

Review

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Pathophysiology of cell phone radiation: oxidative stress and carcinogenesis with focus on male reproductive system

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Abstract

Hazardous health effects stemming from exposure to radiofrequency electromagnetic waves (RF-EMW) emitted from cell phones have been reported in the literature. However, the cellular target of RF-EMW is still controversial. This review identifies the plasma membrane as a target of RF-EMW. In addition, the effects of RF-EMW on plasma membrane structures (i.e. NADH oxidase, phosphatidylserine, ornithine decarboxylase) and voltage-gated calcium channels are discussed. We explore the disturbance in reactive oxygen species (ROS) metabolism caused by RF-EMW and delineate NADH oxidase mediated ROS formation as playing a central role in oxidative stress (OS) due to cell phone radiation (with a focus on the male reproductive system). This review also addresses: 1) the controversial effects of RF-EMW on mammalian cells and sperm DNA as well as its effect on apoptosis, 2) epidemiological, in vivo animal and in vitro studies on the effect of RF-EMW on male reproductive system, and 3) finally, exposure assessment and dosimetry by computational biomodeling.

Background

The health effects of radiofrequency electromagnetic waves (RF-EMW) emitted from cell phones have been debated greatly [1,2]. Researchers initially were concerned about how microwave radiation affected human biological systems by increasing tissue temperature--in other words, its thermal effects [3]. To protect the public from excessive exposure to RF-EMW, limits were established by international organizations such as the ICNIRP (International Commission on Non Ionizing Radiation Protection) [4]. For example, the limit of radiation exposure from a mobile phone in the United States and Europe is 1.6 Watts/kg and 2.0 Watts/kg, respectively [1,4].

Recent studies demonstrated microwaves emitted from the cell phone, i.e., RF-EMW do not produce thermal effect at specific absorption rate (SAR) of 1.6 Watts/Kg [5-7]. However, researchers have demonstrated that RF-EMW from commercially available cell phones have non-thermal effects [8,9]. The literature contains controversial reports on the effects of RF-EMW on, mitochondria, apoptosis pathway, heat shock proteins, free radical metabolism, cell differentiation, DNA damage and the plasma membrane [1,9-16].

Among the effect of RF-EMW on various body organs, effect of RF-EMW on brain is the most researched area [17-24]. Additionally, recent studies suggest that RF-EMW emitted from cell phones can reduce the fertilizing poten-

tial of men [25]. It is important to note that many men carry their cell phones in a trouser pocket (or clipped to their belts on waist) while using a hands-free device such as Bluetooth. This technology exposes the testes to more high power density cell phone radiation than a cell phone would in the 'Stand by mode' in a trouser pocket. Due to this reason, investigating effect of RF-EMW on male fertility is also important.

In this article, we review the effects of RF-EMW on free radical metabolism and carcinogenesis as well as the epidemiological, *in vitro* animal and *in vitro* human studies that have assessed the effect of RF-EMW on male fertility. We also briefly discuss the novel computational biomodeling for *in vitro* study on human semen currently being performed at our center.

Discussion

Biological effects of microwave radiation emitted from cellular phones: Is the plasma membrane a target of RF-EMW?

The literature contains controversial reports on the effects of RF-EMW on various cellular organelles. Of particular note, researchers have consistently demonstrated that RF-EMW affects the plasma membrane in cells [8,11,12,26]. In 1996, Cleary *et al* suggested that RF-EMW may affect the plasma membrane signal transduction, independent of thermal effect [12]. Capri *et al* later showed that exposure to 900 MHz RF-EMW was associated with a significant increase in annexin V-positive human lymphocytes (without any changes in apoptotic cells) and in the mitochondrial membrane potential [11]. They suggested that these findings might be due to the effect of RF fields on transient phosphatidylserine flip-flop movement. Another study using annexin V as a marker of apoptosis also found a significant increase in annexin V staining after exposure to RF-EMW [26]. However, studies using other markers of apoptosis (i.e., caspase, mitochondrial membrane potential) failed to show an increase in apoptosis [27-34]. This suggests that RF-EMW might stimulate apoptosis by acting on the plasma membrane (Annexin receptor). However, cells might not go under apoptosis depending upon their DNA repair capability (no change in caspase, mitochondrial membrane potential).

Recently, Friedman *et al*. reported that RF-EMW stimulates plasma membrane NADH oxidase (of HeLa cells) and causes production of extracellular superoxide [8]. This effect of RF-EMW on NADH oxidase can lead to oxidative stress (OS) and subsequent carcinogenesis (the molecular bases are discussed later in this paper).

Rao *et al* recently provided new evidence supporting the theory that RF-EMW affects the plasma membrane [35]. They studied the effects of RF-EMW on calcium dynamics

in stem cell-derived neuronal cells and discovered a significant increase in intracellular calcium spikes in response to non-thermal RF-EMW.

These studies suggest that the plasma membrane might be the target of RF-EMW and that other observed effects of RF-EMW might be secondary to the effect on the plasma membrane.

Cell phone radiation and oxidative stress

Free radicals are a group of highly reactive molecules consisting of unpaired electrons in the outer orbit. Free radicals that are derived from oxygen metabolism are known as reactive oxygen species (ROS) [36]. ROS are continuously neutralized by antioxidants present in body tissues [36]. Whenever production of ROS exceeds the scavenging capacity of antioxidants, it leads to OS [36]. In 1992, researchers found that electromagnetic fields increase free radical activity in cells [37]. Within the last decade, *in vivo* animal studies have shown that OS develops in response to cell phone radiation [38-42]. RF-EMW might disturb ROS metabolism by increasing production of ROS or by decreasing antioxidant enzyme activity. Studies have also demonstrated that antioxidants such as melatonin, caffeic acid phenyl ester, vitamin C and vitamin E prevent oxidative stress or apoptosis caused by RF-EMW in animal tissues [40-42]. Chronic exposure to RF-EMW decreases the activity of catalase, superoxide dismutase (SOD) and glutathione peroxidase (GSH-Px), and thus decreases the total antioxidant capacity. However, studies designed to measure malonaldehyde (MDA) levels and SOD activity have shown conflicting results [38,41,43-45]. Recent studies on human semen also suggested increased ROS production in human semen due to cell phone radiation [46,47].

We suggest that RF-EMW induces NADH oxidase enzyme stimulation [8], which might play a key role in the various cellular adverse effects observed in *in vitro* studies. As a consequence of increased levels of free radicals, various cellular and physiological processes can be affected including gene expression, release of calcium from intracellular storage sites, cell growth, and apoptosis.

