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# The biological responses to resveratrol and other polyphenols from alcoholic beverages

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# Abstract

Although excessive consumption of ethanol in alcoholic beverages causes multi-organ damage, moderate consumption, particularly of red wine, is protective against all-cause mortality. These protective effects could be due to one or many components of the complex mixture of bioactive compounds present in red wine including flavonols, monomeric and polymeric flavan-3-ols, highly coloured anthocyanins as well as phenolic acids and the stilbene polyphenol, resveratrol. The therapeutic potential of resveratrol, firstly in cancer chemoprevention and then later for cardioprotection, has stimulated many studies on the possible mechanisms of action. Further indications for resveratrol have been developed, including the prevention of age-related disorders such as neurodegenerative diseases, inflammation, diabetes and cardiovascular disease. These improvements are remarkably similar yet there is an important dichotomy: low doses improve cell survival as in cardio- and neuro-protection yet high doses increase cell death as in cancer treatment. Fewer studies have examined the responses to other components of red wine, but the results have, in general, been similar to resveratrol. If the non-alcoholic constitutents of red wine are to become therapeutic agents, their ability to get to the sites of action needs to be understood. This mini-review summarizes recent studies on the possible mechanisms of action, potential therapeutic uses and bioavailability of the non-alcoholic constitutents of alcoholic beverages, in particular resveratrol and other polyphenols.

# INTRODUCTION

Alcohol-related disorders account for an enormous part of the global mortality and morbidity (Rehm et al., 2003). Chronic alcohol abuse may lead to organ damage and carcinoma, especially to the liver and pancreas, but also damage to the brain (Harper, 2007) and immune system (Cook, 1998) as two of many examples. While the role of alcohol (ethanol) in the development of alcohol-related diseases has been intensively studied over the past three decades, fewer investigators have studied the role of non-alcoholic constituents. However, alcoholic beverages contain numerous non-alcoholic compounds. For example, more than 2,000 organic and inorganic compounds have been found in beer, and more than 1,000 in wine. At least on gastric acid secretion, it has been convincingly

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demonstrated that alcohol and non-alcoholic constituents of alcoholic beverages have markedly different effects (Teyssen et al. 1999).

On the other hand, epidemiological studies indicate that light to moderate consumption of alcoholic beverages reduces all-cause mortality (De Lorimier, 2000). In particular, red wine consumption is inversely associated with mortality due to cardiovascular diseases (Gronbaek, 2002), and the protective effects are observed for consumption levels up to 300 ml wine per day (Rotondo et al., 2001). The reductions in risk from low to moderate consumption of alcoholic beverages are greater for red wine compared to white wine, beer and most spirits (Gronbaek, 2002). The potential association between red wine consumption and risk of cardiovascular disease mortality has been highlighted by the `French Paradox' which refers to the finding that people in France suffer from a relatively low incidence of coronary heart disease, despite their diet being rich in saturated fats (Renaud & de Lorgeril, 1992). This concept has been intensely dissected with Ferrières (2004) concluding "that the promotion of primary prevention, based on an optimal diet rich in fruit and vegetables, regular physical exercise, and life without smoking, is worthwhile."

This phenomenon was first noted by the Irish physician Samuel Black (1819) who attributed the observation to the French habit of drinking red wine with meals, while the first scientific study was published by Cabot (1904), and the first comparison of coronary heart disease mortality between wine-drinking countries and those where beer or spirits were consumed was made 75 years later (St. Leger et al., 1979). The effect has since been ascribed to a number of red wine components including ethanol, which at low doses has beneficial effects on cardiovascular disease risk, and various polyphenols derived from the grape skins, seeds and stems or during maturation in oak barrels (Booyse et al., 2007).

Red wine contains a complex mixture of bioactive compounds that are predominantly phenolic in nature (Soleas et al, 1997). These include flavonols such as myricetin, kaempferol and the predominant quercetin, the flavan-3-ol monomers catechin and epicatechin, the oligomeric and polymeric flavan-3-ols or proanthocyanidins, various highly coloured anthocyanins, various phenolic acids (gallic acid, caftaric acid, caffeic acid, pcoumaric acid) and the stilbene resveratrol (figure 1). These compounds occur in red wine but are virtually absent from white wine because the skins, seeds and stems are present during the fermentation of red wine but not white wine. Red wine is one of the richest sources of polyphenols in human diets. Highly tannic red wines can contain up to 3 g of total polyphenols per litre, and moderate red wine drinkers will consume polyphenols at levels well above the population average (Waterhouse, 2002). Urinary resveratrol measurements have been proposed as a robust marker of wine intake (Zamora-Ros et al., 2009). Resveratrol is synthesized in the skins of grapes as a response to fungal infection. Once present, the compound acts as a phytoalexin, preventing the proliferation of pathogens. Rich natural sources of resveratrol include Polygonum cuspidatum (Japanese knot-weed, 0.524 mg/g, red wines (0.1 – 14.3 mg/l), red grape skins, berries such as blueberries, as well as peanuts and other nuts (Baur & Sinclair, 2006).

There is a substantial literature on the biological activities of phenolic compounds that are present in red wine. Much of the early interest centred on quercetin, a potent anti-oxidant that is present as a mixture of glycosides and the aglycone in red wine (Burns et al., 2000). Quercetin has been shown, *in vitro* and in animal models, to possess a range of biological activities relevant to protecting against cardiovascular diseases (Perez-Vizcaino et al., 2006) and against cancer (Birt et al., 2001; Moon et al., 2006a).

There has been considerable interest in resveratrol and other polyphenols in red wine in relation to human health. This mini-review summarizes the current knowledge about the

various beneficial effects of resveratrol and other polyphenols from wine in the prevention of ageing-related disorders such as cancer, neurodegenerative disorders, inflammation and cardiovascular disease; further, the possibility that resveratrol may be useful in the treatment of these diseases will be mentioned. A critical issue for resveratrol and other polyphenols as potential therapeutic agents is whether adequate concentrations can be achieved at the sites of action; thus, the bioavailability and efficacy of these compounds in humans are outlined at the end of this article.

#### Resveratrol: the evidence for potential benefits to human health

There is substantial evidence that resveratrol prevents or delays the onset of chronic diseases such as diabetes, inflammation, Alzheimer's disease and cardiovascular disease that increase in prevalence with age as well as inducing neuroprotection and inhibiting proliferation of human cancer cell lines (Aggarwal *et al*, 2004; Baur & Sinclair, 2006; Das & Maulik, 2006; Das & Das, 2007b; Opie & Lecour, 2007; Vidavalur *et al*, 2006; Harikumar & Aggarwal, 2008; Raval *et al*, 2008; Saiko *et al*, 2008). The breadth of the therapeutic potential of resveratrol is shown by the extension of the lifespan and improved motor function in mice fed a high-calorie diet (Baur *et al*, 2006). This impressive study indicates new approaches for treating obesity-related diseases and the diseases of ageing. The prevention or treatment of cardiovascular disease has been a major research theme in resveratrol studies. The potential benefits cover the range of cardiovascular problems, including myocardial infarction, arrhythmias, hypertension, hypertrophy, inflammation leading to fibrosis, atherosclerosis and thrombosis.

Resveratrol extends the lifespan of species such as yeast, worms and flies by activation of the silent information regulator 2 (SIR2), one of the sirtuins (Baur & Sinclair, 2006). The sirtuins as class III histone deacetylases regulate key pathways by removing acetyl groups from the  $\varepsilon$ -amino group of lysine to silence transcription. Multiple metabolic pathways are regulated by SIRT1, the mammalian analogue of SIR2, as this enzyme is involved in lifespan regulation, the cellular response to stress, glucose homeostasis, insulin secretion and lipid mobilization (Jiang, 2008). SIRT1 is a key regulator of vascular endothelial homeostasis controlling angiogenesis, vascular tone and endothelial dysfunction (Potente & Dimmler, 2008). The cardiac responses to low (2.5 fold) or moderate (7.5 fold) overexpression of SIRT1 in transgenic mouse hearts included attenuation of age-dependent increases in heart weight, apoptosis, fibrosis and cardiac dysfunction while high (12.5 fold) overexpression increased apoptosis, heart weight and decreased cardiac function (Alcendor *et al*, 2007).

