Online Submissions: http://www.wjgnet.com/1007-9327office wjg@wjgnet.com doi:10.3748/wjg.v16.i5.583

World J Gastroenterol 2010 February 7; 16(5): 583-587 ISSN 1007-9327 (print) © 2010 Baishideng. All rights reserved.

BRIEF ARTICLE

Crypt abscess-associated microbiota in inflammatory bowel disease and acute self-limited colitis

Harry Sokol, Nadia Vasquez, Nadia Hoyeau-Idrissi, Philippe Seksik, Laurent Beaugerie, Anne Lavergne-Slove, Philippe Pochart, Philippe Marteau

Harry Sokol, Philippe Seksik, Laurent Beaugerie, Gastroenterology and Nutrition Department, Saint Antoine Hospital, Pierre et Marie Curie-Paris6 University, AP-HP, 184 rue du Faubourg Saint-Antoine, 75571, Paris, France

Nadia Vasquez, Philippe Pochart, Philippe Marteau, Laboratoire de Biologie EA 3199, CNAM, 2 rue Conte, 75003, Paris. France

Nadia Hoyeau-Idrissi, Pathology Department, Saint Antoine Hospital, Pierre et Marie Curie-Paris6 University, AP-HP, 184 rue du Faubourg Saint-Antoine, 75571, Paris, France

Anne Lavergne-Slove, Pathology Department, Lariboisière Hospital & Paris 7 University, 2 rue Ambroise-Paré, 75475, Paris, France

Philippe Marteau, Medicosurgical Department of Digestive Diseases, Lariboisière Hospital, AP-HP & Paris 7 University, 2 rue Ambroise-Paré, 75475, Paris, France

Author contributions: Sokol H, Marteau P and Lavergne-Slove A designed the research; Sokol H and Vasquez N performed the research; Sokol H, Vasquez N, Hoyeau-Idrissi N, Seksik P, Beaugerie L, Lavergne-Slove A, Pochart P and Marteau P analyzed the data; Sokol H and Marteau P wrote the paper.

Supported by The Association François Aupetit

Correspondence to: Harry Sokol, MD, PhD, Gastroenterology and Nutrition Department, Saint Antoine Hospital, Pierre et Marie Curie-Paris6 University, AP-HP, 184 rue du Faubourg Saint-Antoine, 75571, Paris, France, 1881 harry@yahoo.fr

Telephone: +33-1-49283162 Fax: +33-1-49283188 Received: July 22, 2009 Revised: August 28, 2009

Accepted: September 4, 2009 Published online: February 7, 2010

Abstract

AIM: To evaluate whether crypt abscesses from inflammatory bowel disease (IBD) patients contain bacteria and to establish their nature.

METHODS: We studied 17 ulcerative colitis patients, 11 Crohn's disease patients, 7 patients with acute self-limited colitis (ASLC) and normal colonic biopsies from 5 subjects who underwent colonoscopy for colon cancer

screening. A fluorescent *in situ* hybridization technique was applied to colonic biopsies to assess the microbiota composition of the crypts and crypt abscesses.

RESULTS: Crypts colonized by bacteria were observed in 42.9% and 3.6% of ASLC and IBD patients, respectively (P = 0.019). Crypt abscesses colonized by bacteria were observed in 28.6% and 0.0% of ASLC and IBD patients, respectively (P = 0.035).

CONCLUSION: These results do not support the hypothesis that crypt abscesses in IBD are the result of localized dysbiosis arising from persistence of living bacteria colonizing the crypts.

© 2010 Baishideng. All rights reserved.

Key words: Inflammatory bowel diseases; Crohn's disease; Ulcerative colitis; Crypt abscess; Microbiota

Peer reviewers: Dr. John K Marshall, Associate Professor of Medicine, Division of Gastroenterology (4W8), McMaster University Medical Centre, 1200 Main Street West, Hamilton, Ontario L8N 3Z5, Canada; Dr. Daniel R Gaya, Gastrointestinal Unit, Molecular Medicine Centre, School of Molecular and Clinical Medicine, University of Edinburgh, Western General Hospital, Crewe Road, Edinburgh EH4 2XU, United Kingdom

