Immune Mechanisms in Thyroid Eye Disease

Geniece M. Lehmann,¹ Steven E. Feldon,² Terry J. Smith,^{3–5} and Richard P. Phipps^{1,2}

Thyroid eye disease (TED) is an inflammatory condition of the orbit closely associated with Graves' disease. During the course of TED, fibrosis can develop around the extraocular muscles, and excess extracellular matrix and fat accumulates in the periorbital space. This dramatic remodeling results in protrusion of the eye, also known as exophthalmos. Current treatments are sometimes effective in alleviating the symptoms of the disease, but there remains a demand for treatments that prevent or reverse the pathological alterations of orbital tissues. Such treatments may become available as a result of research aimed at understanding the mechanism by which Graves' disease leads to specific remodeling of orbital tissues. Recent findings have uncovered the importance of intercellular communication between autoreactive T cells and orbital fibroblasts. When orbital fibroblasts are activated, possibly by Graves' disease–related autoantibodies, they release T cell chemoattractants, initiating an interaction in which these cells activate each other. These interactions ultimately result in fibroblasts. Although the mechanisms underlying these processes are not completely understood, several currently available therapeutic strategies might interrupt the signaling between B and T cells and fibroblasts, thereby treating the clinical manifestations of TED.

Introduction

RAVES' DISEASE IS AN AUTOIMMUNE condition in which J the thyroid-stimulating hormone receptor (TSH-R) displayed on thyrocytes is targeted by autoantibodies, inducing the production of excess thyroid hormone (1). Besides hyperthyroidism, up to 60% of patients with Graves' disease develop a manifestation localizing to the orbit called Graves' ophthalmopathy or thyroid eye disease (TED) (2,3). TED occurs more frequently in patients older than 50 years of age and in males, smokers, and those with microvascular disease (2,4). TED affects many eye functions, including wetting of the ocular surface, eye motility, optic nerve function, and eyelid anatomy (2,5). The most dramatic pathological findings in end-stage TED include glycosaminoglycan (GAG) deposition (accompanied by dramatic swelling resulting from the waterbinding capacity of these macromolecules) and fibrosis affecting the extraocular muscles and fat accumulation in the orbit (5-8). The increased volume of orbital connective tissue leads to protrusion of the eye (exophthalmos) (2,9). To date, there are no effective means of preventing the disease or reliably altering its course. Current therapeutic options include corticosteroids, external beam radiation, and surgery. These interventions are aimed at the consequences of the disease rather than targeting its cause, which remains poorly understood (5,6,10,11). They do nothing to prevent or reverse pathological remodeling of orbital tissues (10). Understanding the pathogenetic mechanisms underlying TED should yield the identification of predictive biomarkers of progressive disease and effective and specific treatments.

The close association of TED with Graves' disease of the thyroid provides clues to its etiology. The hyperthyroid condition itself does not impact directly on the accumulation of connective tissue within the orbit (1,2). The risk for developing TED is similar for euthyroid and hyperthyroid Graves' disease patients (2,12). Rather, the orbit constitutes a second target of autoimmunity. Therefore, understanding the role of the immune system in orbital inflammation is critical to developing treatment strategies for TED patients.

Cell Types Implicated in TED

From the little we know about TED, there are at least three cell types involved in the development and progression of the disease: B cells, T cells, and orbital fibroblasts. As in the hyperthyroidism of Graves' disease, B cells are important in

⁴The Jules Stein Eye Institute, Los Angeles, California.

Departments of ¹Environmental Medicine and the Lung Biology and Disease Program and ²Ophthalmology, School of Medicine and Dentistry, University of Rochester, Rochester, New York.

³Division of Molecular Medicine, Department of Medicine, Harbor-University of California Los Angeles Medical Center, Torrance, California.

⁵David Geffen School of Medicine at University of California Los Angeles, Los Angeles, California.

the early stages of TED, producing antibodies against at least one self-antigen (1,13). However, development of TED is postulated to require a second initiating event that results in recruitment of activated T cells to the orbit (1). These T cells take on the role of amplifying the B cell response and are thought to be major contributors to disease progression (14). Cells (namely, orbital fibroblasts) that recruit the T cells into the orbit are critical players in the establishment of inflammation (15). Fibroblasts provoke T cell recruitment and then engage them in a cycle of brief reciprocal activation ending in tissue remodeling characteristic of TED (14). This review will explore the interactions among these three cell types that contribute to the disease phenotype and which may provide targets for therapeutic strategies.

