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# **Insight into Environmental Effects on Bonding and Redox Properties of [4Fe-4S] Clusters in Proteins**

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## **Abstract**



The large differences in redox potentials between the HiPIPs and ferredoxins are generally attributed to hydrogen bonds and electrostatic effects from the protein and solvent. Recent ligand K-edge Xray absorption studies by Solomon and co-workers show that the Fe-S covalencies of [4Fe-4S] clusters in the two proteins differ considerably apparently because of hydrogen bonds from water, indicating electronic effects may be important. However, combined density function theory (DFT) and photoelectron spectroscopy studies by our group and Wang and co-workers indicate that hydrogen bonds tune the potential of [4Fe-4S] clusters by mainly electrostatics. The DFT studies here rationalize both results, namely that the observed change in the Fe-S covalency is due to differences in ligand conformation between the two proteins rather than hydrogen bonds. Moreover, the ligand conformation affects the calculated potentials by  $\sim$  100 mV, and thus is a heretofore unconsidered means of tuning the potential.

> A classical question about oxidation-reduction potentials in proteins is why the [4Fe-4S] clusters in the high potential iron-sulfur proteins (HiPIPs) have a 1-/2- reduction at 100 to 450 mV vs. NHE, while the ferredoxins (Fds) have a 2-/3- reduction at −100 to −645 mV.<sup>1</sup> The differences are generally attributed to hydrogen bonding and electrostatic effects from the surrounding protein and solvent. $2-5$  Recent ligand K-edge X-ray absorption spectroscopy

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(XAS) experiments find large differences in the Fe-S covalency between HiPIPs and Fds apparently due to hydration,<sup>6</sup> and attribute the redox potential differences to electronic effects of water hydrogen bonds based on a correlation between electrochemical redox potentials and metal-ligand bond covalencies in iron-sulfur complexes.<sup>7</sup> However, our combined density functional theory (DFT) and photoelectron spectroscopy (PES) studies indicate hydrogen bond effects are actually primarily electrostatic since the electronic structure and bonding of the clusters are not significantly affected.<sup>8</sup> Here, we present a new mechanism for redox tuning, namely the ligand conformation, which reconciles the seemingly conflicting experimental results.

Due to the complexity of the protein environment, the effects of hydration and ligand conformation on Fe-S covalencies and redox properties are separated using broken symmetry DFT (BS-DFT)<sup>9</sup> calculations of the analog  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup> (Et = ethyl), which is a good model for the  $[Fe_4S_4(Cys)_4]^n$  found in proteins.<sup>8</sup> Redox energies in the gas phase were calculated at the B3LYP/6-31( $++$ )<sub>S</sub>G\*\*//B3LYP/6-31G\*\* level and Fe-L covalencies as measured by the percent ligand (%L) character mixing in the Fe 3d orbitals were obtained from natural bond orbital (NBO) analysis<sup>10</sup> of geometries at the B3LYP/6-31G<sup>\*\*</sup> level. Thus, the calculations focus on a single effect at a time without approximations for the environment and take advantage of our previous calibrations of a variety of clusters against experiment.<sup>11,12</sup> Moreover, using these methods, vertical detachment energies (VDE) correlate well with total %L character for a series of  $[Fe_4S_4L_4]^2$  clusters in the gas phase with different ligands<sup>12</sup>, in agreement with the above mentioned XAS experiments.<sup>6,7</sup> However, the vertical reduction energies (VRE), which are defined similarly to the VDE as the energy to add an electron without allowing relaxation, have a more complicated dependence on the total %L character.

To elucidate hydration effects,  $[Fe_4S_4(SEt)_4]^2$  was compared to hydrated complexes in which a water molecule was placed near each terminal or bridging sulfur ([Fe<sub>4</sub>S<sub>4</sub>(SEt)<sub>4</sub>]<sup>2−</sup>-HB(t) and  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup>-HB(b), respectively), resulting in a total of four water molecules per cluster. In the optimized structures, each water forms two hydrogen bonds, with  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup>-HB (t) slightly more favorable by  $\sim 0.9$  eV. Although VDE and VRE change by  $\sim 0.7$  eV upon hydration, the change in %L character is small and accounts for less than ~0.1 eV of the VDE and VRE differences based on the correlation for other ligands (Figure 1). Thus, the major contribution to the redox potential differences appears to be the electrostatic effect of the hydrogen bonds, consistent with PES/DFT studies<sup>8</sup> and protein calculations.<sup>13</sup>-15

