

NIH Public Access

Author Manuscript

Arterioscler Thromb Vasc Biol. Author manuscript; available in PMC 2010 February 1.

Published in final edited form as:

Arterioscler Thromb Vasc Biol. 2009 February ; 29(2): 162–168. doi:10.1161/ATVBAHA.107.156752.

Calcific Aortic Stenosis: Lessons Learned from Experimental and Clinical Studies

Nalini M. Rajamannan, MD

From the Division of Cardiology and Department of Medicine, Feinberg School of Medicine

Abstract

Calcific aortic stenosis is the most common indication for surgical valve replacement in the United States. For years this disease has been described as a passive degenerative process during which serum calcium attaches to the valve surface and binds to the leaflet to form nodules. Therefore, surgical treatment of this disease has been the approach towards relieving outflow obstruction in these patients. Recent studies demonstrate an association between atherosclerosis and its risk factors for aortic valve disease. In 2008, there are increasing number of epidemiology and experimental studies to provide evidence that this disease process is not a passive phenomena. There is an active cellular process that develops within the valve leaflet and causes a regulated bone formation to develop. If the atherosclerotic hypothesis is important in the initiation of aortic stenosis than treatments used in slowing the progression of atherosclerosis may be effective in patients with aortic valve disease. This review will review the pathogenesis and the potential for medical therapy in the management of patients with calcific aortic stenosis by examining the lessons provided from the experimental research.

Keywords

Valvular Heart Disease; Lipids; Pathophysiology Atherosclerosis; Experimental Models

Introduction

Calcific aortic stenosis is the most common indication for surgical valve replacement in the United States¹. With the decline of acute rheumatic fever, calcific aortic stenosis has become the most common indication for valvular disorder in the US. Landmark epidemiologic studies identified risk factors for the aortic valve which are similar to those of vascular atherosclerosis, such as smoking, male gender, hypertension, elevated cholesterol levels, and renal failure^{2–} 4 . For years this disease process was thought to be due to build-up of nodules along the valve surface to induce a mechanical stenosis in the valve⁵. Furthermore, surgical therapy for severe symptomatic aortic stenosis is the only treatment option in 2008 as defined in a landmark study from 1968⁵. This study also defined the classic triad of symptoms which include chest pain, shortness of breath and lightheadedness. Finally, this research demonstrated that the life expectancy of this patient population is reduced significantly, if they do not have surgical valve replacement at the onset of symptoms⁵. Currently, it is a Class I indication for surgical valve replacement according to the American Heart Association and American College of Cardiology guidelines for valvular heart disease⁶. Over the past decade, there are a growing number of studies evaluating human disease tissues to define the cellular pathways important in this

Address correspondence to: Nalini M. Rajamannan M.D., Valve Director Bluhm Cardiovascular Institute, Northwestern University Feinberg School of Medicine, 303 E Chicago, Tarry 12-717, Chicago, IL 60611, n-rajamannan@northwestern.edu, Phone number 312-695-0067.

calcific aortic stenosis. This review will bring together the basic science and clinical science to develop a unified approach towards treating this disease.

Aortic Valve Calcification

The presence of calcification in the aortic valve is responsible for valve stenosis. Recent descriptive studies from patient specimens have demonstrated the hallmark features of aortic valve disease: including early atherosclerosis, cell proliferation and osteoblast \exp ession^{7–} 10 . To understand aortic valve disease, three interrelated events responsible for the development of valve calcification to consider: 1) classical cardiovascular risk factors, 2) genetic factors and 3) valve biology. The interrelationship of these events results in the final common pathway of this disease: a calcifying osteoblast phenotype. The evidence for these three pathways leading to the development of human aortic valve calcification can be found in the experimental and clinical studies outlined in this review.

Traditional Cardiovascular Atherosclerotic Risk Factors

In the past decade, landmark studies^{2, 3}, has described the risk factors for calcific aortic stenosis as identified by large epidemiologic cohort studies which include lipids, hypertension, male gender, renal failure, and diabetes. Many population science papers have subsequently confirmed these findings^{2, 11, 12,13}. These studies have implicated the traditional risk factors for cardiovascular atherosclerosis important in the development of calcific aortic stenosis. The role of lipids as a risk factor for vascular atherosclerosis has been defined in the literature for years. Atherosclerosis is a complex multifactorial process which produces a lesion composed of lipids^{14, 15}, macrophages¹⁶, and proliferating smooth muscle cells¹⁷ and apoptosis¹⁸ which is regulated by endothelial nitric oxide synthase^{19–23} which over time causes occlusion of the vessel diameter. The understanding of these clinical risk factors are providing the foundation for the cellular studies and the potential for targeted medical therapies for this disease similar to vascular atherosclerosis.

