

NIH Public Access

Author Manuscript

Nat Prod Res. Author manuscript; available in PMC 2009 November 12.

Published in final edited form as:

Nat Prod Res. 2007 August; 21(10): 872-876. doi:10.1080/14786410600929576.

Ipomoeassin F, a new cytotoxic macrocyclic glycoresin from the leaves of *Ipomoea squamosa* from the Suriname rainforest[†]

SHUGENG CAO[‡], ANDREW NORRIS[‡], JAN H. WISSE[§], JAMES S. MILLER[¶], RANDY EVANS[¶], and DAVID G. I. KINGSTON^{*,‡}

[‡]Department of Chemistry, M/C 0212, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24061 USA

§Bedrijf Geneesmiddelen Voorziening Suriname, Commissaris Roblesweg 156, Geyersvlijt, Suriname

[¶]Missouri Botanical Garden, P.O. Box 299, St. Louis, Missouri 63166-0299, USA

Abstract

A new cytotoxic macrocyclic glycoresin, ipomoeassin F (6), has been isolated from the leaves of *Ipomoea squamosa*. The structure was elucidated by the interpretation of spectral data. Compound 6 was strongly active in the A2780 (human ovarian cancer cell line) assay with an IC₅₀ value of 0.036 μ M.

Keywords

ipomoeassin F; glycoresin; Ipomoea squamosa; Convolvulaceae; cytotoxicity

1. Introduction

We have previously reported the isolation of the five cytotoxic macrocyclic glycoresins ipomoeassins A-E (1–5) from the leaves of *Ipomoea squamosa* Choisy (Convolvulaceae) from the Suriname rainforest [2]. Subsequent to this work we re-examined a fraction from the final HPLC separation which had eluted just after ipomoeassin A (1). This fraction had originally been assumed to consist of ipomoeassin A, based on its ¹H NMR spectrum, which was very similar to that of 1, and its R_f value on TLC (silica-gel, CH₂Cl₂:CH₃OH; 40:1), which was identical to that of 1. Further chromatography of the fraction by HPLC (C8, 75% CH₃CN/ H₂O) indicated however that the fraction contained a new compound (6), with a different retention time (t_R : 15.2 min) from that of compound 1 (t_R : 11.8 min).

2. Results and Discussion

Compound **6** was obtained as a colorless oil, and its HR FABMS gave a pseudomolecular ion $[M+1]^+$ at m/z 831.4211 (calcd for $C_{44}H_{63}O_{15}$ 831.4167), 28 amu more than that of ipomoeassin A (**1**). Its IR, UV, and ¹H NMR spectra were nearly identical to those of compound **1**. The major difference between their ¹³C NMR spectra was located in the high field region (10 to 50 ppm), and it was thus deduced that compound **6** was a homolog of **1** with a C16 rather than a C14 hydroxyacid side chain, with two more methylenes than in **1**. The oxygenated

[†]See Ref. [1]

^{*}Corresponding author. Fax: (540) 231-3255. Tel.: (540) 231-6570; dkingston@vt.edu; URL: http://www.kingston.chem.vt.edu/.

methine must be located at position 11 since H-11 showed HMBC correlations to C-12 and C-13, and H₃-16 to C-14, and C-15. The chemical shifts of C-1 to C-6 were close to those of

1, and the signals for C-7 to C-16 matched those of 11-hydroxyhexadecanoic acid [3], which confirmed the above deduction. The spin systems of **6** were determined from COSY and TOCSY spectra. The connectivities from H-1 of fucose to C-11, H-1 of glucose to C-2 of fucose, H₂-6 of glucose to C-1 of the side chain, H-4 of fucose to the acetyl carbonyl, and H-3 and H-4 of glucose to the tigloyl and *E*-cinnamoyl carbonyls, respectively, were established from HMQC and HMBC spectra. The configurations at position-11 and the sugar moieties were determined by analysis of its ROESY spectrum (Figure 1). Hence, the structure of **6** was determined as shown.

Compound **6** exhibited potent cytotoxic activity against A2780 human ovarian cancer cell lines with an IC₅₀ value of 0.036 μ M, which was much more active than compound **1** (0.5 μ M) [1]. Here we can see again that sometimes a slight modification of the structure will enhance the activity dramatically.

3. Experimental

3.1 General Procedures

General experimental procedures, cytotoxicity bioassays, plant material, and isolation procedures were as previously described [2].

3.2 Isolation

The crude extract of E940631 (10 g, IC₅₀: 8.0 µg/mL) was separated into five fractions and fraction III was purified by HPLC over C18 using 75% MeCN/H₂O to yield ipomoeassins A-E (**1-5**) as described previously [2]. A fraction collected immediately after compound **1** was examined by ¹H NMR and normal phase TLC (silica-gel, CH₂Cl₂:CH₃OH; 40:1, $R_f = 0.3$), and appeared to be identical with or very similar to compound **1**. Further chromatography of this fraction was carried out using a semi-preparative C8 Varian Dynamax HPLC column (5 µ, 250×10 mm) with 75% CH₃CN/H₂O as eluents to afford compound **6** (1.4 mg, t_R : 15.2 min), which had a different retention time from that of compound **1** (t_R : 11.8 min).

3.3 Ipomoeassin F (6)

colorless oil; $[\alpha]_D^{22}$ -54° (*c* 0.16, EtOH); IR (film) ψ_{max} 3400, 2931, 2857, 1719, 1636, 1450, 1378, 1247, 1154, 1071, 1017; UV (EtOH) λ_{max} (log ε) 278 (4.18) nm; ¹H NMR (500 MHz, C₆D₆) and ¹³C NMR (125 MHz, C₆D₆) data, see Table 1; HRESIMS *m*/*z* 831.4211 (calcd for C₄₄H₆₃O₁₅ 831.4167).

