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## Comprehensive analysis of the impact of SNPs and CNVs on human microRNAs and their regulatory genes

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### Abstract

Human microRNAs (miRNAs) are potent regulators of gene expression and thus involved in a broad range of biological processes. The objective of this study was to update the properties of human miRNAs and to search for SNPs and CNVs with potential effects on them. Based on the latest miRBase 13.0 database, we identified 380 (53.9%) precursor miRNAs (pre-miRNAs) embedded in gene loci that are enriched in biological processes such as “Neuronal activities”, “Cell Cycle” and “Protein phosphorylation” (Bonferroni  $p < 0.05$ ). Gene lengths of the pre-miRNA host genes are significantly larger than other genes in the genome ( $p < 2.2E-16$ ). Using data mining public resources, we performed a genome-scale search for the regulatory polymorphisms in the loci of pre-miRNAs and their related genes. Altogether, we found 187 SNPs in the pre-miRNAs, 497 consensus SNPs in the seed-matching untranslated regions of target genes, 385 CNVs harboring pre-miRNA precursors and 9 CNVs covering important miRNA processing genes. We also noticed that minimum free energy changed by pre-miRNA-residing SNPs could be ranked by the order from low to high as the SNPs in the loop domain, the SNPs in the adjacent stem and basal stem domains, and the SNPs in mature miRNA and its complementary sequence domains ( $p = 0.0065$ ). With a full list of miRNA-related polymorphisms, this study will facilitate future association studies between the genetic polymorphisms in miRNA targets or pre-miRNAs and the disease susceptibility or therapeutic outcome.

### Keywords

microRNA; miRNA; gene; pathway; gene ontology; SNP; CNV

### Introduction

As a new class of abundantly distributed small non-coding RNA molecules, miRNAs are initially transcribed as primary miRNA (pri-miRNAs) that are further processed through stem-loop pre-miRNAs into a single-stranded mature form.<sup>1</sup> Generally, miRNAs partially complement the 3'-untranslated regions (UTR) of target mRNAs, and subsequently invoke a series of posttranscriptional silencing events on the target genes. These include intervention of translational initiation and elongation, induction of deadenylation and interruption of mRNA and protein synthesis.<sup>2,3</sup> The core of miRNA's pairing sequence is termed “seed”, which is usually conserved across multiple species.<sup>4</sup> Since this seed sequence is on average 8 nucleotides in length and critical for the miRNA-target binding, the miRNAs are estimated to have anywhere from several to thousands of targets.<sup>4</sup> Therefore, mature miRNAs as potent

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regulators of gene expression have been implicated in various biological development processes and disease progression.<sup>5,6</sup>

SNPs within the sequences of human miRNAs and their targets have been shown to have impact on various phenotypes including blood pressure,<sup>7</sup> drug resistance,<sup>8</sup> outcomes of therapeutic intervention,<sup>9</sup> abnormal psychiatry disorder,<sup>10</sup> the development of gastric mucosal atrophy,<sup>11</sup> the risk of diseases that consist of asthma,<sup>12</sup> diabetes,<sup>13</sup> Parkinson,<sup>14</sup> cancers of colorectal,<sup>15</sup> breast,<sup>16</sup> bladder,<sup>17</sup> papillary thyroid,<sup>18,19</sup> lung<sup>20,21</sup> and esophageal.<sup>22</sup> Moreover, miRNAs and their target genes may be located at the genomic regions of high instability, a feature often observed in cancer and other genetic diseases. To note, specific deletions of key enzymes such as Dicer1 (dicer 1, ribonuclease type III) may cause global impairment of miRNA processing leading to severe abnormality.<sup>23</sup>

As shown in Table 1, several researchers have uncovered links between genomic variations and miRNAs.<sup>9,15,16,18–22,24–36</sup> However, a systematic update of the genetic factors that influence miRNA activities according to the most recent miRBase is not yet in place. Taking advantage of the latest miRNA databases and bioinformatics tools, we analyzed human miRNA for their biological properties and performed a genome-wide scan for functional genetic variations that may potentially affect human miRNA processing and targeting.

## Results

### Human miRNAs

As shown in the Figure 1, there are a total of 718 loci for human pre-miRNA genes in the genome based on the miRBase 13.0. The pre-miRNAs with multiple copies include mir-1184 (3 copies), mir-1233 (2 copies), mir-1244 (4 copies), mir-1302-2 (4 copies), mir-1972 (2 copies), mir-1974 (2 copies), mir-1977 (2 copies) and mir-1978 (2 copies). These 706 pre-miRNAs can be processed into 885 mature miRNAs (including mirR\*products) with lengths ranging from 17 to 27 nucleotides. Based on the common seed sequence of mature miRNA products, the pre-miRNAs may be further grouped into families. In the current released version of miRBase, there are altogether 381 miR-families (644 members) comprising of up to 42 members in humans. The largest pre-miRNA families include mir-515 (n = 42), mir-548 (n = 31), mir-154 (n = 19), mir-506 (n = 18) and let-7 (n = 12).

### Pre-miRNA host genes

There are 380 human pre-miRNA genes residing in the loci of 340 protein-coding-genes (PCGs) (Suppl. Table 1). The genes hosting the most pre-miRNAs include *HTR2C* (5 pre-miRNAs), *RTL1* (5 pre-miRNAs) and *LARP7* (5 pre-miRNAs). By comparing the length of gene-spanning regions between pre-miRNA host PCGs and other PCGs in humans, our results showed that the pre-miRNA-host PCGs have significant larger gene-spanning regions (Fig. 2A, Kolmogorov-Smirnov test,  $D = 0.38$ ,  $p < 2.2E-16$ ). In addition, the host genes were analyzed for enriched categories in the Gene Ontology (GO)<sup>37</sup> and Kyoto Encyclopedia of Genes and Genomes (KEGG)<sup>38</sup> pathways. As shown in Table 2, the host genes are significantly enriched in the pathways of “Insulin/IGF pathway-mitogen activated protein kinase kinase/ MAP kinase cascade” and “Coenzyme A biosynthesis”, the biological processes of “Neuronal activities”, “Protein phosphorylation”, “Cell cycle” and “Cell structure and motility”, and the molecular function of “Kinase” (Bonferroni corrected  $p < 0.05$ ). Among these pre-miRNA host genes, only 33 genes are not annotated and these are significantly less than those found in all NCBI annotated genes ( $p < 0.0001$ ,  $df = 1$ ,  $\chi^2 = 67.98$ ,  $OR = 0.25$ ). In addition, the category of “unclassified” for both the pathway and GO analysis are significantly underrepresented for miRNA host genes (Bonferroni  $p < 0.05$ ), implying that important functions are implicated for miRNA host genes. To note, the pre-miRNA host PCGs trend to

have larger gene length than the rest PCGs in human genome, thus they are more likely (by chance) to harbor miRNAs. And this may potentially cause some biased observations for certain GO terms.

### SNPs and CNVs in pre-miRNAs

In the present study, we performed a genome-scale search for both SNPs and CNVs that may potentially affect miRNA processing or targeting. Our results revealed that only 188 SNPs are located at 138 pre-miRNA regions (Table 3). In contrast, on average there are over 300 SNPs per 100 bps in the flanking regions of all the pre-miRNAs (Fig. 2B). This observation agrees with the previous finding based on the miRNAs in the earlier miRBase released version.<sup>31,36</sup> Among the SNPs in the hairpins of pre-miRNAs, there are 16 SNPs in adjacent stem, 77 SNPs in the basal stem, 17 SNPs in the loop, 44 SNPs in mature miRNA, 54 SNPs in the complementary sequence of mature miRNA (Table 3). We use RNAfold web server to determine the minimum free energy (MFE) of hairpin structures for all the SNP-residing pre-miRNAs.<sup>39</sup> The changes of minimum free energy (MFE) by the SNPs in the pre-miRNAs are also given in the Table 3. Significantly more MFE changes are caused by SNPs in mature miRNA and its complementary sequence domains than the SNPs in the adjacent stem and basal stem domains that are followed by SNPs in the loop domain (Fig. 3, Kruskal-Wallis test,  $\chi^2 = 14.25$ ,  $df = 4$ ,  $p = 0.0065$ ).

We also evaluated the known CNV coverage of human pre-miRNA genes using the CNVs deposited in the Database of Genomic Variants (DGV).<sup>40</sup> We found that 193 pre-miRNAs were located in the regions covered by 385 CNV markers (Table 4 and Suppl. Table 2). No significant difference for the distribution of pre-miRNAs in PCGs ( $n = 109$ ) and in the intergenic regions ( $n = 84$ ,  $\chi^2 = 0.71$ ,  $df = 1$ ,  $p = 0.39$ ).

### Polymorphisms with potential effects on miRNA targeting

Using the predicted targets by TargetScanS<sup>41</sup> and PITA,<sup>42</sup> we found 1,238 and 4,235 SNPs located in the putative seed-matching regions of targeting genes (Suppl. Tables 3 and 4). As shown in Table 5, eleven 3'-UTR SNPs may disrupt the miRNA-target regulation that has been supported by experimental evidence maintained in TarBase.<sup>43</sup> A total of 497 overlapping SNPs are found in the same seed-matching regions of 434 target genes by both TargetScanS and PITA (Suppl. Table 5). As shown in Table 6, the 434 overlapping target genes are significantly enriched in the KEGG pathways of "Angiogenesis" (Bonferroni  $p = 4 \times 10^{-6}$ ) and "T cell activation" (Bonferroni  $p = 0.004$ ) as well as the GO biological processes of "Developmental processes" (Bonferroni  $p = 3 \times 10^{-21}$ ), "Signal transduction" (Bonferroni  $p = 5 \times 10^{-12}$ ), "mRNA transcription regulation" (Bonferroni  $p = 2 \times 10^{-6}$ ), "Neurogenesis" (Bonferroni  $p = 2 \times 10^{-6}$ ) and "Oncogenesis" (Bonferroni  $p = 8 \times 10^{-5}$ ). Focusing on both the pre-miRNA host genes and the miRNA target genes with SNPs in the consensus target sites, the pathway and GO analysis show some similar pathways and GO categories for these two lists, such as "Neuronal activities", "Cell cycle", "Protein phosphorylation", and "Cell structure and motility" in the biological processes, and the "kinase" in the shared molecular function. These suggest a potential involvement of miRNAs in these biological activities.

### Variations at important miRNA-processing genes

Given these delicate processes in miRNA biogenesis (Fig. 4), any alterations of the key proteins involved in the miRNA processing and targeting will potentially lead to a global deregulation of the miRNA-mediated posttranscriptional silencing. Altogether, we found 3,921 SNPs in these miRNA-processing genes. A total of 83 SNPs may change the coding sequence of protein products. However, there are only 35 SNPs with allele frequency reports (Suppl. Table 6). Their contributions to gene functions remain to be discovered. A further analysis between CNVs and 13 miRNA-related genes shows that there are deletion in the loci of *SNIP1*,

*RNASEN*, *DICER1* and *DGCR8*, suggesting a potentially disrupted miRNA-processing pathway in those CNV carriers (Table 7).