B and T Cells

The most important cells for adaptive immunity, and thus for autoimmunity, are B and T cells. B cells, including two major subsets (B1 and B2), migrate and proliferate extensively and develop into antibody-secreting plasma cells when they encounter foreign antigen in the proper costimulatory context. Dual signals are required for B cell activation, differentiation, and the production of antibodies. One signal is provided by antigen binding to the B cell receptor. The second signal can be provided through interaction between CD40 on the surface of the B cell and CD40 ligand (CD40L, CD154) on the surface of a T cell (16–18). The interaction with CD40L stimulates T cell secretion of cytokines such as IL-4 that are critical to B cell activation and immunoglobulin class switching (19). Initially, differentiated B cells produce antibodies in the form of IgM, and the CD40–CD40L interaction is required for class switching to IgG or IgE. T cells, like B cells, are migratory and highly proliferative in response to antigen. However, T cells are more diverse than B cells, differentiating into many different types of effector cells, including CD8+ cytotoxic T cells and CD4+ helper T cells of the Th1, Th2, Th17, and Treg subsets (20). The context in which T cells recognize antigen is more constrained. Detection of antigen occurs through surface T cell receptors (TCRs) that recognize specific peptides presented in association with major histocompatibility complex (MHC) proteins on the surface of antigen-presenting cells (APCs). These include macrophages, dendritic cells, and B cells. T cells also require dual signals for activation. The first signal is provided by antigen/MHC binding to the TCR (1,21). The second signal is often a B7 molecule on the surface of an APC, possibly a B cell, which binds to CD28 on the T cell surface (1,17,21,22). CD40L can interact with CD40 on the surface of the APC, resulting in increased B7 expression. This in turn can lead to increased interaction with CD28 and further T cell activation (17,21,22). T cell activation results in proliferation and differentiation to an effector cell type. If one signal is present in the absence of another, the T cell will fail to become activated and may become refractory to future activation (1,23). This phenomenon, known as "peripheral anergy," is one of several tolerance mechanisms preventing the development of autoimmunity (23,24).

Tolerance mechanisms sometimes fail and autoimmunity develops. Cells of the Th17 subset of T cells have been implicated in the onset of many autoimmune and inflammatory diseases, including arthritis, multiple sclerosis, psoriasis, and inflammatory bowel disease (20). It may be hypothesized that autoreactive Th17 cells infiltrate the orbit, where they activate B cells to secrete autoantibodies. Th17 cells are defined by their expression of IL-23R (20), and variants in the IL-23R gene have been shown to be strongly associated with the occurrence of TED (25). Further work is required to firmly identify the role of the Th17 subset in TED initiation. However, by an unknown mechanism, production of autoantibodies by autoreactive B cells does occur in TED patients (1,13). The TSH-R has been implicated as an autoantigen targeted in TED, largely based on the close association of this condition with the hyperthyroidism of Graves' disease (26). TSH-R is expressed on the surface of orbital fibroblasts, and TSH-R immunoreactivity has been demonstrated specifically in TED orbits. Insulin-like growth factor-1 receptor (IGF-1R) represents another self-antigen with a pathogenic role in TED (8,27). This receptor is displayed by orbital fibroblasts from patients with TED at a higher level than those from normal orbits (28). Further, antibody directed against the IGF-1R (a) provokes the synthesis of hyaluronan in orbital fibroblasts from TED, but not in those from normal orbits, and (b) induces orbital fibroblasts from patients with TED to produce the T cell chemoattractants IL-16 and RANTES (8,28). Patients with Graves' disease exhibiting anti-TSH-R antibody production may also develop antibodies against the IGF-1R and other orbital antigens. This may occur as a result of extraocular muscle inflammation and damage, as is the case with antibodies to extraocular muscle proteins, which are present in TED patients, but do not contribute to disease progression (29,30). Alternatively, production of autoantibodies against additional self-antigens may result from a process known as determinant spreading (31). Immunoglobulin synthesized by autoreactive B cells can bind an epitope of a macromolecular complex. Then, acting as an APC, the B cell processes peptides from this complex for presentation to specific T cells. Because it processed a macromolecular complex, the B cell may present specific peptides that do not replicate the antigen to which it originally bound, or the antigen against which its antibodies will be directed. However, T cells specific for the antigen will be able to activate the B cell. In this way, T cells can activate multiple B cell clones and produce several autoantibodies. This phenomenon may underlie the large variety of autoantibodies detected in patients with autoimmune disease. Since TED is an autoimmune condition, therapeutic strategies designed to block the loss of immune tolerance and the inappropriate activation of B and T cells may constitute treatments for the condition in its earliest stages. For example, there are agents currently available that disrupt the B7-CD28 and CD40-CD40L interactions, as well as antibody therapies that deplete B and T cells (13,32-34). Rituximab (RTX), a monoclonal antibody directed against the B cell surface antigen CD20, is one such B cell-depleting agent (13). Case reports and one open pilot study demonstrate a marked amelioration of eye symptoms in TED patients treated with RTX (35-37), supporting a role for B cells in the pathogenesis of TED.

