Cell-surface glycoproteins of human sarcomas: Differential expression in normal and malignant tissues and cultured cells

(mesenchyme/transformation/immunohistochemistry/monoclonal antibody/Thy-1 antigen)

Wolfgang J. Rettig^{*}, Pilar Garin-Chesa[†], H. Richard Beresford^{*}, Herbert F. Oettgen^{*}, Myron R. Melamed[†], and Lloyd J. Old^{*}

*Laboratory of Human Cancer Immunology and [†]Department of Pathology, Memorial Sloan-Kettering Cancer Center, New York, NY 10021

Contributed by Lloyd J. Old, December 28, 1987

ABSTRACT Normal differentiation and malignant transformation of human cells are characterized by specific changes in surface antigen phenotype. In the present study, we have defined six cell-surface antigens of human sarcomas and normal mesenchymal cells, by using mixed hemadsorption assays and immunochemical methods for the analysis of cultured cells and immunohistochemical staining for the analysis of normal tissues and >200 tumor specimens. Differential patterns of F19 (M_r, 120,000/95,000 glycoprotein), F24 (M_r, 95,000 glycoprotein), G171 (M_r, 75,000 glycoprotein), G253 $(M_r, 90,000 \text{ glycoprotein})$, S5 $(M_r, 120,000 \text{ glycoprotein})$, and Thy-1 (M_r , 25,000 glycoprotein) antigen expression were found to characterize (i) subsets of cultured sarcoma cell lines, (ii) cultured fibroblasts derived from various organs, (iii) normal resting and activated mesenchymal tissues, and (iv) sarcoma and nonmesenchymal tumor tissues. These results provide a basic surface antigenic map for cultured mesenchymal cells and mesenchymal tissues and permit the classification of human sarcomas according to their antigenic phenotypes.

Distinct pathways and stages of cellular differentiation are associated with specific patterns of cell-surface antigen expression. This principle was first established through analysis of normal and malignant cells of hematopoietic origin and has been extended to the neuroectodermal lineage (1-7). The identification of an ordered progression of surface phenotypic changes during normal differentiation has permitted classification of leukemias and lymphomas, melanomas, astrocytomas, and neuroblastomas into subsets that show antigenic similarity to normal cells at distinct stages of hematopoietic or neuroectodermal differentiation (2-4, 6, 7). In contrast to the hematopoietic and neuroectodermal systems. little is known about surface antigenic phenotypes of mesenchymal cells and changes in antigen expression that accompany normal differentiation or malignant transformation of these cells. The present study describes six cellsurface glycoproteins that are differentially expressed during normal development, proliferative activation, or malignant transformation of mesenchymal cells and tissues.

MATERIALS AND METHODS

Cell Lines. Cell lines Hs 913T, TE-85, HT-1080, Saos-2, A-204, A673, RD, SK-ES-1, and WI-38 VA13 subline 2RA were obtained from the American Type Culture Collection. Additional cell lines and normal cell cultures (4, 8) were from the cell bank at Sloan-Kettering Institute.

Antibodies and Serological Procedures. Monoclonal antibody (mAb) G171 (IgG2a) was derived from a mouse immunized with human SK-GS-1 tumor cells as described (7). mAbs F19, F24, G253, S5, and K117 have been described (7, 9, 10). Mixed hemadsorption rosetting assays for detection of antigens on cultured cells were carried out as described (10).

Immunohistochemical Procedures. Tissues were obtained at autopsy or from surgical specimens and quick-frozen as described (11) or, for some experiments with mAb K117, paraffin-embedded by the AMeX method (12); 5 μ m-sections were cut, mounted on gelatin-coated slides, air-dried, and fixed in cold acetone. The avidin-biotin immunoperoxidase and indirect immunofluorescence procedures were carried out as described (11, 13).

Immunochemical Procedures. Cultured cells were metabolically labeled with [35 S]methionine or [3 H]glucosamine, lysed in extraction buffer (0.01 M Tris·HCl, 0.15 M NaCl, 0.01 M MgCl₂, and 0.002 M phenylmethylsulfonyl fluoride) containing 0.5% Nonidet P-40, fractionated on ConA-Sepharose (Pharmacia), and used for immunoprecipitation experiments, followed by NaDodSO₄/polyacrylamide gel electrophoresis and fluorography, as described (4).