## Discussions

In the present study, we evaluated the properties of human pre-miRNAs based on the miRBase database (release 13). Our survey shows that 53.9% of pre-miRNAs are located in PCGs. We found that pre-miRNA host genes have longer spanning regions than other genes in the genome and pre-miRNA host genes are more likely to be annotated with functional descriptions as compared with other genes in the genome ( $p < 0.0001$ ,  $OR = 4.05$ ), although this may be biased by a trend in the published studies of pre-miRNAs. A total of 193 pre-miRNAs (27.4%) are located in regions with genome instability. Interestingly, there are 10 out of 12 *let-7* family members found in CNV regions (Suppl. Table 2). Given that *let-7* plays a role in tumorigenesis, this finding suggests a non-random connection between pre-miRNA, CNVs and cancer development which is in agreement with previous findings.<sup>44</sup>

PCGs may host multiple pre-miRNAs and thus have potential to network with others. For example, *HTR2C*, a host gene for 5 pre-miRNAs, is a G-protein coupled receptor and mediates the signaling of neuronal activities.<sup>45</sup> *RTL1*, a host gene of 5 pre-miRNAs is a reverse transcriptase and aspartic protease<sup>46</sup> that is involved in angiogenesis, apoptosis and pathway-mitogen activated protein kinase kinase/MAP kinase cascade. Harboring 5 pre-miRNAs, *LARP7* is a ribonucleoprotein and plays an important role in tRNA metabolism.<sup>47</sup>

Given the extensive variation found in the human genome, miRNA-mediated functions may be affected by polymorphisms in the miRNA target loci, pre-miRNA gene loci and/or the miRNA regulating gene loci.<sup>48</sup> The polymorphism-driven alterations of miRNA activity are observed in numerous association studies.<sup>43</sup> Several studies have catalogued the SNPs at the human pre-miRNAs and mature miRNA binding sites based on the earlier miRBase version (Table 1). In the present study, we also evaluated the roles of genomic variants, including both SNPs and CNVs, which may influence the miRNA-associated biological functions. Using the 718 genomic coordinates of 705 human pre-miRNAs (not including mir-941-4), we found 188 SNPs in the 138 pre-miRNAs with various numbers of SNPs in different pre-miRNA domains. The SNP-residing domains with the trend of MFE changes from low to high are loop, stem (including adjacent stem and basal stem), mature miRNA and their complementary sequences (Fig. 3). SNPs in pre-miRNAs could potentially change the stem-loop structures and thus may influence the miRNA processing and maturation. SNPs in the mature miRNAs, especially in the seed regions, are likely to affect the specificity for gene silencing.

Bioinformatics databases along with experimental evidence implicate eleven SNPs in miRNA target sites (Table 5). In our analysis, we found three SNPs with allele frequencies reported in NCBI dbSNP, including rs3783620, rs1621 and rs17620927, that may affect miRNA:gene regulation recorded in TarBase v5.0.<sup>49</sup> For example, the miR-126 was reported to repress the translational level of VCAM1 by binding to the 3'-UTR of the gene<sup>50</sup> and this regulation might be influenced by SNP rs3783620 in the seed-matching region on the 3'-UTR of *VCAM1*. To note, *VCAM1* is an important gene in regulating leukocyte trafficking to sites of inflammation.<sup>51</sup> In addition, Kim et al. reported that miR-199a\* mediated the downregulation of *MET* gene by targeting at the 3'-UTR.<sup>52</sup> However, SNP rs1621 in the seed-matching sequence of *MET* could affect this activity. Another example is miR-373 and *MKRN1* gene. Using a microarray assay, the expression of miR-373 was identified to be inversely associated with the expression of *MKRN1*.<sup>53</sup> SNP rs17620927, located in the miR-373 target site for *MKRN1* likely affects miRNA-mediated gene silencing.

It is notable that the pre-miRNAs with multiple genomic copies, including mir-1244 (4 copies) and mir-1302-2 (4 copies) reside in CNV regions. This implies that the multiple copies of these pre-miRNAs are likely to be generated by genomic instability. The copy number changes may affect both the miRNAs and their host genes. Among the pre-miRNAs and their host genes in the CNV regions, some of them are implicated with important biological functions. Among them, mir-200b was reported to mediate gene silencing of ZFHX1B, a gene that is important in the TGFbeta signaling pathway.<sup>54</sup> As shown in the Table 4, mir-555 is found to be located in a CNV loss region and it is embedded in *ASH1L*, a gene that is identified as a histone methyltransferase regulating gene transcription.<sup>55</sup> AATK, a tyrosine kinase involved in apoptosis,<sup>56</sup> is also located in the CNV loss region and harbors three pre-miRNAs including mir-657, mir-338 and mir-1250. These structural variants of the pre-miRNAs may be implicated with important biological effects in humans. Eleven out of 95 individuals were found to carry a deletion that covers mir-199a-1 and its host PCG (*DNM2*). *DNM2* encodes a member of GTPases and it was a candidate gene of dominant intermediate Charcot-Marie-Tooth disease<sup>57</sup> and autosomal dominant centronuclear myopathy.<sup>58</sup> *HTR2C* that harbors 5 pre-miRNAs was found to be located in a CNV with genotypes of 4 gains and 9 losses. *HTR2C* gene encodes a serotonin receptor that is associated with mental disorders<sup>59,60</sup> and side effects induced by antipsychotic drugs.<sup>61,62</sup> Since little evidence is available about functional connection between the pre-miRNAs and their host PCGs, the current study provides researchers with a comprehensive list for future analysis.

As shown in Figure 4, pre-miRNAs have been shown to be produced through either alternative splicing from their host genes or processing by Drosha (RNASEN, nuclear ribonuclease type III).<sup>2</sup> Subsequently, these intermediate precursors are exported by Exportin-5 (XPO5) and Ran-GTP (RAN) from nucleus to the cytoplasm, where Dicer (DICER1) excises stem-loop pre-miRNAs into single-stranded mature miRNA.<sup>2,3,63</sup> These mature miRNA will be mediated by the argonaute family proteins (EIF2C1, EIF2C2)<sup>64</sup> and other proteins (TARBP2, TNRC6A, PIWIL1)<sup>65,66</sup> to evoke a series of posttranscriptional silencing of target genes. Besides these well known genes, recent evidence shows that SNIP1,<sup>67</sup> and DGCR8,<sup>68</sup> along with Drosha are involved in the initiation of miRNA biogenesis. Gemin3 (DDX20) and Gemin4 (GEMIN4) are two other important genes for miRNA function. They can form novel complex ribonucleoproteins to perform gene silencing function with miRNA and eIF2C2, a member of the Argonaute protein family.<sup>69</sup>

In this study, we evaluated the contribution of CNVs to 13 miRNA-processing pathway genes. SNIP1 and DGCR8 are important proteins in the initiation of the pri-miRNA transcription. EIF2C2 is in the Argonaut family of genes that play an important role in gene silencing. The CNVs with genomic loss suggest that other proteins may rescue their biological functions. Interestingly, we also noticed there is a 350 bp loss in the *DICER1* loci, which will cause a truncated form of DICER1. Given the severe abnormality caused by the Dicer1 gene deletion,<sup>23</sup> we speculate that this truncated form of DICER maintains the function of DICER1. Since the CNV frequencies at most genes except *DGCR8* are relatively low, more evidence is needed for an in-depth exploration.

In summary, we have performed an in-depth analysis of human miRNAs based on multiple association studies. Compared with previous studies, we focused on both SNPs and CNV information that may potentially affect miRNA processing and maturation. Since aberrant miRNA expressions have been implicated in oncogenesis and other diseases,<sup>70</sup> the miRNA-related polymorphisms provided by us will facilitate future studies and increase our understanding of the role of miRNAs in human gene regulation.

## Materials and Methods

### Searching for host genes for the pre-miRNAs

The chromosomal coordinates for 705 (not including mir-941-4) pre-miRNAs and over 30,000 human genes were obtained from the miRBase version 13.0 and NCBI dbGene. We compared the genomic start and end positions of over 30,000 genes with those of 705 pre-miRNAs. A host gene of a pre-miRNA is defined when their loci overlap with each other. All of the genomic coordinates in the present study were based on hg18 (March, 2006).

### Searching for SNPs in the pre-miRNAs, mature miRNAs and miRNA target sites

The genomic coordinates of mature miRNAs were inferred from the pre-miRNAs by the sequence matching. The miRNA target seed-matching regions predicted by TargetScan v4.1 and PITA were downloaded from the UCSC genome browser.<sup>71</sup> Genomic positions of over 10 million SNPs were retrieved from NCBI dbSNP129 and then they were used to search for SNPs within the start and end positions of pre-miRNAs, mature miRNAs and miRNA target sites.

### Searching for CNVs covering pre-miRNAs

The genomic coordinates of CNVs (after excluding inversions) were downloaded from Database of Genomic Variants (DGV) version 7.<sup>40</sup> The genomic start and end positions of over 20,000 CNVs with those of 705 pre-miRNAs were compared to see whether there were overlapping regions between them.

### GO and KEGG pathway analysis

PANTHER database<sup>72</sup> was used to identify enriched functional annotation categories for pre-miRNA host genes and the genes with SNPs in the miRNA target sites. KEGG pathway and two GO terms (biological process and molecular function) were evaluated. Uncorrected  $p < 0.01$  was considered statistically significant.

### Web resources

NCBI dbSNP: <http://www.ncbi.nih.gov/SNP/>

Database of Genomic Variants: <http://projects.tcag.ca/variation/>

UCSC genome browser: <http://genome.ucsc.edu/>

miRBase: <http://microrna.sanger.ac.uk/sequences/>

TarBase: <http://diana.cslab.ece.ntua.gr/tarbase/>

TargetScanS miRNA target database: [http://www.targetscan.org/vert\\_42/](http://www.targetscan.org/vert_42/)

PITA miRNA target database: [http://genie.weizmann.ac.il/pubs/mir07/mir07\\_data.html](http://genie.weizmann.ac.il/pubs/mir07/mir07_data.html)

RNAfold web server: <http://rna.tbi.univie.ac.at/cgi-bin/RNAfold.cgi>

### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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## References