Orbital fibroblasts

TED occurs with the hyperthyroidism of Graves' disease, but the correlation is not perfect (2). Many patients never develop clinically important orbital disease. Some patients with TED fail to develop hyperthyroidism. These variations

IMMUNE MECHANISMS IN THYROID EYE DISEASE

in clinical presentation suggest that development of TED requires multiple "hits." The initial hit could be the generation of an autoantibody within the orbit (26). However, this may be inadequate for triggering disease in some patients. Another autoantibody may represent a second hit. Additional factors might include the genetic predisposition of the individual, environmental factors such as tobacco smoke or infection (1,2,4,8). Once sufficient hits occur, orbital fibroblasts can be activated to initiate inflammation characteristic of TED. Greater understanding of these provocative factors should enhance our development of effective treatment strategies.

Fibroblasts were once viewed as mere structural elements of the microenvironment. They were thought only to produce extracellular matrix components, but otherwise to play no role in tissue homeostasis. Now, it has become apparent that fibroblasts are highly interactive "sentinel cells," capable of detecting danger signals and communicating these to cells of the "professional" immune system. Fibroblasts respond directly to these signals by proliferating and differentiating into effector cells (1,38,39). They respond to immune stimulation and actively participate in inflammation pathways through their synthesis of chemokines, cytokines, and lipid mediators. Orbital fibroblasts are not only responsible for generating GAGs and other macromolecules, but they also differentiate to either myofibroblasts (i.e., scar-forming cells) or to lipofibroblasts (i.e., adipocytes) (40-43). These attributes implicate fibroblasts as effectors in the progression of TED. The earliest stage of TED involves infiltration of the orbit by T cells (5,6,10,11,44,45). Evidence supports the concept that infiltrating T lymphocytes activate fibroblasts. In doing so, they promote the synthesis of extracellular matrix, differentiation to adipocytes, and myofibroblast proliferation (14). T cellfibroblast interactions are mediated through costimulatory molecules, adhesion molecules, and cytokines, including IFN γ , IL-1 β , and TNF α (14,46). GAG and prostaglandin synthesis in orbital fibroblasts is upregulated by CD40 engagement (39,47,48). However, fibroblasts might also play a role in initiating the early stage T cell infiltration of the orbit. Stimulated fibroblasts secrete multiple cytokines, including IL-6, which stimulates B cell differentiation. IL-16 and RANTES are chemoattractants that initiate T cell migration (15,48-50). Fibroblasts can also function as APCs, providing a second signal for lymphocyte activation. Those fibroblasts from patients with TED constitutively express MHC II and CD40 (39,51), both of which are upregulated by IFN γ (46,48). They can also drive proliferation of autologous T cells (52). T cell responsiveness to fibroblasts may mediate T cell infiltration in early disease and amplify inflammation.

Fibroblasts differ from one tissue to another (39,41,47,48, 53). Those present in a particular tissue, such as the orbit,



FIG. 1. Fibroblasts can be stimulated to differentiate into myofibroblasts or lipofibroblasts. This potential may depend on expression of Thy-1 (CD90). TGF- β triggers the differentiation of Thy-1–positive fibroblasts into myofibroblasts, while PPAR γ activation induces adipocyte differentiation of mainly Thy-1–negative cells.

might derive from one of several sources. They could transdifferentiate from epithelial cells, muscle cells, or even endothelium (54). Alternatively, they could be recruited from the bone marrow as circulating "fibrocytes" (55,56). The origin of activated fibroblasts involved in the pathogenesis of TED remains uncertain. Orbital fibroblasts exist as multiple subsets (53,57). Each can differ in morphology, proliferation, biosynthetic capacity, and cell surface marker expression. Two major subsets of orbital fibroblast exist (41,49,53) based on their expression of the surface protein Thy-1 (CD90). The balance between Thy-1-negative and Thy-1-positive fibroblasts in the orbit is essential for normal regulation of inflammation since these subsets have distinct biosynthetic capabilities, differing in cytokine, collagen, and prostaglandin E₂ (PGE₂) production (49). However, this balance may also underlie the development and progression of TED. As shown in Figure 1, depending on their pericellular environment and phenotype, fibroblasts can differentiate into myofibroblasts or lipofibroblasts (40,41). Myofibroblasts participate in normal wound healing and in the pathogenesis of fibrosis, such as that occurring in orbital connective tissue in TED (42). The presence of lipofibroblasts is usually indicative of pathology. In TED, they result in excess orbital fat deposition (58). The potential for terminal differentiation in fibroblasts depends on the expression of Thy-1. TGF- β triggers the differentiation of Thy-1–positive fibroblasts into myofibroblasts, identified by their relatively high level of alpha-smooth muscle actin (α -SMA) expression (40). Thy-1–negative fibroblasts preferentially differentiate into adipocytes, as evidenced by staining with oil red O (40,41). Lipid accumulation is driven by the ligation of peroxisome proliferator activated receptor γ (PPAR γ) with an agonist.