RESULTS

Three types of antigenic maps were defined for the six mAb-defined human cell-surface glycoproteins included in this study. (i) For the cultured cell map, 12 established sarcoma cell lines and normal fibroblast cultures derived from 20 individuals were tested by the mixed hemadsorption rosetting assay (Fig. 1) and immunoprecipitation assays (Fig. 2) to define surface phenotypes of cultured mesenchymal cells. (ii) For the normal tissue map, a wide range of normal adult tissues (Table 1 and Figs. 3 and 4) and several fetal tissues (skin, chest wall, kidney, colon, and lung; at 12-20 weeks of gestation) were tested by immunohistochemical procedures. (iii) For the tumor tissue map, tumor samples obtained from >200 patients with sarcomas or other malignancies (Table 2 and Fig. 5) were tested by immunohistochemistry. The following is a description of the most characteristic patterns of antigen expression in vitro and in vivo.

F19 Glycoprotein. mAb F19 defines M_r 120,000 and 95,000 glycoproteins (gp120/95 or the F19 antigen) expressed on cultured fibroblasts and a proportion of sarcoma cell lines but not on simian virus 40 (SV40)-transformed fibroblasts (Figs. 1 and 2). In normal adult tissues, F19 antigen was restricted to occasional fibroblasts and to a subset of pancreatic islet cells, presumably A cells (Table 1). F19 was more widely expressed in fetal mesenchymal tissues, including fibroblasts in the dermis (Fig. 3D), renal capsule, perichondrium and peritoneum. F19 was not found in the stromal fibroblasts of fetal kidney, colon, and lung or in fetal cartilage and skeletal muscle.

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked "*advertisement*" in accordance with 18 U.S.C. §1734 solely to indicate this fact.

Abbreviations: mAb, monoclonal antibody; SV40, simian virus 40; gp, glycoprotein.



Cultured human cells

FIG. 1. Cell-surface phenotypes of established tumor cell lines and short-term cultures of normal fibroblasts. Results of mixed hemadsorption rosetting assay titration experiments (reciprocal titration end points) are shown in histograms. mAb dilutions tested (ascites fluid) are indicated on the right: 1:500, 1:2400, 1:12,000, 1:60,000, 1:300,000, and 1:1,500,000. Cell line origin is indicated as follows: OS, osteogenic sarcoma; RMS, rhabdomyosarcoma; FS, fibrosarcoma; ES, Ewing sarcoma; SV40-WI38, SV40-transformed fetal lung fibroblasts; Nb, neuroblastoma; Rb, retinoblastoma; Ast, astrocytoma; and Mel, melanoma. For fibroblasts, numbers of cultures derived from various individuals are given in brackets. Typing results with mAbs F19, F24, S5, G253, and K117 on additional cell lines have been reported (7, 9); analysis of an expanded cell line panel (8) with mAb G171 identified gp75 expression on 1 of 6 ovarian cancers (A10), 2 of 8 lung cancers, (SK-LC-8 and -17), 1 of 12 astrocytomas (SK-GS-1), and a medulloblastoma (TE-671). All 10 renal, 5 breast, 8 bladder, and 8 colon cancers, 14 melanomas, and 20 leukemias tested were G171⁻, as were short-term cultures of normal kidney epithelial cells, keratinocytes, and skin melanocytes.