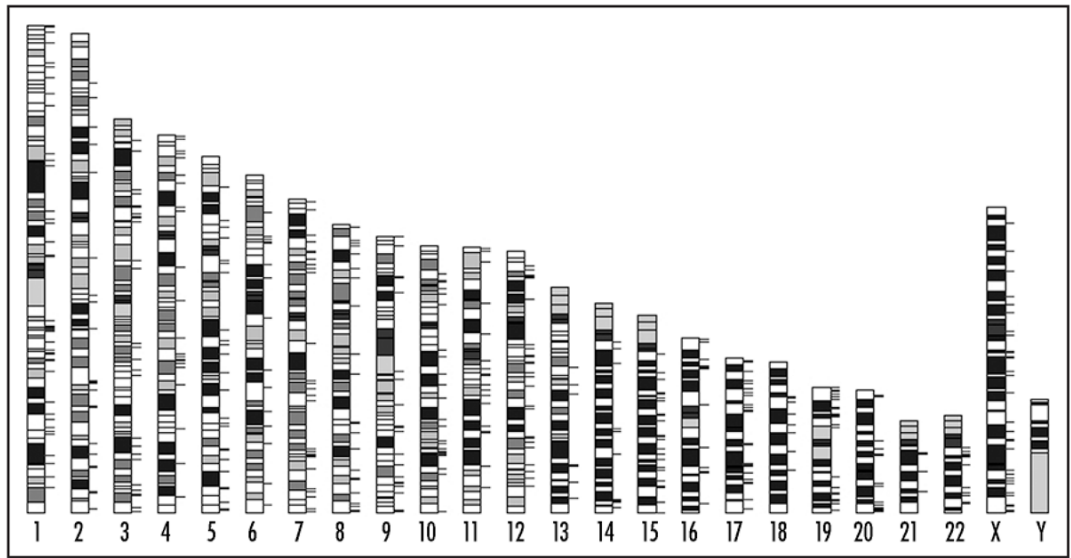
1. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism and function. *Cell* 2004;116:281–97. [PubMed: 14744438]
2. Roush S, Slack FJ. The let-7 family of microRNAs. *Trends Cell Biol* 2008;18:505–16. [PubMed: 18774294]
3. Liu J. Control of protein synthesis and mRNA degradation by microRNAs. *Curr Opin Cell Biol* 2008;20:214–21. [PubMed: 18329869]
4. Lewis BP, Burge CB, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 2005;120:15–20. [PubMed: 15652477]
5. Croce CM, Calin GA. miRNAs, cancer and stem cell division. *Cell* 2005;122:6–7. [PubMed: 16009126]
6. Bicker S, Schrott G. microRNAs: Tiny Regulators of Synapse Function in Development and Disease. *J Cell Mol Med* 2008;12:1466–76. [PubMed: 18624757]
7. Sethupathy P, Borel C, Gagnebin M, Grant GR, Deutsch S, Elton TS, et al. Human microRNA-155 on chromosome 21 differentially interacts with its polymorphic target in the AGTR1 3' untranslated region: a mechanism for functional single-nucleotide polymorphisms related to phenotypes. *Am J Hum Genet* 2007;81:405–13. [PubMed: 17668390]
8. Mishra PJ, Humeniuk R, Mishra PJ, Longo-Sorbello GS, Banerjee D, Bertino JR. A miR-24 microRNA binding-site polymorphism in dihydrofolate reductase gene leads to methotrexate resistance. *Proc Natl Acad Sci USA* 2007;104:13513–8. [PubMed: 17686970]
9. Hu Z, Chen J, Tian T, Zhou X, Gu H, Xu L, et al. Genetic variants of miRNA sequences and non-small cell lung cancer survival. *J Clin Invest* 2008;118:2600–8. [PubMed: 18521189]
10. Jensen KP, Covault J, Conner TS, Tennen H, Kranzler HR, Furneaux HM. A common polymorphism in serotonin receptor 1B mRNA moderates regulation by miR-96 and associates with aggressive human behaviors. *Mol Psychiatry* 2009;14:381–9. [PubMed: 18283276]
11. Arisawa T, Tahara T, Shibata T, Nagasaka M, Nakamura M, Kamiya Y, et al. A polymorphism of microRNA 27a genome region is associated with the development of gastric mucosal atrophy in Japanese male subjects. *Dig Dis Sci* 2007;52:1691–7. [PubMed: 17546506]
12. Tan Z, Randall G, Fan J, Camoretti-Mercado B, Brockman-Schneider R, Pan L, et al. Allele-specific targeting of microRNAs to HLA-G and risk of asthma. *Am J Hum Genet* 2007;81:829–34. [PubMed: 17847008]
13. Lv K, Guo Y, Zhang Y, Wang K, Jia Y, Sun S. Allele-specific targeting of hsa-miR-657 to human IGF2R creates a potential mechanism underlying the association of ACAA-insertion/deletion polymorphism with type 2 diabetes. *Biochem Biophys Res Commun* 2008;374:101–5. [PubMed: 18602895]
14. Wang G, van der Walt JM, Mayhew G, Li YJ, Zuchner S, Scott WK, et al. Variation in the miRNA-433 binding site of FGF20 confers risk for Parkinson disease by overexpression of alpha-synuclein. *Am J Hum Genet* 2008;82:283–9. [PubMed: 18252210]
15. Landi D, Gemignani F, Naccarati A, Pardini B, Vodicka P, Vodickova L, et al. Polymorphisms within micro-RNA-binding sites and risk of sporadic colorectal cancer. *Carcinogenesis* 2008;29:579–84. [PubMed: 18192692]
16. Brendle A, Lei H, Brandt A, Johansson R, Enquist K, Henriksson R, et al. Polymorphisms in predicted microRNA-binding sites in integrin genes and breast cancer: ITGB4 as prognostic marker. *Carcinogenesis* 2008;29:1394–9. [PubMed: 18550570]
17. Yang H, Dinney CP, Ye Y, Zhu Y, Grossman HB, Wu X. Evaluation of genetic variants in microRNA-related genes and risk of bladder cancer. *Cancer Res* 2008;68:2530–7. [PubMed: 18381463]

18. Jazdzewski K, Murray EL, Franssila K, Jarzab B, Schoenberg DR, de la Chapelle A. Common SNP in pre-miR-146a decreases mature miR expression and predisposes to papillary thyroid carcinoma. *Proc Natl Acad Sci USA* 2008;105:7269–74. [PubMed: 18474871]
19. Jazdzewski K, Liyanarachchi S, Swierniak M, Pachucki J, Ringel MD, Jarzab B, et al. Polymorphic mature microRNAs from passenger strand of pre-miR-146a contribute to thyroid cancer. *Proc Natl Acad Sci USA* 2009;106:1502–5. [PubMed: 19164563]
20. Tian T, Shu Y, Chen J, Hu Z, Xu L, Jin G, et al. A Functional Genetic Variant in microRNA-196a2 Is Associated with Increased Susceptibility of Lung Cancer in Chinese. *Cancer Epidemiol Biomarkers Prev* 2009;18:1183–7. [PubMed: 19293314]
21. Chin LJ, Ratner E, Leng S, Zhai R, Nallur S, Babar I, et al. A SNP in a let-7 microRNA complementary site in the KRAS 3' untranslated region increases non-small cell lung cancer risk. *Cancer Res* 2008;68:8535–40. [PubMed: 18922928]
22. Ye Y, Wang KK, Gu J, Yang H, Lin J, Ajani JA, et al. Genetic variations in microRNA-related genes are novel susceptibility loci for esophageal cancer risk. *Cancer Prev Res (Phila Pa)* 2008;1:460–9. [PubMed: 19138993]
23. Kumar MS, Lu J, Mercer KL, Golub TR, Jacks T. Impaired microRNA processing enhances cellular transformation and tumorigenesis. *Nat Genet* 2007;39:673–7. [PubMed: 17401365]
24. Iwai N, Naraba H. Polymorphisms in human pre-miRNAs. *Biochem Biophys Res Commun* 2005;331:1439–44. [PubMed: 15883035]
25. Chen K, Rajewsky N. Natural selection on human microRNA binding sites inferred from SNP data. *Nat Genet* 2006;38:1452–6. [PubMed: 17072316]
26. Clop A, Marcq F, Takeda H, Pirottin D, Tordoir X, Bibe B, et al. A mutation creating a potential illegitimate microRNA target site in the myostatin gene affects muscularity in sheep. *Nat Genet* 2006;38:813–8. [PubMed: 16751773]
27. Adams BD, Furneaux H, White BA. The micro-ribonucleic acid (miRNA) miR-206 targets the human estrogen receptor-alpha (ERalpha) and represses ERalpha messenger RNA and protein expression in breast cancer cell lines. *Mol Endocrinol* 2007;21:1132–47. [PubMed: 17312270]
28. Bao L, Zhou M, Wu L, Lu L, Goldowitz D, Williams RW, et al. PolymiRTS Database: linking polymorphisms in microRNA target sites with complex traits. *Nucleic Acids Res* 2007;35:51–4.
29. Duan R, Pak C, Jin P. Single nucleotide polymorphism associated with mature miR-125a alters the processing of pri-miRNA. *Hum Mol Genet* 2007;16:1124–31. [PubMed: 17400653]
30. Martin MM, Buckenberger JA, Jiang J, Malana GE, Nuovo GJ, Chotani M, et al. The human angiotensin II type 1 receptor +1166 A/C polymorphism attenuates micror-na-155 binding. *J Biol Chem* 2007;282:24262–9. [PubMed: 17588946]
31. Saunders MA, Liang H, Li WH. Human polymorphism at microRNAs and microRNA target sites. *Proc Natl Acad Sci USA* 2007;104:3300–5. [PubMed: 17360642]
32. Yu Z, Li Z, Jolicoeur N, Zhang L, Fortin Y, Wang E, et al. Aberrant allele frequencies of the SNPs located in microRNA target sites are potentially associated with human cancers. *Nucleic Acids Res* 2007;35:4535–41. [PubMed: 17584784]
33. Landi D, Gemignani F, Barale R, Landi S. A catalog of polymorphisms falling in microRNA-binding regions of cancer genes. *DNA Cell Biol* 2008;27:35–43. [PubMed: 17941804]
34. Mishra PJ, Mishra PJ, Banerjee D, Bertino JR. MiRSNPs or MiR-polymorphisms, new players in microRNA mediated regulation of the cell: Introducing microRNA pharmacogenomics. *Cell Cycle* 2008;7:853–8. [PubMed: 18414050]
35. Reumers J, Conde L, Medina I, Maurer-Stroh S, Van Durme J, Dopazo J, et al. Joint annotation of coding and non-coding single nucleotide polymorphisms and mutations in the SNPeff and PupaSuite databases. *Nucleic Acids Res* 2008;36:825–9.
36. Quach H, Barreiro LB, Laval G, Zidane N, Patin E, Kidd KK, et al. Signatures of purifying and local positive selection in human miRNAs. *Am J Hum Genet* 2009;84:316–27. [PubMed: 19232555]
37. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet* 2000;25:25–9. [PubMed: 10802651]
38. Kanehisa M, Araki M, Goto S, Hattori M, Hirakawa M, Itoh M, et al. KEGG for linking genomes to life and the environment. *Nucleic Acids Res* 2008;36:480–4.