While the pharmacological actions of resveratrol have been linked to antioxidant actions, the possible link between sirtuin activation and redox regulation by resveratrol is not yet clear. Hypoxia-induced apoptosis in embryonic rat heart-derived H9c2 cells was greatly decreased following treatment with resveratrol by activation of SIRT1 increasing the ability of FOXO1 (Forkhead box factors regulated by insulin/Akt) to induce cell cycle arrest (Chen et al, 2009). The ability of the FOXO transcription factor FOXO3 to induce cell cycle arrest and resistance to oxidative stress was increased by SIRT1 (Brunet et al, 2004). Moderate expression of SIRT1 protected the heart from oxidative stress and increased expression of antioxidant enzymes but high expression increased oxidative stress (Alcendor *et al*, 2007). Cilostazol, a selective inhibitor of phosphodiesterase III, dose-dependently increased SIRT1 expression to protect human endothelial cells after ischemic vascular damage through increased production of nitric oxide (Ota et al, 2008). These studies suggest that sirtuins are important in the resistance to oxidative stress induced by resveratrol, but the details need to be determined.

The breadth of the therapeutic potential for resveratrol hides a contradiction: how can the same compound improve the survival of cells as a cardioprotective or neuroprotective agent, yet kill cancer cells? Although the limited literature makes comparison of doses difficult, the general trend is that higher doses are needed to kill cancer cells than to improve cell survival. Possible reasons include dose-dependent effects, either at the level of the molecular mechanisms as described above for SIRT1, or by activation of different pathways at different resveratrol doses. In the heart, resveratrol at lower doses (5 mg/kg) activated survival signals by up-regulating the antiapoptotic and redox proteins Akt and Bcl-2 while higher doses of 25 mg/kg potentiated a death signal by down-regulating redox proteins and up-regulating proapoptotic proteins (Dudley et al, 2008). Although resveratrol is usually considered as an antioxidant, primarily by increasing NO bioavailability, resveratrol can also exhibit pro-oxidant properties in the presence of transition metal ions such as copper, leading to oxidative breakage of cellular DNA (Alarcón de la Lastra & Villegas, 2007). These authors suggest that this pro-oxidant action is the common mechanism for anticancer and chemopreventive properties of plant polyphenols.

#### **Resveratrol and chemoprevention**

The initial reports of anti-cancer responses to resveratrol led to the increased interest in this compound from 1992 (Baur & Sinclair, 2006). Resveratrol inhibited or retarded the growth of a wide range of human cancer cells in culture as well as implanted tumors usually in mice (Aggarwal et al, 2004; Athar et al, 2007). The compound inhibited experimental tumorigenesis in a wide range of animal models by targeting many components of intracellular signaling pathways including pro-inflammatory mediators, regulators of cell survival and apoptosis, and tumor angiogenic and metastatic switches by modulating a distinct set of upstream kinases, transcription factors and their regulators (Kundu & Surh, 2008; Pirola & Fröjdö, 2008). However, the role of sirtuin activation by resveratrol has not been defined. Khan et al. (2008) have presented the key findings from studies on the effects of dietary antioxidants including tea polyphenols, curcumin, genistein, resveratrol, lycopene, pomegranate and lupeol against cell lines from human skin, prostate, breast, lung and liver cancers. Many of the same compounds, including resveratrol, curcumin and epigallocatechin gallate, modulated the effects of deregulated cell cycle checkpoints, and this could contribute to the prevention of cancer (Meeran & Katiyar, 2008). However, it should be emphasized that these results are measurements of resveratrol responses on human cancer cells in culture, or taken as conclusions from epidemiological studies, rather than the results from clinical trials with cancer patients. No clinical trials with resveratrol in human cancers have been reported although 5 trials on human cancers are underway or finished (http://www.clinicaltrials.gov/ct2/results?term=resveratrol).

#### **Resveratrol and inflammation**

Inflammation is a pervasive cause of disease, implicated in diseases as diverse as metabolic syndrome (Dandona et al, 2005), rheumatoid arthritis and neurodegenerative disorders (Stolp & Dziegielewska, 2008). The cardioprotective and cancer preventive actions of resveratrol may be attributable to anti-inflammatory effects such as the inhibition of synthesis and release of pro-inflammatory mediators, modifications of eicosanoid synthesis, inhibition of some activated immune cells, or inhibiting enzymes such as cyclooxygenase-1 (COX-1) or cyclooxygenase-2 (COX-2) through to the inhibitory effect of resveratrol on transcription factors such as Nuclear Factor-kappaB (NF- $\kappa$ B) or Activator Protein-1 (AP-1) (Das & Das, 2007a; Udenigwe et al., 2008). Numerous studies have confirmed that resveratrol suppresses TNF- $\stackrel{\alpha}{.}$  activation (Das et. al., 2006b). Some preclinical as well as clinical studies have shown significant suppression of IL-1, IL-6 and IL-8 with resveratrol (Aggarwal et. al., 2004, Marier et. al., 2005, Donnelly et al., 2004). In an acute *in vivo* study, resveratrol inhibited the generation of adhesion molecules (Das et al., 2006b).

#### **Resveratrol and diabetes**

In rat models of diabetes, resveratrol reduced blood glucose and triglyceride concentrations following streptozotocin treatment (Su et al, 2006) and prevented the increase in blood pressure and cardiac hypertrophy while restoring the mesenteric and cardiac eNOS activity and decreasing oxidative stress in fructose-fed rats (Miatello *et al*, 2005). In obese Zucker rats, a genetic model of type-2 diabetes, resveratrol increased GLUT-4 expression and reduced endothelin expression and cardiac apoptosis in ischaemic-reperfused hearts (Lekli *et al*, 2008). Resveratrol increased insulin sensitivity by lowering the blood glucose level in high calorie diet mice with reduced insulin-like growth factor-1 (IGF-I) and increased AMP-activated protein kinase (AMPK) playing a major role (Baur et al, 2006). Resveratrol attenuated diabetic nephropathy in rats (Sharma et. al., 2006a), reduced thermal hyperalgesia and cold allodynia in streptozotocin-induced diabetic rats (Sharma et. al., 2006b) and attenuated diabetic neuropathic pain through changes in NO and TNF- $\alpha$  (Sharma et. al., 2007).

#### **Resveratrol and gastrointestinal diseases**

*Helicobacter pylori* causes peptic ulcer and gastrointestinal cancers. Cancer prevention requires control of the infection and the subsequent inflammatory processes. Plant products including resveratrol inhibited *H. pylori* colonization, decreased gastric inflammation by inhibiting cytokine and chemokine release, and repressed precancerous changes by inhibiting NF- $\kappa$ B DNA binding, inducing profuse levels of apoptosis and inhibiting mutagenesis (Lee et al., 2008).

#### **Resveratrol and neuroprotection**

The first evidence that resveratrol crosses the blood-brain barrier was published by Virgili & Contestabile (2000). In this study in young adult rats, chronic administration of resveratrol protected against the damage caused by the systemic injection of the excitotoxin kainic acid in the olfactory cortex and the hippocampus. Resveratrol effectively protected the brain from traumatic brain injury (Ates et al., 2006) and also inhibited the excitatory synaptic transmission in rat hippocampus (Gao et al., 2006). Resveratrol preconditioning and protection of brain cells *in vitro* from ischemic injury was mediated by SIRT1 (Raval et al., 2006). A similar study showed the neuroprotective effect of resveratrol by inhibiting the voltage-activated potassium currents in hippocampal neurons (Gao and Hu., 2005). In a study of spinal cord ischemia model, resveratrol-induced neuroprotection was mediated by decreased oxidative stress and increased NO release (Kiziltepe et al., 2004). Zamin et al. (2006) suggested that PI3-kinase/Akt pathway but not mitogen-activated protein kinase (MAPK) is involved in the neuroprotection.