In a series of >200 malignant tumors tested by immunohistochemical procedures (Table 2), F19 antigen was found in most types of sarcomas tested, except for those with the "small round cell" phenotype (embryonal rhabdomyosarcoma, Ewing sarcoma, or mesenchymal chondrosarcoma). Neuroectodermal tumors, carcinomas, and lymphomas were consistently F19⁻, but strong induction of F19 was seen in the reactive stroma surrounding many of these F19⁻ tumors (Fig. 5D). To determine whether other modes of mesenchymal activation also induce F19, we tested three skin samples with scar formation following surgical incisions. In



FIG. 2. Immunoprecipitation analysis of human cell-surface antigens. ConA-bound fraction of [³⁵S]methionine-labeled IARC-EW1 cell extract tested with mAb G171 (lane A) or unrelated control mAb (lane B). [³H]Glucosamine-labeled Hs 74 fetal bone marrow fibroblasts tested with mAbs F19 (lane C), F24 (lane D), or G253 (lane E). [³H]Glucosamine-labeled KD adult skin fibroblasts tested with mAbs F19 (lane F), F24 (lane G), or G253 (lane H). [³H]Glucosamine-labeled SV40-transformed fibroblasts tested with mAb F19 (lane I), F24 (lane J), or G253 (lane K). [³H]Glucosamine-labeled Hut 14 fibroblasts tested with mAb S5 (lane M) or control mAb (lane L). Immunoprecipitates were separated on Na DodSO₄/polyacrylamide gels under reducing conditions (extraction buffer containing dithiothreitol at 12 mg/ml). Molecular weights ($M_r \times 10^{-3}$) of immunoprecipitated proteins are indicated on the right. each case, strong F19 expression was seen in the scar but not in adjacent normal dermis.

F24 Glycoprotein. mAb F24 defines a M_r 95,000 glycoprotein (gp95 or the F24 antigen) expressed on cultured fibroblasts but not on most sarcoma cell lines or SV40-transformed fibroblasts (Fig. 1). gp95 comigrated on NaDod-SO₄/polyacrylamide gels with the lower-molecular-weight band of the F19 glycoprotein (Fig. 2) and may be related to this molecule; however, the serological reactivities of mAbs F24 and F19 differed significantly. Both mAbs reacted equally with cultured fibroblasts (Figs. 1 and 2), but mAb F24 was unreactive with most F19⁺ sarcoma cell lines and did not show detectable reactivity with normal tissues or most tumors

 Table 1.
 Distribution of mAb-defined human cell surface antigens in normal adult tissues

Antigen	Antigen-expressing cell types			
F19	Occasional fibroblasts; pancreatic islet (A) cells			
G171	Capillary endothelium; adrenal medullary cells; myoepithelial cells; pneumocytes (weak)			
S5	Visceral smooth muscle; vascular endothelium (weak)			
Thy-1	CNS white and gray matter; peripheral nerves; PNS ganglion and glial cells; myoepithelial cells; mammary ducts; renal tubules (pars convoluta); adrenal endocrine cells; visceral smooth muscle; vascular endothelium; fibroblasts			

Acetone-fixed frozen sections were tested by the avidin-biotin immunoperoxidase procedure. Tissues included were brain, spinal cord, autonomic ganglia, peripheral nerves, skin, mammary gland, parotid gland, tongue, esophagus, stomach, colon, liver, pancreas, bronchus, lung, kidney, urinary bladder, testis, ovary, prostate, uterus, lymph node, spleen, thymus, adrenal gland, thyroid gland, and skeletal and cardiac muscle. CNS, central nervous system; PNS, peripheral nervous system.



FIG. 3. Immunoperoxidase staining of normal human tissues with mAbs to cell surface antigens. mAb S5 tested with smooth muscle of adult urinary bladder (A), adult skeletal muscle (B), and fetal skeletal muscle (C). (D) mAb F19 tested with fetal dermis. Note reactivity with fibroblasts but not epithelial cells of hair follicle (upper left corner). (E) mAb G171 tested with fetal dermis. Note reactivity limited to fibroblasts at the epithelial-mesenchymal junction of the hair follicle. (F) mAb K117 (anti-Thy-1) tested with adult spleen. Note reactivity with connective tissue fibers but not lymphoid cells (germinal center shown in top left corner). Avidin-biotin immunoperoxidase procedure with hematoxylin counterstaining. (A-E) Acetone-fixed frozen tissues. (E) AMeX/paraffin-embedded tissue (12). (For A, $\times 220$; for B-F, $\times 140$.)

tested. Only a single, low-grade fibrosarcoma was found to show strong F24 immunostaining.