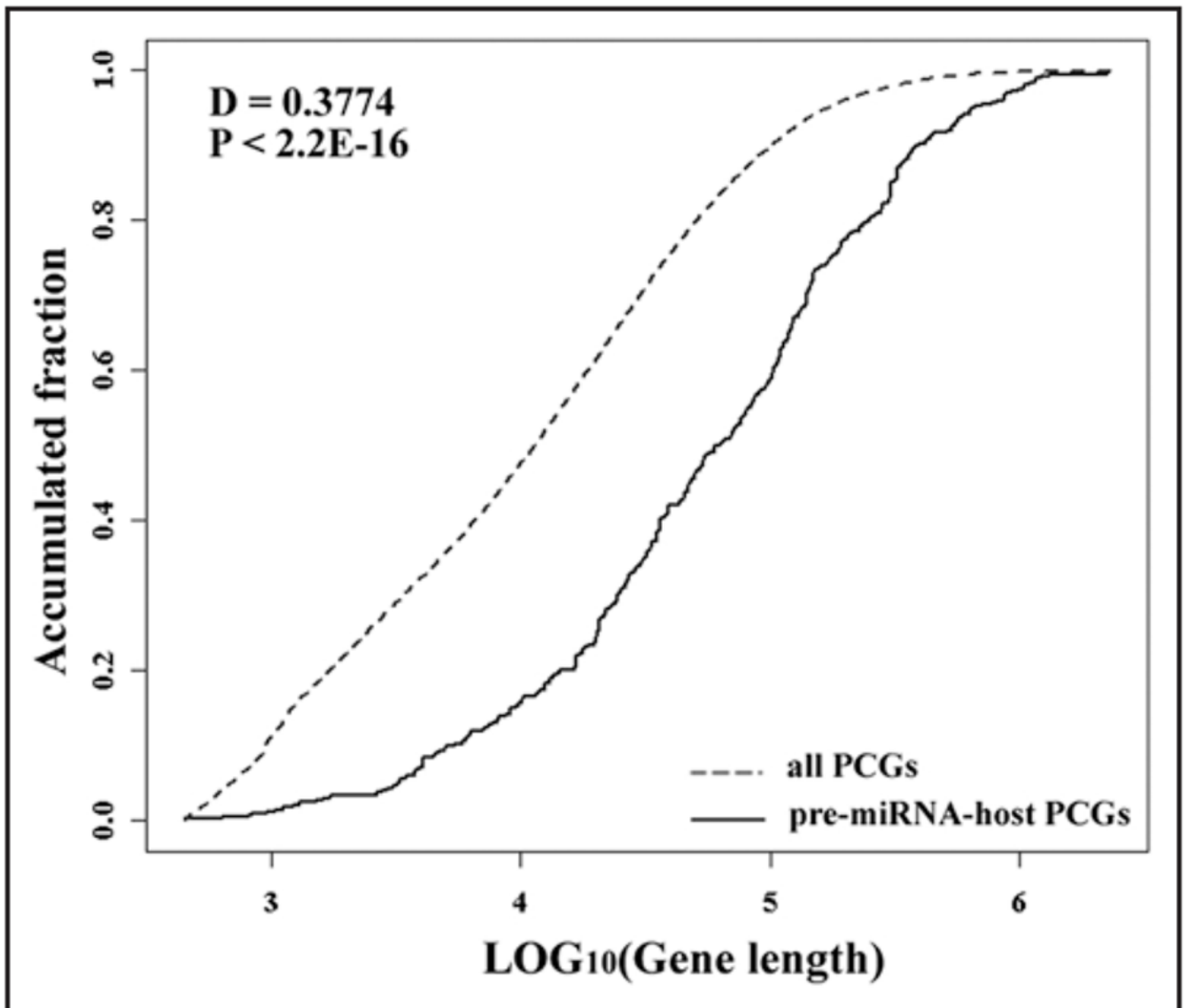


39. Zuker M, Stiegler P. Optimal computer folding of large RNA sequences using thermodynamics and auxiliary information. *Nucleic Acids Res* 1981;9:133–48. [PubMed: 6163133]
40. Iafraite AJ, Feuk L, Rivera MN, Listewnik ML, Donahoe PK, Qi Y, et al. Detection of large-scale variation in the human genome. *Nat Genet* 2004;36:949–51. [PubMed: 15286789]
41. Grimson A, Farh KK, Johnston WK, Garrett-Engele P, Lim LP, Bartel DP. MicroRNA targeting specificity in mammals: determinants beyond seed pairing. *Mol Cell* 2007;27:91–105. [PubMed: 17612493]
42. Kertesz M, Iovino N, Unnerstall U, Gaul U, Segal E. The role of site accessibility in microRNA target recognition. *Nat Genet* 2007;39:1278–84. [PubMed: 17893677]
43. Papadopoulos GL, Reczko M, Simossis VA, Sethupathy P, Hatzigeorgiou AG. The database of experimentally supported targets: a functional update of TarBase. *Nucleic Acids Res*. 2009In press
44. Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Nat Acad Sci USA* 2004;101:2999–3004. [PubMed: 14973191]
45. Drago A, Serretti A. Focus on HTR2C: A possible suggestion for genetic studies of complex disorders. *Am J Med Genet B Neuropsychiatr Genet*. 2008In press
46. Seitz H, Youngson N, Lin SP, Dalbert S, Paulsen M, Bachellerie JP, et al. Imprinted microRNA genes transcribed antisense to a reciprocally imprinted retrotransposon-like gene. *Nature Genet* 2003;34:261–2. [PubMed: 12796779]
47. He N, Jahchan NS, Hong E, Li Q, Bayfield MA, Marais RJ, et al. A La-related protein modulates 7SK snRNP integrity to suppress P-TEFb-dependent transcriptional elongation and tumorigenesis. *Mol Cell* 2008;29:588–99. [PubMed: 18249148]
48. Borel C, Antonarakis SE. Functional genetic variation of human miRNAs and phenotypic consequences. *Mamm Genome* 2008;19:503–9. [PubMed: 18787897]
49. Sethupathy P, Corda B, Hatzigeorgiou AG. TarBase: A comprehensive database of experimentally supported animal microRNA targets. *RNA* 2006;12:192–7. [PubMed: 16373484]
50. Harris TA, Yamakuchi M, Ferlito M, Mendell JT, Lowenstein CJ. MicroRNA-126 regulates endothelial expression of vascular cell adhesion molecule 1. *Proc Nat Acad Sci USA* 2008;105:1516–21. [PubMed: 18227515]
51. Mebius RE. Organogenesis of lymphoid tissues. *Nat Rev Immunol* 2003;3:292–303. [PubMed: 12669020]
52. Kim S, Lee UJ, Kim MN, Lee EJ, Kim JY, Lee MY, et al. MicroRNA miR-199a\* regulates the MET proto-oncogene and the downstream extracellular signal-regulated kinase 2 (ERK2). *J Biol Chem* 2008;283:18158–66. [PubMed: 18456660]
53. Lim LP, Lau NC, Garrett-Engele P, Grimson A, Schelter JM, Castle J, et al. Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. *Nature* 2005;433:769–73. [PubMed: 15685193]
54. Christoffersen NR, Silahtaroglu A, Orom UA, Kauppinen S, Lund AH. miR-200b mediates post-transcriptional repression of ZFX1B. *RNA* 2007;13:1172–8. [PubMed: 17585049]
55. Gregory GD, Vakoc CR, Rozovskaia T, Zheng X, Patel S, Nakamura T, et al. Mammalian ASH1L is a histone methyltransferase that occupies the transcribed region of active genes. *Mol Cell Biol* 2007;27:8466–79. [PubMed: 17923682]
56. Gaozza E, Baker SJ, Vora RK, Reddy EP. AATYK: a novel tyrosine kinase induced during growth arrest and apoptosis of myeloid cells. *Oncogene* 1997;15:3127–35. [PubMed: 9444961]
57. Zuchner S, Noureddine M, Kennerson M, Verhoeven K, Claeys K, De Jonghe P, et al. Mutations in the pleckstrin homology domain of dynamin 2 cause dominant intermediate Charcot-Marie-Tooth disease. *Nature Genet* 2005;37:289–94. [PubMed: 15731758]
58. Bitoun M, Maugendre S, Jeannot PY, Lacene E, Ferrer X, Laforet P, et al. Mutations in dynamin 2 cause dominant centronuclear myopathy. *Nature Genet* 2005;37:1207–9. [PubMed: 16227997]
59. Tecott LH, Sun LM, Akana SF, Strack AM, Lowenstein DH, Dallman MF, et al. Eating disorder and epilepsy in mice lacking 5-HT<sub>2c</sub> serotonin receptors. *Nature* 1995;374:542–6. [PubMed: 7700379]
60. Gurevich I, Tamir H, Arango V, Dwork AJ, Mann JJ, Schmauss C. Altered editing of serotonin 2C receptor pre-mRNA in the prefrontal cortex of depressed suicide victims. *Neuron* 2002;34:349–56. [PubMed: 11988167]

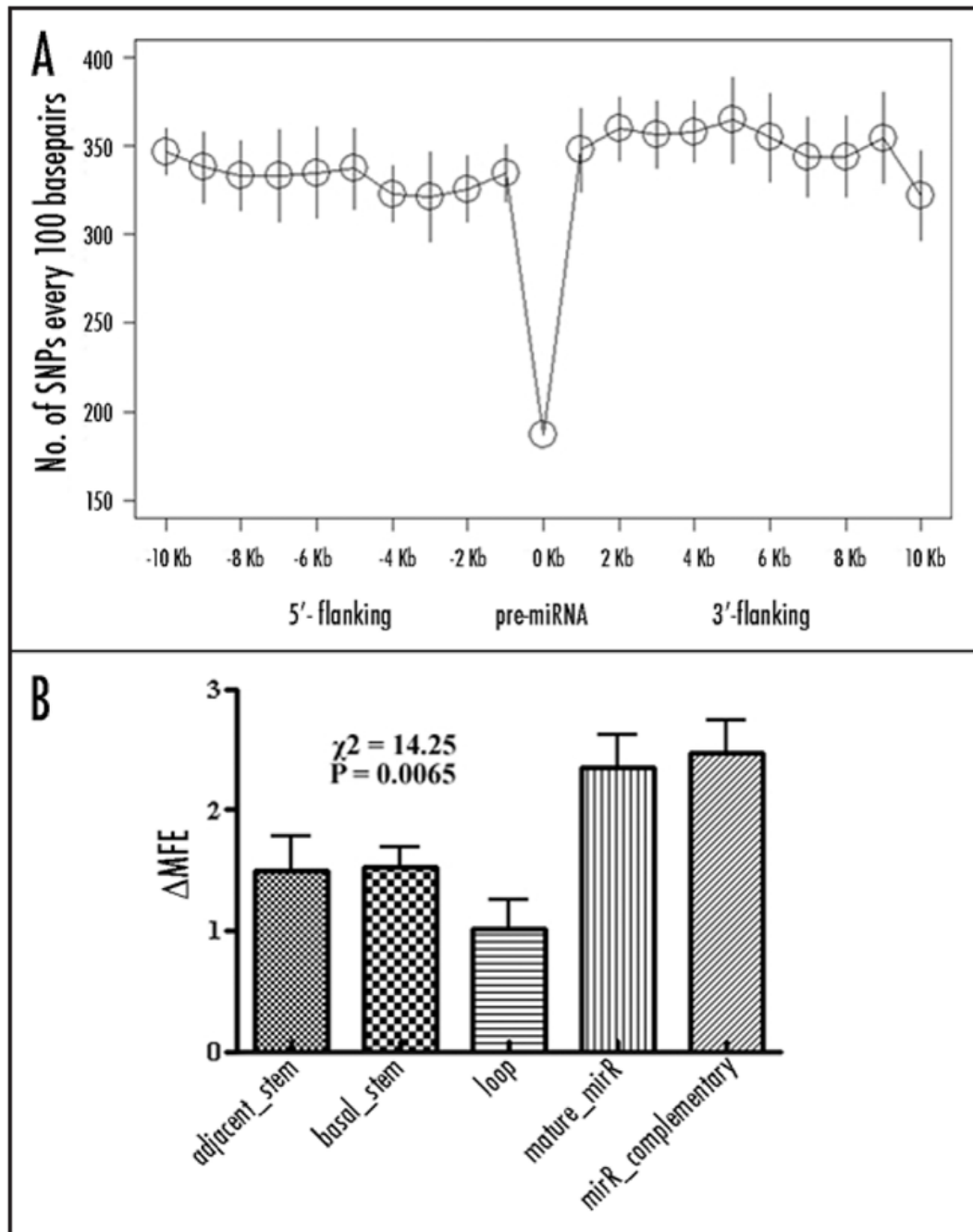
61. Ji SP, Zhang Y, Van Cleemput J, Jiang W, Liao M, Li L, et al. Disruption of PTEN coupling with 5-HT<sub>2C</sub> receptors suppresses behavioral responses induced by drugs of abuse. *Nat Med* 2006;12:324–9. [PubMed: 16474401]
62. Reynolds GP, Zhang ZJ, Zhang XB. Association of antipsychotic drug-induced weight gain with a 5-HT<sub>2C</sub> receptor gene polymorphism. *Lancet* 2002;359:2086–7. [PubMed: 12086765]
63. Lund E, Guttinger S, Calado A, Dahlberg JE, Kutay U. Nuclear export of microRNA precursors. *Science* 2004;303:95–8. [PubMed: 14631048]
64. Janowski BA, Huffman KE, Schwartz JC, Ram R, Nordsell R, Shames DS, et al. Involvement of AGO1 and AGO2 in mammalian transcriptional silencing. *Nat Struct Mol Biol* 2006;13:787–92. [PubMed: 16936728]
65. Peters L, Meister G. Argonaute proteins: mediators of RNA silencing. *Mol Cell* 2007;26:611–23. [PubMed: 17560368]
66. Grivna ST, Pyhtila B, Lin H. MIWI associates with translational machinery and PIWI-interacting RNAs (piRNAs) in regulating spermatogenesis. *Proc Nat Acad Sci USA* 2006;103:13415–20. [PubMed: 16938833]
67. Yu B, Bi L, Zheng B, Ji L, Chevalier D, Agarwal M, et al. The FHA domain proteins DAWDLE in *Arabidopsis* and SNIP1 in humans act in small RNA biogenesis. *Proc Nat Acad Sci USA* 2008;105:10073–8. [PubMed: 18632581]
68. Landthaler M, Yalcin A, Tuschl T. The human DiGeorge syndrome critical region gene 8 and its *D. melanogaster* homolog are required for miRNA biogenesis. *Curr Biol* 2004;14:2162–7. [PubMed: 15589161]
69. Mourelatos Z, Dostie J, Paushkin S, Sharma A, Charroux B, Abel L, et al. miRNPs: a novel class of ribonucleoproteins containing numerous microRNAs. *Genes Dev* 2002;16:720–8. [PubMed: 11914277]
70. Papagiannakopoulos T, Kosik KS. MicroRNAs: regulators of oncogenesis and stemness. *BMC Med* 2008;6:15. [PubMed: 18577221]
71. Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, Zahler AM, et al. The human genome browser at UCSC. *Genome Res* 2002;12:996–1006. [PubMed: 12045153]
72. Mi H, Lazareva-Ulitsky B, Loo R, Kejariwal A, Vandergriff J, Rabkin S, et al. The PANTHER database of protein families, subfamilies, functions and pathways. *Nucleic Acids Res* 2005;33:284–8.
73. Zeng Y, Cullen BR. Structural requirements for pre-microRNA binding and nuclear export by Exportin 5. *Nucleic Acids Res* 2004;32:4776–85. [PubMed: 15356295]
74. Horikawa Y, Wood CG, Yang H, Zhao H, Ye Y, Gu J, et al. Single nucleotide polymorphisms of microRNA machinery genes modify the risk of renal cell carcinoma. *Clin Cancer Res* 2008;14:7956–62. [PubMed: 19047128]