#### Resveratrol and myocardial injury

Preconditioning describes the development of resistance to damage caused by prolonged ischemia and reperfusion to the heart (Bolli, 2007). Coordinated upregulation of iNOS– VEGF–KDR–eNOS by resveratrol has been suggested by Das et al (2005a) as the preconditioning mechanism in rat hearts. Resveratrol likely activates both adenosine A<sub>1</sub> and A<sub>3</sub> receptors which phosphorylate PI-3 kinase, which then phosphorylates protein kinase B (Akt) and thus preconditions the heart by producing NO as well as by the activation of antioxidant Bcl-2 (Das et al, 2005a, 2005b). Activation of adenosine A<sub>3</sub> receptors could also precondition the heart by a survival signal through the cAMP response element-binding protein (CREB) phosphorylation via PI-3 Kinase–Akt and via MEK (Mitogen-activated extracellular signal-regulated protein kinase)–CREB pathways (Das et al., 2005c). Recent studies have also demonstrated that NO can induce the expression of heme oxygenase-1. Das et al. (2006a) showed that tin protoporphyrin (SnPP), a heme oxygenase-1 inhibitor,

abolished the increased cardiac function parameters, reduced myocardial infarct size and decreased cardiomyocyte apoptosis that characterise the cardioprotection with resveratrol. The heme oxygenase-1 mediated mechanisms were related to the p38MAP kinase and Akt survival signaling, but independent of NF-κB activation. Being a polyphenol, resveratrol protects the heart by its anti-oxidative properties through various redox signalling mechanisms. The ability of resveratrol to modulate redox signalling has been extensively reviewed (Das & Maulik, 2006; Vidavalur et al, 2006) but the role of sirtuin activation in the heart is yet to be determined. White wine lacking polyphenols but containing antioxidant compounds such as caffeic acid and tyrosol mediated cardioprotection against ischemia-reperfusion injury in rat hearts by a similar survival pathway involving Akt/FOXO3a/NF-κB (Thirunavukkarasu et al, 2008).

#### **Resveratrol and hypertension**

The concept that reactive oxygen species contribute to the cause and the development of complications of hypertension is well supported by many clinical and experimental studies (Li & Shah, 2004). However, systolic blood pressure was not lowered by resveratrol in young stroke-prone spontaneously hypertensive rats (SHRSP) although oxidative damage to DNA and glyoxidative damage *in vivo* were decreased (Mizutani et al, 2001). Consistent with these findings, administration of quercetin in the diet of young SHR did not delay or lessen the onset or severity of cardiovascular complications, including hypertension (Carlstrom et al, 2007). In contrast, rats with abdominal aortic constriction showed anti-hypertensive and anti-hypertrophic responses to quercetin administered in the food, but without changes in vascular and myocardial function (Jalili et al, 2006).

#### **Resveratrol and angiogenesis**

The formation of new blood vessels is a potential mechanism to reduce ischemic damage to the heart. Ischemia induces VEGF release to encourage the development of collateral coronary circulation. Reactive oxygen species may have an important role in activating this process of myocardial angiogenesis (Maulik, 2006). Resveratrol enhanced angiogenesis both *in vivo* and *in vitro* by induction of VEGF which was regulated by thioredoxin-1 and heme oxygenase-1 (Kaga et al, 2005). This could be an important mechanism mediating cardioprotection in the chronic ischemic myocardium.

#### Resveratrol and the endothelium

Endothelial dysfunction is a characteristic finding in coronary heart disease. Resveratrol appears to protect endothelial cells at multiple targets. In a rat model of injured aorta, a lower dose of resveratrol (10 mg/kg) increased the proliferative, migrative and adhesive activities of endothelial progenitor cells, enhanced eNOS expression and accelerated the repair of the injured artery; however, a higher dose (50 mg/kg) had minimal effects (Gu et al, 2006). Chemokines in endothelial cells such as fractalkine are critical in the initiation, maintenance and resolution of inflammation. Resveratrol inhibited monocyte adhesion to human endothelial cells induced by TNF- $\stackrel{\alpha}{\cdot}$  and then suppressed arterial endothelial fractalkine expression in organs such as the heart (Moon et al, 2006b). The expression of the atherogenic cytokine, monocyte chemoattractant protein-1, was suppressed by about 45% at a resveratrol concentration of 50 µmol/l; this suppression was not altered by heme oxygenase-1 inhibition or knockdown indicating a distinct signalling pathway (Cullen et al., 2007). At submicromolar resveratrol concentrations, TNF-α-induced NF-κB activation, inflammatory gene expression and monocyte adhesiveness to human coronary arterial endothelial cells were inhibited; these anti-inflammatory actions on endothelial cells could lead to cardioprotection (Csiszar et al, 2006).

As the cells lining blood vessels, endothelial cells are sensitive to increased levels of reactive oxygen species in the blood, or produced in the vasculature by NADPH oxidase. The stimulation of this enzyme by oxidized LDL in vascular endothelial cells was inhibited by resveratrol by reducing the membrane association of two of the proteins in the active enzyme complex, gp91 (phox) and Rac1 (Chow et al, 2007). Resveratrol pre-treatment prevented the increase in caspase 3/7 activity produced by oxidized LDL and TNF- $^{\alpha}$  in endothelial cells; this effect was attenuated by inhibition of the antioxidant systems, glutathione peroxidase and heme oxygenase-1 (Ungvari et al., 2007). The oxidant species, peroxynitrite, induced apoptotic changes in aortic endothelial cells in culture; resveratrol concentrations from 1–25 µmol/l caused a concentration, transmigration and target cell adhesion following ischemiareperfusion are important in the development of graft rejection following transplantation. Following tissue allotransplantation, rats treated with resveratrol showed a moderate survival prolongation and reduction of lymphocytic infiltration and necrosis (Hsieh et al, 2007).

Flavanoids such as quercetin and wine polyphenols prevented endothelial dysfunction and reduced blood pressure, oxidative stress and end-organ damage in hypertensive animals, while flavanoid-rich foods improved endothelial function in cardiovascular patients (Perez-Vizcaino et al, 2006). An acute improvement in flow-mediated vasodilatation was measured after administration of a red grape extract containing epicatechin, catechin, resveratrol, rutin and other polyphenols to patients with coronary heart disease (Lekakis et al, 2005).

#### **Resveratrol and collagen deposition**

Resveratrol inhibited cardiac fibroblast proliferation and differentiation to the hypersecretory myofibroblast phenotype in rat cardiac fibroblasts in culture; these are two critical steps in cardiac collagen deposition (Olson et al, 2005). Fibrosis is the final result of an inflammatory challenge and thus inflammation plays a key role in organ damage in chronic cardiovascular disease. Resveratrol interacts at multiple sites within the inflammatory pathways to decrease inflammation (Das & Das, 2007a).

Multiple actions of dietary polyphenols have been proposed for reducing inflammation as well as in the processing of lipids in the body to improve the blood lipid profile to eventually reduce the risk of coronary heart disease (Zern & Fernandez, 2005). The processes of initiation, progression and rupture of atherosclerotic plaques offer many targets to delay or prevent cardiovascular disease. There is much experimental evidence that the phenolic compounds in wine alter these processes as part of their cardioprotective actions (Szmitko & Verma, 2005).

#### **Resveratrol and blood coagulation**

Tissue factor (TF) is the primary initiator of the blood coagulation cascade. Thrombus formation at the site of complicated atherosclerotic lesions following exposure to TF is a major cause of death in industrialized countries. In human umbilical vein endothelial cells and mononuclear cells, resveratrol and quercetin reduced the expression of TF mRNA by

bacterial lipopolysaccharide, interleukin-1<sup> $\beta$ </sup> or TNF-<sup> $\alpha$ </sup> (Di Santo et al, 2003). Several resveratrol derivatives were 2–10-fold more effective in inhibiting TF production in human peripheral blood mononuclear cells than the parent compound (Kaur et al, 2007).

The prevention of excessive or inappropriate aggregation of platelets decreases thrombus formation, reduces the subsequent blockage of small blood vessels and thereby reduces the risk of stroke and myocardial infarction. Many studies, reviewed by Baur & Sinclair (2006),

have shown that resveratrol prevents platelet aggregation *in vitro*, probably by selective inactivation of cyclooxygenase-1.

One of the potential mechanisms to explain this wide range of cardioprotective and vascular protective actions is the removal of highly reactive free radicals such as superoxide following increased expression of heme oxygenase-1 (Huang et al, 2005; Juan et al, 2005). Low concentrations of resveratrol (1–10  $\mu$ mol/l) likely to be physiologically relevant increased heme oxygenase-1 expression, at least partly through actions on the NF- $\kappa$ B pathway in human aortic smooth muscle cells in culture (Huang et al, 2005; Juan et al, 2005). However, the increased expression of heme oxygenase-1 has been attributed to regulation by MAP kinase and Akt survival signalling in the rat heart, not by activation of the NF- $\kappa$ B pathway (Das et al, 2006a).