Surgical pathological studies have demonstrated the presence of LDL and atherosclerosis in calcified valves, suggesting a common cellular mechanism^{24, 25}. Furthermore, patients who have the diagnosis of familial hypercholesterolemia develop aggressive peripheral vascular disease, coronary artery disease, as well as aortic valve lesions which calcify with age. The first descriptions of atherosclerotic aortic valve disease have been in patients with Familial Hypercholesterolemia(FH) who have an early atherosclerotic lesion along the aortic surface of the valve leaflet^{26–28,29}. The discovery of atherosclerosis in the aortic valve in the FH patient population provides the initial proof of principal for the potential treatment of lipids to slow the progression of aortic valve disease.

Experimental Models of Valvular Heart Disease

If atherosclerotic risk factors are important in the development of valvular heart disease, than experimental models of atherosclerosis are important in the understanding this disease process. Studies in mice and rabbits have confirmed that experimental hypercholesterolemia causes both atherosclerosis and calcification in the aortic valves^{10, 30–34}. The experimental hypercholesterolemia diet has been used for over one hundred years for to evaluate the mechanisms of vascular disease. Table 1 demonstrates the *in vivo* rabbit and mouse models of valvular heart disease. The first study to describe early endothelial abnormalities in the aortic valves was in experimental hypercholesterolemia rabbits $35-38$. This hypothesis has been further developed in the rabbit model to test for multiple markers of the atherosclerotic process within the valve which are critical steps towards the development of valvular calcification process. The initial disease markers described in valve atherosclerosis include cell proliferation and apoptosis³⁰. The early valve lesion of aortic valve sclerosis was also shown in a rabbit

model of hypercholesterolemia³⁹. This model was extended using atorvastatin to modify gene markers to define the bone and atherosclerotic pathways and to demonstrate attenuation with a statin¹⁰. Atorvastatin attenuated the early gene markers of bone formation and macrophage infiltration along the aortic surface of the experimental hypercholesterolemia aortic valve¹⁰. The rabbit model of cholesterol with and without atorvastatin was extended to three and six months duration, to determine the timing of calcification in the valves. Three months of cholesterol induced the early mineralization and eNOS modification in the valve as shown by MicroCT and standard eNOS assays 21 .

The next experimental proof of principle in the rabbit model, was to demonstrate complex calcification with chronic duration of cholesterol diet to prove the bone differentiation mechanism. 6 months of cholesterol diet treatment induced marked thickening and complex calcification in the aortic valve leaflets with pharmacologic attenuation with atorvastatin by MicroCT analysis40. The effects of statins were also confirmed by testing *in vitro* for the inhibition of extracellular matrix $41, 42$. Finally, the most recent experimental model tested the effect of an angiotensin receptor blocker(ARB) on the inhibition of atherosclerotic pathways in the rabbit model of hypercholesterolemia⁴³. These findings demonstrate experimentally the beneficial effects of HMG CoA Reductase Agents and ARB's *in vivo* and *in vitro*.

Hemodynamic studies have evolved to study the noninvasive evidence for the development of aortic stenosis³¹, in rabbits treated with cholesterol + vitamin D model. These studies demonstrate the presence of early stenosis with an increase in the pulse wave Doppler velocity across the aortic valve leaflet. This was further confirmed in the LDLR^{$-/-$} mouse model of hypercholesterolemia³⁴. The next hemodynamic study demonstrated severe stenosis and calcification with chronic cholesterol treatment for 24 months of cholesterol.44 The study tested the genetic knock-out mouse which lacks the receptor for the Low-density lipoprotein receptor and expresses only the receptor for the human apoB100 (LDLr^{-/-}apoB^{100/100}) in an aging genetic mouse model which developed mineralization in the valve. The imaging experiments have been taken one step further to measure the development of the osteoblast phenotype in the aortic valve by using multimodality imaging³³. The authors hypothesize that the flexion area of the aortic leaflets near the attachment of the aortic root (commissure) encounters the highest mechanical forces, which might induce endothelial cell activation/injury and expression of adhesion molecules such as VCAM-1, intracellular adhesion molecules-1, and E-selectin. Molecular imaging of the earliest stagesof calcification may identify high-risk valves while disease is silent and may enable the monitoring of valvular osteogenic activity during therapeutic interventions such as lipid lowering.

An important mechanistic study, demonstrated in native aortic valves in hypercholesterolemic mice that ten percent of cells are bone marrow derived cells within the atherosclerotic lesion45. These investigators hypothesize that likely both altered lipid metabolism and aging are essential for the development of murine aortic sclerosis, which potentially causes functional stenosis and regurgitation. Their findings suggest that some of the smooth muscle-like and osteoblast-like cells in degenerative valves might derive from bone marrow. It is likely that growth factors expressed in the endothelium with abnormal oxidative stress may play a role, at least in part, in the recruitment and homing of bone-marrow-derived cells to the site of valvular remodeling. Future studies evaluating mechanisms of the myofibroblast differentiation process, stem cell homing experiments, other medical therapies will provide further proof of the mechanisms for valvular heart disease and further understanding of the osteoblast differentiation cascade.

