

Parkinsonism Relat Disord. Author manuscript; available in PMC 2010 January 14.

Published in final edited form as:

Parkinsonism Relat Disord. 2008; 14(Suppl 2): S130–S134. doi:10.1016/j.parkreldis.2008.04.017.

Dopamine/adenosine interactions related to locomotion and tremor in animal models: Possible relevance to parkinsonism*

John D. Salamone^{a,*}, Keita Ishiwari^{a,b}, Adrienne J. Betz^{a,c}, Andrew M. Farrar^a, Susana M. Mingote^{a,d}, Laura Font^{a,e}, Jörg Hockemeyer^f, Christa E. Müller^f, and Mercè Correa^{a,d}

^aBehavioral Neuroscience Division, Department of Psychology, University of Connecticut, Storrs, CT 06269-1020, USA ^bBioAnalytics Laboratory, Department of Chemistry, National University of Ireland, Maynooth, Co. Kildare, Ireland ^cDepartment of Psychiatry, Yale University, Connecticut Mental Health Center, 34 Park Street, New Haven, CT 06508, USA ^dDepartment of Psychiatry and Molecular Therapeutics, Columbia University, 1051 Riverside Drive, NYSPI Unit 62, New York, NY 10032-2695, USA ^eArea de Psicobiologia, Universitat Jaume I, 12071 Castelló, Spain ^fPharmazeutisches Institut, Pharmazeutische Chemie I, Universität Bonn, Bonn, Germany

Abstract

Adenosine A_{2A} antagonists can exert antiparkinsonian effects in animal models. Recent experiments studied the ability of MSX-3 (an adenosine A_{2A} antagonist) to reverse the locomotor suppression and tremor produced by dopamine antagonists in rats. MSX-3 reversed haloperidol-induced suppression of locomotion, and reduced the tremulous jaw movements induced by haloperidol, pimozide, and reserpine. Infusions of MSX-3 into the nucleus accumbens core increased locomotion in haloperidol-treated rats, but there were no effects of infusions into the accumbens shell or ventrolateral neostriatum. In contrast, MSX-3 injected into the ventrolateral neostriatum reduced pimozide-induced tremulous jaw movements. Dopamine/adenosine interactions in different striatal subregions are involved in distinct aspects of motor function.

Keywords

Dopamine; Basal ganglia; Neostriatum; Caudate; Putamen; Nucleus accumbens; Antipsychotic; Parkinson's disease; Motor; Motivation

1. Introduction

Interactions between diverse neurotransmitter systems in the basal ganglia are thought to regulate aspects of motor function related to parkinsonism. In addition to dopamine (DA), considerable research has implicated several other basal ganglia neurotransmitters, including acetylcholine, serotonin, glutamate and GABA, in aspects of motor function and dysfunction [1-4]. More recently, brain adenosine neurons have also been implicated in regulating the motor functions of the basal ganglia [5-7]. Anatomical studies have demonstrated that the adenosine A_{2A} receptor subtype is highly expressed in DA-rich striatal regions [7-11]. Adenosine A_{2A}

^{*}This article is based on a presentation given at the LIMPE Seminars 2007 "Experimental Models in Parkinson's Disease" held in September 2007 at the "Porto Conte Ricerche" Congress Center in Alghero, Sardinia, Italy.

^{© 2008} Elsevier Ltd. All rights reserved.

^{*}Corresponding author. Tel.: +1 860 486 4302; fax: +1 860 486 2760. *E-mail address*: john.salamone@uconn.edu (J.D. Salamone).. **Conflict of interest**: The authors have declared no conflicts of interest.

receptors in the striatum are largely expressed on enkephalin-positive striatopallidal neurons, which also contain DA D_2 receptors [7-10]. Adenosine A_{2A} receptor antagonists produce motor effects in animal models, and it has been widely suggested that adenosine A_{2A} antagonists could be used as non-dopaminergic treatments for parkinsonian symptoms [4,5,11-17]. For all these reasons, it is important to characterize the effects of adenosine A_{2A} antagonists in animal models [18].