DNA strand break and apoptosis

The effects of RF-EMW on DNA damage have been reported in various studies in the last decade [13,48-51]. Lai and Singh reported an increase in single and double-strand DNA breaks in the brain cells of rats that were exposed for 2 hrs to a 2450 MHz field at 0.6-1.2 W/kg. They also found that EMW exposure caused DNA-protein and DNA-DNA crosslinks and increased apoptosis in biological samples from rats [13,52-54].

Lai and Singh demonstrated that treating rats with free radical scavengers blocked the effects of EMW on DNA [55]. This suggests that EMW enhance free radical activity in cells, which in turn leads to DNA damage (single or double stranded DNA breaks). More recently, Paulraj and Behari (2006) reported an increase in single strand DNA breaks in the developing brain cells of rats that were exposed for 35 days to 2.45 and 16.5 GHz fields at 1 and 2.01 W/kg [56]. Nikolova et al. (2005) reported a low and transient increase in DNA double strand breaks in mouse embryonic stem cells after acute exposure to a 1.7-GHz field [57].

However, the results of more recent studies are controversial regarding the effects of RF-EMW on DNA [29,51,58-61]. Tice et al [60] reported that exposing human blood leukocytes and lymphocytes to RF-EMW at a SAR of 5-10 W/kg for 24 hours induced chromosomal damage while exposure to 3 hours of RF-EMW at a SAR of 5-10 W/kg or exposure to a lower SAR did not. Chromosomal damage was evident as the number of micronucleated cells among lymphocytes increased. Later on, Ramondini *et al* reported that human endothelial cells showed changes in the expression of several genes after exposure to 900 MHz but not 1800 MHz [62]. Schwarz *et al* reported that 1950 MHz RF-EMW exposure for 24 hour at a SAR of 0.05 W/kg induced genotoxic effects *in vitro* in human fibroblasts but not in lymphocytes [63]. In a study on human fibroblasts and rat granulosa cells exposed to mobile phone signals (1800 MHz; SAR 1.2 or 2 W/kg; during 4, 16 and 24 h), Diem et al reported single and double stranded DNA breaks in these cells [59]. On the other hand, many studies also demonstrated no significant effect of RF-EMW on DNA damage. Hook et al failed to find any significant effect of RF-EMW on Molt-4 T lymphoblastoid cells at various SAR [29]. Recent studies by Huang et al and Sannino et al on human fibroblast and Jurkat T-cells, respectively failed to show any significant genotoxic effects of RF-EMW [64,65]. Thus, DNA damage might depend upon the cell type as well as the experimental set up (duration of exposure, frequency of RF-EMW, specific absorbance rate, etc). Recent review by Verschaeve suggested increased frequency of genetic damage due to RF-EMW in many studies, however, RF dosimetry approach was lacking in all studies [66].

DNA damage in cells may have an important implication as it is cumulative. Normally, DNA is capable of repairing itself. Through a homeostatic mechanism, cells maintain a delicate balance between DNA damage and repair. DNA damage accumulates if this balance is altered. Most cells can repair single-strand DNA breaks. However, DNA double strand breaks, if not properly repaired, are known to lead to cell death or apoptosis.

It has been suggested that RF-EMW might affect the apoptosis pathway. However, the effect of RF-EMW on apoptosis is also controversial [5,28,41,67]. As we discussed before, studies suggest that RF-EMW might act on the plasma membrane annexin receptors [11,26]. However, induction of apoptosis might depend upon the cell type as well as the type and duration of RF-EMW exposure.

Effect on male reproductive system

A number of recent reports have suggested a possible link between cell phone use and male infertility [25,46,68,69].

An initial study from our group involving 361 men who had attended an infertility clinic suggested that the use of cell phones adversely affects semen quality by decreasing sperm count, motility, viability and morphology, which might contribute to male infertility [25]. Similarly, Fejes *et al.* studied 371 men undergoing infertility evaluations and reported that the duration of possession and the daily transmission time of cell phones correlated negatively with the proportion of rapidly progressive motile spermatozoa, suggesting that prolonged use of cell phones might have negative effects on sperm motility [70]. Davoudi *et al.* (2002), in a small prospective study involving 13 men with normal semen analysis, also found that using cell phones for 6 h a day for 5 days decreased the rapid progressive motility of spermatozoa [71]. Thus, recent epidemiological studies have highlighted the role of cell phone exposure on sperm motility, morphology and viability, thus proposing a reduction in the fertilizing potential of males. However, the impact of these studies is low due to a lack of a control population (men who do not use cell phones), which would be extremely difficult to create. Additionally, an *in vivo* human exposure study to investigate the effects of cell phone radiation on semen parameters is not feasible due to ethical issues.

In addition to the epidemiological studies, the effects of RF-EMW are well studied in animal studies and "in vitro" studies on human semen. Many studies have indicated that EMW decreases the size of the testicular organs. A decrease in the diameter of the seminiferous tubules [43,44] has been reported after exposure to radiofrequency radiations. Ozguner *et al.* demonstrated a decrease in seminiferous tubular diameter and epithelium thickness after applying RF-EMW of 869 to 894 MHz [72]. These results support the study by Saunders and Kowalczuk that also showed that microwave radiation of 50 mW/cm² at a frequency of 2.45 GHz for 30-40 minutes resulted in significant degeneration of the seminiferous epithelium in mice [73]. However, a recent study by Ribeiro et al. (2007) and follow up study by Dasdag et al could not find any significant adverse effect of cellular phones (1835-1850 MHz) on the rat testis [28,45]. Wang et al suggested that RF-EMW might change the permeability of

the blood-testis barrier [74]. RF-EMW-mediated ROS formation can lead to heat shock protein (hsp) and phosphorylation, which can alter the secretion of growth factors. This, in turn, can increase the permeability of the blood-brain barrier [8,9]. The same mechanism might be involved in the RF-EMW mediated increase in the blood-testis barrier as suggested in Figure 1. However, the rat model is not a good mock-up for studies on reproductive system because of its small size and ability of its testes to migrate freely between the abdomen and scrotum [75].