G171 Glycoprotein. mAb G171 defines a M_r 75,000 glycoprotein (gp75 or the G171 antigen) expressed on a proportion of sarcoma cell lines and on lung fibroblasts (Figs. 1 and 2); fibroblasts from other normal tissues (skin, kidney, colon, and bone) were consistently G171⁻, independent of the passage levels tested (range: primary culture to >15 passages). In normal adult tissues, G171 antigen was restricted to very few cell types (Table 1), including capillary endothelium and endocrine cells in the adrenal medulla. In the fetus, G171 was expressed in some additional mesenchymal tissues, such as skeletal muscle and cartilage. Fetal perichondrium and renal capsule are G171⁻, but fetal fibroblasts located at epithelial–mesenchymal junctions in skin (Fig. 3*E*), kidney, lung, and colon were G171⁺.

Among the >200 tumors tested by immunohistochemical procedures, G171 was expressed in a large proportion of sarcomas and in meningiomas as well as subsets of Schwannomas, lung and ovarian cancers, and teratocarcinomas (Table 2). Other neuroectodermal tumors, carcinomas, and



FIG. 4. Indirect immunofluorescence staining of normal adult kidney with mAb G253 (A) or anti-Thy-1 mAb K117 (B). Note G253 reactivity with glomerulus and vascular smooth muscle and Thy-1 reactivity restricted to convoluted portions of the renal tubules (upper left, glomerulus; lower left corner, straight portions of tubules). (\times 110.)

lymphomas were typed G171⁻. Similar to the F19 antigen, G171 was found in the reactive stroma surrounding many antigen-negative carcinomas and lymphomas and also in the F19⁺ dermal scars.

S5 Glycoprotein. mAb S5 defines a M_r 120,000 glycoprotein (gp120 or the S5 antigen) expressed on cultured fibroblasts and sarcomas (Figs. 1 and 2). Immunohistochemical analysis (Tables 1 and 2) showed that S5 antigen expression *in vivo* was much more restricted than was suggested by the wide distribution among cultured cells. In normal adult tissues, only visceral smooth muscle cells were strong S5 expressors (Fig. 3A); fibromuscular stroma of prostate and cervix showed weaker S5 immunoreactivity and vascular smooth muscle, cardiac, and skeletal muscle were S5⁻ (Fig. 3B). In the fetus, both visceral smooth muscle and skeletal muscle were S5⁺ (Fig. 3C), but all other cell types tested were S5⁻. S5 expression in tumor tissues was restricted to a subset of leiomyosarcomas that generally showed weaker immunoreactivity than normal visceral smooth muscle.

G253 Glycoprotein. mAb G253 defines a M_r 90,000 glycoprotein (gp90 or the G253 antigen) expressed on cultured sarcomas and SV40-transformed fibroblasts but not, or only weakly, on cultured normal fibroblasts (Figs. 1 and 2). These results suggested that G253 antigen was a transformation-specific marker of mesenchymal cells, and we used immunohistochemical procedures to test 40 sarcomas for G253 expression. However, none of the tumor tissues showed antigen expression. In contrast, G253 was readily detected in normal vascular and visceral smooth muscle and in kidney glomeruli (Fig. 4A), demonstrating that lack of G253 reactivity with sarcoma tissues was not due to failure of the mAb to detect antigen in acetone-fixed frozen tissue sections.

Thy-1. mAb K117 recognizes human Thy-1 (10), a M_r 25,000 glycoprotein (gp25) expressed on cultured fibroblasts and sarcomas (Fig. 1). Immunohistochemical analysis showed Thy-1 expression in a variety of cell types (Table 1), including fibroblasts, muscularis mucosae of the gastrointestinal tract, fibromuscular stroma of prostate and cervix, connective tissue fibers (but not lymphoid cells) in spleen and lymph nodes (Fig. 3F), and specific portions of the renal tubules (Fig. 4B). In contrast, the muscularis propria of stomach and colon, smooth muscle of bladder and corpus uteri, smooth muscle of arterial blood vessels, skeletal and

