ANTIGENIC RELATIONSHIP OF THE ENDOSPORES OF BACILLUS CEREUS-LIKE INSECT PATHOGENS TO BACILLUS CEREUS AND BACILLUS ANTHRACIS¹

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Aerobic sporeforming bacilli pathogenic for insects are known. In their appearance and cultural characteristics they are much like Bacillus cereus, except for their habit of depositing during sporulation crystals of protein known as parasporal bodies. Heimpel and Angus (1958) have reviewed the taxonomic status of these crystalliferous spore bearing bacilli. The present study has had as its purpose the revelation of the possible existence of agglutinogenic relationships between the endospores of the insect pathogenic bacilli and the endospores of B. cereus and Bacillus anthracis. Since agglutinin cross reactions have been demonstrated between the spores of B. cereus and B. anthracis (Lamanna and Eisler, 1960), it is of interest to know whether these cross reactions extend to the insect pathogenic sporeforming organisms.

MATERIAL AND METHODS

Strains. Dr. E. A. Steinhaus kindly supplied the following representative strains of the insect pathogenic crystalliferous spore bearers: 0-3-30, Bacillus thuringiensis var. thuringiensis Berliner; 16-2-2, Bacillus thuringiensis var. thuringiensis Berliner; 0-24-3, Bacillus thuringiensis var. alesti Toumanoff and Vago; 0-32-2, Bacillus thuringiensis var. alesti (Anduze strain); 0-28-1, Bacillus thuringiensis var. sotto Aoki and Chigasaki; 57-1-1 Bacillus entomocidus var. entomocidus Heimpel and Angus; and 58-1-1 Bacillus entomocidus var. subtoxicus Heimpel and Angus.

The origins of the *B. cereus* and *B. anthracis* strains employed are listed in the paper by Lamanna and Eisler (1960).

Preparation of spore antigens and agglutination studies. With only minor modifications, preparation of spore antisera free of vegetative bacilli

¹ This work was sponsored by the Bureau of Yards and Docks, U. S. Navy, and the Office of Naval Research under a contract with the Regents of the University of California. agglutinins and agglutination tests were performed as outlined by Lamanna and Eisler (1960). The one notable change was the employment as diluent for the agglutination tests of 0.85% NaCl solution with 0.5% bovine serum albumin added. The presence of the albumin was found to improve the appearance of serological agglutination of spores, and to reduce both the degree of spontaneous agglutination and the tendency of spores to be trapped in a meniscus.

RESULTS AND DISCUSSION

Antisera prepared against the endospores of the insect pathogens were used for agglutination tests with the results recorded in Table 1. Cross reactions occur among the insect pathogenic strains and between these organisms and B. cereus and B. anthracis. That the antigenic relationship between the insect pathogens and B. anthracis may be more remote than for B. cereus is suggested both by the notably fewer number of anthrax strains yielding any agglutination at all and by the lower order of titers obtained with positively reacting strains.

Agglutination tests with the insect pathogens using antisera against spores of strains of B. cereus and B. anthracis were performed to seek confirmation of the existence of spore agglutinogens held in common (Table 2). The data make it obvious that the specificity of antisera induced by injection of spores is not homologous with the species from which the spores are obtained for injection of rabbits. Clearly the spore agglutination test provides no sharp lines of demarcation for separating the three kinds of organisms.

