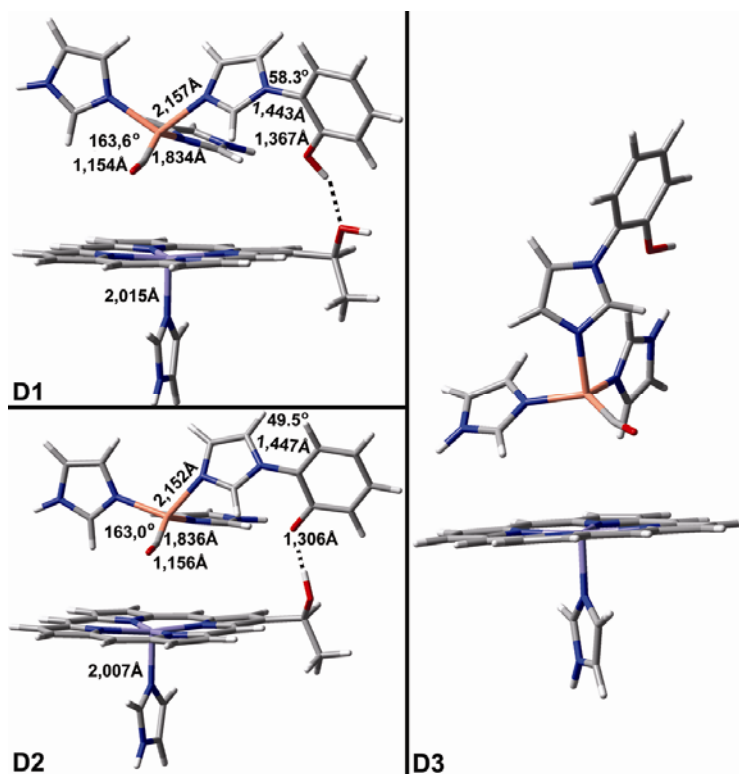


Figure 6. DFT models of diverse heme-iron/ Cu_B -CO complexes of group D (figure taken from *J. Phys. Chem. B* 2007, 111, 10502-10509, [42]).



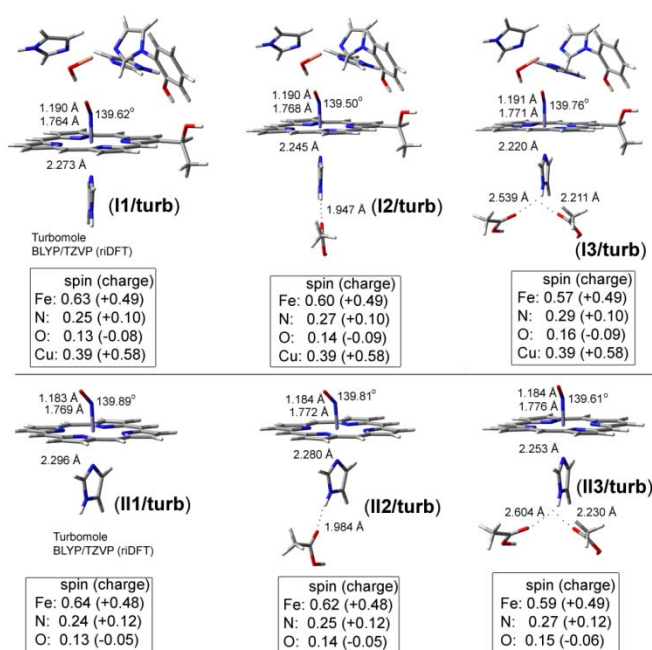
As of the importance of electrostatic interactions in M-C-O systems, DFT calculations by Blomberg L. M. *et al.* [1] have shown that Cu_B in CcO has a similar effect on the binding energy of CO on Fe as the distal histidine in Mb. In addition, CO binding studies on mutants of Mb by Kozłowski and Spiro [53] have revealed that the steric hindrance by the distal histidine is worth only ca. 1 kcal/mol, while electrostatic interaction by H-bonding is the main reason (85%) [54] for CO/O₂ binding discrimination.

In the Cu_B -CO models the changes in the bond lengths of the CO-bound and, thus, the calculated frequency shifts, can be attributed to the increase of electron density on the metal center: as the electron density on the metal centre increases (deprotonation increases the flow of electron density to Cu_B as discussed above) more electron density donation to the CO ligand takes place. This increases the M-CO bond strength making it more double-bond-like (M=C=O) which in turn, further weakens the C-O bond by increasing the electron density into the carbonyl antibonding orbitals. Trans-ligands to a carbonyl can have a particularly large effect on the ability of the CO ligand to effectively π -backbond to the metal. For example, two trans π -backbonding ligands will partially compete for the same d-orbital electron density, weakening each others net π -backbonding. Trans-ligand which are a σ -donors can increase the M-CO bond strength (more M=C=O character) by allowing unimpeded metal to CO π -backbonding.