2. Studies of locomotor activity in rats

Adenosine A_{2A} antagonists have been assessed for their motor effects using a number of tasks that are suitable for rodents. Haloperidol-induced rigidity was reversed by the A_{2A} antagonist SCH 58261 [19]. Hauber et al. [20] observed that catalepsy induced by DA antagonists could be reversed by the selective A_{2A} antagonist MSX-3. In addition, several studies have focused on the effects of adenosine A_{2A} antagonists on locomotor activity. The adenosine A_{2A} antagonist KW-6002 reversed the hypolocomotion induced by the DA-depleting agent reserpine [17]. The impairment of locomotion shown by D_2 receptor-deficient mice was rescued by the adenosine A_{2A} antagonist KW-6002 [21]. Systemic injections of the adenosine A_{2A} antagonist KF17837 (5.0–20.0 mg/kg) reversed the suppression of locomotion induced by subchronic injections of haloperidol [22].

In order to understand more fully the brain mechanisms mediating the effects of drugs acting on adenosine, it is important to identify the specific brain locus at which adenosine A_{2A} receptor antagonists act to increase locomotion in animals with impaired dopaminergic function. Adenosine A_{2A} receptors are present throughout the striatal complex, which includes the caudate/putamen (i.e., neostriatum) and also the nucleus accumbens [6,7,9,10]. Previous studies have indicated that adenosine A_{2A} receptors in the nucleus accumbens may be important for mediating the locomotor effects of drugs that act on adenosine A_{2A} receptors [23-25]. Moreover, there is considerable evidence indicating that interference with DA transmission in the nucleus accumbens leads to a suppression of spontaneous locomotion [26-29].

Recently, experiments were conducted to study the ability of systemic or intra-accumbens injections of the selective adenosine A_{2A} antagonist MSX-3 to reverse the locomotor effects of acute or subchronic administration of haloperidol in rats [30]. Haloperidol is a DA antagonist that is known to suppress locomotion in rats (e.g., Ref. [22]) and to produce parkinsonian sideeffects in humans [31,32]. MSX-3 is a water-soluble pro-drug that is rapidly cleaved by phosphatases in vivo into MSX-2, which is the active antagonist of A_{2A} receptors [33]. Therefore, we studied the ability of systemic injections of MSX-3 to reverse the suppression of locomotion induced by acute or repeated subchronic administration of 0.5 mg/kg haloperidol [30]. Repeated administration of haloperidol was used because this procedure has been employed previously for studies of adenosine A_{2A} antagonists [22] and because repeated administration mimics the conditions seen when antipsychotic drugs such as haloperidol are used clinically. Additional experiments have studied the ability of intracranial injections of MSX-3 to increase locomotion in haloperidol-treated rats [30]. Three brain areas were studied: nucleus accumbens core, dorsomedial nucleus accumbens shell and ventrolateral neostriatum (VLS). The nucleus accumbens was investigated because this brain area is involved in the regulation of locomotor activity [23,24,26-28,30]. Although earlier studies examined the effects of intra-accumbens injections of MSX-3 on locomotor activity, these studies did not differentiate between core and shell subregions, and they did not assess the effects of A_{2A} antagonism in the presence of a DA antagonist. The VLS site was chosen as a control striatal site because this striatal subregion is thought to be involved in motor functions such as tremor (see below) and skilled motor control [34-36], but is not thought to be important for locomotion [28,37,38].

In these studies, systemic injections of MSX-3 in a dose range of 2.5–10.0 mg/kg were capable of reversing the suppression of locomotion induced by either acute or repeated (i.e., 14 day) administration of haloperidol 0.5 mg/kg [30]. Bilateral infusions of MSX-3 into the nucleus accumbens core (2.5-5.0 µg per side) produced a dose-related increase in locomotor activity in rats treated with 0.5 µg/kg haloperidol either acutely or repeatedly [30]. There was no overall significant effect of MSX-3 infused into either the dorsomedial shell or the VLS. In addition, there were no significant effects of systemic or intra-accumbens injections of MSX-3 (10.0 mg/kg and 5.0 µg per side, respectively) in rats that were not treated with haloperidol. These results indicate that antagonism of adenosine A2A receptors can reverse the locomotor suppression produced by DA antagonism and that a critical site for this effect is the nucleus accumbens core. Although Parkinson's disease is generally associated with depletions of DA in the neostriatum [39], this disorder is also characterized by nucleus accumbens DA depletions [40,41]. As with rodents, the nucleus accumbens of primates is also involved in locomotion [42]. Thus, it is possible that DA/adenosine interactions in the nucleus accumbens may be important for regulating behavioural functions, including locomotion, that are impaired in parkinsonism.