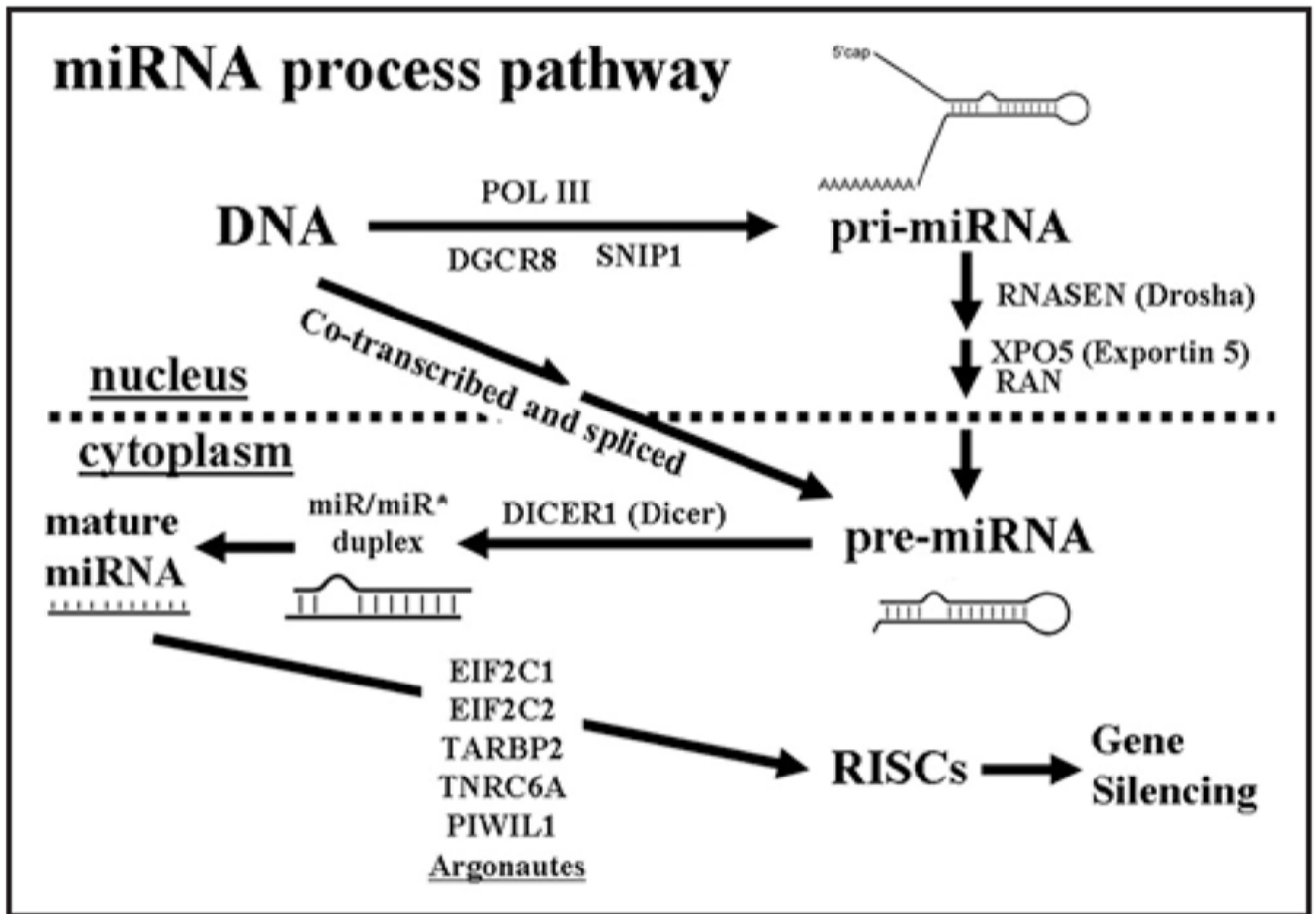
**Figure 1.** Genomic distribution of the pre-miRNAs in humans. The ticks in the right of ideogram are the locations of pre-miRNAs. The darker bands in the ideogram are AT-rich, while the lighter bands are GC-rich.



**Figure 2.**  
Gene length comparison between all PCGs and miRNA host PCGs.



**Figure 3.**  
(A) SNP densities in the human pre-miRNA loci. (B) MFE changes in miRNA domains.



**Figure 4.** miRNA process pathway. The figure is recreated according to refs.<sup>2,3,64–69,73</sup>

**Table 1**

The studies on genetic variants related to miRNAs in humans

Discoveries of genetic variants related to miRNAs	Reference
10 SNPs in miRNA precursors	24
339 SNPs in conserved seed-matching regions of target gene	25
483 SNPs in conserved seed-matching regions of target gene	26
1 SNP in the seed-matching region of target gene	27
A database with 22758 SNPs in the miRNA target sites or with potential ability to create novel miRNA target sites	28
12 SNPs in miRNA precursors	29
1 SNP in the seed-matching region of target gene	30
About 400 SNPs in the miRNA target sites; 250 SNPs with the potential ability to create novel miRNA target sites	31
265 SNPs in the miRNA target sites	32
79 SNPs in the seed-matching regions of target genes; 7 SNPs in miRNA precursors; 1 SNP in the mature miRNA	33
57 SNPs in the seed-matching regions of target genes	15
1 SNP in the seed-matching region of target gene	8
A joint database with SNPs in the miRNAs and their targets	35
One SNP (rs11614913) in hsa-mir-196a2	9, 20
One functional SNP in a miRNA target site.	16
41 genetic variations in 26 microRNA-related genes.	22
One SNP in a let-7 microRNA complementary site.	21
23 SNPs in 11 genes in the miRNA biogenesis pathway, 7 SNPs in 7 pre-miRNAs, and 10 SNPs in 8 pri-miRNAs	74
One SNP (rs2910164) in pre-miR-146a	18, 19
27 SNPs in hairpin structures of pre-miRNAs	36
188 SNPs in the miRNA precursors; 497 consensus SNPs in the seed-matching regions of target genes; 385 CNVs harboring miRNA precursors; 9 CNVs covering important miRNA processing genes.	Present study

Table 2

Pathways and GO analysis of pre-miRNA host genes

Categories	Total	Observed	Expected	+/-	Bonferroni P
<b>Pathways</b>					
Insulin/IGF pathway-mitogen activated protein kinase kinase/MAP kinase cascade	49	8	0.58	+	0.0000306
Coenzyme A biosynthesis	7	3	0.08	+	0.0149
Unclassified	22436	246	267.32	-	0.0418
<b>Biological Process</b>					
Biological process unclassified	11321	99	134.89	-	0.00053
Neuronal activities	569	17	6.78	+	0.0178
Protein phosphorylation	660	20	7.86	+	0.0316
Cell cycle	1009	24	12.02	+	0.0366
Cell structure and motility	1148	26	13.68	+	0.0459
<b>Molecular Function</b>					
Molecular function unclassified	10934	96	130.27	-	0.00102
Kinase	684	18	8.15	+	0.048