T Cell–Fibroblast Interactions

PPAR γ is a nuclear receptor that functions as a transcription factor. It binds lipid ligands, dimerizes with retinoid-X receptor alpha (RXRα), translocates to the nucleus, and alters the transcription of genes containing peroxisome proliferator responsive elements (59). These genes include those that dampen inflammation by decreasing TNFα, IL-6, and IL-8 production (60). Other PPAR γ genes are also important to adipogenesis. PPAR γ ligands trigger the differentiation of fibroblasts to lipofibroblasts (61–65). These agonists promote glucose uptake; thus, they have a therapeutic value in Type 2 diabetes. Insulin-sensitizing drugs, such as ciglitazone,



FIG. 2. One current model focuses on TED being triggered by activation of orbital fibroblasts by autoantibodies. These autoantibodies could be specific for antigens such as TSH-R and/or IGF-1R. Activated orbital fibroblasts release chemokines, including IL-16 and RANTES, which traffic T cells into the orbit. These lymphocytes then interact with fibroblasts, potentially activating each other, further promoting cytokine production (IFN γ , PGD₂, and 15d-PGJ₂) and secretion of T cell-activating factors by the fibroblasts (IL-1 α , IL-8, and products of Cox-2 activity). Fibroblasts are also stimulated to secrete IL-6, which stimulates B cell differentiation. The interactions of fibroblasts with T cells result in the deposition of extracellular matrix molecules, fibroblast proliferation, and differentiation.

IMMUNE MECHANISMS IN THYROID EYE DISEASE

pioglitazone, and rosiglitazone, bind to and activate PPAR γ (61–64,66). Naturally occurring PPAR γ ligands include lysophosphatidic acid, some fatty acids, prostaglandin D₂ (PGD₂), and 15-deoxy-prostaglandin J₂ (15d-PGJ₂) (60,62,66,67). PGD₂ and 15d-PGJ₂ are derived from arachidonic acid (68). Their synthesis is dependent on the activity of cyclooxygenase-2 (Cox-2) and PGD synthase enzymes (68,69). Activated T cells express both Cox-2 and PGD synthase; they also produce 15d-PGJ₂ (58,70,71).

The response of orbital fibroblasts to PPAR γ agonists provides a potentially important link between these cells and T cell activation. 15d-PGJ₂ treatment of Thy-1–negative orbital fibroblasts from patients with TED initiates adipocytic differentiation (40). T cells from these patients constitutively express Cox-2 and secrete high levels of 15d-PGJ₂ (58). It is thus possible to envision a mechanism through which T cell infiltration of the orbit could result in adipocytic differentiation of fibroblasts. In fact, coculture of these fibroblasts with activated T cells results in accumulation of cytoplasmic lipid droplets in the fibroblasts. Understanding these complex interactions may lead to the development of new therapeutic strategies for TED. Blockade of PPAR γ dependent adipogenesis may alleviate fat expansion in this disease (72).