Previously, we discussed that RF-EMW can lead to OS. Human spermatozoa are highly susceptible to OS due to high contents of poly unsaturated fatty acids (PUFA) in their cell membrane [36]. RF-EMW mediated OS in semen might be responsible for decline in motility and viability of spermatozoa, as we had demonstrated in our pilot study on ejaculated human semen [46]. Eroglu et al had also reported decreased sperm parameters in semen samples exposed to RF-EMW [76]. In support of these studies, De Iuliis et al reported increase in ROS formation and higher DNA damage due to RF-EMW [47]. Previously, Aitken et al. (2005) reported significant damage to the mitochondrial and nuclear genome in epididymal spermatozoa of mice exposed to RF-EMW (900 MHz) for 12 hrs a day for 7 days [58]. However, our pilot study failed to show significant DNA damage (by TUNEL assay) after one hour of cell phone exposure [46]. Therefore, we suggest that RF-EMW may stimulate extracellular superoxide production in semen by its action on plasma membrane enzyme (Figure 1), which can lead to decrease in sperm motility and viability. However, DNA damage might depend on SAR, duration of exposure and experimental set up. It is important to note that none of the above mentioned studies on reproductive system used RF dosimetry in their experiment.

Effect on protein kinase C and calcium homeostasis

PKC is a multifunctional protein kinase and is structurally a group of proteins with at least 10 isoenzymes identified thus far [77]. PKC isoenzymes have been reported to differ in their structure, biochemical properties, tissue distribution, subcellular localization, and substrate specificity. PKC has been involved in the regulation of a variety of cellular events including modulation of receptor functions for major hormones and certain enzymes such as adenylate cyclase and ornithine decarboxylase (in the brain). It also plays key role in mediating cellular responses to extracellular stimuli involved in proliferation, differentiation, apoptosis, and exocytotic release in a number of non-neuronal and sperm cells [78-81]. The PKC enzyme complex is present in rat seminiferous tubules and leydig cells [81]. PKC modulates ion conductance by phosphorylating membrane proteins such as channels, pumps, and ion exchange proteins. The activation of this enzyme is

thought to be biochemically dependent on Ca^{2+} [82,83]. In fact, Calcium and cAMP are thought to be the two pivotal regulators of sperm flagellar motility. PKC might also play a crucial role in regulating sperm motility together with cAMP and Ca^{2+} [78]

PKC is highly concentrated in the equatorial segment, suggesting a specific compartmentalized role for PKC in human sperm physiology. Studies have reported declines in sperm motility with decreases in PKC activity [79,84]. Recently, Paulraj et al reported decreases in the activity of PKC in developing rat brains that were exposed to 2.45 GHz of microwaves [56]. The authors reported that the microwave damage was more extensive in the hippocampus, which may cause memory loss. Thus, the PKC enzyme is important in sperm motility, and RF-EMW have been reported to decrease PKC activity. We suggest that RF-EMW can cause declines in sperm motility by affecting PKC. Mobile phone-altered PKC activity has been linked with various types of malignancies [85,86]. Different levels of PKC and differential activation of various PKC isozymes have resulted in testicular and brain tumor cells, which is discussed in detail in the next section.

RF-EMW may also alter intracellular calcium homeostasis by acting on plasma membrane calcium channels [87]. Rao et al recently suggested RF-EMW can increase intracellular calcium spikes. Increases in intracellular calcium levels can trigger cell proliferation and differentiation as well modify activities of various enzymes such as ODC [88] and PKC [78].

Does mobile phone exposure promote carcinogenesis?

Previous reports have suggested that electromagnetic fields alter the proliferation rate of cells as well as the rate of DNA, RNA, and protein synthesis [89,90]. However, evidence from epidemiological studies on the effects of RF-EMW on carcinogenesis are controversial [91,92].

We suggest the possible pathways of carcinogenesis by RF-EMW.

As we have discussed, the plasma membrane may be a target of RF-EMW. RF-EMW might trigger uncontrolled cell proliferation by its action on various plasma membrane enzymes and receptors. Short-term exposure to RF-EMW can lead to increase in the activity of plasma membrane NADH oxidase enzyme, which increases ROS formation [8]. The ROS then activates MMPs (matrix metalloproteinases), which would release epidermal growth factor and activate extracellular signal regulated kinases (ERKs). Chronic exposure to RF-EMW induces stress kinases, which activate p38 MAP (mitogen activated protein) kinase. P38 MAP kinase stimulates phosphorylation of heat shock proteins, which inhibit the apoptosis pathway