3. Studies of tremulous jaw movements

There is considerable uncertainty about the neurochemical mechanisms that underlie tremor generation, despite the fact that resting tremor is one of the primary symptoms of parkinsonism. A few studies have examined the effects of adenosine antagonists on parkinsonian tremor in humans, and some positive effects have been reported [14,43]. One of the rodent procedures used as a model of parkinsonian resting tremor is drug-induced tremulous jaw movements (TJMs). TJMs are rapid vertical deflections of the lower jaw that resemble chewing but are not directed at any stimulus [44]. Studies using slow-motion or freeze-frame video analyses, as well as electromyographic methods, have shown that these movements occur largely in the 3to 7-Hz range that is also characteristic of parkinsonian resting tremor [44-47]. TJMs can be induced by striatal DA depletions [37,45] and by centrally acting cholinomimetic drugs [2, 44,48-51]. They are also induced by typical antipsychotics, such as haloperidol [22,52], pimozide [46,48] and reserpine [47], but not by atypical antipsychotics [52,53]. Although chronic administration of antipsychotic drugs can result in oral movements that may be related to other movement disorders, such as tardive dyskinesia, considerable evidence indicates that the chewing-like jaw movements induced by acute or subchronic administration of typical antipsychotic drugs share many characteristics with parkinsonian symptoms [2,44,46,47,51]. TJMs have been used as a rodent model of parkinsonian tremor for assessing antiparkinsonian drugs with various pharmacological profiles [4,47,48,51]. The adenosine A_{2A} antagonist KF17837 (10.0–20.0 mg/kg) suppressed haloperidol-induced TJMs [22], and the TJMs induced by the acetylcholinesterase inhibitor tacrine were reduced by systemic or intrastriatal injections of the adenosine A_{2A} antagonists SCH 58261 and SCH BT2 [54].

In a recent series of experiments, the potential antiparkinsonian effects of the selective adenosine A_{2A} antagonist MSX-3 were assessed by using acute or subchronic administration of antipsychotic drugs to induce TJMs [55]. In the first group of studies, pimozide (Orap) was used to induce motor impairments. Pimozide is a typical antipsychotic drug, which has been shown to produce motor side-effects in patients with schizophrenia and to exacerbate the symptoms of Parkinson's disease [46,56]. Moreover, pimozide has been reported to be more likely to produce parkinsonian tremor compared with other typical antipsychotics [56]. In recent papers, it was demonstrated that pimozide could induce TJMs with acute or subchronic administration (i.e., 1, 7 or 13 days of injections) at doses up to 1.0 mg/kg [46], and that the TJMs induced by repeated pimozide were blocked by the antiparkinsonian anti-cholinergic drug atropine [48]. Based on these previous experiments, the first group of studies assessed the ability of adenosine A_{2A} antagonism to suppress tremulous movements and increase motor

activity in pimozide-treated rats [55]. In these studies, rats were injected with 1.0 mg/kg of pimozide for 7 days, and on the eighth day they received injections of pimozide plus various doses of the A_{2A} antagonists KW-6002 or MSX-3. After receiving these drug treatments, the rats were assessed with a battery of motor tests that included observations of TJMs, catalepsy and locomotor activity. Administration of both KW-6002 and MSX-3 suppressed pimozideinduced TJMs, and also reduced catalepsy and increased locomotion in the pimozide-treated rats [55]. Additional studies showed that MSX-3 suppressed the TJMs induced by haloperidol, as well as the DA-depleting agent reserpine [55]. An additional experiment investigated the effects of intracranial injections of MSX-3 into the VLS, in order to determine whether local injections of an adenosine A_{2A} antagonist could reverse the TJMs induced by pimozide [55]. The VLS was chosen because this brain area, which is thought to be the homologue of the ventral putamen in primates, has been strongly implicated in the control of TJM activity [34, 37,44,49,54,57]. This experiment demonstrated that injections of MSX-3 into the VLS were able to suppress pimozide-induced TJMs [55], which was consistent with an earlier study showing that injections of an adenosine A_{2A} antagonist into the VLS could reduce the TJMs induced by the cholinomimetic drug tacrine [54].