·	Antigen expression			
T				
Tumor type	F19	61/1	33	1 ny-1
Sarcomas				
Fibrosarcoma	5/5	4/5	0/5	5/5
MFH	4/4	4/4	0/4	3/4
Leiomyosarcoma	8/10	3/10	6/10	7/10
Osteosarcoma	2/4	4/4	0/4	3/4
Chondrosarcoma	2/2	2/2	0/2	2/2
Liposarcoma	3/4	3/4	0/4	3/4
Synovial sarcoma	1/6	5/6	0/6	6/6
ERMS	0/6	1/6	0/6	2/6
Ewing sarcoma	0/8	0/8	0/8	0/8
Mes. chondrosarcoma	0/2	1/2	0/2	0/2
Rhabdomyosarcoma	0/2	1/1	0/1	2/2
Undifferentiated	2/3	2/3	0/2	3/3
Neuroectodermal tumors				
Melanoma	0/12	0/12	0/12	1/12
Astrocytoma	0/12	0/12	0/12	12/12
Schwannoma	2/7	2/6	0/4	6/7
Neuroblastoma	0/5	0/5	0/5	5/5
Meningioma	0/5	5/5	0/5	5/5
Carcinomas	·	•		•
Neuroendocrine	0/7	0/7	0/7	4/7
Colorectal	0/18	0/18	0/18	0/18
Gastric	0/6	0/6	0/5	0/6
Skin	0/8	0/8	0/8	0/8
Lung	0/10	2/10	0/8	0/8
Breast	0/13	0/13	0/13	0/13
Ovarian	0/21	7/21	0/21	7/20
Testicular	0/5	4/5	0/5	3/4
Kidney	0/9	0/9	0/9	0/9
Bladder	0/6	0/6	0/6	0/6
Others	0/10	0/10	0/10	0/10
Lymphomas		-,	-, -,	-,
Hodgkin	0/5	0/4	0/4	5/5
Non-Hodgkin	0/12	0/12	0/11	8/14
	~, ~ ~	•,	v, ++	•/ ⊥

 Table 2.
 Distribution of mAb-defined human cell surface

 antigens in tumor tissues

Acetone-fixed frozen sections were tested by the avidin-biotin immunoperoxidase procedure. Numbers indicate the proportion of tumors obtained from various patients that express the respective antigens (number antigen-positive/total number tested). Strong intratumoral heterogeneity in antigen expression was only seen for G171 in the lung cancers listed as G171⁺. MFH, malignant fibrous histiocytoma; ERMS, embryonal rhabdomyosarcoma; mes. chondrosarcoma, mesenchymal chondrosarcoma.

cardiac muscle were Thy-1⁻ in the adult. Thy-1 expression in the fetus paralleled the adult pattern with two exceptions: basal keratinocytes of the skin and renal glomeruli expressed Thy-1 in the fetus but were Thy- 1^{-} in the adult. Contrary to previous suggestions (14), we did not find any Thy-1 expression in fetal skeletal muscle. Thy-1 expression in tumor tissues (Table 2) mirrored its distribution in normal tissues: Most sarcomas and neuroectodermal tumors (Fig. 5 G and H) were Thy- 1^+ , whereas most carcinomas were Thy- 1^- . Notable exceptions were the presence of Thy-1 in a proportion of ovarian carcinomas, the absence of Thy-1 from melanomas [consistent with the Thy-1⁻ phenotype of most cultured melanoma cell lines (9)], and the lack of Thy-1 expression in Ewing sarcomas, embryonal rhabdomyosarcomas, and mesenchymal chondrosarcomas (Fig. 5F), a group of tumors distinguished by their "small round cell" phenotype.

DISCUSSION

Mesenchymal tissues comprise a range of cell types that differ in embryological derivation, morphology, and function. At present, little is known about the surface antigenic phenotypes of normal or neoplastic mesenchymal cells. In this study, we define distinct patterns of expression for six cell-surface glycoproteins that distinguish (*i*) normal mesenchymal cells of various lineages, (*ii*) mesenchymal cells at distinct stages of development or differentiation, and (*iii*) subsets of mesenchymal tumors.

