## Microbiological Characteristics of Community-Associated *Staphylococcus aureus* Causing Uncomplicated Bacteremia and Infective Endocarditis<sup>7</sup>†

Chia-Yu Chi,<sup>1,2,3</sup> Shih-Min Wang,<sup>4</sup> Chia-Chun Lin,<sup>1</sup> and Ching-Chuan Liu<sup>3</sup>\*

Division of Clinical Research, National Health Research Institutes, Tainan, Taiwan<sup>1</sup>; Graduate Institute of Clinical Medicine, National Cheng Kung University, Tainan, Taiwan<sup>2</sup>; and Departments of Pediatrics<sup>3</sup> and Emergency Medicine,<sup>4</sup> College of Medicine, National Cheng Kung University and Hospital, Tainan, Taiwan

Received 10 September 2009/Returned for modification 22 September 2009/Accepted 9 October 2009

*Staphylococcus aureus* is an important cause of community-associated bacteremia (SAB) and infective endocarditis (IE). No significant differences in distribution or frequency of genes encoding virulence factors, including genes encoding adhesins, were found between isolates from the IE and SAB groups (12 IE and 10 SAB patients).

Staphylococcus aureus bacteremia (SAB) is a serious infection in both adult and pediatric patients (21). One of the most devastating complications of SAB is infective endocarditis (IE), which carries a high risk of morbidity and mortality despite aggressive therapy (12). S. aureus produces many extracellular toxins and virulence factors that elicit immune responses. Exotoxins, such as Panton-Valentine leukocidin (PVL), and exfoliative toxins, toxic shock syndrome toxin 1 (TSST-1), and enterotoxins (SEs) with superantigenic properties cause severe diseases (2, 14). Circulating S. aureus can adhere to the undamaged endothelium either directly through adhesion-receptor interactions or by bridging ligands of plasma and extracellular matrix proteins fibrinogen, fibronectin, and collagen. Attached bacteria may then cause further aggregation of platelets, leading to an enlargement of the infected valvular vegetation (15). S. aureus produces several adhesins, including collagen-, fibrinogen-, and fibronectin-binding proteins. Collagen-binding adhesin (CNA) is important in the development of endocarditis in animal models (11). Clumping factors A and B (ClfA and -B) for fibrinogen- and fibronectin-binding proteins (FnBP) are mediators of S. aureus adherence to platelets (10, 18). Both Clf and FnBP have also been shown to play critical roles in experimental S. aureus IE (19).

This study was designed to determine whether there are differences in the distributions of *S. aureus* genes encoding toxins and virulence factors and antimicrobial resistance patterns in patients with uncomplicated SAB compared to IE.

The population consisted of patients with community-associated *S. aureus* bacteremia who were admitted to the National Cheng Kung Hospital from 2000 to 2007. Demographic and clinical information was obtained from hospital records by using a standardized form. Uncomplicated SAB was defined as the isolation of *S. aureus* from the blood of patients without endocarditis, evidence of hematogenous spread or an overt source of infection. *S. aureus* IE was defined according to the modified Duke criteria (13). Community-associated bacteremia was defined as a positive blood culture for *S. aureus* obtained at the time of or within 48 h of admission. Patients were excluded if they had any established hospital-associated risk factors (9). To focus as much as possible on the influence of genetic microbial factors in the pathogenesis of IE, we excluded patients with underlying heart disease.

Antimicrobial susceptibility was determined by the agar dilution method performed according to Clinical and Laboratory Standards Institute (CLSI) guidelines (17). Erythromycin-resistant strains were further studied for inducible clindamycin resistance.

The synthetic oligonucleotide primers for the PCR amplification of genes encoding the toxin, adherence, and virulence factors were used as described elsewhere (16). PCR for *mecA* and macrolide-lincosamide-streptogramin resistance genes (*ermA*, *ermB*, and *ermC*) was performed using published sequences and temperature conditions (1). The characterization of the staphylococcal cassette chromosome *mec* (SCC*mec*) element was performed according to a published multiplex PCR method (3).