Genetics of Aortic Valve Disease

A growing number of studies are providing further evidence towards the genetic predisposition for aortic valve disease. Two of the studies have correlated genetic lipoprotein abnormalities in patients predisposed to the development of calcific aortic stenosis^{46, 47}. The initial genetic study demonstrated an association of the vitamin D receptor polymorphism with calcific aortic valve stenosis. These investigators found that the B allele of the vitamin D receptor is more common in patients with calcific aortic valve stenosis⁴⁸. In this study, the investigators found an association between the B allele predisposes the carriers to a decrease in calcium absorption and therefore an increase in bone loss. The discovery of this polymorphism further confirms the potential abnormal bone signaling pathways important in the development of this disease.

The next landmark discovery is the loss of function mutation in the Notch1 receptor in patients with calcific aortic stenosis⁴⁹. These patients were identified in the Texas Heart Study as having valvular heart disease and accelerated calcification. It is important to note that the kindred of patients also had associated congenital heart abnormalities present in individual family members. Thus, implicating Notch1 in the development of congenital heart abnormalities as well as accelerated valve calcification. Another study demonstrated that the Pvull polymorphisms in the estrogen receptor alpha gene is related to both the presence of aortic stenosis in postmenpausal women and to lipid levels in adolescent females, suggesting that this polymorphism may influence the risk of aortic stenosis by affecting gender and lipid levels⁵⁰. An interesting study⁵¹, demonstrated a familial aggregation for calcific aortic valve disease in the western part of France. These investigators found that the geographic distribution of calcific aortic valve disease is highly heterogeneous, with an average frequency of operated calcific aortic valve disease of 1.13 per 1000 inhabitants but up to 9.38 per 1000 in specific parishes. These genetic and familial studies show that lipids, Vitamin D, Estrogen receptor and Notch1 signaling in addition to a familial aggregation have important implications in the development of aortic stenosis and that an early atherosclerotic lesion secondary to genetic lipid abnormalities are important in the early initiation of this disease. Future genetic testing in the development of calcification in patients without traditional risk factors may play an important role in the treatment of this disease.

Osteoblast Phenotype is the Final Common Pathway for Aortic Valve Calcification

The presence of calcification in the aortic valve is responsible for hemodynamic progression of aortic valve stenosis. Recent descriptive studies from patient specimens have demonstrated the critical features of aortic valve calcification: including osteoblast expression, cell proliferation and atherosclerosis^{7–10}. These studies define the biochemical and histological characterization of these valve lesions. Furthermore, these studies have also shown that specific bone cell phenotypes are present in calcifying valve tissue from human specimens^{52, 53}. The vascular biologists have performed the initial studies^{54,55} which demonstrate the ability of calcifying vascular cells have the multipotential ability to differentiate to calcifying phenotypes. Calcification in the aortic valve is the final common pathway that leads to aortic valve stenosis. This was confirmed in a landmark echocardiographic study⁵⁶, demonstrating severe aortic stenosis and severe calcification have a worse prognosis than patients with mild calcification and severe aortic stenosis. The data further corroborates the evidence that calcification is the defining feature clinically for prognostic future prognostic implications for this patients population.

Studies have shown that cardiovascular calcification is composed of hydroxyapatite deposited on a bone-like matrix of collagen, osteopontin (OP), and other minor bone matrix proteins 8 , ⁵⁷, 58. This was confirmed histologically with the presence of osteoblast bone formation in

calcified aortic valves removed from surgical valve replacement^{8, 9}. In addition, osteopontin expression has been demonstrated in the mineralization zones of heavily calcified aortic valves obtained at autopsy and surgery^{7, 8}. This discovery has been extended in a study which shows by RTPCR analysis, histomorphometry and microCT that an osteoblast-like cellular phenotype is present in calcified aortic valves removed at the time of surgical valve replacement⁹. The increased gene expression of osteopontin, bone sialoprotein, and Cbfa1 (the osteoblast specific transcription factor for bone formation) were increased in the calcified aortic valves as compared to the control valves from heart transplantation. This is the first to provide the evidence for the gene differentiation pathway in this calcifying tissue and an upregulation of the osteoblast gene program. These discoveries are the foundation for the hypothesis that the cells residing in the aortic valve have the potentiality to differentiate into a bone forming cell which over time mineralizes and expresses an ossification phenotype.

To test the osteoblast hypothesis further, evidence for signaling pathways are important in the development of this disease. The Lrp5 pathway was discovered to regulate bone formation in different diseases of the bone^{59, 60}. There are three studies which have confirmed the regulation of the Lrp5/Wnt pathway in cardiovascular calcification in experimental models of calcification^{40,52,61}. The Lrp5 pathway is one of many signaling pathways important in the development of bone formation. The Lrp5 receptor and other signaling pathways are important in the development of calcific aortic stenosis. Many of these signaling factors are similar to those found in vascular atherosclerosis and bone formation. Matrix MetalloProteinases (MMP) 53, 62, Interleukin 1^{63} , Transforming growth factor-beta(TGF-beta)⁶⁴, purine nucleotides^{42,} 65 , RANK 66 , osteoprotegrin(OPG) 66 , and TNF alpha⁶⁷, have all been identified as signaling pathways important in the development of this disease process. Evidence, for the angiotensin converting enzyme pathway expressed and colocalize with LDL in calcified aortic valves also plays a role in future potential medical therapy68. Recent studies have shown that an increased expression of elastolytic cathepsins S, K, and V and their inhibitor Cystatin C in stenotic aortic valves69. These signaling studies from *ex vivo* human calcified aortic valves are the critical links between the experimental and translational implications for the future treatment of valvular heart disease. Figure 1, demonstrates the signaling pathways and cellular events important in the development of this disease. In the presence of lipids, the aortic valve endothelium is activated and abnormal oxidation state develops. The myofibroblast cells then begin to proliferate and synthesize extracellular bone matrix proteins with the upregulation of the various signaling pathways outlined in this review. These proteins overtime mineralize and calcify. ACE inhibitors and statins have the potential to modify this disease process and slow the progression of this disease.

