

Resveratrol attenuates oxidative stress and histological alterations induced by liver ischemia/reperfusion in rats

Ercan Gedik, Sadullah Girgin, Hayrettin Ozturk, Basra Deniz Obay, Hulya Ozturk, Huseyin Buyukbayram

Ercan Gedik, Sadullah Girgin, Department of General Surgery, Medical School, Dicle University, Diyarbakir 21280, Turkey

Hayrettin Ozturk, Department of Pediatric Surgery, Medical School, Abant Izzet Baysal University, Bolu, Turkey

Basra Deniz Obay, Department of Physiology, Medical School, Dicle University, Diyarbakir 21280, Turkey

Hulya Ozturk, Department of Pediatric Surgery, Medical School, Duzce University, Bolu, Turkey

Huseyin Buyukbayram, Department of Pathology, Medical School, Dicle University, Diyarbakir 21280, Turkey

Author contributions: All of the authors contributed equally to this work.

Correspondence to: Ercan Gedik, Assistant Professor in General Surgery, Department of General Surgery, Medical School, Dicle University, Diyarbakir 21280,

Turkey. ercan.gedik@yahoo.com.tr

Telephone: +90-412-2488001-4679 Fax: +90-412-2488440

Received: September 18, 2008 Revised: October 13, 2008

Accepted: October 20, 2008

Published online: December 14, 2008

Abstract

AIM: To investigate the effects of resveratrol on liver ischemia/reperfusion (I/R) injury in rats.

METHODS: A total of 40 male Sprague-Dawley rats weighing 240-290 g were randomized into four groups of ten: (1) controls: data from unmanipulated animals; (2) sham group: rats subjected to the surgical procedure, except for liver I/R, and given saline; (3) I/R group: rats underwent liver ischemia for 45 min followed by reperfusion for 45 min; (4) I-R/Resveratrol group: rats pretreated with resveratrol (10 μ mol/L, iv). Liver tissues were obtained to determine antioxidant enzyme levels and for biochemical and histological evaluation.

RESULTS: Plasma aminotransferase activities were higher in the I/R group than in the I-R/Resveratrol group. Malondialdehyde levels and the hepatic injury score decreased, while superoxide dismutase, catalase, and glutathione peroxidase levels increased in group 4 compared to group 3. In group 4, histopathological changes were significantly attenuated in resveratrol-treated livers.

CONCLUSION: These results suggest that resveratrol has protective effects against hepatic I/R injury, and is a potential therapeutic drug for ischemia reperfusion-related liver injury.

Key words: Injury; Ischemia/reperfusion; Liver; Resveratrol

Peer reviewers: Dr. Martin Hennenberg, Dipl-Biol, Medizinische Klinik & Poliklinik I, Uni-Klinik Bonn, Sigmund-Freud Str. 25, 53105 Bonn, Germany; Adriana M Torres, Professor of Pharmacology, Suipacha 531, Rosario 2000, Argentina

Gedik E, Girgin S, Ozturk H, Obay BD, Ozturk H, Buyukbayram H. Resveratrol attenuates oxidative stress and histological alterations induced by liver ischemia/reperfusion in rats. *World J Gastroenterol* 2008; 14(46): 7101-7106 Available from: URL: <http://www.wjgnet.com/1007-9327/14/7101.asp> DOI: <http://dx.doi.org/10.3748/wjg.14.7101>

INTRODUCTION

Hepatic injury caused by ischemia/reperfusion (I/R) has been proposed as a key clinical problem associated with liver transplantation and major liver surgery^[1]. Additionally, hepatic I/R injury also occurs in diverse situations, including heart failure, liver trauma, and blood occlusion to the liver^[2,3]. The production of reactive oxygen species (ROS), including superoxide, hydrogen peroxide, and hydroxyl radical, has been demonstrated in reperfusion injury^[4]. I/R activates Kupffer cells (KC), the resident macrophages in the liver, to generate ROS, proinflammatory cytokines, and chemokines and to upregulate inducible nitric oxide synthase^[5].

Resveratrol (3,4,5 tri-hydroxystilbene) is a naturally occurring phytoalexin present in high concentrations in the skin and seeds of grapes^[6]. It occurs naturally in a trans- or cis-isoform^[7-9]. Resveratrol has been reported to have several biologic effects such as a potent antioxidative effect *via* prevention of lipid peroxidation^[10,11], anti-platelet activity^[12], an estrogenic activity^[13], and anti-inflammatory activity attributed to cyclooxygenase inhibition^[6,14]. Previously, it was suggested that resveratrol stimulated nitric oxide production in endothelial cells, has a vasodilatory effect on blood vessels^[15], and improves the energy metabolism system after spinal cord trauma^[16].

The aim of this study was to evaluate the possible protective effect of resveratrol against I/R-induced hepatic injury, using biochemical and histological parameters.