However, the %L character of the HiPIPs and the Fd are significantly different in the XAS studies, which must have an underlying cause. A possibility is suggested by another conformation of  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup> with different  $\chi_3(C-S_t-Fe-S_b)$ , where  $S_b$  is the bridging sulfur on the opposite layer and  $S_t$  is the terminal sulfur ligand) torsions (Figure 2) that has very similar VDE and VRE but only 92% of calculated total %L character of  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup> (Figure 1), apparently due to the minority spin exchange states of the two antiferromagnetically coupled layers that comprise the cubane.<sup>9</sup> Specifically,  $[Fe_4S_4(SEt)_4]^2$ , which is like experimental conformational structure,<sup>16</sup> has all four  $\chi_3 \approx 50^\circ$  and the layers each have a minority spin delocalized to create a symmetric  $Fe^{2.5+}Fe^{2.5+}$  pair. On the other hand, the other conformer is characterized by two  $\chi_3 \approx 85^\circ$  and two  $\chi_3 \approx 60^\circ$  and has one layer in which the minority spin becomes localized to create a Fe<sup>3+</sup>-Fe<sup>2+</sup> pair, henceforth referred to as  $[Fe_4S_4(SEt)_4]^{2-}SL$ (Figure 2). Similar spin state have been found in the fission transition state of [4Fe-4S] clusters<sup>17</sup> and in the hetero-ligand  $[Fe_4S_4(SCH_3)_2L_2]^{2-}$  (L = Cl, H).<sup>18</sup>

Two more conformations of  $[Fe_4S_4(SEt)_4]^{2-}$  (Figure 3) were optimized from X-ray structures of redox sites of *Thermochromatium tepidum* (Tt)19 HiPIP at 0.80 Å resolution ([Fe4S4(SEt)4] <sup>2</sup>−-HiPIP), and *Bacillus thermoproteolyticus* (Bt)20 Fd at 0.92 Å resolution, ([Fe<sub>4</sub>S<sub>4</sub>(SEt)<sub>4</sub>]<sup>2–</sup>-Fd). Interestingly, [Fe<sub>4</sub>S<sub>4</sub>(SEt)<sub>4</sub>]<sup>2–</sup>-HiPIP has two Fe<sup>2.5+</sup>-Fe<sup>2.5+</sup> layers with

 $\gamma_3 = -65$  and +68° for one and  $\gamma_3 = -153$  and 172° for the other, and its cubane structure, spin state, and redox properties are similar to those of  $[Fe_4S_4(SEt)_4]^2$ , in very good agreement with Mössbauer of analogs and HiPIPs.<sup>21</sup> On the other hand,  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup>-Fd has one Fe<sup>3+</sup>-Fe<sup>2+</sup> layer with  $\chi_3 \approx 84$  and 74° and one Fe<sup>2.5+</sup>-Fe<sup>2.5+</sup> layer with  $\chi_3 \approx -76$  and +66° and its cubane structure, spin state, and redox properties are similar to those of  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup>-SL. The Fe<sup>2.5+</sup> in Fd versus the Fe<sup>3+</sup> in HiPIP also appears consistent with Mössbauer results in which the  $[Fe_4S_4(Cys)_4]^2$  cluster in Fd has less quadrupole splitting ( $\Delta E_Q$ ) than in HiPIP.<sup>21</sup>, <sup>22</sup> Remarkably, the calculated total Fe-S covalencies of  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup>-Fd and [Fe<sub>4</sub>S<sub>4</sub>(SEt)<sub>4</sub>]<sup>2–</sup>-HiPIP are 92% and 99% of [Fe<sub>4</sub>S<sub>4</sub>(SEt)<sub>4</sub>]<sup>2–</sup> values, respectively, in good agreement with the experimental results that *Bt* Fd and *Chromatium vinosum* HiPIP are 85% and 96% of  $[Fe_4S_4(SEt)_4]^{2-6}$ 

In summary, BS-B3LYP calculations show dramatic changes in redox energies of  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup> due to hydration with only slight changes in the Fe-S covalency. On the other hand, the conformational changes in thiolate ligands of  $[Fe_4S_4(SEt)_4]^{2-}$ -SL and  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup>-Fd induce minority spin localization, leading to large decreases in Fe-S covalency with only small changes in redox energies relative to  $[Fe_4S_4(SEt)_4]^{2-}$  and  $[Fe_4S_4(SEt)_4]^2$ <sup>-</sup>-HiPIP. These findings suggest that the large change in the Fe-S covalency between Fd and HiPIP observed by XAS is due differences in the conformations of the cysteinyl ligands of the cluster. Furthermore, the resulting differences in redox energies of ~100 mV are significant; thus, ligand conformation is important to consider in understanding redox properties of iron-sulfur proteins.

## **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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Niu and Ichiye Page 5



## **Figure 1.**

The correlation of the calculated total Fe-S/L covalency (%L) with VDE (open square) and VRE (open triangle) of [Fe4S4L4] <sup>2</sup>− (left to right, L=Cl, SH, S-*t*-butyl, SCH3, SeCH3, H, PH<sub>2</sub>, and P(CH<sub>3</sub>)<sub>2</sub> and with VDE (solid square) and VRE (solid triangle) of  $[Fe_4S_4(SEt)_4]^{2-}$ for the spin-delocalized state (SDL), the spin-localized state (SL), with waters hydrogen bonded to the bridging  $(HB_b)$  and terminal  $(HB_t)$  sulfurs, and in the Fd- and HiPIP-like conformations.





The schematic spin-delocalized  $[Fe_4S_4(SEt)_4]^2$  and spin-localized  $[Fe_4S_4(SEt)_4]^2$ -SL structures.



**Figure 3.**

The calculated Fd- and HiPIP-like  $[Fe<sub>4</sub>S<sub>4</sub>(SEt)<sub>4</sub>]<sup>2–</sup>$  structures.