4. Discussion

Taken together, the results of these experiments indicate that adenosine A_{2A} antagonism can reverse locomotor suppression and tremulous movements induced by typical antipsychotics [22,30,55]. These effects are consistent with the hypothesis that blockade of adenosine A_{2A} receptors can produce antiparkinsonian effects in animal models. Adenosine A_{2A} antagonists may be useful clinically for their tremorolytic effects, and may help in treating both idiopathic and antipsychotic-induced parkinsonian symptoms [22,30,55]. Moreover, these experiments indicate that different striatal subregions are involved in distinct aspects of motor function. This principle has been demonstrated clearly in the substantial literature showing that DA depletions or antagonism can have regionally specific effects [28,37,58], and it has important implications for understanding the anatomical mechanisms underlying the motor effects of antiparkinsonian drugs, including adenosine A2A antagonists. Although antiparkinsonian drugs are typically given systemically, with the intention of producing an improvement in several different motor symptoms, it is nevertheless reasonable to suggest that different therapeutic effects (i.e., increase in locomotion, decrease in rigidity or tremor) are related to actions on distinct striatal subregions. In addition to studying these specific aspects of motor function, future research should also investigate the potential role of adenosine A_{2A} receptors in motivational functions that are impaired in parkinsonism, such as psychomotor activation and effort-related processes [59].

Acknowledgments

This research was supported by a grant to J.D.S. from the US NIH/NIMH (Grant numbers: NIH NS47261-2 and MH078023-01A1).

Role of the funding source Funds from the NIH/NINDS and NIMH were used to provide support for the basic research described here.

References

- [1]. DeLong MR. Primate model of movement disorders of basal ganglia origin. Trends Neurosci 1990;13:281–5. [PubMed: 1695404]
- [2]. Salamone J, Correa M, Carlson B, Wisniecki A, Mayorga A, Nisenbaum E, et al. Neostriatal muscarinic receptor subtypes involved in the generation of tremulous jaw movements in rodents. Implications for cholinergic involvement in parkinsonism. Life Sci 2001;68:2579–84. [PubMed: 11392629]

[3]. Wichmann T, Kliem MA, DeLong MR. Antiparkinsonian and behavioral effects of inactivation of the substantia nigra pars reticulata in hemiparkinsonian primates. Exp Neurol 2001;167:410–24. [PubMed: 11161630]