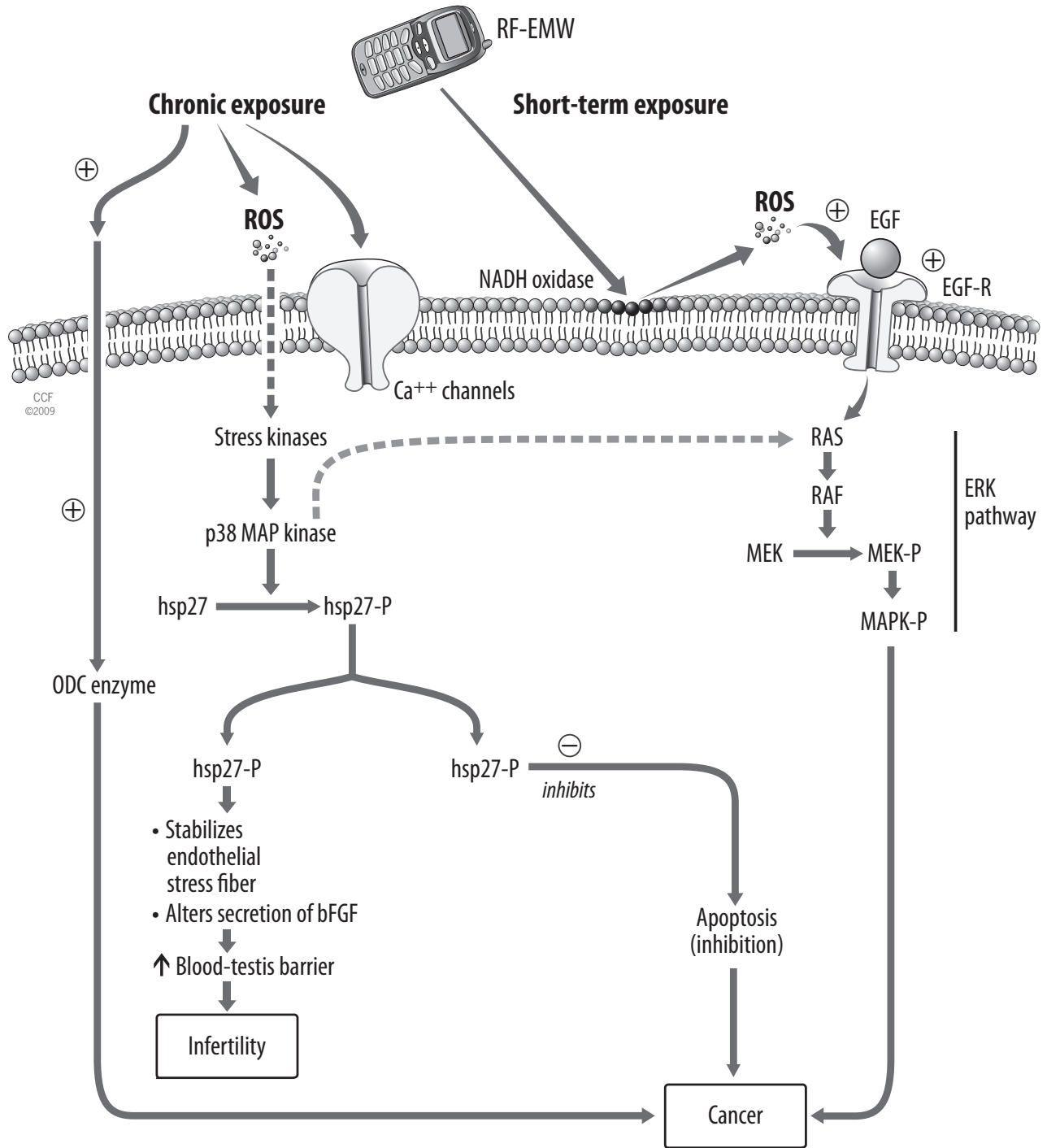


Figure 1 (see legend on next page)

Figure 1 (see previous page)

The figure shows various cellular targets of radiofrequency electromagnetic waves (RF-EMW). Acute (short term) exposure to RF-EMW can stimulate plasma membrane NADH oxidase which can increase reactive oxygen species (ROS) formation. Increase in ROS can stimulate endothelial growth factor (EGF) receptor which in turn activates extra cellular signal regulated kinase (ERK) pathway. ERK pathway consist of subsequent activation of Ras, Raf proteins and mitogen-activated protein kinase (MAPK). MAPK pathway has tumor promoting role. Chronic exposure to ROS can activate various stress kinase (p38 MAP kinase). Activation of p38 MAP kinase can stimulate ERK pathway and also lead to phosphorylation of heat shock proteins (Hsp) which inhibits apoptosis. Inhibition of apoptosis might promote carcinogenesis by prolonging survival of cell with damaged DNA. Hsp also stabilizes endothelial stress fiber and alters secretion of bFGF. This can lead to increase in permeability of blood testis barrier and causes infertility. RF-EMW can also promote cancer by stimulating ornithine decarboxylase (ODC), a rate limiting enzyme in polyamine synthesis pathway as well as by interfering with plasma membrane calcium channels.

[9]. Thus, cell phone radiation can cause DNA damage to accumulate in the cell and trigger uncontrolled cell proliferation.

Moreover, various reports have suggested that ODC may be a target for microwave radiations [93-95]. ODC is the rate limiting enzyme that participates in polyamine synthesis; compounds required for cell division. Over expression of ODC is linked to progression of cancer [96]. ODC activity is modulated by membrane-mediated signals, and RF-EMW might stimulate ODC directly or through its action on the membrane [95].

Although RF-EMW exposure is known to affect PKC isoenzyme expression, the literature contains controversial reports regarding the role of change in PKC activities in carcinogenesis [85,97]. Phorbol ester is a widely known tumor-promoting agent, and acute exposure to phorbol ester stimulates PKC. However, chronic exposure to phorbol ester decreases the activity of PKC delta. Lu et al and Hornia et al found that depletion of PKC delta was associated with tumor promotion [85,86]. As we discussed above, RF-EMW might be associated with a decline in PKC activity. Thus, we suggest that chronic RF-EMW exposure leading to declines in PKC activity might be associated with carcinogenesis.

Changes in intracellular calcium levels and activities of ODC and PKC are interrelated as well as can be secondary to RF-EMW exposure. The literature also contains a report of ROS-mediated changes in the activities of PKC and ODC [98]. Thus, RF-EMW mediated increases in ROS production can trigger cell differentiation by its action on MAPK kinase, hsp, PKC and ODC.

Future studies: A computational approach

Carrying a cell phone in a pant pocket (or cell phone clipped to a belt on the waist) exposes the testes to high-power density mobile radiation. Similarly, this occurs when a person is talking on a Bluetooth headset (or any other hands-free accessories) with the cell phone in the

pant pocket. Recent use of hands-free accessories might decrease radiation exposure to the human head [99] but it might also put the male reproductive system at risk. In real life, the device and the male reproductive organs are separated by multiple tissue layers. Therefore, designing experimental conditions to simulate life-like radiofrequency dosimetry is very important. Thus, to determine the specific distance that is nearly equal to the distance of cell phone (in trouser pocket) and testes, we designed a two-dimensional anatomical-computational model of the tissue to extrapolate the effects seen in "in vitro" condition to real-life conditions (this is a software model made on a computer-linux based system; not any physical-anatomical model). RF dosimetry (radiation dosimetry is the calculation of the absorbed dose in matter and tissue resulting from the exposure to radiation) analysis using finite difference time domain (FDTD) will be performed. We will also design a model of the experimental set up that will calculate the distance between the RF source and the semen sample. This set up will best represent SAR conditions *in vivo*.

As per WHO research agenda for radiofrequency fields, in vitro studies are important for the health risk assessment due to RF-EMW [1]. We have designed this study based on our recently published pilot study [46] and biomodeling study by Bit-Babik et al [99]. We will also examine the effects of specific distances on sperm parameters and ROS formation (in neat semen samples).

Conclusion

We have reviewed the literature to better understand the effects of cell phone radiation on human health, especially on fertility and in relation to cancer. Commercially available cellular phones might affect cell function via non-thermal effects. We hypothesized that the plasma membrane might be the target of cell phone radiation. RF-EMW can increase ROS formation by increasing the activity of plasma membrane NADH oxidase. Prolonged exposure to RF-EMW can also cause DNA damage (by prolonged OS), which may accelerates neuronal and sper-

matozoal cell death and promote neurodegenerative processes as well as promote brain and testicular carcinogenesis. Any tumor promoting effects of RF-EMW might be due to the effect it has on PKC, ODC, intra cellular calcium spikes and stimulation of stress kinase. Stimulation of plasma membrane NADH oxidase might play central role in above mentioned effects.

OS and changes in PKC activity might lead to the RF-EMW related infertility observed in numerous studies. Hence, RF-EMW from commercially available cell phones might affect the fertilizing potential of spermatozoa. Therefore, the SAR limit (maximum acceptable exposure limit) should be lowered for cellular phones. However, more studies are necessary to provide definitive evidence against cell phone radiation, which can be provided by in vitro studies combined with computational biomodeling.