MATERIALS AND METHODS

Forty male Sprague-Dawley rats weighing 240-290 g

were used in the study. All of the experimental protocols were performed according to the guidelines for the ethical treatment of experimental animals.

Animals and experimental protocol

The rooms where the animals were housed were windowless and were under controlled temperature ($22 \pm 2^\circ\text{C}$) and lighting conditions. The rats were housed individually in cages, and allowed free access to standard rat chow and water before and after the experiments. The animals were fasted overnight before the experiments, but were given free access to water. They were anesthetized using 100 mg/kg ketamine and 20 mg/kg xylazine body weight (ip). The right femoral vein was cannulated to administer drugs and saline.

The animals were randomized into four groups ($n = 10$ in each group): (1) controls: Unmanipulated animals, rats not subjected to any surgical procedure or liver manipulation; (2) sham group: Rats subjected to the surgical procedures described below, except for liver I/R, and administered saline vehicle and maintained under anesthesia for an equivalent duration (i.e. 45 min plus 45 min); (3) I/R group: Rats subjected to the surgical procedures described below and underwent liver ischemia for 45 min followed by reperfusion for 45 min ($n = 10$); (4) I-R/Resveratrol group: Rats received resveratrol 10 mg/kg dissolved in 2 mL 50% alcohol (Sigma Chemicals, St. Louis, MO, USA) by infusion pump 15 min before liver reperfusion.

Liver ischemia/reperfusion

As described previously^[17], the ligament attachments connecting the liver, diaphragm, abdominal wall, and neighboring organs were divided. After the organ was carefully isolated, the liver hilus was exposed to find the common hepatic artery and portal vein. A vascular microclamp was used to interrupt the blood supply to three-quarters of the liver for 45 min, and this was followed by 45 min of reperfusion. Other rats were subjected to a sham operation (sham-operated), which was identical to the surgical procedure used for the I/R group rats without clamping; the rats were kept under anesthesia for the same length of time. At the end of the experiments, the rats were killed with an overdose of sodium pentobarbital.

Measuring serum liver enzymes

The abdominal aorta was punctured and 5 mL of blood was taken and placed in heparinized tubes. Plasma was separated by centrifugation (3000 r/min for 10 min at room temperature) for biochemical studies. The activities of alanine aminotransferase (ALT, a specific marker for hepatic parenchymal injury), and aspartate aminotransferase (AST, a nonspecific marker for hepatic injury) in plasma were determined in units per liter using standard auto-analyzer methods on an Abbott Aeroset (Abbott Laboratories, Abbott Park, IL, USA). Just before the rats were sacrificed, the livers were removed for histopathological evaluation.

Histopathological study

The livers were divided into two pieces. One was

immediately placed in 10% formalin solution, left overnight, and then embedded in paraffin blocks. The blocks were cut into 4- μm sections and stained with hematoxylin-eosin, using standard protocols. The severity of hepatic injury in the sections was evaluated using a point-counting method on an ordinal scale as follows: grade 0, minimal or no evidence of injury; grade 1, mild injury consisting of cytoplasmic vacuolation and focal nuclear pyknosis; grade 2, moderate to severe injury with extensive nuclear pyknosis, cytoplasmic hyper eosinophilia, and loss of intercellular borders; and grade 3, severe necrosis with disintegration of hepatic cords, hemorrhage, and neutrophil infiltration^[18].

Biochemical analyses

The other piece of liver was washed in ice-cold 0.9% saline solution, weighed, and stored at -70°C . Tissue homogenates were prepared as 0.1 g/mL in 250 mmol/L sucrose, 1 mmol/L EDTA, 1 mmol/L DL-dithiothreitol, and 15 mmol/L Tris HCl (pH 7.4), using an all-glass Potter Elvehjem homogenizer (Selecta, Barcelona, Spain). Each homogenate was centrifuged for 20 min at 800 r/min. The resulting supernatant fraction was used to determine enzyme activities. The protein concentrations in the supernatant were determined using the Bradford method^[19].

Malondialdehyde determination

Liver malondialdehyde (MDA) levels were determined using the method of Wasowicz *et al*^[20] based on the reaction of MDA with thiobarbituric acid at 95 to 100°C . Fluorescence intensity was measured in the upper *n*-butanol phase using fluorescence spectrophotometry (F-4010; Hitachi, Tokyo, Japan) adjusted for excitation at 525 nm and emission at 547 nm. The arbitrary values obtained were compared with a series of standard solutions (1,1,3,3-tetramethoxypropane). The results are given in nanomoles per gram of wet tissue.