Sequence types (ST) were assigned with reference to the multilocus sequence typing database website (http://www.mlst .net). Pulsed-field gel electrophoresis was performed using the enzyme SmaI. The derived patterns were analyzed using GelCompar software (Applied Maths, Kortrijk, Belgium). Results were analyzed using the unweighted-pair group method for arithmetic averages and Dice coefficient with 1.2% band tolerance (6).

The Mann-Whitney U test was used for continuous variables without normal distribution and the  $\chi^2$  or Fisher exact test for dichotomous variables (SPSS software, version 11.5). A *P* value of <0.05 was considered to be statistically significant.

Twenty-two patients were identified as having either *S. aureus* IE (n = 12) or uncomplicated SAB (n = 10) through blood culture and comprehensive echocardiography examina-

<sup>\*</sup> Corresponding author. Mailing address: Department of Pediatrics, College of Medicine, National Cheng Kung University and Hospital, 138 Sheng-Li Road, Tainan 704, Taiwan. Phone: 886-6-2353535. Fax: 886-6-2753083. E-mail: liucc@mail.ncku.edu.tw.

<sup>†</sup> Supplemental material for this article may be found at http://jcm .asm.org/.

<sup>&</sup>lt;sup>v</sup> Published ahead of print on 21 October 2009.

 TABLE 1. Genes encoding toxins and virulence factors in isolates of community-associated *Staphylococcus aureus* obtained from patients with IE and uncomplicated SAB<sup>a</sup>

Gene	No. (%) of indicated cases with gene sequence		P value
	IE $(n = 12)$	SAB $(n = 10)$	
sea	4 (33)	1 (10)	0.22
seb	3 (25)	6 (60)	0.11
sec	1 (8)	1 (11)	0.71
seg	1 (8)	0	0.55
sei	1 (8)	0	0.55
seg and sei	1 (8)	0	0.55
eta	0	0	
etb	0	0	
pvl	2(17)	2 (20)	0.63
fnbA	12 (100)	9 (90)	0.46
fnbB	2 (17)	1 (10)	0.57
cna	5 (42)	2 (20)	0.27
clfA	10 (83)	8 (80)	0.63
clfB	12 (100)	10 (100)	
tst	1 (8)	0	0.55

<sup>a</sup> The following genes were not identified among any of the staphylococci: *sed*, *see*, *seh*, and *sej* genes.

tion. The mean ages were similar between groups (P > 0.05). Nine patients in the IE group received surgical intervention. The duration of hospital stay was significantly longer for the IE group (36.1 versus 6.8 days; P < 0.001).

The frequencies of carriage of the detected virulence genes, including fnb and clf, did not differ significantly between groups (Table 1). FnBPs and Clf are S. aureus microbial surface components reacting with adherence matrix molecules that are thought to participate in the development of experimental endocarditis (19). However, some reference strains of S. aureus were found to be invasion deficient because of regulatory defects (20). Cheung et al. demonstrated that the inactivated staphylococcal accessory regulator gene (sar) decreased infectivity in the rabbit model of endocarditis because the inactivated mutant has decreased expression of surface FnBPs (4). Therefore the emergence of S. aureus strains with certain adhesion genes is not the only factor responsible for S. aureus IE. The systems involved in the regulation of transcription of the virulence genes may play a more important role in the pathogenesis of S. aureus IE. Exfoliative toxins A and B (ETA and ETB), encoded by eta and etb, respectively, are frequently associated with staphylococcal scalded skin syndrome (SSSS) (5). None of the isolates in this study harbored an *eta* or *etb* gene. This indicates that exfoliative toxins do not play a role in S. aureus bloodstream infections.

Only four (18%) isolates in this study carried *pvl*, and all were methicillin (meticillin)-resistant *S. aureus* (MRSA) strains. Community-acquired MRSA (CA-MRSA) containing *pvl* is associated with severe infections, such as primary skin infections, necrotizing pneumonia, and nonmenstrual staphylococcal toxic shock syndrome (5, 14), but not associated with bacteremia (7, 8). Our finding also supports that *pvl* has no special association with the development of bloodstream infection or its complications, such as IE.