To determine whether some systematic, readily discernible pattern of distribution of agglutinogens exists among the species, a series of cross adsorption trials was undertaken. The results obtained when spores of two strains of B. anthracis were employed to adsorb anti-

TABLE 1 Agglutinin titers of antisera prepared against the endospores of insect pathogenic bacilli employing endospores from the indicated species as agglutinogens

Spore Agglutinin Species and Strain		Insect Pathogenic Species																		
	В	Bacillus thuringiensis					illus ocidus	Bacillus cereus						Bacillus anthracis						
	var. thuring- iensis		var. alesti			var. en- tomo- cidus	en- sub- tomo- toxi-				ø						8			
	0-3-30	16-2-2	0-24-3	0-32-2	0-28-1	57-1-1	58-1-1	Ba2(22)	Ba2(16)	NRS201	NRS1256	NRS617	NRS232	NRS244	NRS793	NRS249	Carbozoo	380	385	391
B. thuringiensis var. thuringiensis																				
0-3-30	2048	4096	128			0	64	64	512		32	1024	256	0	4096	4096		0	0	16
16-2-2	4096	4096	64	128	512	64	64	64	128	128	32	1024	256	0	4096	4096	32	SA*	0	4
var. alesti																				
0-24-3	64	128	1024	1	512	0	128	0	256		32		2048		2048		0	256	0	0
0-32-2	0	0	256	2048	512	64	256	0	0	1024	8	64	128	0	128	16	128	64	0	0
var. <i>sotto</i> 0-28-1	8	0	256	4096	1024	0	512	0	1024	2048	128	32	128	256	2048	32	32	0	32	0
B. entomocidus																				
var. entomocidus 57-1-1	0	32	128	256	64	1024	32		1024	210	1024	90	2048	1004	256	8	32	10		
07-1-1 Var. subtoxicus	1	34	128	400	04	1024	32	4	1024	012	1024	32	2048	1024	200	ð	32	16	4	SA*
58-1-1	16	16	128	512	256	8	256	o	0	32	0	16	128	16	256	8	8	0	16	-4

Titers for the homologous spore agglutinogen are recorded in bold face. In addition to the strains of *B. anthracis* listed, 17 other strains were tested. With 6 of these, agglutination was not obtained; 4 gave only a low titer agglutination with 1 antiserum; 7 showed either no agglutination or spontaneous agglutination with the different antisera.

* Spontaneous agglutination.

TABLE 2

Agglutinin titers with endospores of insect pathogens employing spore antisera prepared against strains of Bacillus cereus and Bacillus anthracis and adsorbed with homologous vegetative bacilli

		Heterologous Spore Agglutinogen									
Spore Antisera	Homol- ogous Titer		Bacil	Bacillus entomocidus							
		0-3-30	16-2-2	0-24-3	0-32-2	0-28-1	57-1-1	58-1-1			
B. cereus											
NRS201	>1024	0	256	512	128	>1024	0	>1024			
NRS946	>1024	0	0	0	128	0	0	0			
NRS232	>1024	64	128	512	>1024	>1024	>1024	>1024			
NRS244	1024	128	64	256	>1024	512	>1024	>1024			
NRS617	1024	>1024	>1024	64	256	64	32	32			
Ba2(16)	>1024	8	8	256	>1024	128	1024	512			
NRS1256	>1024	128	256	64	256	32	256	16			
Ba2(19)	>1024	>1024	0	4	0	0	0	0			
NRS793	>1024	512	512	512	>1024	>1024	64	512			
Ba2(23)	>1024	0	0	>1024	>1024	>1024	512	>1024			
NRS847	SA	>1024	>1024	0	64	0	16	0			
Ba2(22)	256	0	0	0	32	0	0	0			
B. anthracis											
Carbozoo	2048	8	64	8	64	64	0	8			
400	1280	128	256	32	256	128	32	128			
390	1024	0	16	0	32	16	0	8			

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TABLE 3

Agglutinin titers of antisera against spores of insect pathogens adsorbed with spores of Bacillus anthracis