Superoxide dismutase, catalase, and glutathione peroxidase determination

Superoxide dismutase (SOD) activity was measured using the xanthine-oxidase-cytochrome *c* method, as described by McCord and Fridovich^[21]. The final concentrations in the cuvettes were 50 mmol/L potassium phosphate (pH 7.8), 0.1 mmol/L EDTA, 10 mmol/L cytochrome *c*, 50 mmol/L xanthine, 50 or 2 mmol/L cyanide, 1 U catalase (CAT), and 0.05-0.1 mg of tissue. The reaction was initiated by adding 1 U xanthine-oxidase. The inhibition of xanthine-oxidase was followed spectrophotometrically at 550 nm. One unit of SOD activity was defined as the amount of enzyme that produced 50% inhibition of the control rate of cytochrome *c* reduction.

CAT activity was assayed according to the method of Beers and Sizer^[22]. The final concentrations in the cuvettes were 500 mmol/L potassium phosphate (pH 7), 100 mmol/L H_2O_2 , and 0.05-0.1 mg of tissue. The decrease in the absorbance at 240 nm after adding the substrate was followed spectrophotometrically.

GSH activity was assayed using a coupled enzyme system in which oxidized glutathione (GSSG) reduction

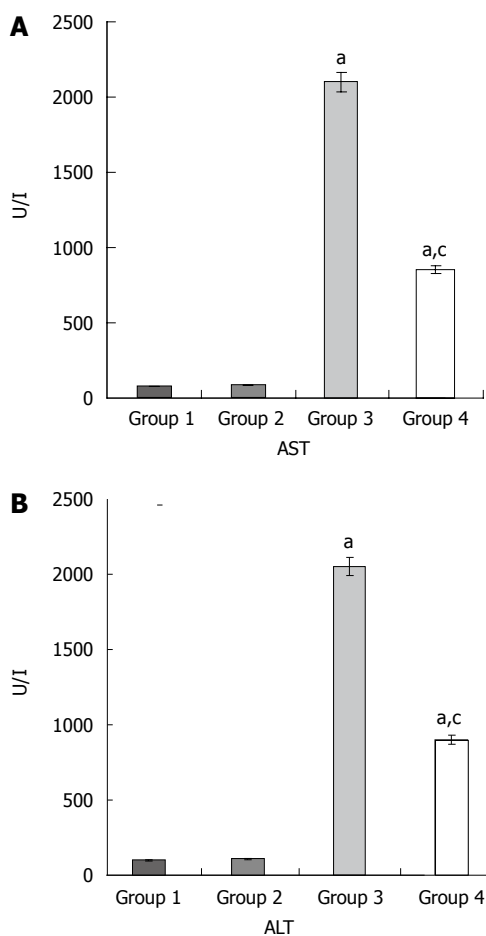


Figure 1 Effects of liver I/R and resvera-trol on liver function. A: The AST values; B: The ALT values ^a $P < 0.05$ compared with Group 1 and 2; ^c $P < 0.05$ compared with Group 3. The values are the mean \pm SE. ALT: Alanine aminotransferase; AST: Aspartate aminotransferase.

was coupled to NADPH oxidation by glutathione reductase^[23]. The assay mixture contained 50 mmol/L potassium phosphate (pH 7.5), 1 mmol/L EDTA, 1 mmol/L NaN₃, 1 mmol/L reduced glutathione (GSH), 0.2 mmol/L NADPH, 1 U glutathione reductase, and tissue (0.05–0.2 mg). After a 5-min preincubation period (20–25°C), the reaction was initiated by adding 0.25 mmol/L H₂O₂. The decrease in absorbance at 340 nm was followed spectrophotometrically.

Protein assays

The protein content of the homogenates was determined using the procedure of Lowry *et al.*^[24].

Statistical analysis

Data were entered and analyzed on an IBM-compatible personal computer using SPSS version 9.0. All values were expressed as the mean \pm SE. The significance of the data obtained was evaluated using analysis of variance (ANOVA). Differences between means were analyzed using the post-ANOVA test (Tukey's *b*). $P < 0.05$ was considered significant.

RESULTS

The ALT and AST levels were significantly increased

in groups 3 and 4 in comparison with groups 1 and 2 ($P < 0.05$ in all cases). Moreover, the ALT and AST levels were significantly decreased in group 4 compared to group 3 ($P < 0.05$) (Figure 1).

The MDA, SOD, CAT, and GSH values for the different groups are shown in Figure 2. In group 3, MDA was significantly increased compared with groups 1, 2, and 4 ($P < 0.05$ in all cases). In addition, SOD, CAT, and GSH significantly decreased in group 3 compared with groups 1, 2, and 4 ($P < 0.05$ in all cases).

The liver histopathological scores were 0.1 ± 0.2 , 0.1 ± 0.2 , 2.8 ± 0.1 , and 1.1 ± 0.2 in groups 1 to 4, respectively. The histopathological scores were higher in groups 3 and 4 than in groups 1 and 2 ($P < 0.05$ in all cases). Moreover, the histopathological score was lower in group 4 than in group 3 ($P < 0.05$). Normal findings were obtained on histological examination of rats in groups 1 or 2 (Figure 3A and B). In group 3, focal necrosis with sinusoidal congestion and neutrophil accumulation were observed at the midzonal region (Figure 3C). In contrast, these changes were markedly attenuated in resveratrol-treated livers (Figure 3D).