Conclusions

A summary of our current model for the pathogenesis of TED is depicted in Figure 2. The key elements involve binding and activation of orbital fibroblasts by autoantibodies, produced by autoreactive B cell-derived plasma cells (13). Trafficking of T cells to the orbit ultimately leads to complex interactions of fibroblasts with these T cells and the deposition of extracellular matrix molecules. Proliferation and differentiation of fibroblast subsets into either fat-laden adipocytes (14) or scar-forming myofibroblasts culminates in the increased orbital connective tissue volume and remodeling that underlie the clinical manifestations of TED (9). Although the disease mechanisms through which TED develops are not fully elucidated, several potential therapeutic strategies might interrupt fibroblast, T cell, and B cell activation in the orbit. General suppression of immune function, such as occurs with corticosteroids, has produced inconsistent results and is associated with undesirable side effects (5,6,10). To avoid these side effects, it is important to target more selective aspects of the disease. The development of autoantibodies against IGF-1R may be prevented through the use of agents that deplete B cell and T cell numbers (13,34) or that disrupt the B7-CD28 and CD40-CD40L interactions that are essential to T and B cell activation (32,33,48). Once orbital fibroblasts have been activated and the amplification of the disease process has begun, antagonists to cytokine and chemokine receptors may be used to interrupt communications between fibroblasts and lymphocytes that drive morphological changes (73). Finally, PPARy signaling may be blocked to prevent fibroblast adipogenesis and fat accumulation, either by inhibiting Cox-2 activity in the T cells (58,71,74), thus eliminating release of natural PPAR γ agonists, or through the use of direct PPAR γ antagonists (72). Targeting more specific aspects of the disease based on expanded understanding of its pathophysiology should provide many opportunities to prevent or alleviate progression of TED.

963

Acknowledgments

This research was supported by EY014564, EY017123, EY011708, DE011390, EY08976, DK063121, T32 HL66988, and the Research to Prevent Blindness Foundation. The continued support of the Bell Charitable Foundation is gratefully acknowledged.

References

- Prabhakar BS, Bahn RS, Smith TJ 2003 Current perspective on the pathogenesis of Graves' disease and ophthalmopathy. Endocr Rev 24:802–835.
- Burch HB, Wartofsky L 1993 Graves' ophthalmopathy: current concepts regarding pathogenesis and management. Endocr Rev 14:747–793.
- Heufelder AE, Weetman AP, Ludgate M, Bahn RS 2000 Pathogenesis of Graves' ophthalmopathy. In: Prummel MF, Wiersinga WM, Mourits MP, Heufelder AE (eds.) Recent Developments in Graves' Ophthalmopathy. Kluwer Academic Publishers, Boston, pp 15–37.
- Bartalena L, Marcocci C, Pinchera A 2002 Graves' ophthalmopathy: a preventable disease? Eur J Endocrinol 146:457– 461.
- Bahn RS, Heufelder AE 1993 Pathogenesis of Graves' ophthalmopathy. N Engl J Med 329:1468–1475.
- Feldon SE, Weiner JM 1982 Clinical significance of extraocular muscle volumes in Graves' ophthalmopathy: a quantitative computed tomography study. Arch Ophthalmol 100:1266–1269.
- Nunery WR, Nunery CW, Martin RT, Truong TV, Osborn DR 1997 The risk of diplopia following orbital floor and medial wall decompression in subtypes of ophthalmic Graves' disease. Ophthal Plast Reconstr Surg 13:153–160.
- Smith TJ, Hoa N 2004 Immunoglobulins from patients with Graves' disease induce hyaluronan synthesis in their orbital fibroblasts through the self-antigen, insulin-like growth factor-I receptor. J Clin Endocrinol Metab 89:5076–5080.
- Hatton MP, Rubin PA 2002 The pathophysiology of thyroidassociated ophthalmopathy. Ophthalmol Clin North Am 15:113–119.
- Liu D, Feldon SE 1992 Thyroid ophthalmopathy. Ophthalmol Clin North Am 5:597–622.
- Wiersinga WM, Prummel MF 2001 Pathogenesis of Graves' ophthalmopathy—current understanding. J Clin Endocrinol Metab 86:501–503.
- Kim JM, LaBree L, Levin L, Feldon SE 2004 The relation of Graves' ophthalmopathy to circulating thyroid hormone status. Br J Ophthalmol 88:72–74.
- El Fassi D, Nielsen CH, Hasselbalch HC, Hegedus L 2006 The rationale for B lymphocyte depletion in Graves' disease. Monoclonal anti-CD20 antibody therapy as a novel treatment option. Eur J Endocrinol 154:623–632.
- 14. Feldon SE, Park DJ, O'Loughlin CW, Nguyen VT, Landskroner-Eiger S, Chang D, Thatcher TH, Phipps RP 2005 Autologous T-lymphocytes stimulate proliferation of orbital fibroblasts derived from patients with Graves' ophthalmopathy. Invest Ophthalmol Vis Sci 46:3913–3921.
- Pritchard J, Horst N, Cruikshank W, Smith TJ 2002 Igs from patients with Graves' disease induce the expression of T cell chemoattractants in their fibroblasts. J Immunol 168:942–950.
- Noelle RJ, Roy M, Shepherd DM, Stamenkovic I, Ledbetter JA, Aruffo A 1992 A 39-kDa protein on activated helper T cells binds CD40 and transduces the signal for cognate activation of B cells. Proc Natl Acad Sci USA 89:6550–6554.