Table 3

SNPs in the hairpin structures of pre-miRNAs

pre_name	SNP	Allele	SNP_position	Change of MFE (% of changes) <sup>d</sup>	Location	Reference mature_miR
mir-663b	rs62165009	A/G	chr2: 132731049	2.5 (5.20%)	Adjacent_stem	miR-663b
mir-559	rs58450758	C/T	chr2: 47458370	1.8 (3.05%)	Adjacent_stem	miR-559
mir-1324	rs7614638	C/T	chr3: 75762634	0.1 (0.31%)	Adjacent_stem	miR-1324
mir-1324	rs3008994	C/G	chr3: 75762635	3 (10.17%)	Adjacent_stem	miR-1324
mir-1303	rs34889453	-/A	chr5: 154045576	1.9 (4.52%)	Adjacent_stem	miR-1303
mir-1303	rs33982250	-/A	chr5: 154045578	3.55 (8.80%)	Adjacent_stem	miR-1303
mir-183	rs41281222	C/T	chr7: 129202042	1.8 (4.49%)	Adjacent_stem	miR-183
mir-1299	rs62555121	A/T	chr9: 68292091	2.32 (7.25%)	Adjacent_stem	miR-1299
mir-612	rs12803915	A/G	chr11: 64968555	0.8 (1.59%)	Adjacent_stem	miR-612
mir-5481	rs11020790	C/T	chr11: 93839358	0.4 (0.96%)	Adjacent_stem	miR-5481
mir-620	rs5801168	-/AT	chr12: 115070809	0.7 (2.50%)	Adjacent_stem	miR-620
mir-300	rs12894467	C/T	chr14: 100577480	0 (0.00%)	Adjacent_stem	miR-300
mir-1233	rs347882	C/G	chr15: 32461618	0.7 (1.53%)	Adjacent_stem	miR-1233
mir-1233	rs347882	C/G	chr15: 32607839	0.7 (1.53%)	Adjacent_stem	miR-1233
mir-1282	rs11269	G/T	chr15: 41873201	0.5 (1.27%)	Adjacent_stem	miR-1282
mir-27a	rs11671784	A/G	chr19: 13808296	0.8 (2.13%)	Adjacent_stem	miR-27a
mir-1302-2	rs11266858	A/G	chr1: 20239	0.5 (0.81%)	basal_stem	miR-1302
mir-1302-2	rs4248191	G/T	chr1: 20274	0.1 (0.16%)	basal_stem	miR-1302
mir-1302-2	rs422363	C/T	chr1: 20352	0.1 (0.16%)	basal_stem	miR-1302
mir-1977	rs9783068	C/T	chr1: 5560668	0.1 (0.49%)	basal_stem	miR-1977
mir-1977	rs41453547	A/G	chr1: 5560669	2.2 (12.02%)	basal_stem	miR-1977
mir-1302-3	rs2441622	A/G	chr2: 114057020	0.1 (0.18%)	basal_stem	miR-1302
mir-1302-3	rs7589328	C/T	chr2: 114057098	1.1 (1.99%)	basal_stem	miR-1302
mir-1302-3	rs6542147	A/G	chr2: 114057133	0.5 (0.89%)	basal_stem	miR-1302
mir-149	rs2292832	C/T	chr2: 241044176	3.9 (7.51%)	basal_stem	miR-149
mir-558	rs35999329	-TGTG	chr2: 32610742	3.8 (9.31%)	basal_stem	miR-558
mir-216a	rs41291179	A/T	chr2: 56069594	1.5 (3.83%)	basal_stem	miR-216a
mir-5481-1	rs34864809	-/G	chr3: 126992064	0 (0.00%)	basal_stem	miR-5481
mir-1324	rs3008993	A/G	chr3: 75762696	0 (0.00%)	basal_stem	miR-1324
mir-577	rs34115976	C/G	chr4: 115797446	4.9 (11.11%)	basal_stem	miR-577
mir-943	rs3034718	-/CT	chr4: 1957986	2.6 (6.09%)	basal_stem	miR-943
mir-943	rs35401110	-/CT	chr4: 1957987	0.6 (1.43%)	basal_stem	miR-943
mir-943	rs1077020	C/T	chr4: 1957991	0.9 (2.12%)	basal_stem	miR-943
mir-1289-2	rs35296450	C/G	chr5: 132791194	1.3 (3.78%)	basal_stem	miR-1289
mir-1289-2	rs35731356	-/G	chr5: 132791203	0.7 (2.07%)	basal_stem	miR-1289
mir-585	rs62376934	A/G	chr5: 168623190	2.9 (5.28%)	basal_stem	miR-585
mir-9-2	rs41265488	A/T	chr5: 87998503	0.7 (1.78%)	basal_stem	miR-9
mir-583	rs10697860	-/ATAAA	chr5: 95440607	0 (0.00%)	basal_stem	miR-583
mir-339	rs13232101	G/T	chr7: 1029100	0 (0.00%)	basal_stem	miR-339-3p
mir-595	rs4909237	C/T	chr7: 158018264	0.6 (1.90%)	basal_stem	miR-595
mir-489	rs35930643	-/A	chr7: 92951259	3.2 (8.99%)	basal_stem	miR-489
mir-1208	rs56863230	C/G	chr8: 129231548	0 (0.00%)	basal_stem	miR-1208
mir-1208	rs2648841	G/T	chr8: 129231615	3.5 (15.84%)	basal_stem	miR-1208
mir-486	rs59908561	-/G	chr8: 41637116	0 (0.00%)	basal_stem	miR-486-3p
mir-1302-2	rs11266858	A/G	chr9: 20154	0.5 (0.81%)	basal_stem	miR-1302
mir-1302-2	rs4248191	G/T	chr9: 20189	0.1 (0.16%)	basal_stem	miR-1302
mir-1302-2	rs422363	C/T	chr9: 20267	0.1 (0.16%)	basal_stem	miR-1302
mir-202	rs12355840	C/T	chr10: 134911103	0.3 (0.51%)	basal_stem	miR-202
mir-605	rs2043556	C/T	chr10: 52729412	2.6 (4.97%)	basal_stem	miR-605
mir-1908	rs174561	C/T	chr11: 61339284	3.2 (7.08%)	basal_stem	miR-1908
mir-194-2	rs11231898	A/G	chr11: 64415412	1.9 (3.85%)	basal_stem	miR-194
mir-612	rs550894	A/C	chr11: 64968516	0.2 (0.39%)	basal_stem	miR-612

pre_name	SNP	Allele	SNP_position	Change of MFE (% of changes) <sup>d</sup>	Location	Reference mature_miR
miR-548c	rs17120527	A/G	chr12: 63302567	0 (0.00%)	basal_stem	miR-548c-3p
miR-141	rs34385807	-/C	chr12: 6943605	4.6 (9.35%)	basal_stem	miR-141
miR-617	rs12815353	C/G	chr12: 79750457	0 (0.00%)	basal_stem	miR-617
miR-492	rs2289030	C/G	chr12: 93752417	1.7 (4.27%)	basal_stem	miR-492
miR-622	rs59274393	C/T	chr13: 89681529	N.A.	basal_stem	miR-622
miR-18a	rs41275866	C/G	chr13: 90801010	0.3 (1.36%)	basal_stem	miR-18a*
miR-329-1	rs34557733	-/A	chr14: 100562882	0 (0.00%)	basal_stem	miR-329
miR-1185-2	rs11844707	A/G	chr14: 100580366	3.3 (9.37%)	basal_stem	miR-1185
miR-624	rs57264777	A/T	chr14: 30553694	N.A.	basal_stem	miR-624
miR-1260	rs28909969	-/T	chr14: 76802381	3.1 (17.67%)	basal_stem	miR-1260
miR-1302-2	rs4222363	C/T	chr15: 100318199	0.1 (0.16%)	basal_stem	miR-1302
miR-1302-2	rs4248191	G/T	chr15: 100318277	0.1 (0.16%)	basal_stem	miR-1302
miR-1302-2	rs11266858	A/G	chr15: 100318312	0.5 (0.81%)	basal_stem	miR-1302
miR-211	rs34520022	-/G	chr15: 29144537	0.8 (1.77%)	basal_stem	miR-211
miR-147b	rs56073218	C/G	chr15: 43512547	6 (21.05%)	basal_stem	miR-147b
miR-7	rs41276930	C/T	chr15: 86956077	1.7 (3.62%)	basal_stem	miR-7
miR-140	rs7205289	A/C	chr16: 68524506	2.4 (4.56%)	basal_stem	miR-140-3p
miR-423	rs6505162	A/C	chr17: 25468309	0 (0.00%)	basal_stem	miR-423-3p
miR-1253	rs7217038	A/T	chr17: 2598127	0 (0.00%)	basal_stem	miR-1253
miR-193a	rs60406007	G/T	chr17: 26911146	4 (8.55%)	basal_stem	miR-193a-5p
miR-365-2	rs351143473	-/T	chr17: 26926632	1.5 (3.89%)	basal_stem	miR-365
miR-187	rs41274312	A/G	chr18: 31738790	0.81 (1.72%)	basal_stem	miR-187
miR-639	rs35149836	A/G	chr19: 14501439	0 (0.00%)	basal_stem	miR-639
miR-1302-2	rs11266858	A/G	chr19: 22983	0.5 (0.81%)	basal_stem	miR-1302
miR-1302-2	rs4248191	G/T	chr19: 23018	0.1 (0.16%)	basal_stem	miR-1302
miR-1302-2	rs4222363	C/T	chr19: 23096	0.1 (0.16%)	basal_stem	miR-1302
miR-1283-1	rs71111412	A/G	chr19: 58883555	4.8 (12.24%)	basal_stem	miR-1283
miR-521-2	rs13382089	G/T	chr19: 58911666	4.3 (14.14%)	basal_stem	miR-521
miR-516b-2	rs10583889	-/TT	chr19: 58920586	0.9 (2.41%)	basal_stem	miR-516b*
miR-518a-1	rs61636451	A/G	chr19: 58926153	N.A.	basal_stem	miR-518a-3p
miR-516a-1	rs2569389	A/G	chr19: 58951814	4.1 (8.99%)	basal_stem	miR-516a-3p
miR-220b	rs1053262	C/G	chr19: 6447045	0.2 (0.53%)	basal_stem	miR-220b
miR-663	rs7266947	A/C	chr20: 26136912	1.1 (2.19%)	basal_stem	miR-663
miR-499	rs7267163	C/T	chr20: 33041937	2.2 (3.66%)	basal_stem	miR-499-3p
miR-646	rs6513496	C/T	chr20: 58316929	2.4 (7.02%)	basal_stem	miR-646
miR-1-1	rs6122014	C/T	chr20: 60561960	0.9 (3.00%)	basal_stem	miR-1
miR-941-1	rs56202554	C/T	chr20: 62021328	1.9 (3.63%)	basal_stem	miR-941
miR-941-1	rs5795631	C/T	chr20: 62021321	0 (0.00%)	basal_stem	miR-941
miR-941-1	rs6089780	A/G	chr20: 62021324	4.1 (8.17%)	basal_stem	miR-941
miR-650	rs5966397	C/G	chr22: 21495340	0.8 (2.22%)	basal_stem	miR-650
miR-548j	rs4822739	C/G	chr22: 25281185	3.1 (6.13%)	basal_stem	miR-548j
miR-146a	rs61270459	C/G	chr5: 159849984	N.A.	loop	miR-146a
miR-1274a	rs3180339	C/T	chr5: 41511523	3.5 (17.95%)	loop	miR-1274a
miR-581	rs788517	A/G	chr5: 53283143	0 (0.00%)	loop	miR-581
miR-96	rs41274239	A/G	chr7: 129201810	1 (2.91%)	loop	miR-96
miR-1307	rs7911488	A/G	chr10: 105144079	0 (0.00%)	loop	miR-1307
miR-2110	rs17091403	C/T	chr10: 115923895	1.5 (4.40%)	loop	miR-2110
miR-1265	rs11259096	C/T	chr10: 14518624	1 (2.00%)	loop	miR-1265
miR-620	rs10549054	-/TA	chr12: 115070798	0.3 (1.09%)	loop	miR-620
miR-656	rs58834075	C/T	chr14: 100602846	0.6 (2.60%)	loop	miR-656
miR-1233	rs347881	C/T	chr15: 32461601	1.6 (3.44%)	loop	miR-1233
miR-27a	rs895819	C/T	chr15: 32607822	1.6 (3.44%)	loop	miR-27a*
miR-639	rs45556632	C/G	chr19: 13808292	0 (0.00%)	loop	miR-639
miR-516b-2	rs10670323	-/AAAGA	chr19: 14501403	1.7 (4.09%)	loop	miR-516b
			chr19: 58920554	2.1 (5.65%)	loop	