DISCUSSION

Hepatic ischemia and reperfusion injury is observed following major liver surgery, transplantation, trauma and sepsis and may cause metabolic and structural hepatic damage^[25,26]. This remains a significant problem in surgical procedures, and is a limitation of liver transplantation^[27]. Several mechanisms have been considered to explain I/R injury of the liver. Activation of KC with enhanced formation of ROS and secretion of inflammatory cytokines and proteolytic enzymes are considered to play an important role in liver reperfusion injury^[28,29]. Additionally, migration of activated polymorphonuclear leukocytes at the injury site may prolong the injury as a result of the production of inflammatory cytokines, adhesion molecules, and ROS^[30,31]. The formation of ROS may initiate oxidative stress which can lead to lipid peroxidation^[32]. Removal of oxidative stress is the primary intervention to decrease tissue injury. Superoxide dismutase, catalase, glutathione, α -tocopherol, and carotene are known endogenous antioxidants, but none of these endogenous antioxidants are sufficient to compensate oxidative stress. Several antioxidants have been studied and reported to decrease oxidative stress in hepatic and renal tissue in experimental obstructive jaundice, septic shock, and I/R models.

Resveratrol is a polyphenol phytoalexin (trans-3,5,4-trihydroxystilbene) that possesses diverse biochemical and physiological actions, including estrogenic, antiplatelet, and anti-inflammatory properties^[33,34]. Recently, resveratrol was found to be a highly potent antioxidant which could inhibit free radical generation in brain, spinal cord, kidney, liver and red cell membrane^[35–38]. It has been shown that resveratrol inhibits lipid peroxidation^[39], and prevents apoptotic cell death induced by oxidative stress^[40]. The end production of lipid peroxidation includes aldehydes, hydrocarbon gases, and MDA. MDA is a sensitive index of lipid peroxidation^[18,41]. Bertelli *et al.*^[33] suggested that

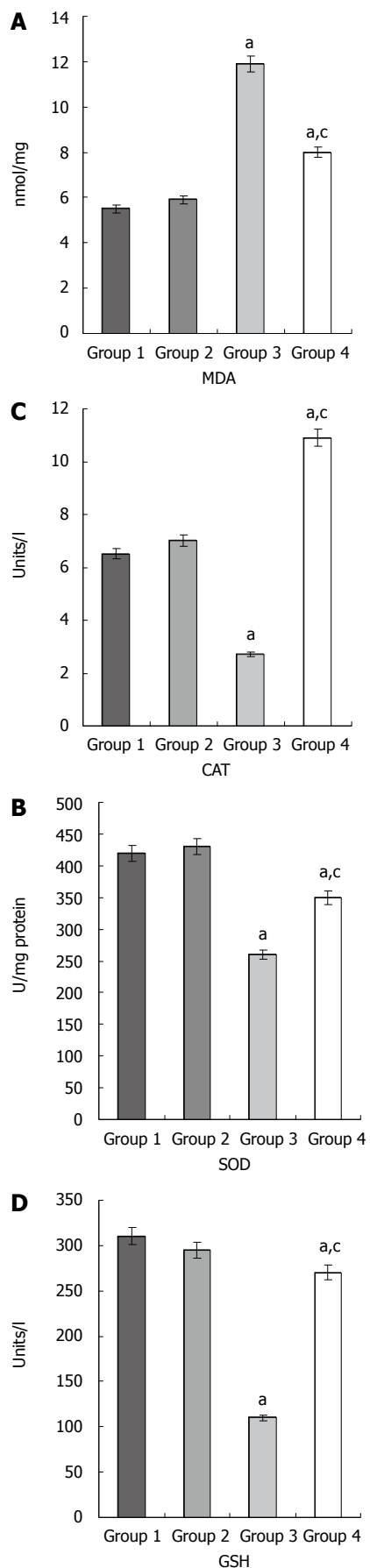


Figure 2 Effects of ischemia/reperfusion and resveratrol on the MDA (A), SOD (B), CAT (C), and GSH (D) levels in liver tissue. ^a*P* < 0.05 compared with groups 1 and 2. ^c*P* < 0.05 compared with group 3. The values are the mean ± SE. MDA: Malondialdehyde; SOD: Superoxide dismutase; CAT: Catalase; GSH: Glutathione peroxidase.