The existence and identity of a reorganization of the Cu_B geometry caused by protonation/deprotonation and/or breakage of one of the Cu-N(His) bonds has been a difficult matter to either prove or disprove since Cu_B is spectrally silent and therefore no definite spectroscopic evidence had been observed. The $\nu(\text{CO})$ frequency of the transient Cu_B -CO complex determines in great amount

the conformational changes (e.g., ligand dissociation) of copper metal site in CcO, while both $\nu(\text{CO})$ and $\nu(\text{Cu-C})$ are reliable criteria for protonation/deprotonation phenomena. Therefore, we can derive structural information for the $\text{Cu}_B\text{-N}$ (His) ligands from the $\nu(\text{CO})$ frequency of the $\text{Cu}_B^{1+}\text{-CO}$ complex. The negative slope in the $d(\text{C-O})$ vs. $d(\text{Cu-C})$ diagram implies the presence of a π -backbonding in the Cu-CO complexes. The calculated $\nu(\text{CO})$ frequencies of Cu_B under different protonation/deprotonation states and/or ligand-detachment indicate that the $\nu(\text{CO})$ frequency depends strongly on the degree of backbonding. A change in the protonation state of one of the His ligands would have significant changes in the back donation, and thus on the frequency of $\nu(\text{CO})$. If one of the His-ligands of Cu_B is capable of cycling through the imidazolate, imidazole and imidazolium states then $\nu(\text{CO})$ is expected to vary. We have shown [42] that no structural change at Cu_B occurs in association with CO binding to and dissociation from heme a_3 in conjunction with the consensus that the pH/pD dependency of the $\text{Cu}_B\text{-C-O}$ vibrational frequencies in heme-copper oxidase is not due to deprotonation of the cross-linked tyrosine demonstrates that the environment of Cu_B does not serve as a proton-labile site.

Figure 7. DFT optimized geometries in heme-iron-NO binuclear complexes. Hydrogen bonding network to the proximal area is variable.



Heme-NO and -CO complexes. Heme-Fe(II)-NO models are treated as doublet neutral species (without Cu_B metal site) or positively charged (+1) with doublet/triplet multiplicity in case of the presence of the Cu_B in oxidation states (I) or (II) respectively. In Figure 7 we present DFT models including both $\text{Cu}_B(\text{II})\text{-OH}$ and $\text{Fe}(\text{II})\text{-NO}$ without (II1), and with one (II2) or two (II3) hydrogen bonds to the proximal coordinating Imidazole (a carboxyl acid provides the O-donor atom). Models III1, III2 and III3 (Figure 8) contain no Cu_B site, but exclusively the heme $\text{Fe}_{a_3}(\text{II})$ site with different hydrogen bonding networks in the proximal area, without (III1), and with one (III2) or two (III3) carboxyl groups in these networks. Model III4 (Figure 8) is derived from III1 by deprotonating the proximal coordinating Imidazole. Figure 7 contains models treated by the ri-blyp/tzvp level of theory by

Turbomole software package (highlighted as **turb**), while Figure 8 by blyp/dnd level of theory in dmol3 module of Materials Studio Suite of programs (highlighted as **dmol3**). We use diverse software packages in order to evaluate the results in different levels of theory.

Figure 8. DFT optimized geometries and theoretical vibrational frequencies in heme-iron-NO complexes. Hydrogen bonding network to the proximal area is variable.

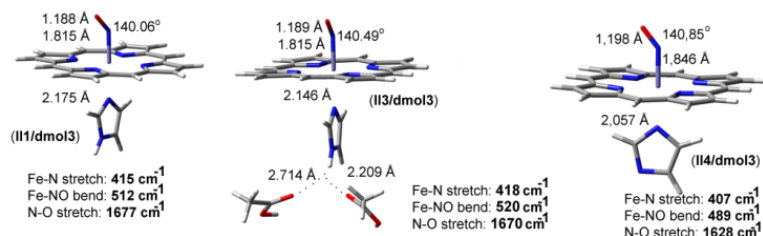
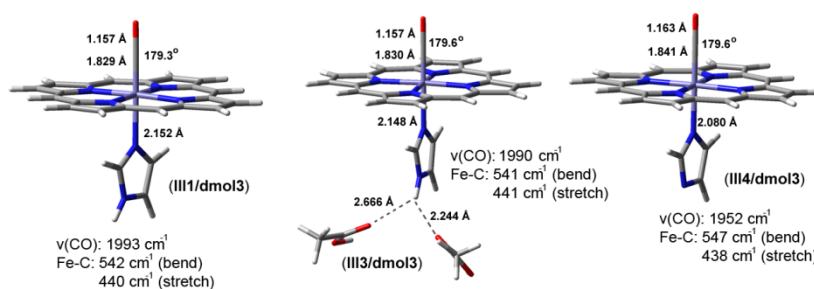