MRSA was isolated from 8/22 (36%) patients. There were no significant differences in antibiotic susceptibility patterns between the two groups (see the supplemental material). Most of the macrolide-resistant isolates were MRSA and possessed the *ermB* gene. One methicillin-susceptible *S. aureus* (MSSA) isolate from the IE group possessed the *ermC* gene and showed inducible clindamycin resistance.

Among eight CA-MRSA isolates, SCCmec type IV was identified in four isolates, and the others were SCCmec type  $V_T$ . On the basis of the interpretable phylogenetic tree, one set of CA-MRSA strains (4/8 [50%]) was clustered, with >80% homology (see the supplemental material). Multilocus ST 59 is the major genetic background of CA-MRSA isolates. This common clone is distinct from other CA-MRSA strains isolated from different countries but is consistent with reports from northern Taiwan (22). They proposed that a single CA-MRSA clone did not spread around the world but that there was simultaneous coevolution of CA-MRSA in different regions.

In conclusion, this study demonstrates that virulence and toxin genes are frequently present in staphylococcal blood isolates. There were no significant differences in frequency of genes encoding such virulence factors studied herein between isolates from patients with IE and those from patients with SAB.

We thank Calvin M. Kunin for providing invaluable suggestions and a critical review of the manuscript. We thank Chao-Ping Liao for collection of clinical isolates.

This study was supported by grants from National Health Research Institutes (NHRI-CL-097-PP08).

## REFERENCES

- Almer, L. S., V. D. Shortridge, A. M. Nilius, J. M. Beyer, N. B. Soni, M. H. Bui, G. G. Stone, and R. K. Flamm. 2002. Antimicrobial susceptibility and molecular characterization of community-acquired methicillin-resistant *Staphylococcus aureus*. Diagn. Microbiol. Infect. Dis. 43:225–232.
- Becker, K., A. W. Friedrich, G. Lubritz, M. Weilert, G. Peters, and E. C. Von. 2003. Prevalence of genes encoding pyrogenic toxin superantigens and exfoliative toxins among strains of *Staphylococcus aureus* isolated from blood and nasal specimens. J. Clin. Microbiol. 41:1434–1439.
- Boye, K., M. D. Bartels, I. S. Andersen, J. A. Moller, and H. Westh. 2007. A new multiplex PCR for easy screening of methicillin-resistant *Staphylococcus aureus* SCCmec types I-V. Clin. Microbiol. Infect. 13:725–727.
- Cheung, A. L., K. J. Eberhardt, E. Chung, M. R. Yeaman, P. M. Sullam, M. Ramos, and A. S. Bayer. 1994. Diminished virulence of a sar-/agr- mutant of Staphylococcus aureus in the rabbit model of endocarditis. J. Clin. Investig. 94:1815–1822.
- Chi, C. Y., S. M. Wang, H. C. Lin, and C. C. Liu. 2006. A clinical and microbiological comparison of *Staphylococcus aureus* toxic shock and scalded skin syndromes in children. Clin. Infect. Dis. 42:181–185.
- Dice, L. R. 1945. Measures of the amount of ecological association between species. Ecology 26:297–302.
- Ellington, M. J., R. Hope, M. Ganner, M. Ganner, C. East, G. Brick, and A. M. Kearns. 2007. Is Panton-Valentine leucocidin associated with the pathogenesis of *Staphylococcus aureus* bacteraemia in the UK? J. Antimicrob. Chemother. 60:402–405.
- Etienne, J. 2005. Panton-Valentine leukocidin: a marker of severity for Staphylococcus aureus infection? Clin. Infect. Dis. 41:591–593.
- Fridkin, S. K., J. C. Hageman, M. Sanza, L. T. Morrison, K. Como-Sabetti, J. A. Jernigan, K. Harriman, L. H. Harrison, R. Lynfield, and M. M. Farley. 2005. Methicillin-resistant *Staphylococcus aureus* disease in three communities. N. Engl. J. Med. 352:1436–1444.
- Grundmeier, M., M. Hussain, P. Becker, C. Heilmann, G. Peters, and B. Sinha. 2004. Truncation of fibronectin-binding proteins in *Staphylococcus aureus* strain Newman leads to deficient adherence and host cell invasion due to loss of the cell wall anchor function. Infect. Immun. 72:7155–7163.
- Hienz, S. A., T. Schennings, A. Heimdahl, and J. I. Flock. 1996. Collagen binding of *Staphylococcus aureus* is a virulence factor in experimental endocarditis. J. Infect. Dis. 174:83–88.
- Hoen, B., F. Alla, C. Selton-Suty, I. Beguinot, A. Bouvet, S. Briancon, J. P. Casalta, N. Danchin, F. Delahaye, J. Etienne, M. Le, V. C. Leport, J. L. Mainardi, R. Ruimy, and F. Vandenesch. 2002. Changing profile of infective endocarditis: results of a 1-year survey in France. JAMA 288:75–81.
- 13. Li, J. S., D. J. Sexton, N. Mick, R. Nettles, V. G. Fowler, Jr., T. Ryan, T.