#### Bioavailability and bioefficacy of resveratrol and other polyphenols in humans

At high doses, orders of magnitude higher than would be achievable through red wine consumption by humans, orally ingested resveratrol is also able to extend lifespan in a number of organisms ranging from mono-cellular yeasts to animals such as mice. These findings are remarkable, with lifespan extension of greater than 50% reported for certain species of yeast, nematode, insect and fish (Howitz et al., 2003; Wood et al., 2004; Valenzano et al., 2006). It has also been demonstrated that high doses of resveratrol can prevent the detrimental effects of a high fat diet in a mouse model of obesity (Baur et al., 2006), another truly remarkable observation with wide-ranging implications for mankind considering the obesity epidemic that is sweeping the western world. But, it must be recognized that all these observations have been made with doses of resveratrol that are well above those achievable in humans though normal diet. Red wine is almost the only source of resveratrol in human diets, and the richest red wines contain about 7 mg resveratrol per litre. To achieve the equivalent dose of resveratrol that was fed to the fish in order to achieve a >50% lifespan extension (Valenzano et al., 2006) and to mice in order to prevent the detrimental effects of a high fat diet (Baur et al., 2006), one would need to consume around 60 litres per day of high resveratrol red wine, which is clearly not feasible. Therefore, these potential effects of resveratrol remain achievable through pharmaceutical means only and not through dietary means. It is interesting to note that other dietary components have been shown to be effective in extending lifespan, including blueberry polyphenols (Wilson et al., 2006).

Flavan-3-ols, or the catechins and their oligo- and poly-meric derivatives the proanthocyanidins, have also attracted considerable interest due to their ability to improve endothelial function, vascular tone and platelet reactivity *in vivo*. In particular, ingestion of cocoa flavan-3-ols has been shown, consistently, to improve vascular function (Schroeter et al., 2006; Heiss et al., 2007) and platelet reactivity (Rein et al., 2000). These observations have been consistent across a number of studies and the required dose of epicatechin has been estimated at 1–2 mg per kg body weight 18 (Schroeter et al., 2006). The concentration of (–)-epicatechin in red wine is in the range of 20–200  $\mu$ M but (+)-catechin is present at 200–500  $\mu$ M (Burns et al., 2000).

#### Absorption from the gut

The ability of a dietary component to influence vascular function and prevent cardiovascular disease is dependent on how much of the oral dose is absorbed from the gut, the influence of metabolism during the first pass (small intestine and liver) and the subsequent exposure that is a function of the excretion rate which is reflected in the plasma half life. It is possible that some wine compounds may accumulate in certain tissues and hence increase the exposure at these specific sites. Although there is some evidence that certain polyphenols accumulate to

higher concentrations in certain body tissues compared to others, there is considerably more evidence (from animal experiments) showing that dietary polyphenols do not become concentrated in specific tissues, besides those of the gut. Nevertheless, there is some evidence that resveratrol accumulates in a time- and dose-dependent manner in the liver (Vitaglione et al., 2008) and brain (El-Mohsen et al., 2006) of rats intragastrically administered resveratrol over a period of days, while another report indicates preferential fixation of <sup>14</sup>C-resveratrol in the organs of absorption and elimination (stomach, intestine, liver, kidney) (Vitrac et al., 2003).

Polyphenols are only partially absorbed and only a fraction is excreted via the urine in humans. The plasma concentrations achieved and the urinary yield as a function of oral dose depends on a number of factors, but by far the most important is the structure of the polyphenol. Almost without exception, phenolics in plants are glycosylated, and this is true of most plant-derived foods and beverages also. Glycosylation can have an enormous influence on the efficiency of absorption and the site of absorption - monoglucoside derivatives are preferentially absorbed from the small intestine and are better absorbed than other glycosides such as rhamnoglucosides, that are absorbed later (probably from the colon) and to a lesser extent. The enhanced absorption of monoglucosides from the small intestine is due to human small intestinal lactase enzyme that has the capacity to hydrolyse flavonoid glucosides and which is required for absorption and metabolism of these compounds (Németh et al., 2003). Some polyphenols are quite well absorbed and a significant proportion is excreted in the urine; examples include gallic acid, the isoflavones genistein and daidzein, and the (citrus) flavanones hesperetin and naringenin (Manach et al., 2005). However, for others such as anthocyanins, much less than 1% of the oral dose is recovered in urine and plasma concentrations peak in the low nanomolar range (Manach et al., 2005). To date, the highest concentrations of polyphenols observed in plasma following oral ingestion by humans have been around  $1-10 \,\mu$ M. These concentrations are only observed for relatively large doses of polyphenols from supplements or polyphenol-rich foods and beverages that contain the more bioavailable flavonoids, such as monomeric flavan-3-ols, isoflavones and flavanones.

#### Metabolism during the first pass

Flavonoids and other polyphenols are extensively metabolized during absorption from the gut, and the typical products are phase-2 conjugates (i.e. glucuronidated, sulfated and methylated derivatives of the parent polyphenol (Kroon et al., 2004)). Indeed, for many polyphenols, the conjugation process is so efficient that the aglycone is not detectable in plasma and urine (Donovan et al., 2006). The first pass includes the small intestine and the liver, and both are capable of metabolizing polyphenols (Manach et al., 2004). The kidney also has significant capacity to modify polyphenols but there is little evidence to support a major role for this organ, probably because it is precluded from contributing by the efficiency of the small intestine and liver in these activities. The major plasma and urinary forms resulting from ingestion of different polyphenols have been summarized by Kroon et al. (2004). Many red wine polyphenols (flavonols, catechin / epicatechin, resveratrol, phenolic acids) are efficiently conjugated and are present in plasma predominantly as phase-2 conjugated metabolites. For example, following oral ingestion, 100% of quercetin and virtually all resveratrol is present as glucuronidated and sulfated conjugates of the aglycone or a methylated derivative (Day et al., 2001; Vitaglione et al., 2005) (see Figure 2).

The flavanol monomers, (+)-catechin and (-)-epicatechin, are also conjugated during absorption. The major products are glucuronidated and sulfated conjugates of the parent aglycone or methylated derivatives (Kroon et al., 2004). However, anthocyanins (the brightly coloured components responsible for the red colour of wine) are often not conjugated during absorption, and in this respect, they are rather unique. Although the

mechanisms of absorption for anthocyanins are not well understood, the fact that they appear in plasma un-metabolized and at extremely low concentrations, and that the urinary yield typically is extremely low (Manach et al., 2005), may indicate that these compounds escape intestinal metabolism by passing between the enterocytes rather than through them.

#### What are effects of metabolism on biological activity?

Due to the lack of commercially-available standards, and the technical difficulties associated with their synthesis, there are only a limited number of reports concerned with the biological activities of authentic polyphenol phase-2 conjugates. Readers are referred to Kroon et al. (2004) and Williamson et al. (2005) that discuss much of the available data. Many of the studies concerned with the effects of conjugation on biological activity have focussed on quercetin, the conjugates of which have been synthesized chemically (Needs & Kroon, 2006). For example, conjugation has been shown to detrimentally affect the ability of quercetin to inhibit xanthine oxidase and lipoxygenase enzyme activities in vitro, although the loss of potency was heavily dependent on the position of conjugation; the 3'- and 4'glucuronides were only slightly less effective inhibitors of xanthine oxidase (Day et al., 2000). Quercetin glucuronides have also been shown to inhibit the N-acetylation of the arylamine carcinogen 2-aminofluorene by human acute myeloid leukaemia (HL-60) cells (Kuo et al., 2002), inhibit lung cancer cell growth via cell cycle arrest and induction of apoptosis (Yang et al., 2006), prevent angiotensin-II-induced vascular cell hypertrophy in cultured rat aortic smooth muscle cells via inhibition of JNK and AP-1 signalling pathways (Yoshizumi et al., 2002) and down-regulate transcription of human cyclooxygenase-2 (COX-2) (O'Leary et al., 2004; de Pascual-Teresa et al., 2004). In contrast, Donnini et al. (2006) provide evidence that whereas quercetin and quercetin-3-glucuronide inhibited vascular endothelial growth factor (VEGF)-induced changes in cell functions and angiogenesis, quercetin-3'-sulfate promoted cell proliferation and angiogenesis. Therefore, phase-2 conjugation is an important factor to consider when assessing the potential biological activities of polyphenols, as the effects can be both profound and unpredictable. To date, there are no reports documenting the relative biological activity of resveratrol phase-2 conjugates, even though these represent the predominant forms in vivo following resveratrol ingestion.