Table 2. Depicts the vibrational frequencies for **II1**, **II2** and **II3**, without the Cu_B site. 7-8 cm^{-1} shifts were calculated in $\nu(\text{N-O})$ and $\nu(\text{Fe-N})$. For the extreme case of proximal Imidazole deprotonation, a 42 cm^{-1} shift in $\nu(\text{N-O})$ is observed.

Fe-NO models	$\nu(\text{N-O})$ in cm^{-1}	$\nu(\text{Fe-N})$ in cm^{-1}	$\delta_{\text{Fe-N-O}}$ in cm^{-1}
II1/dmol3	1677	415	512
II3/dmol3	1670	418	520
II4/dmol3	1628	407	489

Figure 9. DFT optimized geometries and theoretical vibrational frequencies in heme-iron-CO complexes. Hydrogen bonding network to the proximal area is variable.



In contrast to Fe-NO complexes, heme-CO complexes (Figure 9), where proximal Imidazole is found without (**III1**) and with two (**III3**) hydrogen bonds in the proximal area or with a deprotonated Imidazole (**III4**) the shift in $\nu(\text{CO})$ is significant only for the latter case at 40 cm^{-1} , in connection to experimental and theoretical studies [40,55].

Theoretical DFT studies on heme-NO complexes of heme proteins conclude that the distal effect is crucial for recognition and discrimination of ligand molecules by the protein [1,56]. As discussed above, it seems that the proximal effect is not significantly higher to be involved in recognition and/or discrimination mechanisms by such proteins. We cannot, though, exclude that upon binding, conformational changes are communicated through the proximal site.

To summarize, we conclude that, for CO bound in heme groups, changes to the axial proximal ligand, through either hydrogen bonding networks or deprotonation, do not affect to the same extent, compared to Cu_B-CO complexes, the Fe-C/C-O bond energies.

3. Calculation Methods

Density Functional Theory on unbound CO, NO and O₂. For each structure considered, a full geometry optimization was performed by using the density functional B3LYP method. We used the 6–31 g (d,p) double- ζ valence basis set augmented with *p*-PGTOs on H atoms, as implemented in the Gaussian 03 software package [57]. When vibrational frequencies are calculated by electronic structure theory, they can often be improved by scaling, and it is useful to have general scaling factors. Such factors depend on the level of electronic structure theory and the one-electron basis set. It has been established that calculated frequencies may be scaled in various ways [58]. For example, one scaling factor is applied to reproduce the true harmonic frequencies, the true fundamental frequencies, or the zero point energy. To accurately reproduce the experimental $\nu(\text{CO})$ of interaction-free gas CO, a scaling factor of $\times 0.9703$ was used for the theoretically calculated $\nu(\text{CO})$. A scan for $\nu(\text{CO})$ using different basis sets starting from 6–31g (d,p) through 6–311g++(2df, 2pd) showed identical trends ($\pm 10 \text{ cm}^{-1}$) for $\nu(\text{CO})$ concerning the environmental effect. Solvation effects count for only a 5 cm^{-1} theoretical shift and, thus, were not taken into account. Scaling factor of $\times 0.938$ on theoretically calculated stretching vibrational $\nu(\text{O-O})$ frequencies, to reproduce the experimental value of $\nu(\text{O-O})$ in a non-interacting *triplet* O₂ ($1,555 \text{ cm}^{-1}$). While, a scaling factor of $\times 0.942$ was used for the $\nu(\text{N-O})$ vibrational stretching frequencies, based on the $\nu(\text{N-O})$ experimental value of NO gas ($1,876 \text{ cm}^{-1}$). Calculation of the solvent effect (H₂O) and a basis set scan was also performed for both $\nu(\text{O-O})$ and $\nu(\text{N-O})$, with negligible variations for $\nu(\text{X-O})$ once again (X = N, O).