The F19 antigen is expressed in several fetal mesenchymal tissues, many sarcomas, the stroma of F19⁻ carcinomas, scar tissue, and cultured fibroblasts, but is not generally found in normal adult mesenchyme. This pattern suggests that F19 is a cell-surface marker for proliferating mesenchymal cells and that its expression may be induced by normal growth factors or during malignant transformation. Since F19 expression is apparently down-regulated in SV40-transformed cultured fibroblasts, this mode of *in vitro* transformation and spontaneous transformation *in vivo* differ in their effects on surface antigen phenotype. Expression of the F24 antigen is similar in some respects to that of F19, but F24 is more restricted in its distribution and the two antigens differ biochemically.

The G171 antigen differs from F19 and F24 in molecular size and tissue distribution. For example, among cultured fibroblasts, G171 is expressed on only those derived from lung, whereas F19 and F24 are expressed on fibroblasts derived from several organs. Since none of these antigens is generally expressed in normal adult mesenchyme, it seems likely that their expression in the cultured cells is a response to mesenchymal growth or differentiation signals provided by the in vitro culture system and that fibroblasts derived from various organs respond differently to these extrinsic signals. Similarly, differential regulation for G171 and F19 by extrinsic signals in vivo and in vitro could account for other discordancies between the cultured cell maps and tissue maps of these antigens. For example, cultured Ewing sarcoma cells strongly express G171 and subsets of cultured astrocytomas and melanomas strongly express F19 (7), but the corresponding fresh tumor tissues are antigen-negative. Furthermore, cultured cells derived from fibrosarcomas are G171⁻, whereas the corresponding tumor tissues are generally G171⁺. Parallel analysis of tumor specimens and derived cell lines will help determine at which stage changes in antigenic phenotype occur.

The G253 and S5 antigens also exhibit various patterns of expression in tissues and in cultured cell panels. G253 appears to be a marker of transformed mesenchymal cells in vitro. It is strongly expressed in most sarcoma cell lines and in SV40-transformed fibroblasts but was not found in any uncultured sarcomas tested by immunohistochemical methods. S5 is expressed on a wide range of cultured mesenchymal and neuroectodermal cells, as well as on some cultured epithelial and hematopoietic cells (7), but in normal adult tissues the antigen is restricted to visceral smooth muscle. Moreover, among tumors tested by immunohistochemical methods, S5 is restricted to leiomyosarcomas. Analysis of normal fetal tissues reveals that additional cell types express S5 transiently during development (e.g., skeletal muscle). It is thus conceivable that the wide distribution of S5 and G253 on cultured cells reflects specific activation of early developmental traits in the cultures or adaptive changes induced by altered cell-substrate or cell-cell interactions in cells grown on plastic culture surfaces. Since uncultured sarcomas and reactive mesenchyme do not generally express G253 or S5, it is less likely that they are linked to proliferation, as was suggested for F19.

The Thy-1 antigen was first identified as a thymocyte differentiation antigen in mice but has subsequently been found in lymphoid and neural tissues of several species, including humans (15). The present study provides an extensive analysis of Thy-1 expression in human tumors and



FIG. 5. Immunoperoxidase staining of tumor tissues with mAbs to cell surface antigens. mAb F19 tested with a chondrosarcoma (A), osteogenic sarcoma (B), fibrosarcoma (C), and colon cancer (D). Note reactivity with tumor cells in A-C but reactivity in D is limited to the reactive stroma surrounding clusters of colon cancer cells. Mesenchymal chondrosarcoma tested with mAb G171 (E) and parallel section tested with mAb K117 (anti-Thy-1) (F). Note Thy-1 expression in the perivascular connective tissue but not in the tumor cells. mAb K117 tested with a ganglioneuroblastoma (G) and malignant Schwannoma (H). (I) mAb G171 tested with a meningioma. Avidin-biotin immunoperoxidase staining of acetone-fixed frozen sections with hematoxylin counterstaining. $(\times 140.)$

normal tissues. With respect to mesenchymal tissues, we found that subsets of normal cells can be distinguished by their Thy-1 phenotypes (e.g., visceral and vascular smooth muscle in various sites differ in antigen expression) and that sarcomas are generally Thy-1⁺, except for those with "small round cell" phenotype (Ewing sarcoma, embryonal rhabdomyosarcoma, and mesenchymal chondrosarcoma).