List of abbreviations

RF-EMW: Radiofrequency electromagnetic waves; SAR: Specific absorption rate; FDTD: Finite difference time domain; MDA: Malonaldehyde; Hsp: Heat shock protein; PKC: Protein Kinase C; SOD: Superoxide dismutase; GSH-Px: Glutathione peroxidase; CAT: catalase; OS: Oxidative stress; MAPK: Mitogen activated protein kinase; ERK: Extracellular signal regulated kinases; ODC: Ornithine decarboxylase.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

NRD has substantial contribution in designing the article and drafting 50-60% article. KK has substantial contribution in conception and designing article and drafting 20-30% article. AA has critical contribution in drafting article, revision and final approval. All the authors read and approved the final manuscript.

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References

- World Health Organization (WHO): **2006 WHO research agenda for radio frequency fields.** 2006.
- Makker K, Varghese A, Desai NR, Mouradi R, Agarwal A: **Cell phones: modern man's nemesis?** *Reprod Biomed Online* 2009, **18(1)**:148-157.
- Oftedal G, Wilen J, Sandstrom M, Mild KH: **Symptoms experienced in connection with mobile phone use.** *Occup Med (Lond)* 2000, **50(4)**:237-245.
- International Commission on Non-Ionizing Radiation Protection (ICNIRP): **Guidelines for limiting exposure to time-varying electric, magnetic, and electromagnetic fields (up to 300 GHz).** *International Commission on Non-Ionizing Radiation Protection.* *Health Phys* 1998, **74(4)**:494-522.
- Anderson V, Rowley J: **Measurements of skin surface temperature during mobile phone use.** *Bioelectromagnetics* 2007, **28(2)**:159-162.
- Straume A, Oftedal G, Johnsson A: **Skin temperature increase caused by a mobile phone: a methodological infrared camera study.** *Bioelectromagnetics* 2005, **26(6)**:510-519.
- Yan JG, Agresti M, Bruce T, Yan YH, Granlund A, Matloub HS: **Effects of cellular phone emissions on sperm motility in rats.** *Fertil Steril* 2007, **88(4)**:957-964.
- Friedman J, Kraus S, Hauptman Y, Schiff Y, Seger R: **Mechanism of short-term ERK activation by electromagnetic fields at mobile phone frequencies.** *Biochem J* 2007, **405(3)**:559-568.
- Leszczynski D, Joenvaara S, Reivinen J, Kuokka R: **Non-thermal activation of the hsp27/p38MAPK stress pathway by mobile phone radiation in human endothelial cells: molecular mechanism for cancer- and blood-brain barrier-related effects.** *Differentiation* 2002, **70(2-3)**:120-129.
- Blank M: **Do electromagnetic fields interact with electrons in the Na, K-ATPase?** *Bioelectromagnetics* 2005, **26(8)**:677-683.
- Capri M, Scarcella E, Fumelli C, Bianchi E, Salvioli S, Mesirca P, Agostini C, Antolini A, Schiavoni A, Castellani G, Bersani F, Franceschi C: **In vitro exposure of human lymphocytes to 900 MHz CW and GSM modulated radiofrequency: studies of proliferation, apoptosis and mitochondrial membrane potential.** *Radiat Res* 2004, **162(2)**:211-218.
- Cleary SF, Du Z, Cao G, Liu LM, McCrady C: **Effect of isothermal radiofrequency radiation on cytolytic T lymphocytes.** *FASEB J* 1996, **10(8)**:913-919.
- Lai H, Singh NP: **Single- and double-strand DNA breaks in rat brain cells after acute exposure to radiofrequency electromagnetic radiation.** *Int J Radiat Biol* 1996, **69(4)**:513-521.
- Lantow M, Lupke M, Frahm J, Mattsson MO, Kuster N, Simko M: **ROS release and Hsp70 expression after exposure to 1,800 MHz radiofrequency electromagnetic fields in primary human monocytes and lymphocytes.** *Radiat Environ Biophys* 2006, **45(1)**:55-62.
- Lixia S, Yao K, Kaijun W, Deqiang L, Huajun H, Xiangwei G, Baohong W, Wei Z, Jianling L, Wei W: **Effects of 1.8 GHz radiofrequency field on DNA damage and expression of heat shock protein 70 in human lens epithelial cells.** *Mutat Res* 2006, **602(1-2)**:135-142.
- McNamee JP, Bellier PV, Gajda GB, Lavallee BF, Marro L, Lemay E, Thansandote A: **No evidence for genotoxic effects from 24 h exposure of human leukocytes to 1.9 GHz radiofrequency fields.** *Radiat Res* 2003, **159(5)**:693-697.
- Bondy ML, Scheurer ME, Malmer B, Barnholtz-Sloan JS, Davis FG, Il'yasova D, Kruchko C, McCarthy BJ, Rajaraman P, Schwartzbaum JA, Sadetzki S, Schlehofer B, Tihan T, Wiemels JL, Wrensch M, Buffler PA: **Brain tumor epidemiology: consensus from the Brain Tumor Epidemiology Consortium.** *Cancer* 2008, **113(7 Suppl)**:1953-1968.
- Cardis E, Deltour I, Mann S, Moissonnier M, Taki M, Varsier N, Wake K, Wiart J: **Distribution of RF energy emitted by mobile phones in anatomical structures of the brain.** *Phys Med Biol* 2008, **53(11)**:2771-2783.
- Eberhardt JL, Persson BR, Brun AE, Salford LG, Malmgren LO: **Blood-brain barrier permeability and nerve cell damage in rat brain 14 and 28 days after exposure to microwaves from GSM mobile phones.** *Electromagn Biol Med* 2008, **27(3)**:215-229.
- Hardell L, Sage C: **Biological effects from electromagnetic field exposure and public exposure standards.** *Biomed Pharmacother* 2008, **62(2)**:104-109.
- O'Keefe S: **Does the use of cell phones cause brain tumors?** *Clin J Oncol Nurs* 2008, **12(4)**:671-672.
- Pawl R: **Cell phones more dangerous than cigarettes!** *Surg Neurol* 2008, **70(5)**:445-446.
- Sokolovic D, Djindjic B, Nikolic J, Bjelakovic G, Pavlovic D, Kocic G, Krstic D, Cvetkovic T, Pavlovic V: **Melatonin reduces oxidative stress induced by chronic exposure of microwave radiation from mobile phones in rat brain.** *J Radiat Res (Tokyo)* 2008, **49(6)**:579-586.
- Wiholm C, Lowden A, Kuster N, Hillert L, Arnetz BB, Akerstedt T, Moffat SD: **Mobile phone exposure and spatial memory.** *Bioelectromagnetics* 2009, **30(1)**:59-65.
- Agarwal A, Deepinder F, Sharma RK, Ranga G, Li J: **Effect of cell phone usage on semen analysis in men attending infertility clinic: an observational study.** *Fertil Steril* 2008, **89(1)**:124-128.
- Markkanen A, Penttinen P, Naarala J, Pelkonen J, Sihvonen AP, Juutilainen J: **Apoptosis induced by ultraviolet radiation is**