Density Functional Theory on bound complexes. DFT calculations are performed on systems with Cu_B(I) and heme-Fe(II) leading to a *singlet*, positively charged (+1) mononuclear Cu_B(I)-CO complex or a binuclear Cu_B(I)-CO Fe(II) center. For each structure considered, a full geometry optimization was performed using the density functional BLYP method of **dmol3** [59] module in Accelrys Materials Studio 2.21. Double Numerical plus d-functions with a polarization d-function on all non-hydrogen atoms are used for all light elements. For the metals (iron and copper) effective core potential (ECP) from the Stuttgart-Dresden group [60] was used as this is implemented in dmol3 module of Materials Studio (*Accelrys*). The functional GGA/BLYP on a DND basis set used for the calculations is consistent with a BLYP/6-31G(d) level calculation in Gaussian software package. In all cases (convergence tolerance, integration accuracy, SCF tolerance) level fine was selected.

For the **heme-CO or -NO complexes** the ri-blyp functional along with the tzvp triple- ζ valence basis set level of theory has been applied, as implemented in the Turbomole software package or the blyp/dnd level of theory when dmol3 module of Materials Studio is mentioned.

A scaling factor has not been applied for the bound model species, as absolute experimental vibrational frequency values depend highly on the system under study and usually refer to either more complicated model-compounds or enzymes. In contrast, all unbound vibrational frequencies have been scaled as small theoretical models in reference (CO, NO and O₂) have been also studied experimentally as they were designed in theory.

No restrictions in geometry, as well as a Hessian calculation, after the full geometrical optimization of each structure have been applied for all above calculations. Mulliken population analysis was also performed and the results are shown on Figure 7 and 8. All vibrational frequencies stated in this study are harmonic and represent those of the highest intensity.

Unbound (CO, NO and O₂) and bound (heme-CO, Cu_B-CO/heme-NO) complexes are treated differently in the level of theory. As the theoretical model used increases in size (unbound to bound states), we have to decrease the level of theory to converge the calculations in a descent amount of cpu-time. This seemed to be a compromise in accuracy and on this line we compared spectroscopic results between b3lyp/lanl2dz based calculations in Gaussian 98 and blyp/dnd based in dmol3 for selected Cu_B-CO theoretical models in Figure 6. The calculated frequencies (b3lyp) for the Cu-C stretching mode are 380 cm⁻¹ for **A1** and 391 cm⁻¹ for **A5**. Thus, we observe an A1→A5 + 11 cm⁻¹ shift in frequencies in the higher level of theory (b3lyp/lanl2dz) and +7 cm⁻¹ for the blyp/dnd (dmol3) level. Both methods calculate a ν(CO) frequency of 2031 cm⁻¹ for **A1**, whereas in **A5**, there is a 1 cm⁻¹ difference for the ν(CO) frequency. Geometric parameters, such as bond lengths and angles, differ only of a maximum amount of 0.04 Å and 5-6°, respectively, between the two levels of theory. Despite the differences in the absolute values of the vibrational frequencies between the two methods, the calculated frequency shifts follow the same trend. As absolute values usually are of no significant use, trends and vibrational shifts contain important information for experimental spectroscopy.

4. Conclusions

Intracavity dynamics play a major role in the function of protein and enzymes [51,61-68]. Ligand molecules are directed into the active site or internal cavities of a protein and/or through channels. The specificity or potency of an enzymatic catalytic reaction relies strongly on the interactions between the protein matrix and the substrate. Density Functional Theory is a potent tool for probing such interactions, even in the cases where a crystal structure has not been solved. Based on the theoretical studies presented in this review, we can conclude that for small molecules like NO and CO, distal interactions become important for ligand discrimination or recognition by the proteins, while proximal effects exert a significantly lower contribution. Nevertheless, we have to observe that in the case of NO, proximal effects, although weak, become more pronounced than in the case of CO. In addition, inner-cavity interactions play a major role in polarizing gases like O₂, NO and CO. The latter exerts a significantly higher polarizability, as shown by the diversity of ν(CO) vibrational frequencies calculated in models of protein CO-cavity interactions. In the case of CO-bound models of Cu_B CcO it seems that deprotonation/protonation or ligand dissociation events in the first coordination sphere of the metal are sensed by the Cu_B-C-O unit, as they are accompanied by pronounced shifts in both ν(CO) and ν(Cu-C). DFT can be successfully used not only to structurally and spectroscopically probe active sites in proteins, but also cavities. In this way we can link experimental data to an atomistic overview of the systems under study and extract structural information from the experimental data in the most efficient way.