- Yin D, Zhang L, Wang R, Radvanyi L, Haudenschild C, Fang Q, Kehry MR, Shi Y 1999 Ligation of CD28 *in vivo* induces CD40 ligand expression and promotes B cell survival. J Immunol 163:4328–4334.
- Ray DM, Akbiyik F, Bernstein SH, Phipps RP 2005 CD40 engagement prevents peroxisome proliferator-activated receptor gamma agonist-induced apoptosis of B lymphocytes and B lymphoma cells by an NF-kappaB-dependent mechanism. J Immunol 174:4060–4069.
- Kehry MR, Hodgkin PD 1993 Helper T cells: delivery of cell contact and lymphokine-dependent signals to B cells. Semin Immunol 5:393–400.
- 20. Reiner SL 2007 Development in motion: helper T cells at work. Cell **129:33**–36.
- Van Gool SW, Vandenberghe P, de Boer M, Ceuppens JL 1996 CD80, CD86 and CD40 provide accessory signals in a multiple-step T-cell activation model. Immunol Rev 153: 47–83.
- 22. Howland KC, Ausubel LJ, London CA, Abbas AK 2000 The roles of CD28 and CD40 ligand in T cell activation and tolerance. J Immunol **164**:4465–4470.
- 23. Powell JD 2006 The induction and maintenance of T cell anergy. Clin Immunol **120**:239–246.
- 24. Melchers F 2006 Anergic B cells caught in the act. Immunity 25:864–867.
- Huber AK, Jacobson EM, Jazdzewski K, Concepcion ES, Tomer Y 2008 IL-23R is a major susceptibility gene for Graves' ophthalmopathy: the IL-23/Th17 axis extends to thyroid autoimmunity. J Clin Endocrinol Metab 93:1077–1081.
- Bahn RS, Dutton CM, Natt N, Joba W, Spitzweg C, Heufelder AE 1998 Thyrotropin receptor expression in Graves' orbital adipose/connective tissues: potential autoantigen in Graves' ophthalmopathy. J Clin Endocrinol Metab 83:998– 1002.
- 27. Smith TJ 2003 The putative role of fibroblasts in the pathogenesis of Graves' disease: evidence for the involvement of the insulin-like growth factor-1 receptor in fibroblast activation. Autoimmunity 36:409–415.
- Pritchard J, Han R, Horst N, Cruikshank WW, Smith TJ 2003 Immunoglobulin activation of T cell chemoattractant expression in fibroblasts from patients with Graves' disease is mediated through the insulin-like growth factor I receptor pathway. J Immunol 170:6348–6354.
- 29. Khoo TK, Bahn RS 2007 Pathogenesis of Graves' ophthalmopathy: the role of autoantibodies. Thyroid **17:**1013–1018.
- 30. Wu YJ, Clarke EM, Shepherd P 1998 Prevalence and significance of antibodies reactive with eye muscle membrane antigens in sera from patients with Graves' ophthalmopathy and other thyroid and nonthyroid diseases. Thyroid 8:167–174.
- Dai YD, Carayanniotis G, Sercarz E 2005 Antigen processing by autoreactive B cells promotes determinant spreading. Cell Mol Immunol 2:169–175.
- Nogid A, Pham DQ 2006 Role of abatacept in the management of rheumatoid arthritis. Clin Ther 28:1764–1778.
- 33. Huang W, Sinha J, Newman J, Reddy B, Budhai L, Furie R, Vaishnaw A, Davidson A 2002 The effect of anti-CD40 ligand antibody on B cells in human systemic lupus erythematosus. Arthritis Rheum 46:1554–1562.
- Chatenoud L 2004 Anti-CD3 antibodies: towards clinical antigen-specific immunomodulation. Curr Opin Pharmacol 4:403–407.
- 35. El Fassi D, Nielsen CH, Hasselbalch HC, Hegedus L 2006 Treatment-resistant severe, active Graves' ophthalmopathy

successfully treated with B lymphocyte depletion. Thyroid **16:**709–710.