- [4]. Cousins MS, Carriero DL, Salamone JD. Tremulous jaw movements induced by the acetylcholinesterase inhibitor tacrine: effects of antiparkinsonian drugs. Eur J Pharmacol 1997;322:137–45. [PubMed: 9098680]
- [5]. Ferré S, Freidholm BB, Morelli M, Popoli P, Fuxe K. Adenosine-dopamine receptor—receptor interactions as an integrative mechanism in the basal ganglia. Trends Neurosci 1997;20:482–7. [PubMed: 9347617]
- [6]. Ferré S, Popoli P, Gimenez-Llort L, Rimondini R, Müller CE, Stromberg I, et al. Adenosine/dopamine interaction: implications for the treatment of Parkinson's disease. Parkinsonism Relat Disord 2001;7:235–41. [PubMed: 11331192]
- [7]. Svenningsson P, Le Moine C, Fisone G, Fredholm BB. Distribution, biochemistry and function of striatal adenosine A_{2A} receptors. Prog Neurobiol 1999;59:355–96. [PubMed: 10501634]
- [8]. Chen JF, Moratalla R, Impagnatiello F, Grandy DK, Cuellar B, Rubinstein M, et al. The role of the D₂ dopamine receptor (D₂R) in A₂A adenosine-receptor (A₂AR) mediated behavioral and cellular responses as revealed by A₂A and D₂ receptor knockout mice. Proc Natl Acad Sci U S A 2001;98:1970–5. [PubMed: 11172060]
- [9]. Fink JS, Weaver DR, Rivkees SA, Peterfreund RA, Pollack AE, Adler EM, et al. Molecular cloning of the rat A2 adenosine receptor: selective co-expression with D2 dopamine receptors in rat striatum. Brain Res 1992;14:186–95.
- [10]. Hettinger BD, Lee A, Linden J, Rosin DL. Ultrastructural localization of adenosine A_{2A} receptors suggests multiple cellular sites for modulation of GABAergic neurons in rat striatum. J Comp Neurol 2001;431:331–46. [PubMed: 11170009]
- [11]. Wang WF, Ishiwata K, Nonaka H, Ishii S, Kiyosawa M, Shimada J, et al. Carbon-11-labeled KF21213: a highly selective ligand for mapping CNS adenosine A(2A) receptors with positron emission tomography. Nucl Med Biol 2000;27:541–6. [PubMed: 11056367]
- [12]. Fuxe K, Ferré S, Zoli M, Agnati LF. Integrated events in central dopamine transmission as analyzed at multiple levels. Evidence for intra-membrane adenosine A2A/dopamine D2 and adenosine A1/ dopamine D1 receptor interactions in the basal ganglia. Brain Res Rev 1998;26:258–73. [PubMed: 9651540]
- [13]. Jenner P. Istradefylline, a novel adenosine A2A receptor antagonist, for the treatment of Parkinson's disease. Expert Opin Investig Drugs 2005;14:729–38.
- [14]. Mally J, Stone TW. Potential of adenosine A_{2A} antagonists in the treatment of movement disorders. CNS Drugs 1998;10:311–20.
- [15]. Morelli M, Pinna A. Interaction between dopamine and adenosine A_{2A} receptors as a basis for the treatment of Parkinson's disease. Neurol Sci 2001;22:71–2. [PubMed: 11487207]
- [16]. Pinna A, Wardas J, Simola N, Morelli M. New therapies for the treatment of Parkinson's disease: adenosine A_{2A} receptor antagonists. Life Sci 2005;77:3259–67. [PubMed: 15979104]
- [17]. Shiozaki S, Ichikawa S, Nakamura J, Kitamura S, Yamada K, Kuwana Y. Actions of adenosine A_{2A} receptor antagonist KW-6002 on drug-induced catalepsy and hypokinesia caused by reserpine or MPTP. Psychopharmacology 1999;147:90–5. [PubMed: 10591873]
- [18]. Cenci MA, Whishaw IQ, Schallert T. Animal models of neurological deficits: how relevant is the rat? Nat Rev Neurosci 2002;3:574–9. [PubMed: 12094213]
- [19]. Wardas J, Konieczny J, Lorenc-Koci E. SCH 58261, an A(2A) adenosine receptor antagonist, counteracts parkinsonian-like muscle rigidity in rats. Synapse 2001;41:160–71. [PubMed: 11400182]
- [20]. Hauber W, Neuscheler P, Nagel J, Müller CE. Catalepsy induced by a blockade of dopamine D1 or D2 receptors was reversed by a concomitant blockade of adenosine A_{2A} receptors in the caudate putamen of rats. Eur J Neurosci 2001;14:1287–93. [PubMed: 11703457]
- [21]. Aoyama S, Kase H, Borrelli E. Rescue of locomotor impairment in dopamine D2 receptor-deficient mice by an adenosine A_{2A} receptor antagonist. J Neurosci 2000;20:5848–82. [PubMed: 10908627]
- [22]. Correa M, Wisniecki A, Betz A, Dobson DR, O'Neill MF, O'Neill MJ, et al. The adenosine A_{2A} antagonist KF 17837 reverses the locomotor suppression and tremulous jaw movements induced