References and Notes

1. Blomberg, L.M.; Blomberg, M.R.A.; Siegbahn, P.E.M. A theoretical study on the binding of O₂, NO and CO to heme proteins. *J. Inorg. Biochem.* **2005**, *99*, 949-958.
2. Rovira, C.; Kunc, K.; Hutter, J.; Ballone, P.; Parrinello, M. Equilibrium geometries and electronic structure of iron-porphyrin complexes: A density functional study. *J. Phys. Chem. A* **1997**, *101*, 8914-8925.
3. Rovira, C.; Kunc, K.; Hutter, J.; Ballone, P.; Parrinello, M. A comparative study of O₂, CO, and NO binding to iron-porphyrin. *Int. J. Quant. Chem.* **1998**, *69*, 31-35.
4. Sigfridsson, E.; Ryde, U. On the significance of hydrogen bonds for the discrimination between CO and O₂ by myoglobin. *J. Biol. Inorg. Chem.* **1999**, *4*, 99-110.
5. Harvey, J.N. DFT computation of the intrinsic barrier to CO geminate recombination with heme compounds. *J. Am. Chem. Soc.* **2000**, *122*, 12401-12402.
6. Sigfridsson, E.; Ryde, U. Theoretical study of the discrimination between O₂ and CO by myoglobin. *J. Inorg. Biochem.* **2002**, *91*, 101-115.
7. Jensen, K.P.; Roos, B.O.; Ryde, U. O₂-binding to heme: Electronic structure and spectrum of oxyheme, studied by multiconfigurational methods. *J. Inorg. Biochem.* **2005**, *99*, 45-54.
8. Jensen, K.P.; Roos, B.O.; Ryde, U. Erratum to "O₂-binding to heme: Electronic structure and spectrum of oxyheme, studied by multiconfigurational methods". *J. Inorg. Biochem.* **2005**, *99*, 978.
9. Rutkowska-Zbik, D.M.W.; Stochel, G. Theoretical density functional theory studies on interactions of small biologically active molecules with isolated heme group. *J. Comput. Chem.* **2007**, *28*, 825-831.
10. Strickland, N.; Harvey, J.N. Spin-forbidden ligand binding to the ferrous-heme group: Ab initio and DFT studies. *J. Phys. Chem. B* **2007**, *111*, 841-852.
11. Ribas-Ariño, J.; Novoa, J.J. The mechanism for the reversible oxygen addition to heme. A theoretical CASPT2 study. *Chem. Commun.* **2007**, *30*, 3160-3162.
12. Nienhaus, K.; Olson, J.S.; Franzen, S.; Nienhaus, G.U. The origin of stark splitting in the initial photoproduct state of MbCO. *J. Am. Chem. Soc.* **2005**, *127*, 40-41.
13. Lim, M.; Jackson, T.; Anfinrud, P. Binding of CO to myoglobin from a heme pocket docking site to form nearly linear Fe-C-O. *Science* **1995**, *269*, 962-966.
14. Frauenfelder, H.; Sligar, S.; Wolynes, P. The energy landscapes and motions of proteins. *Science* **1991**, *254*, 1598-1603.
15. Ansari, A.; Jones, C.M.; Henry, E.R.; Hofrichter, J.; Eaton, W.A. Conformational relaxation and ligand binding in myoglobin. *Biochemistry* **1994**, *33*, 5128-5145.
16. Lim, M.; Jackson, T.A.; Anfinrud, P.A. Nonexponential protein relaxation: Dynamics of conformational change in myoglobin. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 5801-5804.
17. Nienhaus, K.; Deng, P.; Kriegl, J.M.; Nienhaus, G.U. Structural dynamics of myoglobin: Effect of internal cavities on ligand migration and binding. *Biochemistry* **2003**, *42*, 9647-9658.
18. Koutsoupakis, C.; Soulimane, T.; Varotsis, C. Docking site dynamics of ba₃-cytochrome c oxidase from thermus thermophilus. *J. Biol. Chem.* **2003**, *278*, 36806-36809.