- 36. Salvi M, Vannucchi G, Campi I, Rossi S, Bonara P, Sbrozzi F, Guastella C, Avignone S, Pirola G, Ratiglia R, Beck-Peccoz P 2006 Efficacy of rituximab treatment for thyroid-associated ophthalmopathy as a result of intraorbital B-cell depletion in one patient unresponsive to steroid immunosuppression. Eur J Endocrinol **154**:511–517.
- 37. Salvi M, Vannucchi G, Campi I, Curro N, Dazzi D, Simonetta S, Bonara P, Rossi S, Sina C, Guastella C, Ratiglia R, Beck-Peccoz P 2007 Treatment of Graves' disease and associated ophthalmopathy with the anti-CD20 monoclonal antibody rituximab: an open study. Eur J Endocrinol 156:33–40.
- Smith TJ 2005 Insights into the role of fibroblasts in human autoimmune diseases. Clin Exp Immunol 141:388–397.
- Smith RS, Smith TJ, Blieden TM, Phipps RP 1997 Fibroblasts as sentinel cells. Synthesis of chemokines and regulation of inflammation. Am J Pathol 151:317–322.
- Koumas L, Smith TJ, Feldon S, Blumberg N, Phipps RP 2003 Thy-1 expression in human fibroblast subsets defines myofibroblastic or lipofibroblastic phenotypes. Am J Pathol 163:1291–1300.
- 41. Smith TJ, Koumas L, Gagnon A, Bell A, Sempowski GD, Phipps RP, Sorisky A 2002 Orbital fibroblast heterogeneity may determine the clinical presentation of thyroid-associated ophthalmopathy. J Clin Endocrinol Metab 87:385–392.
- 42. Gabbiani G 1981 The myofibroblast: a key cell for wound healing and fibrocontractive diseases. Prog Clin Biol Res **54**:183–194.
- Kumar S, Coenen MJ, Scherer PE, Bahn RS 2004 Evidence for enhanced adipogenesis in the orbits of patients with Graves' ophthalmopathy. J Clin Endocrinol Metab 89:930–935.
- Weetman AP, Cohen S, Gatter KC, Fells P, Shine B 1989 Immunohistochemical analysis of the retrobulbar tissues in Graves' ophthalmopathy. Clin Exp Immunol 75:222–227.
- 45. Drexhage HA, Weetman AP, Heufelder AE, Feldon SE 2000 Future research in Graves' ophthalmopathy. In: Prummel MF, Wiersinga WM, Mourits MP, Heufelder AE (eds.) Recent Developments in Graves' Ophthalmopathy. Kluwer Academic Publishers, Boston, p 187.
- Heufelder AE, Bahn RS 1993 Detection and localization of cytokine immunoreactivity in retro-ocular connective tissue in Graves' ophthalmopathy. Eur J Clin Invest 23:10–17.
- 47. Cao HJ, Wang HS, Zhang Y, Lin HY, Phipps RP, Smith TJ 1998 Activation of human orbital fibroblasts through CD40 engagement results in a dramatic induction of hyaluronan synthesis and prostaglandin endoperoxide H synthase-2 expression. Insights into potential pathogenic mechanisms of thyroid-associated ophthalmopathy. J Biol Chem 273: 29615–29625.
- Sempowski GD, Rozenblit J, Smith TJ, Phipps RP 1998 Human orbital fibroblasts are activated through CD40 to induce proinflammatory cytokine production. Am J Physiol 274: C707–C714.
- Koumas L, Smith TJ, Phipps RP 2002 Fibroblast subsets in the human orbit: Thy-1+ and Thy-1- subpopulations exhibit distinct phenotypes. Eur J Immunol 32:477–485.
- Sciaky D, Brazer W, Center DM, Cruikshank WW, Smith TJ 2000 Cultured human fibroblasts express constitutive IL-16 mRNA: cytokine induction of active IL-16 protein synthesis through a caspase-3-dependent mechanism. J Immunol 164:3806–3814.
- 51. Heufelder AE, Smith TJ, Gorman CA, Bahn RS 1991 Increased induction of HLA-DR by interferon-gamma in

cultured fibroblasts derived from patients with Graves' ophthalmopathy and pretibial dermopathy. J Clin Endocrinol Metab **73**:307–313.