In summary, further research is required to understand the role of resveratrol and other polyphenols in providing the additional benefits of consumption of low to moderate amounts of red wine, compared to white wine, beer or spirits. However, the various risks associated with drinking alcoholic beverages has so far prevented a fully controlled clinical trial to determine the effects of moderate red wine consumption on cardiovascular disease risk.

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#### REFERENCES

- Aggarwal BB, Bhardwaj A, Aggarwal RS, Seeram NP, Shishodia S, Takada Y. Role of resveratrol in prevention and therapy of cancer: preclinical and clinical studies. Anticancer Res. 2004; 24:2783–2840. [PubMed: 15517885]
- Alarcón de la Lastra CA, Villegas I. Resveratrol as an antioxidant and prooxidant agent: mechanisms and clinical implications. □Biochem Soc Trans. 2007; 35:1156–60. [PubMed: 17956300]
- Alcendor RR, Gao S, Zhai P, Zablocki D, Holle E, Yu X, Tian B, Wagner T, Vatner SF, Sadoshima J. Sirt1 regulates aging and resistance to oxidative stress in the heart. Circ Res. 2007; 100:1512–1521. [PubMed: 17446436]

- Ates O, Cayli S, Altinoz E, Gurses I, Yucel N, Sener M, Kocak A, Yologlu S. Neuroprotection by resveratrol against traumatic brain injury in rats. Mol Cell Biochem. 2007; 294:137–144. [PubMed: 16924419]
- Athar M, Back JH, Tang X, Kim KH, Kopelovich L, Bickers DR, Kim AL. Resveratrol: a review of pre-clinical studies for human cancer prevention. Toxicol Appl Pharmacol. 2007; 224:274–283. [PubMed: 17306316]
- Baur JA, Pearson KJ, Price NL, Jamieson HA, Lerin C, Kalra A, Prabhu VV, Allard JS, Lopez-Lluch G, Lewis K, Pistell PJ, Poosala S, Becker KG, Boss O, Gwinn D, Wang M, Ramaswamy S, Fishbein KW, Spencer RG, Lakatta EG, Le Couteur D, Shaw RJ, Navas P, Puigserver P, Ingram DK, de Cabo R, Sinclair DA. Resveratrol improves health and survival of mice on a high-calorie diet. Nature. 2006; 444:337–342. [PubMed: 17086191]
- Baur JA, Sinclair DA. Therapeutic potential of resveratrol: the in vivo evidence. Nat Rev Drug Discov. 2006; 5:493–506. [PubMed: 16732220]
- Birt DF, Hendrich S, Wang W. Dietary agents in cancer prevention: flavonoids and isoflavonoids. Pharmacol Ther. 2001; 90:157–177. [PubMed: 11578656]
- Black, S. Clinical and pathological reports. Alex Wilkinson; Newry, England: 1819. p. 1-47.
- Booyse FM, Pan W, Grenett HE, Parks DA, Darley-Usmar VM, Bradley KM, Tabengwa EM. Mechanism by which alcohol and wine polyphenols affect coronary heart disease risk. Ann Epidemiol. 2007; 17(Suppl):S24–31. [PubMed: 17478321]
- Bolli R. Preconditioning: a paradigm shift in the biology of myocardial ischemia. Am J Physiol Heart Circ Physiol. 2007; 292:H19–27. [PubMed: 16963615]
- Brito PM, Mariano A, Almeida LM, Dinis TCP. Resveratrol affords protection against peroxynitritemediated endothelial cell death: a role for intracellular glutathione. Chem-Biol Inter. 2006; 164:157–166.
- Brunet A, Sweeney LB, Sturgill JF, Chua KF, Greer PL, Lin Y, Tran H, Ross SE, Mostoslavsky R, Cohen HY, Hu LS, Cheng HL, Jedrychowski MP, Gygi SP, Sinclair DP, Alt FW, Greenberg ME. Stress-dependent regulation of FOXO transcription factors by SIRT1 deacetylase. Science. 2004; 303:2011–2015. [PubMed: 14976264]
- Burns J, Gardner PT, O'Neil J, Crawford S, Morecroft I, McPhail DB, Lister C, Matthews D, MacLean MR, Lean ME, Duthie GG, Crozier A. Relationship among antioxidant activity, vasodilation capacity, and phenolic content of red wines. J Agric Food Chem. 2000; 48:220–230. [PubMed: 10691619]
- Cabot RC. The relation of alcohol to arteriosclerosis. J Am Med Assoc. 1904; 43:774-775.
- Carlstrom J, Symons JD, Wu TC, Bruno RS, Litwin SE, Jalili T. A quercetin supplemented diet does not prevent cardiovascular complications in spontaneously hypertensive rats. J Nutr. 2007; 137:628–633. [PubMed: 17311951]
- Chen C-J, Yu W, Fu Y-C, Wang X, Li J-L, Wang W. Resveratrol protects cardiomyocytes from hypoxia-induced apoptosis through the SIRT1-FoxO1 pathway. Biochem Biophys Res Commun. 2009; 378:389–393. [PubMed: 19059213]
- Chow SE, Hshu YC, Wang JS, Chen JK. Resveratrol attenuates ox-LDL-stimulated NADPH oxidase activity and protects endothelial cells from oxidative functional damages. J Appl Physiol. 2007; 102:1520–1527. [PubMed: 17194732]
- Cook RT. Alcohol abuse, alcoholism, and damage to the immune system a review. Alcoholism: Clinical & Experimental Research. 1998; 22:1927–1942.
- Csiszar A, Smith K, Labinskyy N, Orosz Z, Rivera A, Ungvari Z. Resveratrol attenuates TNF-αinduced activation of coronary arterial endothelial cells: role of NF-κB inhibition. Am J Physiol Heart Circ Physiol. 2006; 291:H1694–H1699. [PubMed: 16973825]
- Cullen JP, Morrow D, Jin Y, Curley B, Robinson A, Sitzmann JV, Cahill PA, Redmond EM. Resveratrol, a polyphenolic phytostilbene, inhibits endothelial monocyte chemotactic protein-1 synthesis and secretion. J Vasc Res. 2007; 44:75–84. [PubMed: 17191021]
- Dandona P, Aljada A, Chaudhuri A, Mohanty P, Garg R. Metabolic syndrome: a comprehensive perspective based on interactions between obesity, diabetes, and inflammation. Circulation. 2005; 111:1448–54. [PubMed: 15781756]