	Antiserum Adsorbed											
				Bacillus thuringiensis								
Spore Agglutinogen	Bacillu	s entomocidu	s 57-1-1		0-3-30		0-24-3					
	Before adsorption		Adsorption with 380	Before adsorption	Adsorption with Carbozoo	Adsorption with 380	Before adsorption	Adsorption with Carbozoo	Adsorption with 380			
Insect pathogen												
0-3-30				1024	0	0	16	0	0			
16-2-2	0	0	0	1024	0	16	16	0	32			
0-32-2	128	0	0	32	0	0	512	0	256			
0-28-1	64	0	0	16	0	0	32	0	64			
0-24-3	64	0	0	16	0	0	512	0	128			
57-1-1	16	0	0	0	0	0	0	0	0			
58-1-1	0	0	0	32	0	0	64	0	32			
B. anthracis												
Carbozoo	32	0	0	32	0	0						
380	256	0	0				512	0	0			
Bacillus cereus												
NRS232	512	0	64	32	8	16	1024	64	64			
NRS244	256	0	256				256	0	0			
NRS249				512	32	128	64	0	256			
NRS793	256	16	16	256	8	32	1024	16	128			
NRS201	256	32	16	16	0	0						
NRS1256	512	64	0	0	0	0	64	0	32			
NRS617	256	0	0	128	0	0						
Ba2(16)	256	128	512	64	0	0	512	64	64			

TABLE 4

Adsorption of antisera against spores of insect pathogens and Bacillus cereus with spores of heterologous strains

Data recorded are the number of strains for which the agglutinin titer was reduced to zero to the total number of strains tested exclusive of the strain used for adsorption.

Adsorbing Spore Antigen	Serum Adsorbed											
		Bacillus th	uringiensis		Bacillus er	ntomocidus	B. cereus					
	0-3	-30	0-2	4-3	57-	1-1	NR	5232	NRS793			
	Species for spore agglutination											
	Insect pathogens	B. cereus	Insect pathogens	B. cereus	Insect pathogens	B. cereus	Insect pathogens	B. cereus	Insect pathogens	B, cereu		
0-3-30			0/4	2/6	3/4	3/7	0/4	1/10	1/5	3/6		
0-24-3	3/5	3/8			3/3	2/6	2/3	2/10	3/5	3/6		
57-1-1	0/6	3/8	0/4	3/6			1/4	4/10	1/6	2/6		
NRS232	1/6	5/6	3/6	4/5	4/4	6/6						

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body from antisera prepared against three strains of insect pathogens are recorded in Table 3. Data are also included on how the adsorption affected titers against spores of eight strains of B. cereus. No simple relationships can be derived from these data which show that results with one strain as adsorbent are not necessarily duplicated by the use of another strain of B. anthracis. Table 4 attempts to summarize results recorded when antisera against spores of insect pathogens and B. cereus were adsorbed with spores from the heterologous species. These adsorption experiments demonstrate the antigenic complexity of the spores and the existence of a complicated pattern of distribution of antigens among the three kinds of organisms. Based on past experience (Lamanna and Eisler, 1960), it might be expected that if a larger battery of strains were subjected to analysis, the existence of interspecies and intraspecies relationships would appear even more complex.

In spite of the antigenic cross reactions demonstrated in this paper and the recent report of Yoder and Nelson (1960) on the existence of bacteriophage which can cross the host range boundary between *B. thuringiensis* Berliner, and *B. anthracis*, we believe it would be premature for taxonomic purposes to consider the organisms under discussion as a single species. Particularly in the case of *B. anthracis*, there are a number of characteristics aside from pathogenicity which justify its continued treatment as a distinct and recognizable species (Burdon, 1956; Leise et al., 1959). In this connection it is of interest to know that neither by laboratory exposure nor in field trials as a live pest control agent against susceptible insect species has *B. thuringiensis* Berliner shown any signs of pathogenicity for man and other animals susceptible to anthrax (Fisher and Rosner, 1959).

SUMMARY

Agglutination and agglutinin adsorption tests have revealed the existence of antigenic relationships among the endospores of the crystalliferous insect pathogenic spore-bearing bacilli and *Bacillus cereus* and *Bacillus anthracis*. It has not proved possible to differentiate among these organisms by means of serological agglutination tests employing endospores as agglutinogens.

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