Sokol H, Vasquez N, Hoyeau-Idrissi N, Seksik P, Beaugerie L, Lavergne-Slove A, Pochart P, Marteau P. Crypt abscess-associated microbiota in inflammatory bowel disease and acute self-limited colitis. *World J Gastroenterol* 2010; 16(5): 583-587 Available from: URL: http://www.wjgnet.com/1007-9327/full/v16/i5/583.htm DOI: http://dx.doi.org/10.3748/wjg.v16.i5.583

INTRODUCTION

Ulcerative colitis (UC) and Crohn's disease (CD) are the 2 most common types of inflammatory bowel disease



(IBD). Although their pathophysiology is still unknown, the gut microbiota is considered to play a crucial role^[1]. The microbiota close to the mucosa differs from the luminal microbiota^[2]. We and others described a luminal and mucosal dysbiosis in IBD^[3-8] including a high proportion of unusual bacteria^[6,8] and a restricted microbial biodiversity^[7,9]. Nevertheless, no difference in the dominant microbiota was observed between inflamed and non-inflamed mucosa in CD and UC^[5,10].

It would be of great importance to establish the early events leading to IBD onset or to perpetuation of inflammation. Crypt abscesses are early lesions observed in IBD, particularly in UC^[11], consisting of dilated crypts containing polymorphonuclear cells. They can also occur in acute self limited colitis (ASLC)^[12], collagenous and lymphocytic colitis^[13], diverticula-associated colitis^[14] and diversion colitis^[15]. Defensins are antimicrobial peptides secreted in intestinal and colonic crypts and recent studies pointed out a defensin secretion defect in IBD patients^[16-20]. The aim of this study was to evaluate if crypt abscesses in UC, CD and ASLC patients contained bacteria and, if so, to establish their nature i.e. investigate localized dysbiosis in this specific ecosystem.

MATERIALS AND METHODS

Patients

We studied 35 patients with acute colitis and crypt abscesses at histological examination of colon biopsy: 17 UC patients, 11 CD patients and 7 patients with ASLC (bacteria involved: *Shigella sonnei*, *Campylobacter sp.* and no identified pathogenic bacteria in the 5 other cases). We also analyzed normal colonic biopsies from 5 subjects who underwent colonoscopy for colon cancer screening. The characteristics of the patients are described in Table 1. Rectocolonic biopsies containing crypt abscesses were analyzed.

Tissues, histological examination and fluorescent in situ hybridization (FISH)

Histological examination and assessment of the bacterial composition of the crypts and crypt abscess microbiota were performed as previously described using FISH with one general probe (Eubacteria) and 6 group-specific probes, [Bacteroides-Prevotella, γ Proteobacteria, Bifidobacterium, Clostridium coccoides, Faecalibacterium prausnitzii (F. prausnitzii) and Lactobacillus-Enterococcus

Histological examination: Colonic biopsy sections were departaffined in xylene and successively rehydrated for 3 min in 100%, 96%, and 70% ethanol. They were then stained with hematoxylin and eosin for morphological assessment.

FISH: Prior to FISH, sections were deparaffinized, rehydrated, and postfixed in 4% paraformaldehyde for 5 min. Fixation was stopped in phosphate-buffered saline (PBS) 3 × and slides were washed twice for 1 min in

Table 1 Characteristics of the patients

	uc	CD	ASLC
n	17	11	8
Male (%)	47.1	54.5	50.0
Mean age (± SE, yr)	42.0 ± 3.3	44.0 ± 3.9	45.6 ± 10.5
Mean disease duration (± SE, mo)	75 ± 23	50 ± 27	NA
Montreal classification (%)			
E1	0	NA	NA
E2	70.6	NA	NA
E3	29.4	NA	NA
L1	NA	0	NA
L2	NA	63.6	NA
L3	NA	36.4	NA
Analyzed segment (%)			
Rectum	64.7	36.4	71.4
Left colon	35.3	45.4	14.3
Right colon	0.0	18.2	14.3
Treatment (%)			
5-aminosalicylic acid	35.3	18.2	0.0
Corticosteroids	35.3	27.3	0.0
Azathioprine	17.6	18.2	0.0
Infliximab	0.0	18.2	0.0

UC: Ulcerative colitis; CD: Crohn's disease; ASLC: Acute self-limited colitis; NA: Not available.