Detailed analysis (cultured cell map, normal tissue map, and tumor tissue map) of six distinct human cell surface glycoproteins has revealed characteristic patterns of antigen expression in normal developing and adult mesenchyme and in mesenchymal tumors. It is apparent that typing with mAbs F19, G171, S5, and K117 identifies subsets of sarcomas with distinctive antigenic phenotypes, and it is possible that these antigenic patterns correlate with differences in histogenesis or biological behavior. We have shown (16) that differential expression of the cell surface receptor for nerve growth factor also distinguishes subsets of human sarcomas. but in contrast to the glycoproteins described here, the receptor for nerve growth factor is selectively expressed in Ewing sarcomas and embryonal rhabdomyosarcomas. Since expression of F19, G171, and S5 is highly restricted in normal tissues, they may be useful targets for immunolocalization or immunotherapy of antigen-expressing tumors.

We are grateful to S. Walker, G. Lark, and D. Josefson for expert technical assistance and to J. Rios for excellent secretarial help. This work was supported in part by grants from the National Cancer Institute (CA-08748 and CA-25803) and by the Oliver S. and Jennie R. Donaldson Charitable Trust.

- 1. Boyse, E. A. & Old, L. J. (1969) Annu. Rev. Genet. 3, 269-290.
- 2. Foon, K. A. & Todd, R. F. (1986) Blood 68, 1-31.
- Houghton, A. N., Eisinger, M., Albino, A. P., Cairncross, J. G. & Old, L. J. (1982) J. Exp. Med. 156, 1755-1766.
- Rettig, W. J., Murty, V. V. V. S., Mattes, M. J., Chaganti, R. S. K. & Old, L. J. (1986) J. Exp. Med. 164, 1581–1599.
- Rettig, W. J., Garin Chesa, P., Jennings, M. T., Spengler, B. A., Melamed, M. R., Oettgen, H. F., Biedler, J. L. & Old, L. J. (1985) Proc. Natl. Acad. Sci. USA 82, 6894-6898.
- Rettig, W. J., Spengler, B. A., Garin Chesa, P., Old, L. J. & Biedler, J. L. (1987) Cancer Res. 47, 1383–1389.
- Rettig, W. J., Garin Chesa, P., Beresford, H. R., Feickert, H.-J., Jennings, M. T., Cohen, J., Oettgen, H. F. & Old, L. J. (1986) Cancer Res. 46, 6406-6412.
- Rettig, W. J., Cordon-Cardo, C., Ng, J. S. C., Oettgen, H. F., Old, L. J. & Lloyd, K. O. (1985) Cancer Res. 45, 815–821.
- Rettig, W. J., Dracopoli, N. C., Garin Chesa, P., Spengler, B. A., Beresford, H. R., Davies, P., Biedler, J. L. & Old, L. J. (1985) J. Exp. Med. 162, 1603-1619.
 Rettig, W. J., Nishimura, H., Yenamandra, A. K., Seki, T.,
- Rettig, W. J., Nishimura, H., Yenamandra, A. K., Seki, T., Obata, F., Beresford, H. R., Old, L. J. & Silver, J. (1987) J. Immunol. 138, 4484-4489.
- Gordon, J., Garin Chesa, P., Nishimura, H., Rettig, W. J., Maccari, J. E., Endo, T., Seravalli, E., Seki, T. & Silver, J. (1987) Cell 50, 445-452.
- Sato, Y., Mukai, K., Watanabe, S., Goto, M. & Shimosato, Y. (1986) Am. J. Pathol. 125, 431–435.
- Garin Chesa, P., Rettig, W. J. & Melamed, M. R. (1986) Am. J. Surg. Pathol. 10, 829-835.
- 14. Walsh, F. & Ritter, M. A. (1981) Nature (London) 289, 60-64.
- 15. Williams, A. F. & Gagnon, J. (1982) Science 216, 696-703.
- 16. Garin Chesa, P., Rettig, W. J., Thomson, T. M., Old, L. J. & Melamed, M. R. J. Histochem. Cytochem., in press.