- by haloperidol in rats: possible relevance to parkinsonism. Behav Brain Res 2004;148:47–54. [PubMed: 14684247]
- [23]. Barraco RA, Martens KA, Parizon M, Normile HJ. Role of adenosine A2A receptors in the nucleus accumbens. Prog Neuro-psychopharmacol Biol Psychiatry 1994;18:545–53.
- [24]. Hauber W, Munkle M. Motor depressant effects mediated by dopamine D₂ and adenosine A_{2A} receptors in the nucleus accumbens and the caudate-putamen. Eur J Pharmacol 1997;323:127–31. [PubMed: 9128830]
- [25]. Nagel J, Shladebach H, Kock M, Schwienbacher I, Müller CE, Hauber W. Effects of an adenosine A_{2A} receptor blockade in the nucleus accumbens on locomotion, feeding, and prepulse inhibition in rats. Synapse 2003;49:279–86. [PubMed: 12827647]
- [26]. Baldo BA, Sadeghian K, Basso AM, Kelley AE. Effects of selective dopamine D1 or D2 receptor blockade within nucleus accumbens subregions on ingestive behavior and associated motor activity. Behav Brain Res 2002;137:165–77. [PubMed: 12445722]
- [27]. Correa M, Carlson BB, Wisniecki A, Salamone JD. Nucleus accumbens dopamine and work requirements on interval schedules. Behav Brain Res 2002;137:179–87. [PubMed: 12445723]
- [28]. Cousins MS, Sokolowski JD, Salamone JD. Different effects of nucleus accumbens and ventrolateral striatal dopamine depletions on instrumental response selection in the rat. Pharmacol Biochem Behav 1993;46:943–51. [PubMed: 8309975]
- [29]. Koob GF, Riley SJ, Smith SC, Robbins TW. Effects of 6-hydroxydopamine lesions of the nucleus accumbens septi and olfactory tubercle on feeding, locomotor activity, and amphetamine anorexia in the rat. J Comp Physiol Psychol 1978;92:917–27. [PubMed: 282297]
- [30]. Ishiwari K, Madson LJ, Farrar AM, Mingote SM, DiGianvittorio MD, Frank LE, et al. Injections of the selective adenosine A_{2A} antagonist MSX-3 into the nucleus accumbens core attenuate the locomotor suppression induced by haloperidol in rats. Behav Brain Res 2007;178:190–9. [PubMed: 17223207]
- [31]. Bezchlibnyk-Butler KZ, Remington GJ. Antiparkinsonian drugs in the treatment of neuroleptic-induced extrapyramidal symptoms. Can J Psychiatry 1994;39:74–84. [PubMed: 7908605]
- [32]. Marsden C, Duvoisin R, Jenner P, Parkes J, Pycock C, Tarsy D. Relationship between animal models and clinical parkinsonism. Adv Neurol 1975;9:165–75. [PubMed: 1146651]
- [33]. Hockemeyer J, Burbiel JC, Muller CE. Multigram-scale syntheses, stability, and photoreactions of A2A adenosine receptor antagonists with 8-styrylxanthine structure: potential drugs for Parkinson's disease. J Org Chem 2004;69:3308–18. [PubMed: 15132536]
- [34]. Cousins MS, Salamone JD. Involvement of ventrolateral striatal dopamine in movement initiation and execution: a microdialysis and behavioral investigation. Neuroscience 1996;70:849–59. [PubMed: 8848171]
- [35]. Salamone JD, Mahan K, Rogers S. Ventrolateral striatal dopamine depletions impair feeding and food handling in rats. Pharmacol Biochem Behav 1993;44:605–10. [PubMed: 8451265]
- [36]. Salamone JD, Kurth PA, McCullough LD, Sokolowski JD, Cousins JD. The role of brain dopamine in response initiation: effects of haloperidol and regionally specific dopamine depletions on the local rate of instrumental responding. Brain Res 1993;628:218–26. [PubMed: 8313150]
- [37]. Jicha G, Salamone JD. Vacuous jaw movements and feeding deficits in rats with ventrolateral striatal dopamine depletions: possible model of parkinsonian symptoms. J Neurosci 1991;11:3822–9. [PubMed: 1744692]
- [38]. Kelley AE, Gauthier AM, Lang CG. Amphetamine microinjections into distinct striatal subregions cause dissociable effects on motor and ingestive behavior. Behav Brain Res 1989;35:27–39. [PubMed: 2803542]
- [39]. Hornykiewicz O. Dopamine in the basal ganglia. Its role and therapeutic implications (including the clinical use of L-DOPA). Brit Med Bull 1973;29:172–8. [PubMed: 4356552]
- [40]. Bokobza B, Ruberg M, Scatton B, Javoy-Agid F, Agid Y. [3H]Spiperone binding, dopamine and HVA concentrations in Parkinson's disease and supranuclear palsy. Eur J Pharmacol 1984;99:167– 75. [PubMed: 6734727]
- [41]. Madras BK, Gracz LM, Fahey MA, Elmaleh D, Meltzer PC, Liang AY, et al. Altropane, a SPECT or PET imaging probe for dopamine neurons: III. Human dopamine transporter in postmortem normal and Parkinson's diseased brain. Synapse 1998;29:116–27. [PubMed: 9593102]

[42]. Annett LE, Ridley RM, Gamble SJ, Baker HF. Social withdrawal following amphetamine administration to marmosets. Psychopharmacology 1989;99:222–9. [PubMed: 2508158]