PBS 1 ×. Tissue sections were incubated 10 min at room temperature with Tris-EDTA buffer containing 10 mg/mL of lysozyme and then washed using the hybridization solution (0.9 mol/L NaCl, 20 mmol/L Tris HCl, pH 8, 0.01% SDS, 30% formamide). Fixed tissue sections were then hybridized with the previous hybridization solution containing 4.5 ng/µL of one of the 5'-end-Cy3-labeled 16S rRNA targeted oligonucleotide probes. Hybridizations were performed at 35°C overnight in a microscope slide incubator and stringent washings were carried out at 37°C (2 × 15 min) in a buffer containing 65 mmol/L NaCl, 20 mmol/L Tris HCl, pH 8.0, 5 mmol/L EDTA, and 0.01% SDS to remove nonspecific binding. The sections were mounted with Vectashield [mounting medium with 4',6'-diamidino-2-phenylindole (DAPI), Vector Laboratories, Burlingame, CA, USA]. DNA was stained with DAPI to visualize all cells.

Detection of crypt abscess-associated-bacteria

Bacteria were visualized with an epifluorescence microscope Leica DMRB using Cy3- and DAPI-specific filters at 100 ×, 400 ×, and 1000 × magnification and images were captured with Leica DFC 300 FX camera and FW 4000 software (Leica microsystemes SAS, Rueil-Malmaison, France). The entire mucosal surface (crypts and crypts abscess) of each colonic biopsy section was examined for the presence of bacteria. Pure cultured bacteria belonging to each group were hybridized as positive and negative controls for the FISH procedure.

Statistical analysis

We first performed a "patient analysis" considering that a patient had colonized crypts or crypt abscesses if at least one of his crypts or crypt abscesses was colonized



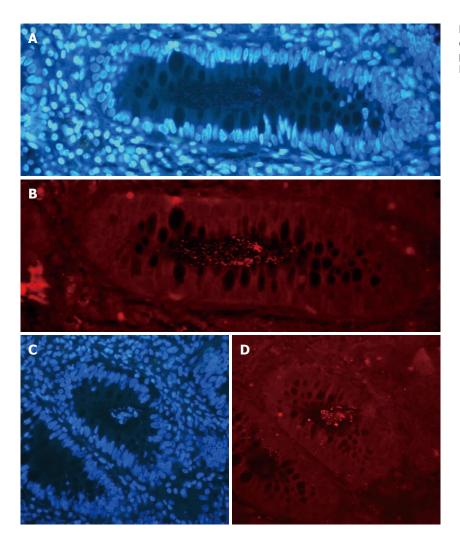


Figure 1 Colonized crypt (A, B) and colonized crypt abscess (C, D). A, C: 4',6'-diamidino-2-phenylindole (DAPI); B, D: Hybridization with the Eubacteria probe.

by bacteria. As colonic biopsies harbored a different number of crypts or crypt abscesses, we also normalized the data by calculating in each patient group the ratio: total number of colonized crypts or crypt abscesses/ total number of crypts or crypt abscesses. The χ^2 test was performed for comparison of qualitative variables.

RESULTS

The FISH technique allowed detection of crypts and crypt abscesses colonized by bacteria as shown in Figure 1. Crypts colonized by bacteria (general probe targeting Eubacteria) were observed in 42.9% and 3.6% of ASLC and IBD patients (only UC patients), respectively (P = 0.019, Figure 2). No colonized crypt was observed in biopsies from CD patients. Crypt abscesses colonized by bacteria were observed in 2 patients with ASLC (28.6% of total ASLC patients) and in no biopsy from the 28 IBD patients (P = 0.035, Figure 2). Neither colonized crypts nor crypt abscesses were observed in colonic biopsies from control patients.

Global and normalized analysis

We observed 2317, 2499 and 2405 crypts in UC, CD and ASLC groups, respectively. None were colonized by bacteria (general probe for Eubacteria) in CD patients

whereas 2 (0.09%) and 46 (1.91%) were colonized in UC and ASLC patients, respectively (Figure 3). We observed 121, 76 and 100 crypt abscesses in UC, CD and ASLC patients, respectively. Among these, 6 were colonized by bacteria in ASLC patients (6.0%) and none in UC and CD patients (Figure 3).