- Das S, Alagappan VK, Bagchi D, Sharma HS, Maulik N, Das DK. Coordinated induction of iNOS-VEGF-KDR-eNOS after resveratrol consumption: a potential mechanism for resveratrol preconditioning of the heart. Vascul Pharmacol. 2005a; 42:281–289. [PubMed: 15905131]
- Das S, Cordis GA, Maulik N, Das DK. Pharmacological preconditioning with resveratrol: role of CREB-dependent Bcl-2 signaling via adenosine A3 receptor activation. Am J Physiol Heart Circ Physiol. 2005b; 288:H328–H335. [PubMed: 15345477]
- Das S, Tosaki A, Bagchi D, Maulik N, Das DK. Resveratrol-mediated activation of cAMP response element-binding protein through adenosine A3 receptor by Akt-dependent and -independent pathways. J Pharmacol Exp Ther. 2005c; 314:762–769. [PubMed: 15879002]
- Das S, Fraga CG, Das DK. Cardioprotective effect of resveratrol via HO-1 expression involves p38 map kinase and PI-3-kinase signaling, but does not involve NFkappaB. Free Radic Res. 2006a; 40:1066–1075. [PubMed: 17015251]
- Das S, Falchi M, Bertelli A, Maulik N, Das DK. Attenuation of ischemia/reperfusion injury in rats by the anti-inflammatory action of resveratrol. Arzneimittel Forschung Drug Res. 2006b; 56:700– 706. [PubMed: 17225566]
- Das S, Das DK. Anti-inflammatory responses of resveratrol. Inflamm Allergy Drug Targets. 2007a; 6:168–173. [PubMed: 17897053]
- Das S, Das DK. Resveratrol: a therapeutic promise for cardiovascular disease. Recent Patents on Cardiovascular Drug Discovery. 2007b; 2:133–138. [PubMed: 18221111]
- Das DK, Maulik N. Resveratrol in cardioprotection: a therapeutic promise of alternative medicine. Mol Interv. 2006; 6:36–47. [PubMed: 16507749]
- Day AJ, Bao Y, Morgan MR, Williamson G. Conjugation position of quercetin glucuronides and effect on biological activity. Free Radic Biol Med. 2000; 29:1234–1243. [PubMed: 11118813]
- Day AJ, Mellon F, Barron D, Sarrazin G, Morgan MR, Williamson G. Human metabolism of dietary flavonoids: identification of plasma metabolites of quercetin. Free Radic Res. 2001; 35:941–952. [PubMed: 11811545]
- de Lorimier AA. Alcohol, wine, and health. Am J Surg. 2000; 180:357–361. [PubMed: 11137687]
- de Pascual-Teresa S, Johnston KL, DuPont MS, O'Leary KA, Needs PW, Morgan LM, Clifford MN, Bao Y, Williamson G. Quercetin metabolites downregulate cyclooxygenase-2 transcription in human lymphocytes ex vivo but not in vivo. J Nutr. 2004; 134:552–557. [PubMed: 14988445]
- Di Santo A, Mezzetti A, Napoleone E, Di Tommaso R, Donati MB, De Gaetano G, Lorenzet R. Resveratrol and quercetin down-regulate tissue factor expression by human stimulated vascular cells. J Thromb Haemost. 2003; 1:1089–1095. [PubMed: 12871381]
- Donnelly LE, Newton R, Kennedy GE, Fenwick PS, Fenwick PS, Leung RH, Ito K, Russell RE, Barnes PJ. Anti-inflammatory effects of resveratrol in lung epithelial cells: molecular mechanisms. Am J Physiol Lung Cell Mol Physiol. 2004; 287:L774–L783. [PubMed: 15180920]
- Donnini S, Finetti F, Lusini L, Morbidelli L, Cheynier V, Barron D, Williamson G, Waltenberger J, Ziche M. Divergent effects of quercetin conjugates on angiogenesis. Br J Nutr. 2006; 95:1016– 1023. [PubMed: 16611395]
- Donovan, JL.; Manach, C.; Faulks, RM.; Kroon, PA. Absorption and metabolism of plant secondary metabolites. In: Crozier, A.; Clifford, MN.; Ashihara, H., editors. Plant Secondary Metabolites: Occurrence, Structure and Role in the Human Diet. Blackwell Publishing; Oxford, UK: 2006. p. 303-351.
- Dudley J, Das S, Mukherjee S, Das DK. Resveratrol, a unique phytoalexin present in red wine, delivers either survival signal or death signal to the ischemic myocardium depending on dose. J Nutr Biochem. 2008 in press, doi:10.1016/j.jnutbio.2008.05.003.
- Abd El-Mohsen M, Bayele H, Kuhnle G, Gibson G, Debnam E, Kaila Srai S, Rice-Evans C, Spencer JP. Distribution of [3H]trans-resveratrol in rat tissues following oral administration. Br J Nutr. 2006; 96:62–70. [PubMed: 16869992]
- Ferrières J. The French paradox: lessons for other countries. Heart. 2004; 90:107–111. [PubMed: 14676260]
- Gao ZB, Hu GY. Trans-resveratrol, a red wine ingredient, inhibits voltage-activated potassium currents in rat hippocampal neurons. Brain Res. 2005; 1056:68–75. [PubMed: 16112093]

- Gao ZB, Chen XQ, Hu GY. Inhibition of excitatory synaptic transmission by trans-resveratrol in rat hippocampus. Brain Res. 2006; 1111:41–47. [PubMed: 16876771]
- Gronbaek M. Alcohol, type of alcohol, and all-cause and coronary heart disease mortality. Ann N Y Acad Sci. 2002; 957:16–20. [PubMed: 12074958]
- Gu J, Wang CQ, Fan HH, Ding HY, Xie XL, Xu YM, Wang BY, Huang DJ. Effects of resveratrol on endothelial progenitor cells and their contributions to reendothelialization in intima-injured rats. J Cardiovasc Pharmacol. 2006; 47:711–721. [PubMed: 16775512]
- Harikumar KB, Aggarwal BB. Resveratrol. A multitargeted agent for age-associated chronic diseases. Cell Cycle. 2008; 7:1020–1035. [PubMed: 18414053]
- Harper C. The neurotoxicity of alcohol. Human & Experimental Toxicology. 2007; 26:251–257. [PubMed: 17439928]
- Heiss C, Finis D, Kleinbongard P, Hoffmann A, Rassaf T, Kelm M, Sies H. Sustained increase in flow-mediated dilation after daily intake of high-flavanol cocoa drink over 1 week. J Cardiovasc Pharmacol. 2007; 49:74–80. [PubMed: 17312446]
- Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, Zipkin RE, Chung P, Kisielewski A, Zhang LL, Scherer B, Sinclair DA. Small molecule activators of sirtuins extend Saccharomyces cerevisiae lifespan. Nature. 2003; 425:191–196. [PubMed: 12939617]
- Hsieh Y-H, Huang S-S, Wei F-C, Huang L-M. Resveratrol attenuates ischemiareperfusion-induced leukocyte-endothelial cell adhesive interactions and prolongs allograft survival across the MHC barrier. Circ J. 2007; 71:423–428. [PubMed: 17322646]
- Huang H-M, Liang Y-C, Cheng T-H, Chen C-H, Juan S-H. Potential mechanism of blood vessel protection by resveratrol, a component of red wine. Ann NY Acad Sci. 2005; 1042:349–356. [PubMed: 15965080]
- Jalili T, Carlstrom C, Kim S, Freeman D, Jin H, Wu T-C, Litwin SE, Symons JD. Quercetinsupplemented diets lower blood pressure and attenuate cardiac hypertrophy in rats with aortic constriction. J Cardiovasc Pharmacol. 2006; 47:531–541. [PubMed: 16680066]
- Jiang W-J. Sirtuins: Novel targets for metabolic disease in drug development. Biochem Biophys Res Commun. 2008; 373:341–344. [PubMed: 18577374]
- Juan S-H, Cheng T-H, Lin H-C, Chu Y-L, Lee W-S. Mechanism of concentration-dependent induction of heme oxygenase-1 by resveratrol in human aortic smooth muscle cells. Biochem Pharmacol. 2005; 69:41–48. [PubMed: 15588712]
- Kaga S, Zhan L, Matsumoto M, Maulik N. Resveratrol enhances neovascularization in the infarcted rat myocardium through the induction of thioredoxin-1, heme oxygenase-1 and vascular endothelial growth factor. J Mol Cell Cardiol. 2005; 39:813–822. [PubMed: 16198371]
- Kaldas MI, Walle UK, Walle T. Resveratrol transport and metabolism by human intestinal Caco-2 cells. J Pharm Pharmacol. 2003; 55:307–312. [PubMed: 12724035]
- Kaur G, Roberti M, Raul F, Pendurthi UR. Suppression of human monocyte tissue factor induction by red wine phenolics and synthetic derivatives of resveratrol. Thromb Res. 2007; 119:247–256. [PubMed: 16507316]
- Khan N, Afaq F, Mukhtar H. Cancer chemoprevention through dietary antioxidants: progress and promise. Antiox. Redox Signal. 2008; 10:475–510.
- Kiziltepe U, Turan NN, Han U, Ulus AT, Akar F. Resveratrol, a red wine polyphenol, protects spinal cord from ischemia-reperfusion injury. J Vasc Surg. 2004; 40:138–145. [PubMed: 15218474]
- Kroon PA, Clifford MN, Crozier A, Day AJ, Donovan JL, Manach C, Williamson G. How should we assess the effects of exposure to dietary polyphenols in vitro? Am J Clin Nutr. 2004; 80:15–21. [PubMed: 15213022]
- Kundu JK, Surh Y-J. Cancer chemopreventive and therapeutic potential of resveratrol: Mechanistic perspectives. Cancer Letters. 2008; 269:243–261. [PubMed: 18550275]
- Kuo HM, Ho HJ, Chao PD, Chung JG. Quercetin glucuronides inhibited 2-aminofluorene acetylation in human acute myeloid HL-60 leukemia cells. Phytomedicine. 2002; 9:625–631. [PubMed: 12487326]
- Lee SY, Shin YW, Hahm KB. Phytoceuticals: mighty but ignored weapons against *Helicobacter pylori* infection. J Dig Dis. 2008; 9:129–139. [PubMed: 18956590]