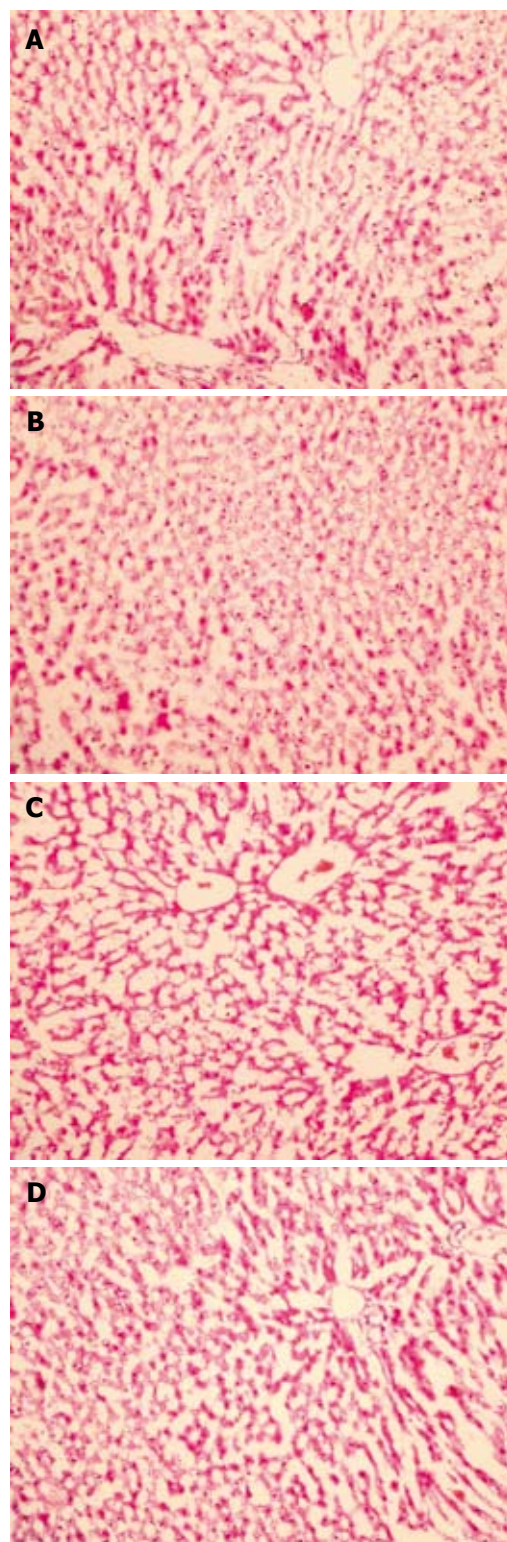


Figure 3 Effect of resveratrol on hepatic tissue injury after I/R evaluated by histological examination. In groups 1 and 2, normal findings were obtained on histological examination (A and B, respectively) (HE, x 200); C: In group 3, swollen hepatocytes with marked vacuolization and congestion were observed (HE, x 200); D: In group 4, the hepatocytes and sinusoids showed normal morphology, reflecting a well preserved liver parenchyma (HE, x 200).

MDA formation was reduced in the ischemic-reperfused myocardium of resveratrol-treated rats. In an experimental study by Kirimlioglu *et al*^[42], MDA levels in liver tissue and plasma were higher in group B subjected to 70% partial hepatectomy than those of group A treated with

resveratrol before and after 70% partial hepatectomy. Although the exact mechanism is not known, it is thought that this was probably due to the antioxidant and inhibitory effects of resveratrol on ROS, which might be a biochemical mechanism related to the anti-inflammatory and anticarcinogenic properties of resveratrol^[43]. To control the detrimental effects of ROS, organisms have developed a variety of antioxidant defense systems, especially the endogenous antioxidant enzymes system including SOD, CAT and GSH. The activities of these enzymes are higher in the liver than in other tissues. CAT is an oxidoreductase enzyme, which transforms H₂O₂ into H₂O and O₂. It can protect cells from damage induced by ischemia reperfusion by scavenging ROS^[44]. GSH is considered to be the principal mitochondrial antioxidant and its depletion markedly enhances the sensitivity of mitochondrial structures to ROS-mediated injury^[45]. A decrease in GSH level after reperfusion could reflect ROS-mediated consumption^[46]. SOD catalyses dismutation of the superoxide anion (O₂⁻) into H₂O₂; H₂O₂ can be transformed into H₂O and O₂ by CAT; GSH is a selenoprotein, which reduces lipidic or nonlipidic hydroperoxides as well as H₂O₂ while oxidizing GSH^[47]. Plin *et al*^[48] showed that resveratrol protects mitochondria and cells against cold preservation-warm reperfusion (CPWR)-induced injury. Resveratrol improved both mitochondrial coupling and ATP synthesis measured after CPWR as demonstrated by the increase in respiratory control ratio and ADP/oxygen values, respectively. These protective effects could be related to the antioxidant properties of the molecule which has been shown to be able to scavenge ROS^[49].