Summary of In vivo, In vitro and Ex vivo Models of Aortic Valve Disease

These models have provided the clues for the development of therapeutic approaches for this disease. Understanding these models and genetics will help our future understanding of this complex disease process. First, the initiating events in vascular disease and valvular disease may be similar, but the outcomes and the understanding of treatment of this disease are different because of the different biologic endpoints for vascular disease as compared to valvular heart disease. Figure 2, shows the interrelated events important in the development of aortic valve disease which is important in the understanding of the complexity for developing clinical trials in this field. Future clinical trials will need to include the atherosclerotic hypothesis in addition to the potential of genetics affecting the development of this disease. Finally, understanding the signaling pathways involved in the development of cell proliferation and osteoblast bone formation will allow for the medical therapy for these patients.

Discussion

Recent epidemiological studies have revealed that the risk factors for arterial atherosclerosis, male gender, smoking, and elevated serum cholesterol, are similar to the risk factors associated with development of aortic valve stenosis. The risk factors, growing number of models of experimental hypercholesterolemia which produce atherosclerosis in the aortic valve are similar to the early stages of vascular atherosclerotic lesion formation. The interplay of genetics, environmental risk factors and cellular biology play a critical role towards the underlying mechanism of this disease process. In summary, these findings suggest that medical therapies may have a potential role in patients in the early stages of this disease process to prolong the time to severe aortic stenosis and to delay the timing of surgery.

Acknowledgments

Sources of Funding: This work was completed with the support of an American Heart Association Grant-in-Aid (0555714Z) and a grant from the National Institute of Health (5K08HL073927-04, 1R01HL085591-01A1). **Contributions:** Dr. Rajamannan would like to thank Sheila Macomber for figure preparation and graphic design. **Conflict of Interest:** Nalini M. Rajamannan is an inventor on a patent for the use of statins in degeneration of aortic valve disease. This patent is owned by the Mayo Clinic and Dr. Rajamannan does not receive any royalties from this patent.

References

- 1. Lindroos M, Kupari M, Heikkila J, Tilvis R. Prevalence of aortic valve abnormalities in the elderly: an echocardiographic study of a random population sample. Journal of the American College of Cardiology 1993;21(5):1220–1225. [PubMed: 8459080]
- 2. Stewart BF, Siscovick D, Lind BK, Gardin JM, Gottdiener JS, Smith VE, Kitzman DW, Otto CM. Clinical factors associated with calcific aortic valve disease. Cardiovascular Health Study. Journal of the American College of Cardiology 1997;29(3):630–634. [PubMed: 9060903]
- 3. Otto CM, Lind BK, Kitzman DW, Gersh BJ, Siscovick DS. Association of aortic-valve sclerosis with cardiovascular mortality and morbidity in the elderly. [comment]. New England Journal of Medicine 1999;341(3):142–147. [PubMed: 10403851]
- 4. Aronow WS, Ahn C, Kronzon I, Goldman ME. Association of coronary risk factors and use of statins with progression of mild valvular aortic stenosis in older persons. American Journal of Cardiology 2001;88(6):693–695. [PubMed: 11564402]
- 5. Ross J Jr, Braunwald E. Aortic stenosis. Circulation 1968;38(1 Suppl):61–67. [PubMed: 4894151]
- 6. Bonow RO, Carabello BA, Kanu C, de Leon AC Jr, Faxon DP, Freed MD, Gaasch WH, Lytle BW, Nishimura RA, O'Gara PT, O'Rourke RA, Otto CM, Shah PM, Shanewise JS, Smith SC Jr, Jacobs AK, Adams CD, Anderson JL, Antman EM, Fuster V, Halperin JL, Hiratzka LF, Hunt SA, Nishimura R, Page RL, Riegel B. ACC/AHA 2006 guidelines for the management of patients with valvular heart disease: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines (writing committee to revise the 1998 Guidelines for the Management of Patients With Valvular Heart Disease): developed in collaboration with the Society of Cardiovascular Anesthesiologists: endorsed by the Society for Cardiovascular Angiography and Interventions and the Society of Thoracic Surgeons. Circulation 2006;114(5):e84–231. [PubMed: 16880336]
- 7. O'Brien KD, Kuusisto J, Reichenbach DD, Ferguson M, Giachelli C, Alpers CE, Otto CM. Osteopontin is expressed in human aortic valvular lesions. [comment]. Circulation 1995;92(8):2163–2168. [PubMed: 7554197]
- 8. Mohler ER 3rd, Gannon F, Reynolds C, Zimmerman R, Keane MG, Kaplan FS. Bone formation and inflammation in cardiac valves. Circulation 2001;103(11):1522–1528. [PubMed: 11257079]
- 9. Rajamannan NM, Subramaniam M, Rickard D, Stock SR, Donovan J, Springett M, Orszulak T, Fullerton DA, Tajik AJ, Bonow RO, Spelsberg T. Human aortic valve calcification is associated with an osteoblast phenotype. Circulation 2003;107(17):2181–2184. [PubMed: 12719282]
- 10. Rajamannan NM, Subramaniam M, Springett M, Sebo TC, Niekrasz M, McConnell JP, Singh RJ, Stone NJ, Bonow RO, Spelsberg TC. Atorvastatin inhibits hypercholesterolemia-induced cellular