- Lekakis J, Rallidis LS, Andreadou I, Vamvakou G, Kazantzoglou G, Magiatis P, Skaltsounis A-L, Kremastinos DT. Polyphenolic compounds from red grapes acutely improve endothelial function in patients with coronary heart disease. Eur J Cardiovasc Prev Rehab. 2005; 12:596–600.
- Lekli I, Szabo G, Juhasz B, Das S, Das M, Varga E, Szendrei L, Gesztelyi R, Varadi J, Bak I, Das DK, Tosaki A. Protective mechanisms of resveratrol against ischemia-reperfusion-induced damage in hearts obtained from Zucker obese rats: the role of GLUT-4 and endothelin. Am J Physiol Heart Circ Physiol. 2008; 294:H859–H866. [PubMed: 18065527]
- Li J-M, Shah AM. Endothelial cell superoxide generation: regulation and relevance for cardiovascular pathophysiology. Am J Physiol Regul Integr Comp Physiol. 2004; 287:R1014–R1030. [PubMed: 15475499]
- Manach C, Williamson G, Morand C, Scalbert A, Remesy C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. Am J Clin Nutr. 2005; 81:230S– 242S. [PubMed: 15640486]
- Marier JF, Chen K, Prince P, Scott G, del Castillo JR, Vachon P. Production of ex vivo lipopolysaccharide-induced tumor necrosis factor-alpha, interleukin-1beta, and interleukin-6 is suppressed by trans-resveratrol in a concentration-dependent manner. Can J Vet Res. 2005; 69:151–154. [PubMed: 15971681]
- Maulik N. Reactive oxygen species drives myocardial angiogenesis? Antioxid Redox Signal. 2006; 8:2161–2168. [PubMed: 17034358]
- Meeran SM, Katiyar SK. Cell cycle control as a basis for cancer chemoprevention through dietary agents. Front Biosci. 2008; 13:2191–2202. [PubMed: 17981702]
- Miatello R, Vázquez M, Renna N, Cruzado M, Ponce Zumino A, Risler N. Chronic administration of resveratrol prevents biochemical cardiovascular changes in fructose-fed rats. Am J Hypertens. 2005; 18:864–870. [PubMed: 15925749]
- Mizutani K, Ikeda K, Kawai Y, Yamori Y. Protective effect of resveratrol on oxidative damage in male and female stroke-prone spontaneously hypertensive rats. Clin Exp Pharmacol Physiol. 2001; 28:55–59. [PubMed: 11153537]
- Moon S-O, Kim W, Sung MJ, Lee S, Kang KP, Kim DH, Lee SY, So J-N, Park SK. Resveratrol suppresses tumor necrosis factor-α-induced fractalkine expression in endothelial cells. Mol Pharmacol. 2006a; 70:112–119. [PubMed: 16614140]
- Moon YJ, Wang X, Morris ME. Dietary flavonoids: effects on xenobiotic and carcinogen metabolism. Toxicol in vitro. 2006b; 20:187–210. [PubMed: 16289744]
- Needs PW, Kroon PA. Convenient syntheses of metabolically important quercetin glucuronides and sulfates. Tetrahedron. 2006; 62:6862–6868.
- Németh K, Plumb GW, Berrin JG, Juge N, Jacob R, Naim HY, Williamson G, Swallow DM, Kroon PA. Deglycosylation by small intestinal epithelial cell beta-glucosidases is a critical step in the absorption and metabolism of dietary flavonoid glycosides in humans. Eur J Nutr. 2003; 42:29–42. [PubMed: 12594539]
- O'Leary KA, de Pascual-Tereasa S, Needs PW, Bao YP, O'Brien NM, Williamson G. Effect of flavonoids and vitamin E on cyclooxygenase-2 (COX-2) transcription. Mutat Res. 2004; 551:245– 254. [PubMed: 15225597]
- Olson ER, Naugle JE, Zhang X, Bomser JA, Meszaros JG. Inhibition of cardiac fibroblast proliferation and myofibroblast differentiation by resveratrol. Am J Physiol. 2005; 288:H1131–H1138.
- Opie LH, Lecour S. The red wine hypothesis: from concepts to protective signalling molecules. Eur Heart J. 2007; 28:1683–1693. [PubMed: 17561496]
- Ota H, Eto M, Kano MR, Ogawa S, Iijima K, Akishita M, Ouchi Y. Cliostazol inhibits oxidative stress-induced premature senescence via upregulation of Sirt1 in human endothelial cells. Arterioscler Thromb Vasc Biol. 2008; 28:1634–1639. [PubMed: 18556572]
- Perez-Vizcaino F, Duarte J, Andriantsitohaina R. Endothelial function and cardiovascular disease: effects of quercetin and wine polyphenols. Free Radic Res. 2006; 40:1054–1065. [PubMed: 17015250]
- Pirola L, Fröjdö S. Resveratrol: one molecule, many targets. IUBMB Life. 2008; 60:323–332. [PubMed: 18421779]

- Potente M, Dimmeler S. Emerging roles of SIRT1 in vascular endothelial homeostasis. Cell Cycle. 2008; 7:2117–2122. [PubMed: 18641460]
- Raval AP, Dave KR, Perez-Pinzon MA. Resveratrol mimics ischemic preconditioning in the brain. J Cereb Blood Flow Metab. 2006; 26:1141–1147. [PubMed: 16395277]
- Raval AP, Lin HW, Dave KR, DeFazio RA, Della Morte D, Kim EJ, Perez-Pinzon MA. Resveratrol and ischemic preconditioning in the brain. Current Medicinal Chemistry. 2008; 15:1545–1551. [PubMed: 18537630]
- Rehm J, Gmel G, Sempos CT, Trevisan M. Alcohol-related morbidity and mortality. Alcohol Res Health. 2003; 27:39–51. [PubMed: 15301399]
- Rein D, Paglieroni TG, Pearson DA, Wun T, Schmitz HH, Gosselin R, Keen CL. Cocoa and wine polyphenols modulate platelet activation and function. J Nutr. 2000; 130:2120S–2126S. [PubMed: 10917933]
- Renaud S, de Lorgeril M. Wine, alcohol, platelets, and the French paradox for coronary heart disease. Lancet. 1992; 339:1523–1526. [PubMed: 1351198]
- Rotondo S, Di Castelnuovo A, de Gaetano G. The relationship between wine consumption and cardiovascular risk: from epidemiological evidence to biological plausibility. Ital Heart J. 2001; 2:1–8. [PubMed: 11214695]
- Saiko P, Szakmary A, Jaeger W, Szekeres T. Resveratrol and its analogs: Defense against cancer, coronary disease and neurodegenerative maladies or just a fad? Mutation Research. 2008; 658:68– 94. [PubMed: 17890139]
- Schroeter H, Heiss C, Balzer J, Kleinbongard P, Keen CL, Hollenberg NK, Sies H, Kwik-Uribe C, Schmitz HH, Kelm M. (–)-Epicatechin mediates beneficial effects of flavanol-rich cocoa on vascular function in humans. Proc Natl Acad Sci U S A. 2006; 103:1024–1029. [PubMed: 16418281]
- Sharma S, Anjaneyulu M, Kulkarni SK, Chopra K. Resveratrol, a polyphenolic phytoalexin, attenuates diabetic nephropathy in rats. Pharmacology. 2006a; 76:69–75. [PubMed: 16286809]
- Sharma S, Kulkarni SK, Chopra K. Resveratrol, a polyphenolic phytoalexin attenuates thermal hyperalgesia and cold allodynia in STZ-induced diabetic rats. Indian J Exp Biol. 2006b; 44:566– 569. [PubMed: 16872046]
- Sharma S, Chopra K, Kulkarni SK. Effect of insulin and its combination with resveratrol or curcumin in attenuation of diabetic neuropathic pain: participation of nitric oxide and TNF-alpha. Phytother Res. 2007; 21:278–283. [PubMed: 17199240]
- Soleas GJ, Diamandis EP, Goldberg DM. Wine as a biological fluid: history, production, and role in disease prevention. J Clin Lab Anal. 1997; 11:287–313. [PubMed: 9292395]
- Leger AS, Cochrane AL, Moore F. Factors associated with cardiac mortality in developed countries with particular reference to the consumption of wine. Lancet. 1979; 1:1017–1020. [PubMed: 86728]
- Stolp HB, Dziegielewska KM. Role of developmental inflammation and blood-brain barrier dysfunction in neurodevelopmental and neurodegenerative diseases. Neuropathology and Applied Neurobiology. published online 11 December 2008, 10.1111/j.1365-2990.2008.01005.x.
- Su HC, Hung LM, Chen JK. Resveratrol, a red wine antioxidant, possesses an insulin-like effect in streptozotocin-induced diabetic rats. Am J Physiol Endocrinol Metab. 2006; 290:E1339–E1346. [PubMed: 16434553]
- Szmitko PE, Verma S. Antiatherogenic potential of red wine: clinician update. Am J Physiol Heart Circ Physiol. 2005; 288:2023–2030.
- Thirunavukkarasu M, Penumathsa SV, Samuel SM, Akita Y, Zhan L, Bertelli AAE, Maulik G, Maulik N. White wine induced cardioprotection against ischemia-reperfusion injury is mediated by life extending Akt/FOXO3a/NFκB survival pathway. J Agric Food Chem. 2008; 56:6733–6739. [PubMed: 18642840]
- Teyssen S, Gonzalez-Calero G, Schimiczek M, Singer MV. Maleic acid and succinic acid in fermented alcoholic beverages are the stimulants of gastric acid secretion. J Clin Invest. 1999; 103:707–713. [PubMed: 10074488]
- Udenigwe CC, Ramprasath VR, Aluko RE, Jones PJH. Potential of resveratrol in anticancer and antiinflammatory therapy. Nutrition Reviews. 2008; 66:445–454. [PubMed: 18667005]