- [43]. Bara-Jimenez W, Sherzai A, Dimitrova T, Favit A, Bibbiani F, Gillespie M, et al. Adenosine A(2A) receptor antagonist treatment of Parkinson's disease. Neurology 2003;61:293–6. [PubMed: 12913186]
- [44]. Salamone JD, Mayorga AJ, Trevitt JT, Cousins MS, Conlan A, Nawab A. Tremulous jaw movements in rats: a model of parkinsonian tremor. Prog Neurobiol 1998;56:591–611. [PubMed: 9871939]
- [45]. Finn M, Jassen A, Baskin P, Salamone JD. Tremulous characteristics of vacuous jaw movements induced by pilocarpine and ventrolateral striatal dopamine depletions. Pharmacol Biochem Behav 1997;57:243–9. [PubMed: 9164578]
- [46]. Ishiwari K, Betz A, Weber S, Felsted J, Salamone JD. Validation of the tremulous jaw movement model for assessment of the motor effects of typical and atypical antipychotics: effects of pimozide (Orap) in rats. Pharmacol Biochem Behav 2005;80:351–62. [PubMed: 15680188]
- [47]. Salamone JD, Baskin PB. Vacuous jaw movements induced by acute reserpine and low-dose apomorphine: possible model of parkinsonian tremor. Pharmacol Biochem Behav 1996;53:179–83. [PubMed: 8848448]
- [48]. Betz AJ, McLaughlin PJ, Burgos M, Weber SM, Salamone JD. The muscarinic receptor antagonist tropicamide suppresses tremulous jaw movements in a rodent model of parkinsonian tremor: possible role of M4 receptors. Psychopharmacology 2007;194:347–59. [PubMed: 17594079]
- [49]. Cousins MS, Finn M, Trevitt J, Carriero DL, Conlan A, Salamone JD. The role of ventrolateral striatal acetylcholine in the production of tacrine-induced jaw movements. Pharmacol Biochem Behav 1999;62:439–47. [PubMed: 10080235]
- [50]. Salamone JD, Johnson CJ, McCullough LD, Steinpreis RE. Lateral striatal cholinergic mechanisms involved in oral motor activities in the rat. Psychopharmacology 1990;102:529–34. [PubMed: 2096410]
- [51]. Salamone JD, Carlson BB, Rios C, Lentini E, Correa M, Wisniecki A, et al. Dopamine agonists suppress cholinomimetic-induced tremulous jaw movements in an animal model of Parkinsonism: tremorolytic effects of pergolide, ropinirole and CY 208-243. Behav Brain Res 2005;156:173–9. [PubMed: 15582103]
- [52]. Trevitt JT, Atherton LA, Aberman J, Salamone JD. Effects of subchronic administration of clozapine, thioridazine and haloperidol on tests related to extrapyramidal motor function. Psychopharmacology 1998;137:61–6. [PubMed: 9631957]
- [53]. Betz A, Ishiwari K, Wisniecki A, Huyn N, Salamone JD. Quetiapine (Seroquel) shows a pattern of behavioral effects similar to the atypical antipsychotics clozapine and olanzapine: studies with tremulous jaw movements in rats. Psychopharmacology 2005;179:383–92. [PubMed: 15619122]
- [54]. Simola N, Fenu S, Baraldi PG, Tabrizi MA, Morelli M. Blockade of adenosine A_{2A} receptors antagonizes parkinsonian tremor in the rat tacrine model by an action on specific striatal regions. Exp Neurol 2004;189:182–8. [PubMed: 15296848]
- [55]. Salamone JD, Betz AJ, Ishiwari K, Felsted J, Madson L, Mirante B, et al. Systemic or intrastriatal injections of adenosine A_{2A} antagonists reverse the oral tremor induced by antipsychotic agents: studies with rodent models of drug-induced parkinsonism. Front Biosci 2008;13:3594–605. [PubMed: 18508458]
- [56]. Sultana A, McMonagle T. Pimozide for schizophrenia or related psychoses. Cochrane Database Syst Rev 2000:3. CD001949.
- [57]. Mayorga AJ, Trevitt JT, Conlan A, Ginutsos G, Salamone JD. Striatal and nigral D₁ mechanisms involved in the antiparkinsonian effects of SKF 82958 (APB): studies of tremulous jaw movements in rats. Psychopharmacology 1999;143:72–81. [PubMed: 10227082]
- [58]. Bakshi VP, Kelley AE. Dopaminergic regulation of feeding behavior: I. Differential effects of haloperidol microinjection in three striatal subregions. Psychobiology 1991;19:223–32.
- [59]. Farrar A, Pereira M, Velasco F, Hockemeyer J, Müller CE, Salamone JD. Adenosine A_{2A} receptor antagonism reverses the effects of dopamine receptor antagonism on instrumental output and effortrelated choice in the rat. Implications for studies of psychomotor slowing. Psychopharmacology 2007;191:579–86. [PubMed: 17072593]