19. Case, D.A.; Karplus, M. Dynamics of ligand binding to heme proteins. *J. Mol. Biol.* **1979**, *132*, 343-368.
20. Tilton, R.F.; Singh, U.C.; Weiner, S.J.; Connolly, M.L.; Kuntz, I.D.; Kollman, P.A.; Max, N.; Case, D.A. Computational studies of the interaction of myoglobin and xenon. *J. Mol. Biol.* **1986**, *192*, 443-456.
21. Elber, R.; Karplus, M. Enhanced sampling in molecular dynamics: Use of the time-dependent Hartree approximation for a simulation of carbon monoxide diffusion through myoglobin. *J. Am. Chem. Soc.* **1990**, *112*, 9161-9175.
22. Bossa, C.; Anselmi, M.; Roccatan, D.; Amadei, A.; Vallone, B.; Brunori, M.; Di Nola, A. Extended molecular dynamics simulation of the carbon monoxide migration in sperm whale myoglobin. *Biophys. J.* **2004**, *86*, 3855-3862.
23. Agmon, N. Coupling of protein relaxation to ligand binding and migration in myoglobin. *Biophys. J.* **2004**, *87*, 1537-1543.
24. Nutt, D.R.; Meuwly, M. CO migration in native and mutant myoglobin: Atomistic simulations for the understanding of protein function. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 5998-6002.
25. Banushkina, P.; Meuwly, M. Free-energy barriers in MbCO rebinding. *J. Phys. Chem. B* **2005**, *109*, 16911-16917.
26. Olson, J.S.; Phillips, G.N., Jr. Kinetic pathways and barriers for ligand binding to myoglobin. *J. Biol. Chem.* **1996**, *271*, 17593-17596.
27. Ostermann, A.; Waschipky, R.; Parak, F.G.; Nienhaus, U.G. Ligand binding and conformational motions in myoglobin. *Nature* **2000**, *404*, 205-208.
28. Carver, T.E.; Rohlfis, R.J.; Olson, J.S.; Gibson, Q.H.; Blackmore, R.S.; Springer, B.A.; Sligar, S.G. Analysis of the kinetic barriers for ligand binding to sperm whale myoglobin using site-directed mutagenesis and laser photolysis techniques. *J. Biol. Chem.* **1990**, *265*, 20007-20020.
29. Vojtechovský, J.; Chu, K.; Berendzen, J.; Sweet, R.M.; Schlichting, I. Crystal structures of myoglobin-ligand complexes at near-atomic resolution. *Biophys. J.* **1999**, *77*, 2153-2174.
30. Hou, S.; Larsen, R.W.; Boudko, D.; Riley, C.W.; Karatan, E.; Zimmer, M.; Ordal, G.W.; Alam, M. Myoglobin-like aerotaxis transducers in Archaea and Bacteria. *Nature* **2000**, *403*, 540-544.
31. Pinakoulaki, E.; Yoshimura, H.; Daskalakis, D.; Yoshioka, S.; Aono, S.; Varotsis, C. Two ligand-binding sites in the O₂-sensing signal transducer HemAT: Implications for ligand recognition/discrimination and signaling. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 14796-14801.
32. Ischiropoulos, H. Biological tyrosine nitration: A pathophysiological function of nitric oxide and reactive oxygen species. *Arch. Biochem. Biophys.* **1998**, *356*, 1-11.
33. Radi, R.; Beckman, J.S.; Bush, K.M.; Freeman, B.A. Peroxynitrite oxidation of sulfhydryls. The cytotoxic potential of superoxide and nitric oxide. *J. Biol. Chem.* **1991**, *266*, 4244-4250.
34. King, P.A.; Anderson, V.E.; Edwards, J.O.; Gustafson, G.; Plumb, R.C.; Suggs, J.W. A stable solid that generates hydroxyl radical upon dissolution in aqueous solutions: Reaction with proteins and nucleic acid. *J. Am. Chem. Soc.* **1992**, *114*, 5430-5432.
35. Goodin, D.B.; McRee, D.E. The Asp-His-iron triad of cytochrome *c* peroxidase controls the reduction potential electronic structure, and coupling of the tryptophan free radical to the heme. *Biochemistry* **1993**, *32*, 3313-3324.