proliferation and bone matrix production in the rabbit aortic valve. Circulation 2002;105(22):2260– 2265.

- 11. Palta S, Pai AM, Gill KS, Pai RG. New insights into the progression of aortic stenosis: implications for secondary prevention. Circulation 2000;101(21):2497–2502. [PubMed: 10831524]
- 12. Aronow WS, Ahn C, Kronzon I. Association of mitral annular calcium with symptomatic peripheral arterial disease in older persons. Am J Cardiol 2001;88(3):333–334. [PubMed: 11472724]
- 13. Briand M, Lemieux I, Dumesnil JG, Mathieu P, Cartier A, Despres JP, Arsenault M, Couet J, Pibarot P. Metabolic syndrome negatively influences disease progression and prognosis in aortic stenosis. J Am Coll Cardiol 2006;47(11):2229–2236. [PubMed: 16750688]
- 14. Desai MY, Rodriguez A, Wasserman BA, Gerstenblith G, Agarwal S, Kennedy M, Bluemke DA, Lima JA. Association of cholesterol subfractions and carotid lipid core measured by MRI. Arterioscler Thromb Vasc Biol 2005;25(6):e110–111. [PubMed: 15923536]
- 15. Subbaiah PV, Gesquiere LR, Wang K. Regulation of the selective uptake of cholesteryl esters from high density lipoproteins by sphingomyelin. J Lipid Res 2005;46(12):2699–2705. [PubMed: 16162942]
- 16. Kim WJ, Chereshnev I, Gazdoiu M, Fallon JT, Rollins BJ, Taubman MB. MCP-1 deficiency is associated with reduced intimal hyperplasia after arterial injury. Biochem Biophys Res Commun 2003;310(3):936–942. [PubMed: 14550294]
- 17. Tanner FC, Boehm M, Akyurek LM, San H, Yang ZY, Tashiro J, Nabel GJ, Nabel EG. Differential effects of the cyclin-dependent kinase inhibitors p27(Kip1), p21(Cip1), and p16(Ink4) on vascular smooth muscle cell proliferation. Circulation 2000;101(17):2022–2025. [PubMed: 10790340]
- 18. Zhang R, Luo D, Miao R, Bai L, Ge Q, Sessa WC, Min W. Hsp90-Akt phosphorylates ASK1 and inhibits ASK1-mediated apoptosis. Oncogene 2005;24(24):3954–3963. [PubMed: 15782121]
- 19. Laufs U, Liao JK. Post-transcriptional regulation of endothelial nitric oxide synthase mRNA stability by Rho GTPase. Journal of Biological Chemistry 1998;273(37):24266–24271. [PubMed: 9727051]
- 20. Venema RC, Sayegh HS, Kent JD, Harrison DG. Identification, characterization, and comparison of the calmodulin-binding domains of the endothelial and inducible nitric oxide synthases. Journal of Biological Chemistry 1996;271(11):6435–6440. [PubMed: 8626444]
- 21. Rajamannan NM, Subramaniam M, Stock SR, Stone NJ, Springett M, Ignatiev KI, McConnell JP, Singh RJ, Bonow RO, Spelsberg TC. Atorvastatin inhibits calcification and enhances nitric oxide synthase production in the hypercholesterolaemic aortic valve. Heart 2005;91(6):806–810. [PubMed: 15894785]
- 22. Charest A, Pepin A, Shetty R, Cote C, Voisine P, Dagenais F, Pibarot P, Mathieu P. Distribution of SPARC During Neovascularization of Degenerative Aortic Stenosis. Heart. 2006
- 23. Ngo DT, Heresztyn T, Mishra K, Marwick TH, Horowitz JD. Aortic stenosis is associated with elevated plasma levels of asymmetric dimethylarginine (ADMA). Nitric Oxide 2007;16(2):197–201. [PubMed: 17126043]
- 24. O'Brien KD, Reichenbach DD, Marcovina SM, Kuusisto J, Alpers CE, Otto CM. Apolipoproteins B, (a), and E accumulate in the morphologically early lesion of 'degenerative' valvular aortic stenosis. Arteriosclerosis, Thrombosis & Vascular Biology 1996;16(4):523–532.
- 25. Olsson M, Thyberg J, Nilsson J. Presence of oxidized low density lipoprotein in nonrheumatic stenotic aortic valves. Arterioscler Thromb Vasc Biol 1999;19(5):1218–1222. [PubMed: 10323772]
- 26. Goldstein JL, Brown MS. Familial hypercholesterolemia: identification of a defect in the regulation of 3-hydroxy-3-methylglutaryl coenzyme A reductase activity associated with overproduction of cholesterol. Proc Natl Acad Sci U S A 1973;70(10):2804–2808. [PubMed: 4355366]
- 27. Sprecher DL, Schaefer EJ, Kent KM, Gregg RE, Zech LA, Hoeg JM, McManus B, Roberts WC, Brewer HB Jr. Cardiovascular features of homozygous familial hypercholesterolemia: analysis of 16 patients. American Journal of Cardiology 1984;54(1):20–30. [PubMed: 6331147]
- 28. Kawaguchi A, Miyatake K, Yutani C, Beppu S, Tsushima M, Yamamura T, Yamamoto A. Characteristic cardiovascular manifestation in homozygous and heterozygous familial hypercholesterolemia 1999 Mar;1999:410–418.
- 29. Rajamannan NM, Edwards WD, Spelsberg TC. Hypercholesterolemic aortic-valve disease. New England Journal of Medicine 2003;349(7):717–718. [PubMed: 12917318]