pre_name	SNP	Allele	SNP_position	Change of MFE (% of changes) <sup>d</sup>	Location	Reference mature_miR
miR-516b-2	rs33953969	-/AAAAGA	chr19: 58920555	2.1 (5.65%)	loop	miR-516b*
miR-521-1	rs2561251	A/G	chr19: 58943749	N.A.	loop	miR-521
miR-650	rs11558654	A/T	chr22: 21495309	0 (0.00%)	loop	miR-650
miR-92b	rs12759620	C/G	chr1: 153431668	3.7 (5.60%)	loop	miR-92b
miR-1977	rs2854138	A/G	chr1: 556124	1.8 (9.63%)	mature_miR	miR-1977
miR-34a	rs35301225	A/C/T	chr1: 9134389	4.7 (10.22%)	mature_miR	miR-34a
miR-1978	rs55723650	A/G	chr2: 149355840	1.8 (13.14%)	mature_miR	miR-1978
miR-1978	rs56489998	C/T	chr2: 149355844	0 (0.00%)	mature_miR	miR-1978
miR-568	rs28632138	G/T	chr3: 115518077	1.5 (4.92%)	mature_miR	miR-568
miR-1324	rs10155043	C/T	chr3: 75762664	0 (0.00%)	mature_miR	miR-1324
miR-1255b-1	rs6841938	A/G	chr4: 36104443	2.5 (17.01%)	mature_miR	miR-1255b
miR-585	rs62376935	G/T	chr5: 168623213	7.1 (14.85%)	mature_miR	miR-585
miR-581	rs1694089	C/T	chr5: 53283151	0.7 (1.84%)	mature_miR	miR-581
miR-581	rs810917	A/G	chr5: 53283157	6.2 (19.07%)	mature_miR	miR-581
miR-449b	rs10061133	A/G	chr5: 54502301	0 (0.00%)	mature_miR	miR-449b
miR-590	rs6971711	C/T	chr7: 73243535	2.4 (8.16%)	mature_miR	miR-590-3p
miR-25	rs41274221	C/T	chr7: 99529136	1.8 (5.00%)	mature_miR	miR-25
miR-1322	rs59878596	C/T	chr8: 10720304	0.1 (0.89%)	mature_miR	miR-1322
miR-596	rs61388742	C/T	chr8: 1752832	2.8 (8.31%)	mature_miR	miR-596
miR-608	rs58078477	C/G	chr10: 102724756	N.A.	mature_miR	miR-608
miR-608	rs4919510	C/G	chr10: 102724768	1.4 (4.23%)	mature_miR	miR-608
miR-938	rs12416605	C/T	chr10: 29931266	1.1 (3.05%)	mature_miR	miR-938
miR-606	rs34610391	-/A	chr10: 76982290	0 (0.00%)	mature_miR	miR-606
miR-5481	rs13447640	A/G	chr11: 93839369	2.5 (6.39%)	mature_miR	miR-5481
miR-431	rs12884005	A/G	chr14: 100417161	0 (0.00%)	mature_miR	miR-431*
miR-379	rs61991156	A/G	chr14: 100558167	0.6 (2.31%)	mature_miR	miR-379
miR-299	rs41286566	C/T	chr14: 100559898	2.9 (7.71%)	mature_miR	miR-299-5p
miR-412	rs61992671	A/G	chr14: 100601607	1.6 (4.61%)	mature_miR	miR-412
miR-208b	rs2754157	A/T	chr14: 22957059	1.5 (5.03%)	mature_miR	miR-208b
miR-154	rs41286570	C/G	chr14: 100595880	0.4 (1.1%)	mature_miR	miR-154
miR-1268	rs28599926	A/T	chr15: 20014635	0.7 (2.73%)	mature_miR	miR-1268
miR-627	rs2620381	A/C	chr15: 40279140	4.7 (9.07%)	mature_miR	miR-627
miR-1276	rs34381260	-/T	chr15: 84114798	4.1 (13.95%)	mature_miR	miR-1276
miR-940	rs35356504	-/C	chr16: 2261821	0 (0.00%)	mature_miR	miR-940
miR-662	rs9745376	A/G	chr16: 760250	0.5 (1.11%)	mature_miR	miR-662
miR-548h-3	rs9913045	A/G	chr17: 13387649	2.5 (4.85%)	mature_miR	miR-548h
miR-423	rs61093106	A/C	chr17: 25468297	N.A.	mature_miR	miR-423-3p
miR-122	rs41292412	C/T	chr18: 54269338	2.5 (5.67%)	mature_miR	miR-122*
miR-125a	rs12975333	G/T	chr19: 56888340	6.1 (14.52%)	mature_miR	miR-125a-5p
miR-520c	rs7255628	C/G	chr19: 58902546	6.6 (14.77%)	mature_miR	miR-520c-5p
miR-518e	rs34416818	-/A	chr19: 58924924	4.4 (10.24%)	mature_miR	miR-518e*
miR-499	rs3746444	A/G	chr19: 58924924	0.4 (0.65%)	mature_miR	miR-499-3p
miR-646	rs6513497	G/T	chr20: 58317000	0.8 (2.40%)	mature_miR	miR-646
miR-124-3	rs34059726	G/T	chr20: 33041912	6.5 (19.23%)	mature_miR	miR-124
miR-941-2	rs34604519	C/G	chr20: 62021613	3.6 (6.63%)	mature_miR	miR-941
miR-941-3	rs35544770	A/G	chr20: 62021716	3.6 (6.63%)	mature_miR	miR-941
miR-941-3	rs12625454	C/G	chr20: 62021725	3.6 (6.63%)	mature_miR	miR-941
miR-1302-2	rs11266859	A/G	chr1: 20279	0.4 (0.64%)	mature_miR	miR-941
miR-1302-2	rs422582	A/C	chr1: 20287	6 (10.60%)	miR_complementary	miR-1302
miR-1977	rs9701099	C/T	chr1: 556076	1.1 (5.37%)	miR_complementary	miR-1977
miR-1302-3	rs2441621	G/T	chr2: 114057085	6 (10.62%)	miR_complementary	miR-1302
miR-1244	rs1804520	A/G	chr2: 232286288	1.3 (7.78%)	miR_complementary	miR-1244
miR-217	rs41291173	A/G	chr2: 56063644	0.4 (1.20%)	miR_complementary	miR-217
miR-570	rs9860655	C/T	chr3: 196911485	4.1 (10.25%)	miR_complementary	miR-570
miR-564	rs2292181	C/G	chr3: 44878438	1.9 (3.61%)	miR_complementary	miR-564

pre_name	SNP	Allele	SNP_position	Change of MFE (% of changes) <sup>a</sup>	Location	Reference mature_miR
mir-1324	rs28620398	G/T	chr3: 75762617	3.1 (10.54%)	mirR_complementary	mir-1324
mir-1255a	rs28664200	C/T	chr4: 102470524	0.3 (0.49%)	mirR_complementary	mir-1255a
mir-1244	rs1804520	A/G	chr5: 118338200	1.3 (7.78%)	mirR_complementary	mir-1244
mir-1294	rs13186787	A/G	chr5: 153706962	0.7 (1.01%)	mirR_complementary	mir-1294
mir-146a	rs2910164	C/G	chr5: 159844996	2.8 (6.95%)	mirR_complementary	mir-146a
mir-1229	rs2291418	C/T	chr5: 179157930	0 (0.00%)	mirR_complementary	mir-1229
mir-548a-1	rs12197631	G/T	chr6: 18680035	2.51 (7.59%)	mirR_complementary	mir-548a-3p
mir-1206	rs2114358	A/G	chr8: 129090361	0.5 (2.65%)	mirR_complementary	mir-1206
mir-939	rs35486628	-G	chr8: 145590185	0 (0.00%)	mirR_complementary	mir-939
mir-1234	rs2291134	C/G	chr8: 145596344	4.2 (8.38%)	mirR_complementary	mir-1234
mir-1302-2	rs11266859	A/G	chr9: 20194	0.4 (0.64%)	mirR_complementary	mir-1302
mir-1302-2	rs422582	A/C	chr9: 20202	6 (10.60%)	mirR_complementary	mir-1302
mir-603	rs11014002	C/T	chr10: 24604659	1.8 (4.44%)	mirR_complementary	mir-603
mir-604	rs2368393	A/G	chr10: 29874004	0.1 (0.37%)	mirR_complementary	mir-604
mir-604	rs2368392	A/G	chr10: 29874009	0.6 (2.25%)	mirR_complementary	mir-604
mir-607	rs12778876	A/T	chr10: 98578485	4.6 (6.79%)	mirR_complementary	mir-607
mir-607	rs12780546	A/T	chr10: 98578486	4.5 (6.64%)	mirR_complementary	mir-607
mir-1304	rs2155248	G/T	chr11: 93106514	6.5 (9.69%)	mirR_complementary	mir-1304
mir-619	rs34651680	-G	chr12: 107754861	0 (0.00%)	mirR_complementary	mir-619
mir-620	rs3043743	-T/T/A	chr12: 115070818	0.4 (1.65%)	mirR_complementary	mir-620
mir-620	rs34380284	-C	chr12: 115070819	2.9 (11.60%)	mirR_complementary	mir-620
mir-1178	rs7311975	C/T	chr12: 118635876	0 (0.00%)	mirR_complementary	mir-1178
mir-1244	rs1804520	A/G	chr12: 12156173	1.3 (7.78%)	mirR_complementary	mir-1244
mir-196a-2	rs11614913	A/G	chr12: 52671866	4.6 (9.87%)	mirR_complementary	mir-196a
mir-618	rs2682818	A/C	chr12: 79853667	3.5 (10.12%)	mirR_complementary	mir-618
mir-1244	rs1804520	A/G	chr12: 9283394	1.3 (7.78%)	mirR_complementary	mir-1244
mir-92a-1	rs9589207	A/G	chr13: 90801590	0 (0.00%)	mirR_complementary	mir-92a
mir-453	rs56103835	C/T	chr14: 100592309	0.3 (1.16%)	mirR_complementary	mir-453
mir-625	rs12894182	A/C	chr14: 65007642	6.5 (9.50%)	mirR_complementary	mir-625
mir-1302-2	rs422582	A/C	chr15: 100318264	6 (10.60%)	mirR_complementary	mir-1302
mir-1302-2	rs11266859	A/G	chr15: 100318272	0.4 (0.64%)	mirR_complementary	mir-1302
mir-631	rs5745925	-CT	chr15: 73433019	6.5 (20.63%)	mirR_complementary	mir-631
mir-1826	rs62030476	A/G	chr16: 33873090	0 (0.00%)	mirR_complementary	mir-1826
mir-1972	rs57629257	C/T	chr16: 68621762	2.7 (7.62%)	mirR_complementary	mir-1972
mir-633	rs17759989	A/G	chr17: 58375343	0.6 (1.80%)	mirR_complementary	mir-633
mir-1181	rs2569788	C/G	chr19: 10375159	2.2 (5.66%)	mirR_complementary	mir-1181
mir-1302-2	rs11266859	A/G	chr19: 23023	0.4 (0.64%)	mirR_complementary	mir-1302
mir-1302-2	rs422582	A/C	chr19: 23031	6 (10.60%)	mirR_complementary	mir-1302
mir-520h	rs56013413	A/G	chr19: 58937600	4.1 (12.57%)	mirR_complementary	mir-520h
mir-663	rs28670321	C/T	chr20: 26136824	0 (0.00%)	mirR_complementary	mir-663
mir-663	rs2019798	G/T	chr20: 26136846	0.5 (1.01%)	mirR_complementary	mir-663
mir-645	rs35645123	-A	chr20: 48635761	0 (0.00%)	mirR_complementary	mir-645
mir-941-1	rs7268785	C/G	chr20: 62021250	3.6 (6.63%)	mirR_complementary	mir-941
mir-941-1	rs2427556	A/G	chr20: 62021268	4.1 (8.17%)	mirR_complementary	mir-941
mir-941-3	rs12625445	C/G	chr20: 62021669	3.6 (6.63%)	mirR_complementary	mir-941
mir-548j	rs12161068	C/T	chr22: 25281215	0.4 (0.75%)	mirR_complementary	mir-548j

<sup>a</sup>N.A. stands for inconsistent allele reports existed.