- enhanced by amplitude modulated radiofrequency radiation in mutant yeast cells. *Bioelectromagnetics* 2004, **25**(2):127-133.
27. Buttiglione M, Roca L, Montemurno E, Vitiello F, Capozzi V, Cibelli G: **Radiofrequency radiation (900 MHz) induces Egr-1 gene expression and affects cell-cycle control in human neuroblastoma cells.** *J Cell Physiol* 2007, **213**(3):759-767.
 28. Dasdag S, Akdag MZ, Ulukaya E, Uzunlar AK, Yegin D: **Mobile phone exposure does not induce apoptosis on spermatogenesis in rats.** *Arch Med Res* 2008, **39**(1):40-44.
 29. Hook GJ, Zhang P, Lagroye I, Li L, Higashikubo R, Moros EG, Straube WL, Pickard WF, Baty JD, Roti Roti JL: **Measurement of DNA damage and apoptosis in Molt-4 cells after in vitro exposure to radiofrequency radiation.** *Radiat Res* 2004, **161**(2):193-200.
 30. Joubert V, Leveque P, Cueille M, Bourthoumieu S, Yardin C: **No apoptosis is induced in rat cortical neurons exposed to GSM phone fields.** *Bioelectromagnetics* 2007, **28**(2):115-121.
 31. Joubert V, Leveque P, Rametti A, Collin A, Bourthoumieu S, Yardin C: **Microwave exposure of neuronal cells in vitro: Study of apoptosis.** *Int J Radiat Biol* 2006, **82**(4):267-275.
 32. Lantow M, Viergutz T, Weiss DG, Simko M: **Comparative study of cell cycle kinetics and induction of apoptosis or necrosis after exposure of human mono mac 6 cells to radiofrequency radiation.** *Radiat Res* 2006, **166**(3):539-543.
 33. Merola P, Marino C, Lovisolo GA, Pinto R, Laconi C, Negroni A: **Proliferation and apoptosis in a neuroblastoma cell line exposed to 900 MHz modulated radiofrequency field.** *Bioelectromagnetics* 2006, **27**(3):164-171.
 34. Falzone N, Huyser C, Fourie F, Toivo T, Leszczynski D, Franken D: **In vitro effect of pulsed 900 MHz GSM radiation on mitochondrial membrane potential and motility of human spermatozoa.** *Bioelectromagnetics* 2008, **29**(4):268-276.
 35. Rao VS, Titushkin IA, Moros EG, Pickard WF, Thatte HS, Cho MR: **Nonthermal effects of radiofrequency-field exposure on calcium dynamics in stem cell-derived neuronal cells: elucidation of calcium pathways.** *Radiat Res* 2008, **169**(3):319-329.
 36. Desai N, Sharma R, Makker K, Sabanegh E, Agarwal A: **Physiologic and pathologic levels of reactive oxygen species in neat semen of infertile men.** *Fertil Steril* 2008 in press.
 37. Grundler W, Kaiser F, Keilmann F, Walleczek J: **Mechanisms of electromagnetic interaction with cellular systems.** *Naturwissenschaften* 1992, **79**(12):551-559.
 38. Balci M, Devrim E, Durak I: **Effects of mobile phones on oxidant/antioxidant balance in cornea and lens of rats.** *Curr Eye Res* 2007, **32**(1):21-25.
 39. Meral I, Mert H, Mert N, Deger Y, Yoruk I, Yetkin A, Keskin S: **Effects of 900-MHz electromagnetic field emitted from cellular phone on brain oxidative stress and some vitamin levels of guinea pigs.** *Brain Res* 2007, **1169**:120-124.
 40. Oktem F, Ozguner F, Mollaoglu H, Koyu A, Uz E: **Oxidative damage in the kidney induced by 900-MHz-emitted mobile phone: protection by melatonin.** *Arch Med Res* 2005, **36**(4):350-355.
 41. Oral B, Guney M, Ozguner F, Karahan N, Mungan T, Comlekci S, Cesur G: **Endometrial apoptosis induced by a 900-MHz mobile phone: preventive effects of vitamins E and C.** *Adv Ther* 2006, **23**(6):957-973.
 42. Ozguner F, Bardak Y, Comlekci S: **Protective effects of melatonin and caffeic acid phenethyl ester against retinal oxidative stress in long-term use of mobile phone: a comparative study.** *Mol Cell Biochem* 2006, **282**(1-2):83-88.
 43. Dasdag S, Ketani MA, Akdag Z, Ersay AR, Sari I, Demirtas OC, Celik MS: **Whole-body microwave exposure emitted by cellular phones and testicular function of rats.** *Urol Res* 1999, **27**(3):219-223.
 44. Dasdag S, Zulkuf Akdag M, Aksen F, Yilmaz F, Bashan M, Mutlu Dasdag M, Salih Celik M: **Whole body exposure of rats to microwaves emitted from a cell phone does not affect the testes.** *Bioelectromagnetics* 2003, **24**(3):182-188.
 45. Ribeiro EP, Rhoden EL, Horn MM, Rhoden C, Lima LP, Toniolo L: **Effects of subchronic exposure to radio frequency from a conventional cellular telephone on testicular function in adult rats.** *J Urol* 2007, **177**(1):395-399.