- 30. Rajamannan NM, Sangiorgi G, Springett M, Arnold K, Mohacsi T, Spagnoli LG, Edwards WD, Tajik AJ, Schwartz RS. Experimental hypercholesterolemia induces apoptosis in the aortic valve. Journal of Heart Valve Disease 2001;10(3):371–374. [PubMed: 11380101]
- 31. Drolet MC, Arsenault M, Couet J. Experimental aortic valve stenosis in rabbits. Journal of the American College of Cardiology 2003;41(7):1211–1217. [PubMed: 12679224]
- 32. Weiss RM, Ohashi M, Miller JD, Young SG, Heistad DD. Calcific aortic valve stenosis in old hypercholesterolemic mice. Circulation 2006;114(19):2065–2069. [PubMed: 17075015]
- 33. Aikawa E, Nahrendorf M, Sosnovik D, Lok VM, Jaffer FA, Aikawa M, Weissleder R. Multimodality molecular imaging identifies proteolytic and osteogenic activities in early aortic valve disease. Circulation 2007;115(3):377–386. [PubMed: 17224478]
- 34. Drolet MC, Roussel E, Deshaies Y, Couet J, Arsenault M. A high fat/high carbohydrate diet induces aortic valve disease in C57BL/6J mice. J Am Coll Cardiol 2006;47(4):850–855. [PubMed: 16487855]
- 35. Sarphie TG. Interactions of IgG and beta-VLDL with aortic valve endothelium from hypercholesterolemic rabbits. Atherosclerosis 1987 Dec;:199–212. [PubMed: 3322301]
- 36. Sarphie TG. A cytochemical study of the surface properties of aortic and mitral valve endothelium from hypercholesterolemic rabbits. Exp Mol Pathol 1986 Jun;:281–296. [PubMed: 3720917]
- 37. Sarphie TG. Anionic surface properties of aortic and mitral valve endothelium from New Zealand white rabbits. Am J Anat 1985 Oct;:145–160. [PubMed: 3840642]
- 38. Sarphie TG. Surface responses of aortic valve endothelia from diet-induced, hypercholesterolemic rabbits. Atherosclerosis 1985 Mar;:283–299. [PubMed: 3994784]
- 39. Cimini M, Boughner DR, Ronald JA, Aldington L, Rogers KA. Development of aortic valve sclerosis in a rabbit model of atherosclerosis: an immunohistochemical and histological study. The Journal of heart valve disease 2005;14(3):365–375. [PubMed: 15974532]
- 40. Rajamannan NM, Subramaniam M, Caira F, Stock SR, Spelsberg TC. Atorvastatin inhibits hypercholesterolemia-induced calcification in the aortic valves via the Lrp5 receptor pathway. Circulation 2005;112(9 Suppl):I229–234. [PubMed: 16159822]
- 41. Osman L, Yacoub MH, Latif N, Amrani M, Chester AH. Role of human valve interstitial cells in valve calcification and their response to atorvastatin. Circulation 2006;114(1 Suppl):I547–552. [PubMed: 16820635]
- 42. Osman L, Chester AH, Amrani M, Yacoub MH, Smolenski RT. A novel role of extracellular nucleotides in valve calcification: a potential target for atorvastatin. Circulation 2006;114(1 Suppl):I566–572. [PubMed: 16820639]
- 43. Arishiro K, Hoshiga M, Negoro N, Jin D, Takai S, Miyazaki M, Ishihara T, Hanafusa T. Angiotensin receptor-1 blocker inhibits atherosclerotic changes and endothelial disruption of the aortic valve in hypercholesterolemic rabbits. J Am Coll Cardiol 2007;49(13):1482–1489. [PubMed: 17397679]
- 44. Ortlepp JR, Pillich M, Schmitz F, Mevissen V, Koos R, Weiss S, Stork L, Dronskowski R, Langebartels G, Autschbach R, Brandenburg V, Woodruff S, Kaden JJ, Hoffmann R. Lower serum calcium levels are associated with greater calcium hydroxyapatite deposition in native aortic valves of male patients with severe calcific aortic stenosis. The Journal of heart valve disease 2006;15(4): 502–508. [PubMed: 16901043]
- 45. Tanaka K, Sata M, Fukuda D, Suematsu Y, Motomura N, Takamoto S, Hirata Y, Nagai R. Ageassociated aortic stenosis in apolipoprotein E-deficient mice. J Am Coll Cardiol 2005;46(1):134– 141. [PubMed: 15992647]
- 46. Avakian SD, Annicchino-Bizzacchi JM, Grinberg M, Ramires JA, Mansura AP. Apolipoproteins AI, B, and E polymorphisms in severe aortic valve stenosis. Clin Genet 2001;60(5):381–384. [PubMed: 11903341]
- 47. Novaro GM, Sachar R, Pearce GL, Sprecher DL, Griffin BP. Association between apolipoprotein E alleles and calcific valvular heart disease. Circulation 2003;108(15):1804–1808. [PubMed: 14530190]
- 48. Ortlepp JR, Hoffmann R, Ohme F, Lauscher J, Bleckmann F, Hanrath P. The vitamin D receptor genotype predisposes to the development of calcific aortic valve stenosis. Heart 2001;85(6):635– 638. [PubMed: 11359741]