In our study, liver tissue MDA levels in the I/R group increased compared with normal and sham groups. With resveratrol administration, liver tissue MDA levels decreased. Additionally, our results suggest that increased SOD, GSH and CAT activities may be attributed to reactive oxygen products such as superoxide anions and H₂O₂ in the resveratrol-treated group. These data indicate that resveratrol may provide protection to the liver during I/R injury, in part by improving activities of the endogenous antioxidant enzymes, which scavenge ROS and reduce their effects. Histopathologically, the resveratrol-treated group showed well preserved liver parenchyma with hepatocytes arranged radially around the central vein. In a study by Hassan-Khabbar *et al*^[50], the effect of trans-resveratrol on liver injury induced by I/R was investigated. While Hassan-Khabbar performed measurements after 3 h of reperfusion, in the present study, measurements were performed after less than 1 h. Thus, the present study suggests that the protective effect of resveratrol may occur more rapidly than previously thought.

In conclusion, this study is the first to evaluate the hepatoprotective effects of resveratrol in hepatic I/R. The protective effects of resveratrol may be associated with its antioxidant activity and free radical scavenging activity which are released during the reperfusion period.

COMMENTS

Background

Hepatic injury caused by ischemia/reperfusion (I/R) has been proposed as a key clinical problem associated with liver transplantation and major liver sur-

gery. Furthermore, hepatic I/R injury also occurs in diverse situations, including heart failure, liver trauma, and blood occlusion to the liver. The production of reactive oxygen species has been demonstrated in reperfusion injury. Resveratrol has been reported to have several biologic effects such as a potent antioxidant effect via prevention of lipid peroxidation. Therefore, the possible protective effects of resveratrol against I/R-induced hepatic injury were investigated.

Research frontiers

This study is the first to evaluate the hepatoprotective effects of resveratrol in hepatic I/R. The protective effects of resveratrol may be associated with its antioxidant activity and free radical scavenging activity which are released during the reperfusion period.

Innovations and breakthroughs

This study explained one of the possible mechanisms of the protective effects of resveratrol. This may be associated with its antioxidant activity and free radical scavenging activity released during the reperfusion period.

Applications

In this study, the implication of ROS in the pathology of hepatic I/R injury was explored. Protective effects of resveratrol were determined. This may represent a novel and attractive approach to determining the protective effects of resveratrol therapy on hepatic injury caused by ischemia/reperfusion

Peer review

In the present study, the authors tested the effect of Resveratrol on ischemia/reperfusion-induced liver injury in rats, and found a protective effect.

REFERENCES

- 1 **Ito K**, Ozasa H, Noda Y, Koike Y, Arai S, Horikawa S. Effect of non-essential amino acid glycine administration on the liver regeneration of partially hepatectomized rats with hepatic ischemia/reperfusion injury. *Clin Nutr* 2008; **27**: 773-780
- 2 **Jaeschke H**, Bautista AP, Spolarics Z, Spitzer JJ. Superoxide generation by neutrophils and Kupffer cells during in vivo reperfusion after hepatic ischemia in rats. *J Leukoc Biol* 1992; **52**: 377-382
- 3 **Tejima K**, Arai M, Ikeda H, Tomiya T, Yanase M, Inoue Y, Nagashima K, Nishikawa T, Watanabe N, Omata M, Fujiwara K. Ischemic preconditioning protects hepatocytes via reactive oxygen species derived from Kupffer cells in rats. *Gastroenterology* 2004; **127**: 1488-1496
- 4 **Lemasters JJ**. The mitochondrial permeability transition and the calcium, oxygen and pH paradoxes: one paradox after another. *Cardiovasc Res* 1999; **44**: 470-473
- 5 **Taniai H**, Hines IN, Bharwani S, Maloney RE, Nimura Y, Gao B, Flores SC, McCord JM, Grisham MB, Aw TY. Susceptibility of murine periportal hepatocytes to hypoxia-reoxygenation: role for NO and Kupffer cell-derived oxidants. *Hepatology* 2004; **39**: 1544-1552
- 6 **Soleas GJ**, Diamandis EP, Goldberg DM. Wine as a biological fluid: history, production, and role in disease prevention. *J Clin Lab Anal* 1997; **11**: 287-313
- 7 **Daniel O**, Meier MS, Schlatter J, Frischknecht P. Selected phenolic compounds in cultivated plants: ecologic functions, health implications, and modulation by pesticides. *Environ Health Perspect* 1999; **107** Suppl 1: 109-114
- 8 **Sobolev VS**, Cole RJ. trans-resveratrol content in commercial peanuts and peanut products. *J Agric Food Chem* 1999; **47**: 1435-1439
- 9 **Fremont L**. Biological effects of resveratrol. *Life Sci* 2000; **66**: 663-673
- 10 **Inoue H**, Umeson K, Nishimori T, Hirata Y, Tanabe T. Glucocorticoid-mediated suppression of the promoter activity of the cyclooxygenase-2 gene is modulated by expression of its receptor in vascular endothelial cells. *Biochem Biophys Res Commun* 1999; **254**: 292-298
- 11 **Sinha K**, Chaudhary G, Gupta YK. Protective effect of resveratrol against oxidative stress in middle cerebral artery occlusion model of stroke in rats. *Life Sci* 2002; **71**: 655-665
- 12 **Inoue H**, Tanabe T, Umeson K. Feedback control of cyclooxygenase-2 expression through PPARgamma. *J Biol Chem* 2000; **275**: 28028-28032