36. Vogel, K.M.; Spiro, T.G. Resonance raman evidence for a novel charge relay activation mechanism of the CO-dependent heme protein transcription factor CooA. *Biochemistry* **1999**, *38*, 2679-2687.
37. Daskalakis, V.; Farantos, S.C.; Varotsis, C. Assigning vibrational spectra of ferryl-oxo intermediates of cytochrome *c* oxidase by periodic orbits and molecular dynamics. *J. Am. Chem. Soc.* **2008**, *130*, 12385-12393.
38. Crane, B.R.; Siegel, L.M.; Getzoff, E.D. Structures of the siroheme- and Fe₄S₄-containing active center of sulfite reductase in different states of oxidation: Heme activation via reduction-gated exogenous ligand exchange. *Biochemistry* **1997**, *36*, 12101-12119.
39. Decatur, S.M.; Belcher, K.L.; Rickert, P.K.; Franzen, S. Hydrogen bonding modulates binding of exogenous ligands in a myoglobin proximal cavity mutant. *Biochemistry* **1999**, *38*, 11086-11092.
40. Franzen, S. Effect of a charge relay on the vibrational frequencies of carbonmonoxy iron porphine adducts: The coupling of changes in axial ligand bond strength and porphine core size. *J. Am. Chem. Soc.* **2001**, *123*, 12578-12589.
41. Hu, C.; Noll, B.C.; Schulz, C.E.; Scheidt, W.R. Proton-mediated electron configuration change in high-spin iron(II) porphyrinates. *J. Am. Chem. Soc.* **2005**, *127*, 15018-15019.
42. Daskalakis, V.; Pinakoulaki, E.; Stavrakis, S.; Varotsis, C. Probing the environment of Cu_B in Heme-copper oxidases. *J. Phys. Chem. B* **2007**, *111*, 10502-10509.
43. Redfern, P.C.; Zapol, P.; Curtiss, L.A. Assessment of Gaussian-3 and density functional theories for a larger experimental test set. *J. Phys. Chem.* **2000**, *112*, 7374-7383.
44. Straub, J.E.; Karplus, M. Molecular dynamics study of the photodissociation of carbon monoxide from myoglobin: Ligand dynamics in the first 10 ps. *Chem. Phys.* **1991**, *158*, 221-248.
45. Nutt, D.R.; Meuwly, M. Theoretical investigation of infrared spectra and pocket dynamics of photodissociated carbonmonoxy myoglobin. *Biophys. J.* **2003**, *85*, 3612-3623.
46. Badger, R.M. The relation between the internuclear distances and force constants of molecules and its application to polyatomic molecules. *J. Chem. Phys.* **1935**, *3*, 710-714.
47. Brewer, S.H.; Franzen, S. A quantitative theory and computational approach for the vibrational Stark effect. *J. Chem. Phys.* **2003**, *119*, 851-858.
48. Das, T.K.; Pecoraro, C.; Tomson, F.L.; Gennis, R.B.; Rousseau, D.L. The post-translational modification in cytochrome *c* oxidase is required to establish a functional environment of the catalytic site. *Biochemistry* **1998**, *37*, 14471-14476.
49. Proshlyakov, D.A.; Pressler, M.A.; Babcock, G.T. Dioxygen activation and bond cleavage by mixed-valence cytochrome *c* oxidase. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 8020-8025.
50. Pinakoulaki, E.; Pfitzner, U.; Ludwig, B.; Varotsis, C. The role of the cross-link His-Tyr in the functional properties of the binuclear center in cytochrome *c* oxidase. *J. Biol. Chem.* **2002**, *277*, 13563-13568.
51. Pinakoulaki, E.; Pfitzner, U.; Ludwig, B.; Varotsis, C. Direct detection of Fe(IV)=O intermediates in the cytochrome *aa*₃ oxidase from *Paracoccus denitrificans*/H₂O₂ reaction. *J. Biol. Chem.* **2003**, *278*, 18761-18766.
52. Hoffmann, R.; Chen, M.M.L.; Thorn, D.L. Qualitative discussion of alternative coordination modes of diatomic ligands in transition metal complexes. *Inorg. Chem.* **1977**, *16*, 503-511.