- 49. Garg V, Muth AN, Ransom JF, Schluterman MK, Barnes R, King IN, Grossfeld PD, Srivastava D. Mutations in NOTCH1 cause aortic valve disease. Nature 2005;437(7056):270–274. [PubMed: 16025100]
- 50. Nordstrom P, Glader CA, Dahlen G, Birgander LS, Lorentzon R, Waldenstrom A, Lorentzon M. Oestrogen receptor alpha gene polymorphism is related to aortic valve sclerosis in postmenopausal women. J Intern Med 2003;254(2):140–146. [PubMed: 12859695]
- 51. Probst V, Le Scouarnec S, Legendre A, Jousseaume V, Jaafar P, Nguyen JM, Chaventre A, Le Marec H, Schott JJ. Familial aggregation of calcific aortic valve stenosis in the western part of France. Circulation 2006;113(6):856–860. [PubMed: 16461814]
- 52. Caira FC, Stock SR, Gleason TG, McGee EC, Huang J, Bonow RO, Spelsberg TC, McCarthy PM, Rahimtoola SH, Rajamannan NM. Human degenerative valve disease is associated with upregulation of low-density lipoprotein receptor-related protein 5 receptor-mediated bone formation. J Am Coll Cardiol 2006;47(8):1707–1712. [PubMed: 16631011]
- 53. Jian B, Jones PL, Li Q, Mohler ER 3rd, Schoen FJ, Levy RJ. Matrix metalloproteinase-2 is associated with tenascin-C in calcific aortic stenosis. Am J Pathol 2001;159(1):321–327. [PubMed: 11438479]
- 54. Demer LL. Cholesterol in vascular and valvular calcification. Circulation 2001;104(16):1881–1883. [PubMed: 11602487]
- 55. Tintut Y, Alfonso Z, Saini T, Radcliff K, Watson K, Bostrom K, Demer LL. Multilineage potential of cells from the artery wall. Circulation 2003;108(20):2505–2510. [PubMed: 14581408]
- 56. Rosenhek R, Binder T, Porenta G, et al. Predictors of outcome in severe asymptomatic aortic stenosis. NEJM 2000;343:611–617. [PubMed: 10965007]
- 57. Mohler ER 3rd, Adam LP, McClelland P, Graham L, Hathaway DR. Detection of osteopontin in calcified human aortic valves. Arteriosclerosis, Thrombosis & Vascular Biology 1997;17(3):547– 552.
- 58. O'Brien KD, Kuusisto J, Reichenbach DD, Ferguson M, Giachelli C, Alpers CE, Otto CM. Osteopontin is expressed in human aortic valvular lesions. Circulation 1995;92(8):2163–2168. [PubMed: 7554197]
- 59. Gong Y, Slee RB, Fukai N, Rawadi G, Roman-Roman S, Reginato AM, Wang H, Cundy T, Glorieux FH, Lev D, Zacharin M, Oexle K, Marcelino J, Suwairi W, Heeger S, Sabatakos G, Apte S, Adkins WN, Allgrove J, Arslan-Kirchner M, Batch JA, Beighton P, Black GC, Boles RG, Boon LM, Borrone C, Brunner HG, Carle GF, Dallapiccola B, De Paepe A, Floege B, Halfhide ML, Hall B, Hennekam RC, Hirose T, Jans A, Juppner H, Kim CA, Keppler-Noreuil K, Kohlschuetter A, LaCombe D, Lambert M, Lemyre E, Letteboer T, Peltonen L, Ramesar RS, Romanengo M, Somer H, Steichen-Gersdorf E, Steinmann B, Sullivan B, Superti-Furga A, Swoboda W, van den Boogaard MJ, Van Hul W, Vikkula M, Votruba M, Zabel B, Garcia T, Baron R, Olsen BR, Warman ML. Osteoporosis-Pseudoglioma Syndrome Collaborative G. LDL receptor-related protein 5 (LRP5) affects bone accrual and eye development. Cell 2001;107(4):513–523. [PubMed: 11719191]
- 60. Boyden LM, Mao J, Belsky J, Mitzner L, Farhi A, Mitnick MA, Wu D, Insogna K, Lifton RP. High bone density due to a mutation in LDL-receptor-related protein 5. N Engl J Med 2002;346(20):1513– 1521. [PubMed: 12015390]
- 61. Shao JS, Cheng SL, Pingsterhaus JM, Charlton-Kachigian N, Loewy AP, Towler DA. Msx2 promotes cardiovascular calcification by activating paracrine Wnt signals. J Clin Invest 2005;115(5):1210– 1220. [PubMed: 15841209]
- 62. Kaden JJ, Vocke DC, Fischer CS, Grobholz R, Brueckmann M, Vahl CF, Hagl S, Haase KK, Dempfle CE, Borggrefe M. Expression and activity of matrix metalloproteinase-2 in calcific aortic stenosis. Z Kardiol 2004;93(2):124–130. [PubMed: 14963678]
- 63. Kaden JJ, Dempfle CE, Grobholz R, Tran HT, Kilic R, Sarikoc A, Brueckmann M, Vahl C, Hagl S, Haase KK, Borggrefe M. Interleukin-1 beta promotes matrix metalloproteinase expression and cell proliferation in calcific aortic valve stenosis. Atherosclerosis 2003;170(2):205–211. [PubMed: 14612199]
- 64. Jian B, Narula N, Li QY, Mohler ER 3rd, Levy RJ. Progression of aortic valve stenosis: TGF-beta1 is present in calcified aortic valve cusps and promotes aortic valve interstitial cell calcification via apoptosis. Ann Thorac Surg 2003;75(2):457–465. [PubMed: 12607654]discussion 465–456