 46. Agarwal A, Desai NR, Makker K, Varghese A, Mouradi R, Sabanegh E, Sharma R: **Effects of radiofrequency electromagnetic waves (RF-EMW) from cellular phones on human ejaculated semen: an in vitro pilot study.** *Fertil Steril* 2008, **92**(4):1318-25. Epub 2008 Sep 20.
 47. De Iulius GN, Newey RJ, King BV, Aitken RJ: **Mobile phone radiation induces reactive oxygen species production and DNA damage in human spermatozoa in vitro.** *PLoS One* 2009, **4**(7):e6446.
 48. Garaj-Vrhovac V, Horvat D, Koren Z: **The effect of microwave radiation on the cell genome.** *Mutat Res* 1990, **243**(2):87-93.
 49. Maes A, Verschaev L, Arroyo A, De Wagter C, Verduyssen L: **In vitro cytogenetic effects of 2450 MHz waves on human peripheral blood lymphocytes.** *Bioelectromagnetics* 1993, **14**(6):495-501.
 50. Sarkar S, Ali S, Behari J: **Effect of low power microwave on the mouse genome: a direct DNA analysis.** *Mutat Res* 1994, **320**(1-2):141-147.
 51. Belyaev IY, Koch CB, Terenius O, Roxstrom-Lindquist K, Malmgren LO, W HS, Salford LG, Persson BR: **Exposure of rat brain to 915 MHz GSM microwaves induces changes in gene expression but not double stranded DNA breaks or effects on chromatin conformation.** *Bioelectromagnetics* 2006, **27**(4):295-306.
 52. Lai H, Singh NP: **Acute low-intensity microwave exposure increases DNA single-strand breaks in rat brain cells.** *Bioelectromagnetics* 1995, **16**(3):207-210.
 53. Lai H, Singh NP: **Acute exposure to a 60 Hz magnetic field increases DNA strand breaks in rat brain cells.** *Bioelectromagnetics* 1997, **18**(2):156-165.
 54. Lai H, Singh NP: **Magnetic-field-induced DNA strand breaks in brain cells of the rat.** *Environ Health Perspect* 2004, **112**(6):687-694.
 55. Lai H, Singh NP: **Melatonin and N-tert-butyl-alpha-phenyl-nitrozone block 60-Hz magnetic field-induced DNA single and double strand breaks in rat brain cells.** *J Pineal Res* 1997, **22**(3):152-162.
 56. Paulraj R, Behari J: **Single strand DNA breaks in rat brain cells exposed to microwave radiation.** *Mutat Res* 2006, **596**(1-2):76-80.
 57. Nikolova T, Czyn J, Rolletschek A, Blyszczuk P, Fuchs J, Jovtchev G, Schuderer J, Kuster N, Wobus AM: **Electromagnetic fields affect transcript levels of apoptosis-related genes in embryonic stem cell-derived neural progenitor cells.** *FASEB J* 2005, **19**(12):1686-1688.
 58. Aitken RJ, Bennetts LE, Sawyer D, Wiklendt AM, King BV: **Impact of radio frequency electromagnetic radiation on DNA integrity in the male germline.** *Int J Androl* 2005, **28**(3):171-179.
 59. Diem E, Schwarz C, Adlkofer F, Jahn O, Rudiger H: **Non-thermal DNA breakage by mobile-phone radiation (1800 MHz) in human fibroblasts and in transformed GFSH-R17 rat granulosa cells in vitro.** *Mutat Res* 2005, **583**(2):178-183.
 60. Tice RR, Hook GG, Donner M, McRee DI, Guy AVW: **Genotoxicity of radiofrequency signals. I. Investigation of DNA damage and micronuclei induction in cultured human blood cells.** *Bioelectromagnetics* 2002, **23**(2):113-126.
 61. Vijayalaxmi , Prihoda TJ: **Genetic damage in mammalian somatic cells exposed to radiofrequency radiation: a meta-analysis of data from 63 publications (1990-2005).** *Radiat Res* 2008, **169**(5):561-574.
 62. Remondini D, Nylund R, Reivinen J, Poullietier de Gannes F, Veyret B, Lagroye I, Haro E, Trillo MA, Capri M, Franceschi C, Schlatterer K, Gminski R, Fitzner R, Tauber R, Schuderer J, Kuster N, Leszczynski D, Bersani F, Maercker C: **Gene expression changes in human cells after exposure to mobile phone microwaves.** *Proteomics* 2006, **6**(17):4745-4754.
 63. Schwarz C, Kratochvil E, Pilger A, Kuster N, Adlkofer F, Rudiger HW: **Radiofrequency electromagnetic fields (UMTS, 1,950 MHz) induce genotoxic effects in vitro in human fibroblasts but not in lymphocytes.** *Int Arch Occup Environ Health* 2008, **81**(6):755-767.
 64. Sannino A, Di Costanzo G, Brescia F, Sarti M, Zeni O, Juutilainen J, Scarfi MR: **Human fibroblasts and 900 MHz radiofrequency radiation: evaluation of DNA damage after exposure and co-exposure to 3-chloro-4-(dichloromethyl)-5-hydroxy-2(5 h)-furanone (MX).** *Radiat Res* 2009, **171**(6):743-751.
 65. Huang TQ, Lee MS, Oh E, Zhang BT, Seo JS, Park WY: **Molecular responses of Jurkat T-cells to 1763 MHz radiofrequency radiation.** *Int J Radiat Biol* 2008, **84**(9):734-741.
 66. Verschaev L: **Genetic damage in subjects exposed to radiofrequency radiation.** *Mutat Res* 2009, **681**(2-3):259-270.