- Otto EA, Ochs K, Hansen C, Wall JR, Kahaly GJ 1996 Orbital tissue-derived T lymphocytes from patients with Graves' ophthalmopathy recognize autologous orbital antigens. J Clin Endocrinol Metab 81:3045–3050.
- Smith TJ, Sempowski GD, Wang HS, Del Vecchio PJ, Lippe SD, Phipps RP 1995 Evidence for cellular heterogeneity in primary cultures of human orbital fibroblasts. J Clin Endocrinol Metab 80:2620–2625.
- Postlethwaite AE, Shigemitsu H, Kanangat S 2004 Cellular origins of fibroblasts: possible implications for organ fibrosis in systemic sclerosis. Curr Opin Rheumatol 16:733–738.
- Lama VN, Phan SH 2006 The extrapulmonary origin of fibroblasts: stem/progenitor cells and beyond. Proc Am Thorac Soc 3:373–376.
- Quan TE, Cowper S, Wu SP, Bockenstedt LK, Bucala R 2004 Circulating fibrocytes: collagen-secreting cells of the peripheral blood. Int J Biochem Cell Biol 36:598–606.
- Fries KM, Blieden T, Looney RJ, Sempowski GD, Silvera MR, Willis RA, Phipps RP 1994 Evidence of fibroblast heterogeneity and the role of fibroblast subpopulations in fibrosis. Clin Immunol Immunopathol 72:283–292.
- Feldon SE, O'Loughlin CW, Ray DM, Landskroner-Eiger S, Seweryniak KE, Phipps RP 2006 Activated human T lymphocytes express cyclooxygenase-2 and produce proadipogenic prostaglandins that drive human orbital fibroblast differentiation to adipocytes. Am J Pathol 169:1183–1193.
- Willson TM, Brown PJ, Sternbach DD, Henke BR 2000 The PPARs: from orphan receptors to drug discovery. J Med Chem 43:527–550.
- Daynes RA, Jones DC 2002 Emerging roles of PPARs in inflammation and immunity. Nat Rev Immunol 2:748–759.
- Rangwala SM, Lazar MA 2004 Peroxisome proliferatoractivated receptor gamma in diabetes and metabolism. Trends Pharmacol Sci 25:331–336.
- Kliewer SA, Lenhard JM, Willson TM, Patel I, Morris DC, Lehmann JM 1995 A prostaglandin J2 metabolite binds peroxisome proliferator-activated receptor gamma and promotes adipocyte differentiation. Cell 83:813–819.
- Barak Y, Nelson MC, Ong ES, Jones YZ, Ruiz-Lozano P, Chien KR, Koder A, Evans RM 1999 PPAR gamma is required for placental, cardiac, and adipose tissue development. Mol Cell 4:585–595.

- 64. Viles-Gonzalez JF, Choi BG, Fuster V, Badimon JJ 2004 Peroxisome proliferator-activated receptor ligands in atherosclerosis. Expert Opin Investig Drugs **13**:1393–1403.
- Harris SG, Padilla J, Koumas L, Ray D, Phipps RP 2002 Prostaglandins as modulators of immunity. Trends Immunol 23:144–150.
- 66. Forman BM, Tontonoz P, Chen J, Brun RP, Spiegelman BM, Evans RM 1995 15-Deoxy-delta 12, 14-prostaglandin J2 is a ligand for the adipocyte determination factor PPAR gamma. Cell 83:803–812.
- Yu K, Bayona W, Kallen CB, Harding HP, Ravera CP, McMahon G, Brown M, Lazar MA 1995 Differential activation of peroxisome proliferator-activated receptors by eicosanoids. J Biol Chem 270:23975–23983.
- Shibata T, Kondo M, Osawa T, Shibata N, Kobayashi M, Uchida K 2002 15-deoxy-delta 12,14-prostaglandin J2. A prostaglandin D2 metabolite generated during inflammatory processes. J Biol Chem 277:10459–10466.
- 69. Smith WL, Langenbach R 2001 Why there are two cyclooxygenase isozymes. J Clin Invest **107**:1491–1495.
- Pablos JL, Santiago B, Carreira PE, Galindo M, Gomez-Reino JJ 1999 Cyclooxygenase-1 and -2 are expressed by human T cells. Clin Exp Immunol 115:86–90.
- Xu L, Zhang L, Yi Y, Kang HK, Datta SK 2004 Human lupus T cells resist inactivation and escape death by upregulating COX-2. Nat Med 10:411–415.
- 72. Rieusset J, Touri F, Michalik L, Escher P, Desvergne B, Niesor E, Wahli W 2002 A new selective peroxisome proliferator-activated receptor gamma antagonist with antiobesity and antidiabetic activity. Mol Endocrinol 16:2628– 2644.
- 73. Ribeiro S, Horuk R 2005 The clinical potential of chemokine receptor antagonists. Pharmacol Ther **107**:44–58.
- Kawamori T, Rao CV, Seibert K, Reddy BS 1998 Chemopreventive activity of celecoxib, a specific cyclooxygenase-2 inhibitor, against colon carcinogenesis. Cancer Res 58:409–412.

Address reprint requests to: Richard P. Phipps, Ph.D. Department of Environmental Medicine University of Rochester School of Medicine and Dentistry 601 Elmwood Ave., Box 850 Rochester, NY 14642

E-mail: richard_phipps@urmc.rochester.edu