- 65. Osman L, Amrani M, Isley C, Yacoub MH, Smolenski RT. Stimulatory effects of atorvastatin on extracellular nucleotide degradation in human endothelial cells. Nucleosides Nucleotides Nucleic Acids 2006;25(9–11):1125–1128. [PubMed: 17065076]
- 66. Kaden JJ, Bickelhaupt S, Grobholz R, Haase KK, Sarikoc A, Kilic R, Brueckmann M, Lang S, Zahn I, Vahl C, Hagl S, Dempfle CE, Borggrefe M. Receptor activator of nuclear factor kappaB ligand and osteoprotegerin regulate aortic valve calcification. J Mol Cell Cardiol 2004;36(1):57–66. [PubMed: 14734048]
- 67. Kaden JJ, Kilic R, Sarikoc A, Hagl S, Lang S, Hoffmann U, Brueckmann M, Borggrefe M. Tumor necrosis factor alpha promotes an osteoblast-like phenotype in human aortic valve myofibroblasts: a potential regulatory mechanism of valvular calcification. Int J Mol Med 2005;16(5):869–872. [PubMed: 16211257]
- 68. O'Brien KD, Shavelle DM, Caulfield MT, McDonald TO, Olin-Lewis K, Otto CM, Probstfield JL. Association of angiotensin-converting enzyme with low-density lipoprotein in aortic valvular lesions and in human plasma. Circulation 2002;106(17):2224–2230. [PubMed: 12390952]
- 69. Helske S, Syvaranta S, Lindstedt KA, Lappalainen J, Oorni K, Mayranpaa MI, Lommi J, Turto H, Werkkala K, Kupari M, Kovanen PT. Increased expression of elastolytic cathepsins S, K, and V and their inhibitor cystatin C in stenotic aortic valves. Arterioscler Thromb Vasc Biol 2006;26(8):1791– 1798. [PubMed: 16728655]

Figure 1. Cellular, Molecular and Genetic Mechanisms of Calific Aortic Stenosis

Model Implicating Lipids in the Development of Calcific Aortic Stenosis and the potential for Future Medical Therapies Targeting this disease at the cellular level in the Aortic Valve.

Figure 2.

Three Factors Responsible for the Development of Calcific Aortic Stenosis: Cardiovascular Risk Factors, Genetic Factors and Osteoblast Regulatory Pathways.

Experimental Animal Models of Valvular Heart Disease **Experimental Animal Models of Valvular Heart Disease**

List of Experimental Models of Aortic Valve Disease. List of Experimental Models of Aortic Valve Disease.