Group/species-specific probe analysis

In order to determine what kind of bacteria was present in colonized crypts and crypt abscesses, we performed FISH analysis using 6 group- or species-specific probes on the sample previously identified to harbor colonized crypts or crypt abscesses. No fluorescent signal was detected in any of the crypt abscesses analyzed with the 6 specific probes. No bacteria-colonizing crypts were recognized by the *F. pransnitzii* or the *Lactobacillus-Enterococcus* probes. The 2 colonized crypts in UC patients contained *Enterobacteria* (detected by the *gamma Proteobacteria* probe) whereas 39.1%, 8.7%, 4.3% and 2.2% of the colonized crypts in ASLC patients contained bacteria from the *Bacteroides (Bacteroides-Prevotella* probe), the *Clostridium coccoides*, the *Enterobacteria* and the *Bifidobacteria* groups, respectively.

DISCUSSION

This study shows for the first time that crypt abscesses



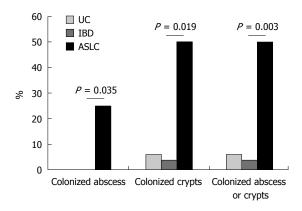


Figure 2 Proportion of patients with at least one colonized crypt abscess, one colonized crypt or at least one crypt abscess or one crypt colonized by bacteria. UC: Ulcerative colitis; IBD: Inflammatory bowel disease; ASLC: Acute self-limited colitis.

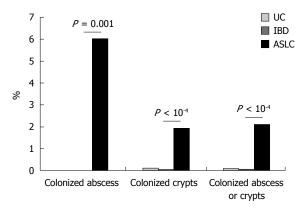


Figure 3 Proportion of crypt abscesses and crypts harboring bacterial colonization.

from IBD patients do not contain bacteria (aseptic abscess); colonic crypts of these patients are less often colonized than those from ASLC patients. These results do not support the hypothesis of the role of localized dysbiosis in the pathogenesis of crypt abscesses.

Dysbiosis is involved in the perpetuation of inflammation in IBD but it is presently not known whether it occurs early in the disease process and whether a localized dysbiosis in a specific ecological niche could trigger early inflammatory events. No previous study searched for bacteria inside crypt abscess in IBD or in other conditions. Swidsinski et al^[22] also looked at colonic crypts and found bacteria inside the crypts more frequently in IBD patients than in control subjects without inflammation or infection. In the current study, we did not find any colonized crypts in healthy subjects, and colonized crypts in IBD patients were infrequent. The differences between these 2 studies could arise from the population studied but also from the method used for the fixation of the colon biopsy. Indeed Swidsinski et al^{22]} used a nonaqueous Carnoy solution which focuses at preserving the mucus layer on the surface epithelium while we used 4% buffered formalin. Nevertheless, in our hands, nonaqueous Carnoy solution and formalin gave similar results (unpublished data).

One can hypothesize that the results may be different

in a cohort of newly diagnosed patients without any antiinflammatory or immunomodulatory therapy. It is not possible to address this question. Nevertheless, the fact that 53.6% of the IBD patients in our cohort received no treatment or only 5-aminosalicyclic acid, and that in 36% the IBD diagnosis was performed in the previous year suggested that the results would be similar in naive patients.

In ASLC, pathogenic bacteria induce an acute and transient breakdown of the intestinal barrier. On the other hand, many pathogenic bacteria involved in ASLC developed a strategy to escape the host immune system [23,24]. Alteration of the intestinal barrier could lead to deep penetration of gut microbiota bacteria (and also pathogenic bacteria) into the crypts where they could proliferate. In the current study, bacteria found in crypts from ASLC patients belonged to various phylogenetic bacterial groups of the normal gut microbiota, suggesting a nonspecific intestinal barrier breakdown in this pathology, allowing the penetration of any bacteria in the crypts.

Our results do not support the hypothesis that crypt abscesses in IBD are the result of persistent localized dysbiosis with a focused reaction against high numbers of living bacteria specifically colonizing the crypts. On the other hand, the absence of entire bacteria in crypt abscesses does not rule out the implication of bacteria in their onset as one may hypothesize that bacteria could stimulate the genesis of crypt abscesses, with recruitment of polymorphonuclear leukocytes which would then destroy them but also contribute to chronic epithelial lesions. The dysbiosis in IBD seems to affect the surface of the whole mucosa (diffuse mucosal dysbiosis) and more studies should be performed to understand its specificity and the ways to influence it.