Acknowledgments

This project was supported by the Fogarty International Center, the National Cancer Institute, the National Science Foundation, the National Heart Lung and Blood Institute, the National Institute of Mental Health, the Office of Dietary Supplements, and the Office of the Director of NIH, under Cooperative Agreement U01 TW000313 with the International Cooperative Biodiversity Groups, and this support is gratefully acknowledged. We also thank Mr. Bill Bebout for obtaining the HRFABMS spectrum, and Mr. Tom Glass for assistance obtaining NMR spectra.

Reference

[1]. Biodiversity Conservation and Drug Discovery in Suriname, Part 18. For Part 17, see Adou E, Williams RB, Schilling JK, Malone S, Meyer J, Wisse JH, Frederik D, Koese D, Werkhoven MCM, Snipes CE, Werk TL, Kingston DGI. Bioorg. Med. Chem 2005;13:6009. [PubMed: 16125394](this is erroneously listed as Part 16)

- [2]. Cao SG, Guza RC, Wisse JH, Miller JS, Evans R, Kingston DGI. J. Nat. Prod 2005;68:487. [PubMed: 15844934]
- [3]. Fürstner A, Müller T. J. Am. Chem. Soc 1999;121:7814.

CAO et al.





CAO et al.



Structures 1-6.

Nat Prod Res. Author manuscript; available in PMC 2009 November 12.

Table 1

¹H and ¹³C NMR data of ipomoeassins F (6) and A (1)^a

no	6	1	6	1	
1			171.5	171.5	-
2	2.39 ddd (17.3, 9.6, 3.2) 2.14 ddd (17.3, 7.4, 3.4)	2.40 ddd (17.4, 9.4, 3.4) 2.14 ddd (17.4, 7.7, 3.5)	37.4 ^b	37.3 ^b	
3	2.65 ddd (16.4, 7.4, 3.2) 2.52 ddd (16.4, 9.6, 3.4)	2.65 ddd (16.1, 7.7, 3.4) 2.52 ddd (16.1, 9.4, 3.5)	29.6 ^{<i>c</i>}	29.7 ^{<i>c</i>}	
4	2102 add (1011, 210, 511)	2022 ddd (1011, 511, 510)	208.3	208.4	
5	2.07 t (7.1)	2.07 t (6.0)	41.6	41.6	
6			23.5	23.8	
7			28.7	28.7	
8			29.4°	29.4^{c}	
9			25.5	25.5	
10			34.3	34.3	
11	3.72^{b} m	3.72^{b} m	79.4	79.0	
12			35.4^{b}	37.6^{b}	
13			25.2	18.7	
14		0.95 t (7.1)	32.3	$14 4^d$	
15			23.1	1	
16	0.92 t (7.1)		$\frac{14}{14} \frac{3^d}{3^d}$		
1'	440 d(76)	4 40 d (7 7)	100.8	100.8	
2'	$3.96^{\circ}dd(9.4,7.6)$	3.96° dd (9.5, 7.7)	84.0	84.0	
3'	3.70^{b} dd $(0.4, 3.7)$	3.70^{b} dd (0.5, 7.7)	72 00	72 7 ^e	
J 1'	5.72 dd (9.4, 5.7) 5.15 hr d (3.7)	5.72 dd $(9.3, 5.7)5.15 dd (3.7, 0.5)$	12.0 72.0	72.7	
	2.11 hr = (6.4)	2.10 = 1(6.4, 0.5)	72.8	72.9	
5	3.11 br q (0.4) 1 11 d (6 d)	3.10 qu (6.4, 0.5)	69.0	69.0	
0	1.11 d (0.4)	1.10 d (0.4)	14.1	14.14	
1"	4.54 d (7.8)	4.52 d (7.9)	106.5	106.6	
2	3.91°dd (9.8, 7.8)	3.91°dd (9.7, 7.9)	/4.8	74.8	
3" 4"	5.40 t (9.8)	5.39 t (9.7)	/6./	/6.4	
4 5"	3.701(9.8)	3.091(9.7)	07.8 72.06	07.8	
5	5.27 bi d (9.8)	5.24 ddd (9.7, 5.2, 1.0)	73.0°	/3.0°	
0	4.67 dd (12.6, 3.0) 4.13 br d (12.6)	4.66 dd (12.6, 3.2) 4.11 dd (12.6, 1.6)	61.5	61.5	
AA-1			170.9	171.0	
AA-2	1.84 s	1.82 s	20.5	20.5	
TA-1			168.7	168.5	
TA-2			128.0	128.0	
TA-3	6.95-7.00 m	6.95 m	139.4	139.2	
TA-4	1.26 d (7.1)	1.23 d (7.1)	16.6	16.6	
TA-5	1.71 br s	1.68 br s	12.0	12.1	
CA-1			165.6	165.6	
CA-2	6.39 d (16.0)	6.39 d (15.9)	117.6	117.6	
CA-3	7.82 d (16.0)	7.81 d (15.9)	146.1	146.1	
CA-4	6.02 + (7.1)	6 80 7 07	154.4	134.5	
CA-5	0.92 d (7.1)	0.89-7.07	128.5	128.3	
CA-0	7.05 d (7.1) 6.95-7.00 m	0.09-7.07	128.9	128.9	
CA-/	0.75-7.00 III	0.09-7.07	150.4	130.4	

 $^{a}\delta$ (ppm), in benzene- d_{6} , 500 MHz for ¹H NMR and 125 MHz for ¹³C NMR; multiplicities; *J* values (Hz) in parentheses. AA = acetoyl; TA = tigloyl; CA = cinnamoyl

b-e overlapped