67. Zhao TY, Zou SP, Knapp PE: **Exposure to cell phone radiation up-regulates apoptosis genes in primary cultures of neurons and astrocytes.** *Neurosci Lett* 2007, **412(1)**:34-38.
68. Baste V, Riise T, Moen BE: **Radiofrequency electromagnetic fields; male infertility and sex ratio of offspring.** *Eur J Epidemiol* 2008, **23(5)**:369-77.
69. Wdowiak A, Wdowiak L, Wiktor H: **Evaluation of the effect of using mobile phones on male fertility.** *Ann Agric Environ Med* 2007, **14(1)**:169-172.
70. Fejes I, Zavaczki Z, Szollosi J, Koloszar S, Daru J, Kovacs L, Pal A: **Is there a relationship between cell phone use and semen quality?** *Arch Androl* 2005, **51(5)**:385-393.
71. Davoudi M, Brossner C, Kuber W: **The influence of electromagnetic waves on sperm motility.** *Urol Urogynacol* 2002, **19(22)**:.
72. Ozguner M, Koyu A, Cesur G, Ural M, Ozguner F, Gokcimen A, Delibas N: **Biological and morphological effects on the reproductive organ of rats after exposure to electromagnetic field.** *Saudi Med J* 2005, **26(3)**:405-410.
73. Saunders RD, Kowalczyk CI: **Effects of 2.45 GHz microwave radiation and heat on mouse spermatogenic epithelium.** *Int J Radiat Biol Relat Stud Phys Chem Med* 1981, **40(6)**:623-632.
74. Wang XW, Ding GR, Shi CH, Zhao T, Zhang J, Zeng LH, Guo GZ: **Effect of electromagnetic pulse exposure on permeability of blood-testicle barrier in mice.** *Biomed Environ Sci* 2008, **21(3)**:218-221.
75. Cairnie AB, Harding RK: **Cytological studies in mouse testis irradiated with 2.45-GHz continuous-wave microwaves.** *Radiat Res* 1981, **87(1)**:100-108.
76. Erogul O, Oztas E, Yildirim I, Kir T, Aydur E, Komesli G, Irkilata HC, Irmak MK, Peker AF: **Effects of electromagnetic radiation from a cellular phone on human sperm motility: an in vitro study.** *Arch Med Res* 2006, **37(7)**:840-843.
77. Larsson C: **New insights into PKC family affairs: three novel phosphorylation sites in PKCepsilon and at least one is regulated by PKCalpha.** *Biochem J* 2008, **411(2)**:e15-16.
78. Kimura K, Katoh N, Sakurada K, Kubo S: **Phospholipid-sensitive Ca²⁺-dependent protein kinase system in testis: localization and endogenous substrates.** *Endocrinology* 1984, **115(6)**:2391-2399.
79. Naor Z, Breitbart H: **Protein kinase C and Mammalian spermatozoa acrosome reaction.** *Trends Endocrinol Metab* 1997, **8(9)**:337-342.
80. Nishizuka Y: **Studies and perspectives of protein kinase C.** *Science* 1986, **233(4761)**:305-312.
81. Nikula H, Naor Z, Parvinen M, Huhtaniemi I: **Distribution and activation of protein kinase C in the rat testis tissue.** *Mol Cell Endocrinol* 1987, **49(1)**:39-49.
82. Crocenzi FA, Sanchez Pozzi EJ, Ruiz ML, Zucchetti AE, Roma MG, Mottino AD, Vore M: **Ca(2+)-dependent protein kinase C isoforms are critical to estradiol 17beta-D-glucuronide-induced cholestasis in the rat.** *Hepatology* 2008, **48(6)**:1885-1895.
83. Kubitz R, Saha N, Kuhlkamp T, Dutta S, vom Dahl S, Wettstein M, Haussinger D: **Ca²⁺-dependent protein kinase C isoforms induce cholestasis in rat liver.** *J Biol Chem* 2004, **279(11)**:10323-10330.
84. White D, de Lamirande E, Gagnon C: **Protein kinase C is an important signaling mediator associated with motility of intact sea urchin spermatozoa.** *J Exp Biol* 2007, **210(Pt 22)**:4053-4064.
85. Hornia A, Lu Z, Sukezane T, Zhong M, Joseph T, Frankel P, Foster DA: **Antagonistic effects of protein kinase C alpha and delta on both transformation and phospholipase D activity mediated by the epidermal growth factor receptor.** *Mol Cell Biol* 1999, **19(11)**:7672-7680.
86. Lu Z, Hornia A, Jiang YW, Zang Q, Ohno S, Foster DA: **Tumor promotion by depleting cells of protein kinase C delta.** *Mol Cell Biol* 1997, **17(6)**:3418-3428.
87. Blackman CF, Benane SG, Elder JA, House DE, Lampe JA, Faulk JM: **Induction of calcium-ion efflux from brain tissue by radiofrequency radiation: effect of sample number and modulation frequency on the power-density window.** *Bioelectromagnetics* 1980, **1(1)**:35-43.
88. Ishizuka J, Bold RJ, Townsend CM Jr, Thompson JC: **Role of calcium in the regulation of ornithine decarboxylase enzyme activity in mouse colon cancer cells.** *Cancer Invest* 1995, **13(2)**:181-187.
89. Fitzsimmons RJ, Strong DD, Mohan S, Baylink DJ: **Low-amplitude, low-frequency electric field-stimulated bone cell proliferation may in part be mediated by increased IGF-II release.** *J Cell Physiol* 1992, **150(1)**:84-89.
90. Goodman R, Henderson AS: **Exposure of salivary gland cells to low-frequency electromagnetic fields alters polypeptide synthesis.** *Proc Natl Acad Sci USA* 1988, **85(11)**:3928-3932.
91. Kundi M, Mild K, Hardell L, Mattsson MO: **Mobile telephones and cancer--a review of epidemiological evidence.** *J Toxicol Environ Health B Crit Rev* 2004, **7(5)**:351-384.
92. Lahkola A, Salminen T, Raitanen J, Heinavaara S, Schoemaker MJ, Christensen HC, Feychting M, Johansen C, Klæboe L, Lonn S, Swerdlow AJ, Tynes T, Auvinen A: **Meningioma and mobile phone use--a collaborative case-control study in five North European countries.** *Int J Epidemiol* 2008, **37(6)**:1304-1313.
93. Hoyto A, Juutilainen J, Naarala J: **Ornithine decarboxylase activity is affected in primary astrocytes but not in secondary cell lines exposed to 872 MHz RF radiation.** *Int J Radiat Biol* 2007, **83(6)**:367-374.
94. Behari J, Paulraj R: **Biomarkers of induced electromagnetic field and cancer.** *Indian J Exp Biol* 2007, **45(1)**:77-85.
95. Byus CV, Kartun K, Pieper S, Adey WR: **Increased ornithine decarboxylase activity in cultured cells exposed to low energy modulated microwave fields and phorbol ester tumor promoters.** *Cancer Res* 1988, **48(15)**:4222-4226.
96. Hogarty MD, Norris MD, Davis K, Liu X, Evageliou NF, Hayes CS, Pawel B, Guo R, Zhao H, Sekyere E, Keating J, Thomas W, Cheng NC, Murray J, Smith J, Sutton R, Venn N, London WB, Buxton A, Gilmour SK, Marshall GM, Haber M: **ODCI is a critical determinant of MYCN oncogenesis and a therapeutic target in neuroblastoma.** *Cancer Res* 2008, **68(23)**:9735-9745.
97. Griner EM, Kazanietz MG: **Protein kinase C and other diacylglycerol effectors in cancer.** *Nat Rev Cancer* 2007, **7(4)**:281-294.
98. Otieno MA, Kensler TW: **A role for protein kinase C-delta in the regulation of ornithine decarboxylase expression by oxidative stress.** *Cancer Res* 2000, **60(16)**:4391-4396.
99. Bit-Babik G, Chou CK, Faraone A, Gessner A, Kanda M, Balzano Q: **Estimation of the SAR in the human head and body due to radiofrequency radiation exposure from handheld mobile phones with hands-free accessories.** *Radiat Res* 2003, **159(4)**:550-557.

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