Bashore, and G. R. Corey. 2000. Proposed modifications to the Duke criteria for the diagnosis of infective endocarditis. Clin. Infect. Dis. 30:633–638.

- Lina, G., Y. Piemont, F. Godail-Gamot, M. Bes, M. O. Peter, V. Gauduchon, F. Vandenesch, and J. Etienne. 1999. Involvement of Panton-Valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. Clin. Infect. Dis. 29:1128–1132.
- Lowy, F. D. 1998. Staphylococcus aureus infections. N. Engl. J. Med. 339: 520–532.
- Martínez-Aguilar, G., A. Valos-Mishaan, K. Hulten, W. Hammerman, E. O. Mason, Jr., and S. L. Kaplan. 2004. Community-acquired, methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* musculoskeletal infections in children. Pediatr. Infect. Dis. J. 23:701–706.
- NCCLS/CLSI. 2009. Performance standards for antimicrobial susceptibility testing: 19th informational supplement. CLSI document M100-S19, 19th ed. Clinical and Laboratory Standards Institute, Wayne, PA.
- O'Brien, L., S. W. Kerrigan, G. Kaw, M. Hogan, J. Penades, D. Litt, D. J. Fitzgerald, T. J. Foster, and D. Cox. 2002. Multiple mechanisms for the activation of human platelet aggregation by *Staphylococcus aureus*: roles for

the clumping factors ClfA and ClfB, the serine-aspartate repeat protein SdrE and protein A. Mol. Microbiol. **44:**1033–1044.

- Que, Y. A., P. Francois, J. A. Haefliger, J. M. Entenza, P. Vaudaux, and P. Moreillon. 2001. Reassessing the role of *Staphylococcus aureus* clumping factor and fibronectin-binding protein by expression in *Lactococcus lactis*. Infect. Immun. 69:6296–6302.
- Sinha, B., P. P. Francois, O. Nusse, M. Foti, O. M. Hartford, P. Vaudaux, T. J. Foster, D. P. Lew, M. Herrmann, and K. H. Krause. 1999. Fibronectinbinding protein acts as *Staphylococcus aureus* invasin via fibronectin bridging to integrin alpha5beta1. Cell. Microbiol. 1:101–117.
- Valente, A. M., R. Jain, M. Scheurer, V. G. Fowler, Jr., G. R. Corey, A. R. Bengur, S. Sanders, and J. S. Li. 2005. Frequency of infective endocarditis among infants and children with *Staphylococcus aureus* bacteremia. Pediatrics 115:e15–e19.
- Wang, C. C., W. T. Lo, M. L. Chu, and L. K. Siu. 2004. Epidemiological typing of community-acquired methicillin-resistant *Staphylococcus aureus* isolates from children in Taiwan. Clin. Infect. Dis. 39:481–487.