- 13 **Huang SS**, Tsai MC, Chih CL, Hung LM, Tsai SK. Resveratrol reduction of infarct size in Long-Evans rats subjected to focal cerebral ischemia. *Life Sci* 2001; **69**: 1057-1065
- 14 **Bloomfield Rubins H**, Davenport J, Babikian V, Brass LM, Collins D, Wexler L, Wagner S, Papademetriou V, Rutan G, Robins SJ. Reduction in stroke with gemfibrozil in men with coronary heart disease and low HDL cholesterol: The Veterans Affairs HDL Intervention Trial (VA-HIT). *Circulation* 2001; **103**: 2828-2833
- 15 **Kimura Y**, Okuda H, Arichi S. Effects of stilbenes on arachidonate metabolism in leukocytes. *Biochim Biophys Acta* 1985; **834**: 275-278
- 16 **Yang YB**, Piao YJ. Effects of resveratrol on secondary damages after acute spinal cord injury in rats. *Acta Pharmacol Sin* 2003; **24**: 703-710
- 17 **Sepodes B**, Maio R, Pinto R, Marques C, Mendes-do-Vale J, McDonald MC, Thiemermann C, Mota-Filipe H. Tempol, an intracellular free radical scavenger, reduces liver injury in hepatic ischemia-reperfusion in the rat. *Transplant Proc* 2004; **36**: 849-853
- 18 **Ozturk H**, Gezici A, Ozturk H. The effect of celecoxib, a selective COX-2 inhibitor, on liver ischemia/reperfusion-induced oxidative stress in rats. *Hepato Res* 2006; **34**: 76-83
- 19 **Bradford MM**. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 1976; **72**: 248-254
- 20 **Wasowicz W**, Neve J, Peretz A. Optimized steps in fluorometric determination of thiobarbituric acid-reactive substances in serum: importance of extraction pH and influence of sample preservation and storage. *Clin Chem* 1993; **39**: 2522-2526
- 21 **McCord JM**, Fridovich I. Superoxide dismutase. An enzymic function for erythropoietin (hemocuprein). *J Biol Chem* 1969; **244**: 6049-6055
- 22 **Beers RF Jr**, Sizer IW. A spectrophotometric method for measuring the breakdown of hydrogen peroxide by catalase. *J Biol Chem* 1952; **195**: 133-140
- 23 **Lawrence RA**, Burk RF. Glutathione peroxidase activity in selenium-deficient rat liver. *Biochem Biophys Res Commun* 1976; **71**: 952-958
- 24 **Lowry OH**, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951; **193**: 265-275
- 25 **Shin T**, Kuboki S, Huber N, Eismann T, Galloway E, Schuster R, Blanchard J, Pritts TA, Lentsch AB. Activation of peroxisome proliferator-activated receptor-gamma during hepatic ischemia is age-dependent. *J Surg Res* 2008; **147**: 200-205
- 26 **van Gulik TM**, de Graaf W, Dinant S, Busch OR, Gouma DJ. Vascular occlusion techniques during liver resection. *Dig Surg* 2007; **24**: 274-281
- 27 **He XS**, Ma Y, Wu LW, Wu JL, Hu RD, Chen GH, Huang JF. Dynamical changing patterns of glycogen and enzyme histochemical activities in rat liver graft undergoing warm ischemia injury. *World J Gastroenterol* 2005; **11**: 2662-2665
- 28 **Shibuya H**, Ohkohchi N, Tsukamoto S, Satomi S. Tumor necrosis factor-induced, superoxide-mediated neutrophil accumulation in cold ischemic/reperfused rat liver. *Hepatology* 1997; **26**: 113-120
- 29 **Suzuki S**, Toledo-Pereyra LH. Interleukin 1 and tumor necrosis factor production as the initial stimulants of liver ischemia and reperfusion injury. *J Surg Res* 1994; **57**: 253-258
- 30 **Hughes H**, Farhood A, Jaeschke H. Role of leukotriene B4 in the pathogenesis of hepatic ischemia-reperfusion injury in the rat. *Prostaglandins Leukot Essent Fatty Acids* 1992; **45**: 113-119
- 31 **Jaeschke H**. Cellular adhesion molecules: regulation and functional significance in the pathogenesis of liver diseases. *Am J Physiol* 1997; **273**: G602-G611
- 32 **Aldemir M**, Bosnak M, Al B, Buyukbayram H, Tacyildiz I. Effects of molsidomine and leixapafant in hepatic ischaemia-reperfusion injury. *Injury* 2004; **35**: 232-237