53. Spiro, T.G.; Kozlowski, P.M. Will the real FeCO please stand up? *J. Biol. Inorg. Chem.* **1997**, *2*, 516-520.
54. Spiro, T.G.; Kozlowski, P.M. Is the CO adduct of myoglobin bent, and does it matter? *Acc. Chem. Res.* **2001**, *34*, 137-144.
55. Müller, J.D.; McMahon, B.H.; Chien, E.Y.; Sligar, S.G.; Nienhaus, G.U. Connection between the taxonomic substates and protonation of histidines 64 and 97 in carbonmonoxy myoglobin. *Biophys. J.* **1999**, *77*, 1036-1051.
56. Ghosh, A. Mettaloporphyrin-NO bonding: Building bridges with organometallic chemistry. *Acc. Chem. Res.* **2005**, *38*, 943-954.
57. Frisch, M.J.; Trucks, G.W.; Schlegel, H.B.; Scuseria, G.E.; Robb, M.A.; Cheeseman, J.R.; Montgomery, J.A., Jr.; Vreven, T.; Kudin, K.N.; Burant, J.C.; Millam, J.M.; Iyengar, S.S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G.A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J.E.; Hratchian, H.P.; Cross, J.B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R.E.; Yazyev, O.; Austin, A.J.; Cammi, R.; Pomelli, C.; Ochterski, J.W.; Ayala, P.Y.; Morokuma, K.; Voth, G.A.; Salvador, P.; Dannenberg, J.J.; Zakrzewski, V.G.; Dapprich, S.; Daniels, A.D.; Strain, M.C.; Farkas, O.; Malick, D.K.; Rabuck, A.D.; Raghavachari, K.; Foresman, J.B.; Ortiz, J.V.; Cui, Q.; Baboul, A.G.; Clifford, S.; Cioslowski, J.; Stefanov, B.B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R.L.; Fox, D.J.; Keith, T.; Al-Laham, M.A.; Peng, C.Y.; Nanayakkara, A.; Challacombe, M.; Gill, P.M.W.; Johnson, B.; Chen, W.; Wong, M.W.; Gonzalez, C.; Pople, J.A. *Gaussian 03, Revision C.02*; Gaussian, Inc.: Wallingford CT, USA, 2004.
58. Pople, J.A.; Scott, A.P.; Wong, M.W.; Radom, L. Scaling factors for obtaining fundamental vibrational frequencies and zero-point energies from HF/6-31G* and MP2/6-31G* harmonic frequencies. *Isr. J. Chem.* **1993**, *33*, 345-350.
59. Delley, B. An all-electron numerical method for solving the local density functional for polyatomic molecules. *J. Chem. Phys.* **1990**, *92*, 508-517.
60. Dolg, M.; Wedig, U.; Stoll, H.; Preuss, H. Energy-adjusted ab initio pseudopotentials for the first row transition elements. *J. Chem. Phys.* **1987**, *86*, 866-872.
61. Pinakoulaki, E.; Stavarakis, S.; Urbani, A.; Varotsis, C. Resonance raman detection of a ferrous five-coordinate nitrosylheme b_3 complex in cytochrome cbb_3 from *Pseudomonas stutzeri*. *J. Am. Chem. Soc.* **2002**, *124*, 9378-9379.
62. Stavarakis, S.; Pinakoulaki, E.; Urbani, A.; Varotsis, C. Fourier transform infrared evidence for a ferric six-coordinate nitrosylheme b_3 complex of cytochrome cbb_3 oxidase from *Pseudomonas stutzeri* at ambient temperature. *J. Phys. Chem. B* **2002**, *106*, 12860-12862.
63. Pinakoulaki, E.; Varotsis, C. Time-resolved resonance Raman and time-resolved step-scan FTIR studies of nitric oxide reductase from *Paracoccus denitrificans*: Comparison of the heme b_3 -Fe_B site to that of the Heme-Cu_B in oxidases. *Biochemistry* **2003**, *42*, 14856-14861.
64. Ohta, T.; Pinakoulaki, E.; Soulimane, T.; Kitagawa, T.; Varotsis, C. Detection of a photostable five-coordinate heme a_3 -Fe-CO species and functional implications of His384/a10 in CO-bound ba_3 -cytochrome c oxidase from *Thermus thermophilus*. *J. Phys. Chem. B* **2004**, *108*, 5489-5491.

65. Pinakoulaki, E.; Koutsoupakis, C.; Stavrakis, S.; Aggelaki, M.; Papadopoulos, G.; Daskalakis, V.; Varotsis, C. Structural dynamics of Heme-copper oxidases and nitric oxide reductases: Time-resolved step-scan FTIR and time-resolved resonance Raman studies. *J. Raman Spectrosc.* **2005**, *36*, 337-349.
66. Pinakoulaki, E.; Ohta, T.; Soulimane, T.; Kitagawa, T.; Varotsis, C. Detection of the primary His-heme Fe²⁺-NO intermediate in the reduction of NO to N₂O by *ba*₃-oxidase from *Thermus thermophilus*. *J. Am. Chem. Soc.* **2005**, *127*, 15161-15167.
67. Pinakoulaki, E.; Yoshimura, H.; Yoshioka, S.; Aono, S.; Varotsis, C. Recognition and discrimination of gases by the oxygen-sensing signal transducer protein HemAT as revealed by FTIR spectroscopy. *Biochemistry* **2006**, *45*, 7763-7766.
68. Varotsis, C.; Ohta, T.; Kitagawa, T.; Soulimane, T.; Pinakoulaki, E. The structure of the hyponitrite species in a heme Fe-Cu binuclear center. *Angew. Chem. Int.* **2007**, *46*, 2210-2214.

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