- Ungvari Z, Orosz Z, Rivera A, Labinskyy N, Zhao X, Olson S, Podlutsky A, Csiszar A. Resveratrol increases vascular oxidative stress resistance. Am J Physiol Heart Circ Physiol. 2007; 292:H2417–H2424. [PubMed: 17220179]
- Valenzano DR, Terzibasi E, Genade T, Cattaneo A, Domenici L, Cellerino A. Resveratrol prolongs lifespan and retards the onset of age-related markers in a short-lived vertebrate. Curr Biol. 2006; 16:296–300. [PubMed: 16461283]
- Vidavalur R, Otani H, Singal PK, Maulik N. Significance of wine and resveratrol in cardiovascular disease: French paradox revisited. Exp Clin Cardiol. 2006; 11:217–225. [PubMed: 18651034]
- Virgili M, Contestabile A. Partial neuroprotection of in vivo excitotoxic brain damage by chronic administration of the red wine antioxidant agent, transresveratrol in rats. Neurosci Lett. 2000; 281:123–126. [PubMed: 10704758]
- Vitaglione P, Sforza S, Galaverna G, Ghidini C, Caporaso N, Vescovi PP, Fogliano V, Marchelli R. Bioavailability of trans-resveratrol from red wine in humans. Mol Nutr Food Res. 2005; 49:495– 504. [PubMed: 15830336]
- Vitaglione P, Ottanelli B, Milani S, Morisco F, Caporaso N, Fogliano V. Dietary trans-resveratrol bioavailability and effect on CCl<sub>4</sub>-induced liver lipid peroxidation. J Gastroenterol Hepatol. Sep 24.2008 [Epub ahead of print; doi:10.1111/j.1440-1746.2008.05598.x)].
- Vitrac X, Desmoulière A, Brouillaud B, Krisa S, Deffieux G, Barthe N, Rosenbaum J, Mérillon JM. Distribution of [14C]-trans-resveratrol, a cancer chemopreventive polyphenol, in mouse tissues after oral administration. Life Sci. 2003; 72:2219–2233. [PubMed: 12628442]
- Waterhouse AL. Wine phenolics. Ann N Y Acad Sci. 2002; 957:1–36. [PubMed: 12074956]
- Williamson G, Barron D, Shimoi K, Terao J. In vitro biological properties of flavonoid conjugates found in vivo. Free Radic Res. 2005; 39:457–469. [PubMed: 16036321]
- Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. Am J Clin Nutr. 2005; 81:2438–2558. [PubMed: 15640487]
- Wilson MA, Shukitt-Hale B, Kalt W, Ingram DK, Joseph JA, Wolkow CA. Blueberry polyphenols increase lifespan and thermotolerance in Caenorhabditis elegans. Aging Cell. 2006; 5:59–68. [PubMed: 16441844]
- Wood JG, Rogina B, Lavu S, Howitz K, Helfand SL, Tatar M, Sinclair D. Sirtuin activators mimic caloric restriction and delay ageing in metazoans. Nature. 2004; 430:686–689. [PubMed: 15254550]
- Yang JH, Hsia TC, Kuo HM, Chao PD, Chou CC, Wei YH, Chung JG. Inhibition of lung cancer cell growth by quercetin glucuronides via G2/M arrest and induction of apoptosis. Drug Metab Dispos. 2006; 34:296–304. [PubMed: 16280456]
- Yoshizumi M, Tsuchiya K, Suzaki Y, Kirima K, Kyaw M, Moon JH, Terao J, Tamaki T. Quercetin glucuronide prevents VSMC hypertrophy by angiotensin II via the inhibition of JNK and AP-1 signaling pathway. Biochem Biophys Res Commun. 2002; 293:1458–1465. [PubMed: 12054679]
- Zamin LL, Dillenburg-Pilla P, Argenta-Comiran R, Horn AP, Simao F, Nassif M, Gerhardt D, Frozza RL, Salbego C. Protective effect of resveratrol against oxygen-glucose deprivation in organotypic hippocampal slice cultures: Involvement of PI3-K pathway. Neurobiol Dis. 2006; 24:170–182. [PubMed: 16860989]
- Zamora-Ros R, Lamuela-Raventós RM, Estruch R, Andres-Lacueva C. Resveratrol, a new biomarker of moderate wine intake? Brit J Nutr. 2009; 101:148. [PubMed: 18782464]
- Zern TL, Fernandez ML. Cardioprotective effects of dietary polyphenols. J Nutr. 2005; 135:2291–2294. [PubMed: 16177184]

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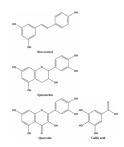
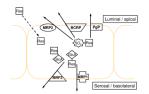


Figure 1. Chemical structures of resveratrol, epicatechin, gallic acid and quercetin



#### Figure 2. Intestinal metabolism and transport of resveratrol

Resveratrol aglycone is taken up by the enterocytes, probably by passive diffusion, and is efficiently conjugated with glucuronic acid and sulfate through the action of UDP-glucuronosyl transferase and aryl-sulfatase enzymes. Methylation has not been observed, presumably due to the lack of a catechol function in the resveratrol molecule, which is a requirement for catechol-O-methyltransferase (COMT). The conjugates are transported out of the enterocytes by as yet unconfirmed processes, but it is thought that efflux transporters of the multi-drug resistance proteins (MRP) family are involved (several are indicated). Resveratrol sulfate is preferentially transported to the apical (luminal) side of polarized enterocytes, while resveratrol glucuronides are preferentially effluxed to the basolateral (serosal) side (Kaldas et al., 2003). Hence, the predominant form of resveratrol in human blood following oral dosing is resveratrol glucuronide (95–99% of total plasma resveratrol). Solid arrows, probable active transport process; Dashed arrows, diffusion; Res, resverastrol; GlcA, glucuronic acid; SO4<sup>-</sup>, sulphate; MRP, multi-drug-resistance protein; BCRP, breast cancer resistance protein; PgP, P-glycoprotein.