- 33 **Bertelli AA**, Giovannini L, Bernini W, Migliori M, Fregoni M, Bavaresco L, Bertelli A. Antiplatelet activity of cis-resveratrol. *Drugs Exp Clin Res* 1996; **22**: 61-63
- 34 **Ferrero ME**, Bertelli AE, Fulgenzi A, Pellegatta F, Corsi MM, Bonfrate M, Ferrara F, De Caterina R, Giovannini L, Bertelli A. Activity in vitro of resveratrol on granulocyte and monocyte adhesion to endothelium. *Am J Clin Nutr* 1998; **68**: 1208-1214
- 35 **Yang YB**, Piao YJ. Effects of resveratrol on secondary damages after acute spinal cord injury in rats. *Acta Pharmacol Sin* 2003; **24**: 703-710
- 36 **Ray PS**, Maulik G, Cordis GA, Bertelli AA, Bertelli A, Das DK. The red wine antioxidant resveratrol protects isolated rat hearts from ischemia reperfusion injury. *Free Radic Biol Med* 1999; **27**: 160-169
- 37 **Olas B**, Wachowicz B. Resveratrol and vitamin C as antioxidants in blood platelets. *Thromb Res* 2002; **106**: 143-148
- 38 **Chander V**, Tirkey N, Chopra K. Resveratrol, a polyphenolic phytoalexin protects against cyclosporine-induced nephrotoxicity through nitric oxide dependent mechanism. *Toxicology* 2005; **210**: 55-64
- 39 **Tadolini B**, Juliano C, Piu L, Franconi F, Cabrini L. Resveratrol inhibition of lipid peroxidation. *Free Radic Res* 2000; **33**: 105-114
- 40 **Chanvitayapongs S**, Draczynska-Lusiak B, Sun AY. Amelioration of oxidative stress by antioxidants and resveratrol in PC12 cells. *Neuroreport* 1997; **8**: 1499-1502
- 41 **Giakoustidis D**, Papageorgiou G, Iliadis S, Kontos N, Kostopoulou E, Papachrestou A, Tsantilas D, Spyridis C, Takoudas D, Botsoglou N, Dimitriadou A, Giakoustidis E. Intramuscular administration of very high dose of alpha-tocopherol protects liver from severe ischemia/reperfusion injury. *World J Surg* 2002; **26**: 872-877
- 42 **Kirimlioglu V**, Karakayali H, Turkoglu S, Haberal M. Effect of resveratrol on oxidative stress enzymes in rats subjected to 70% partial hepatectomy. *Transplant Proc* 2008; **40**: 293-296
- 43 **Jang DS**, Kang BS, Ryu SY, Chang IM, Min KR, Kim Y. Inhibitory effects of resveratrol analogs on unopsonized zymosan-induced oxygen radical production. *Biochem Pharmacol* 1999; **57**: 705-712
- 44 **Shen SQ**, Zhang Y, Xiang JJ, Xiong CL. Protective effect of curcumin against liver warm ischemia/reperfusion injury in rat model is associated with regulation of heat shock protein and antioxidant enzymes. *World J Gastroenterol* 2007; **13**: 1953-1961
- 45 **Fernandez-Checa JC**, Garcia-Ruiz C, Colell A, Morales A, Mari M, Miranda M, Ardite E. Oxidative stress: role of mitochondria and protection by glutathione. *Biofactors* 1998; **8**: 7-11
- 46 **Jaeschke H**. Glutathione disulfide as index of oxidant stress in rat liver during hypoxia. *Am J Physiol* 1990; **258**: G499-G505
- 47 **Michiels C**, Raes M, Toussaint O, Remacle J. Importance of Se-glutathione peroxidase, catalase, and Cu/Zn-SOD for cell survival against oxidative stress. *Free Radic Biol Med* 1994; **17**: 235-248
- 48 **Plin C**, Tillement JP, Berdeaux A, Morin D. Resveratrol protects against cold ischemia-warm reoxygenation-induced damages to mitochondria and cells in rat liver. *Eur J Pharmacol* 2005; **528**: 162-168
- 49 **Leonard SS**, Xia C, Jiang BH, Stinefelt B, Klandorf H, Harris GK, Shi X. Resveratrol scavenges reactive oxygen species and effects radical-induced cellular responses. *Biochem Biophys Res Commun* 2003; **309**: 1017-1026
- 50 **Hassan-Khabbar S**, Cottart CH, Wendum D, Vibert F, Clot JP, Savouret JF, Conti M, Nivet-Antoine V. Postischemic treatment by trans-resveratrol in rat liver ischemia-reperfusion: a possible strategy in liver surgery. *Liver Transpl* 2008; **14**: 451-459