Table 4

Representative pre-miRNAs in the CNV regions

pre-miRNA ID	pre-miRNA location (hg18)	Host gene	CNV ID	CNV position (hg18)	Observed CNVs
mir-200b	chr1:1092347-1092441(+)	-	Variation_30362	chr1:702445-1697636	11 gains
mir-200a	chr1:1093106-1093195(+)	-	Variation_30362	chr1:702445-1697636	11 gains
mir-429	chr1:1094248-1094330(+)	-	Variation_30362	chr1:702445-1697636	11 gains
mir-320b-1	chr1:117015894-117015972(+)	-	Variation_4243	chr1:116927828-117128034	11 gains
mir-555	chr1:153582765-153582860(-)	ASHIL	Variation_6789	chr1:153489907-154184585	39 losses
mir-1302-2	chr1:20229-20366(+)	-	Variation_30360	chr1:1794-115824	25 gains
mir-663b	chr2:132731009-132731123(-)	LOC100133239	Variation_31014	chr2:132726460-132763489	19 gains
mir-570	chr3:196911452-196911548(+)	-	Variation_4491	chr3:196584076-196965419	188 gains
mir-566	chr3:50185763-50185856(+)	SEMA3F	Variation_2435	chr3:50173490-50368468	18 losses
mir-1324	chr3:75762604-75762699(+)	-	Variation_2462	chr3:75474768-76085726	18 losses
mir-218-1	chr4:20138996-20139105(+)	SLIT2	Variation_4380	chr4:20111993-20290054	3 gains, 13 losses
mir-95	chr4:8057928-8058008(-)	ABLJM2	Variation_4373	chr4:8009599-8099960	9 gains, 18 losses
mir-548i-2	chr4:9166887-9167035(-)	-	Variation_2069	chr4:9010036-9203157	10 losses
mir-1236	chr6:32032595-32032696(-)	RDBP	Variation_4492	chr6:31995533-32055579	42 losses
mir-548o	chr7:101833194-101833307(-)	PRKRIP1	Variation_4553	chr7:10167725-102083105	36 losses
mir-1183	chr7:21477201-21477289(+)	SP4	Variation_4527	chr7:21455625-21654541	20 gains
mir-939	chr8:145590172-145590253(-)	CPSF1	Variation_4613	chr8:145536611-145740218	16 losses
mir-1234	chr8:145596284-145596367(-)	CPSF1	Variation_4613	chr8:145536611-145740218	16 losses
mir-548i-3	chr8:7983873-7984021(-)	-	Variation_2116	chr8:7917018-8067760	25 gains, 34 losses
let-7a-1	chr9:95978060-95978139(+)	-	Variation_4645	chr9:95873863-96081830	18 gains
let-7f-1	chr9:95978450-95978536(+)	-	Variation_4645	chr9:95873863-96081830	18 gains
let-7d	chr9:95980937-95981023(+)	LOC158257	Variation_4645	chr9:95873863-96081830	18 gains
mir-202	chr10:134911006-134911115(-)	-	Variation_2896	chr10:134868158-135282675	20 gains, 1 loss
mir-1268	chr15:20014593-20014644(-)	-	Variation_3070	chr15:18403665-21241985	204 gains, 24 losses
mir-1233	chr15:32461562-32461643(-)	-	Variation_7058	chr15:29769358-32654590	47 gains, 48 losses
mir-657	chr17:76713671-76713768(-)	AA TK	Variation_5036	chr17:76600967-76762177	13 losses
mir-338	chr17:76714278-76714344(-)	AA TK	Variation_5036	chr17:76600967-76762177	13 losses
mir-1250	chr17:76721591-76721703(-)	AA TK	Variation_5036	chr17:76600967-76762177	13 losses
mir-199a-1	chr19:10789102-10789172(-)	DNM2	Variation_5087	chr19:10728678-10897044	11 losses
mir-1270	chr19:1767158-1767237(-)	REXO1	Variation_7191	chr19:1271267-1950204	11 losses
mir-1227	chr19:20371080-20371162(-)	FLN44894	Variation_3183	chr19:20360525-20566187	13 gains, 10 losses
mir-220c	chr19:2185061-2185148(-)	PLEKHJ1	Variation_5068	chr19:2138091-2323221	35 losses
mir-150	chr19:53755341-53755423(-)	SLIT2B1	Variation_32261	chr19:53097718-55337070	17 losses
mir-663	chr19:54695854-54695937(-)	LOC100128528	Variation_5111	chr19:54643858-54765745	18 losses
mir-124-3	chr20:26136822-26136914(-)	LOC284801	Variation_31037	chr20:26136626-26139184	25 losses
mir-185	chr20:61280297-61280383(+)	-	Variation_5147	chr20:61234049-61347722	10 gains
mir-649	chr22:18400662-18400743(+)	C22orf25	Variation_2261	chr22:18259187-18435258	10 losses
mir-650	chr22:19718465-19718561(-)	-	Variation_5170	chr22:19664133-19854524	12 gains
mir-1912	chr22:21495270-21495365(+)	IGL@	Variation_2268	chr22:21394879-21570697	35 gains, 32 losses
mir-1264	chrX:113792275-113792354(+)	HTR2C	Variation_7758	chrX:113724807-114050880(+)	4 gains, 9 losses
mir-1298	chrX:113793386-113793454(+)	HTR2C	Variation_7758	chrX:113724807-114050880(+)	4 gains, 9 losses
mir-1911	chrX:113855906-113856017(+)	HTR2C	Variation_7758	chrX:113724807-114050880(+)	4 gains, 9 losses
mir-1911	chrX:113904000-113904079(+)	HTR2C	Variation_7758	chrX:113724807-114050880(+)	4 gains, 9 losses
mir-448	chrX:113964273-113964383(+)	HTR2C	Variation_7758	chrX:113724807-114050880(+)	4 gains, 9 losses

**Table 5**

SNPs in the seed-matching regions of miRNA target genes

SNP <sup>a</sup>	miRNA	Target	Prediction method
rs56788643	let-7g	<i>HMGA2</i>	PITA
rs35180728	miR-1	<i>ARCN1</i>	TargetScanS
rs36076633	miR-1	<i>TAGLN2</i>	TargetScanS
rs8829	miR-101	<i>EZH2</i>	PITA
<b>rs3783620</b>	miR-126	<i>VCAM1</i>	PITA
rs34335657	miR-129	<i>CAMTA1</i>	PITA
rs41286082	miR-141	<i>KLF5</i>	PITA
<b>rs1621</b>	miR-199a*	<i>MET</i>	PITA
rs34954531	miR-30a-3p	<i>VEZT</i>	PITA
rs1051780	miR-34	<i>VAMP2</i>	TargetScanS
<b>rs17620927</b>	miR-373	<i>MKRN1</i>	PITA
rs56788643	miR-98	<i>HMGA2</i>	PITA

<sup>a</sup> SNPs with frequency report are in bold font.

Table 6

Pathways and GO analysis of genes with SNPs in miRNA target sites

Categories	Total	Observed	Expected	+/-	Bonferroni P
<b>Pathway</b>					
Angiogenesis	229	19	3.87	+	0.00000402
Unclassified	22436	340	379.36	-	0.0000077
T cell activation	111	10	1.88	+	0.00426
PDGF signaling pathway	189	13	3.2	+	0.00459
<b>Biological Process</b>					
Developmental processes	2152	104	36.39	+	3.47E-21
Biological process unclassified	11321	107	191.42	-	7.92E-16
Signal transduction	3406	115	57.59	+	5.33E-12
Intracellular signaling cascade	871	46	14.73	+	2.54E-09
mRNA transcription	1914	67	32.36	+	0.0000021
Neurogenesis	587	32	9.93	+	0.00000232
mRNA transcription regulation	1459	56	24.67	+	0.00000251
Mesoderm development	551	30	9.32	+	0.00000506
Ectoderm development	692	34	11.7	+	0.00000702
Nucleoside, nucleotide and nucleic acid metabolism	3343	93	56.53	+	0.0000265
Cell proliferation and differentiation	1028	40	17.38	+	0.0000398
Oncogenesis	472	24	7.98	+	0.0000825
Protein modification	1157	42	19.56	+	0.000559
Neuronal activities	569	25	9.62	+	0.000601
Muscle contraction	198	13	3.35	+	0.00137
Other intracellular signaling cascade	225	15	3.8	+	0.00194
Cell structure and motility	1148	38	19.41	+	0.00248
Receptor protein tyrosine kinase signaling pathway	211	14	3.57	+	0.00402
Protein phosphorylation	660	27	11.16	+	0.00612
Cell cycle	1009	32	17.06	+	0.0187
Cell communication	1213	38	20.51	+	0.0346
Cell surface receptor mediated signal transduction	1638	47	27.7	+	0.0466
<b>Molecular Function</b>					
Molecular function unclassified	10934	105	184.88	-	2.18E-14
Transcription factor	2052	69	34.7	+	0.00000117
Nucleic acid binding	2850	83	48.19	+	0.0000187
Voltage-gated ion channel	145	13	2.45	+	0.000276
Ion channel	357	18	6.04	+	0.00151
Other miscellaneous function protein	427	21	7.22	+	0.00293
Other transcription factor	349	18	5.9	+	0.00631
Miscellaneous function	866	28	14.64	+	0.0286
Other DNA-binding protein	331	16	5.6	+	0.0345
Membrane traffic protein	359	15	6.07	+	0.0415
Kinase	684	23	11.57	+	0.0485

Table 7

Important miRNA-processing genes in the CNV regions

Gene	Entrez ID	Gene location (hg18)	CNV ID	CNV position (hg18)	Observed CNVs <sup>a</sup>
<i>SNIP1</i>	79753	chr1:37774729-37792490(-)	Variation_0006	chr1:37714745-37826968	1 loss
<i>RNA5EN</i>	29102	chr5:31436358-31568039(-)	Variation_3550	chr5:31332034-31505885	N.A.
<i>RNA5EN</i>	29102	chr5:31436358-31568039(-)	Variation_47879	chr5:31412880-31908908	3 losses
<i>XPO5</i>	57510	chr6:43598050-43651642(-)	Variation_9530	chr6:43583452-43604640	1 gain
<i>PIWIL1</i>	9271	chr12:129388567-129422826(+)	Variation_3000	chr12:128577929-129659406	1 gain
<i>PIWIL1</i>	9271	chr12:129388567-129422826(+)	Variation_3901	chr12:128512417-129681529	N.A.
<i>PIWIL1</i>	9271	chr12:129388567-129422826(+)	Variation_8740	chr12:128578742-129654380	1 gain
<i>PIWIL1</i>	9271	chr12:129388567-129422826(+)	Variation_35062	chr12:129417091-129445428	1 gain
<i>DICER1</i>	23405	chr14:94622319-94693512(-)	Variation_5766	chr14:94638002-94644051	1 loss
<i>GEMIN4</i>	50628	chr17:594411-602251(-)	Variation_3136	chr17:595817-897708	2 gains
<i>GEMIN4</i>	50628	chr17:594411-602251(-)	Variation_4021	chr17:568336-1008155	N.A.
<i>GEMIN4</i>	50628	chr17:594411-602251(-)	Variation_5332	chr17:60543-963131	1 gain
<i>DGCR8</i>	54487	chr22:18447834-18479400(+)	Variation_4117	chr22:18406493-18689447	N.A.
<i>DGCR8</i>	54487	chr22:18447834-18479400(+)	Variation_5168	chr22:18267966-18449970	6 losses
<i>DGCR8</i>	54487	chr22:18447834-18479400(+)	Variation_31071	chr22:17399088-19383198	6 gains

<sup>a</sup>N.A. stands for not available.