COMMENTS

Background

Ulcerative colitis (UC) and Crohn's disease (CD) are the 2 most common types of inflammatory bowel disease (IBD). Although their pathophysiology is still unknown, the gut microbiota is considered to play a crucial role. The microbiota close to the mucosa differs from the luminal microbiota. We and others described a luminal and mucosal dysbiosis in IBD. Nevertheless, no difference in the dominant microbiota was observed between inflamed and non-inflamed mucosa in CD and UC.

Research frontiers

It would be of great importance to establish the early events leading to IBD onset or to perpetuation of inflammation. Crypt abscesses are early lesions observed in IBD, particularly in UC, but they can also occur in acute self limited colitis. The aim of this study was to search for a localized dysbiosis in crypt abscesses from UC, CD and acute self-limited colitis patients.

Innovations and breakthroughs

The results do not support the hypothesis that crypt abscesses in IBD are the result of localized dysbiosis arising from persistence of living bacteria colonizing the crypts.

Terminology

Dysbiosis: Breakdown in the balance between putative species of "protective" vs "harmful" intestinal bacteria.

Peer review

This is an interesting and original study, which attempts to characterize dysbiosis associated with cryptitis and crypt abscesses in patients with early IBD. The results suggest that these lesions are in fact sterile.



REFERENCES

- Seksik P, Sokol H, Lepage P, Vasquez N, Manichanh C, Mangin I, Pochart P, Doré J, Marteau P. Review article: the role of bacteria in onset and perpetuation of inflammatory bowel disease. *Aliment Pharmacol Ther* 2006; 24 Suppl 3: 11-18
- 2 Lepage P, Seksik P, Sutren M, de la Cochetière MF, Jian R, Marteau P, Doré J. Biodiversity of the mucosa-associated microbiota is stable along the distal digestive tract in healthy individuals and patients with IBD. *Inflamm Bowel Dis* 2005; 11: 473-480
- 3 Sokol H, Lay C, Seksik P, Tannock GW. Analysis of bacterial bowel communities of IBD patients: what has it revealed? *Inflamm Bowel Dis* 2008; 14: 858-867
- 4 Lepage P, Seksik P, De La Cochetiere MF, Sutren M, Marteau P, Jian R, Dore J. Molecular study of the ileal and colonic mucosa associated flora in IBD. *Gastroenterology* 2003; 124: A477-A478
- 5 Seksik P, Lepage P, de la Cochetière MF, Bourreille A, Sutren M, Galmiche JP, Doré J, Marteau P. Search for localized dysbiosis in Crohn's disease ulcerations by temporal temperature gradient gel electrophoresis of 16S rRNA. J Clin Microbiol 2005; 43: 4654-4658
- 6 Seksik P, Rigottier-Gois L, Gramet G, Sutren M, Pochart P, Marteau P, Jian R, Doré J. Alterations of the dominant faecal bacterial groups in patients with Crohn's disease of the colon. *Gut* 2003; 52: 237-242
- 7 Sokol H, Lepage P, Seksik P, Doré J, Marteau P. Temperature gradient gel electrophoresis of fecal 16S rRNA reveals active Escherichia coli in the microbiota of patients with ulcerative colitis. J Clin Microbiol 2006; 44: 3172-3177
- 8 Sokol H, Seksik P, Rigottier-Gois L, Lay C, Lepage P, Podglajen I, Marteau P, Doré J. Specificities of the fecal microbiota in inflammatory bowel disease. *Inflamm Bowel* Dis 2006; 12: 106-111
- 9 Manichanh C, Rigottier-Gois L, Bonnaud E, Gloux K, Pelletier E, Frangeul L, Nalin R, Jarrin C, Chardon P, Marteau P, Roca J, Dore J. Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. *Gut* 2006; 55: 205-211
- Sokol H, Lepage P, Seksik P, Doré J, Marteau P. Molecular comparison of dominant microbiota associated with injured versus healthy mucosa in ulcerative colitis. *Gut* 2007; 56: 152-154
- 11 **Le Berre N**, Heresbach D, Kerbaol M, Caulet S, Bretagne JF, Chaperon J, Gosselin M, Ramée MP. Histological discrimination of idiopathic inflammatory bowel disease from other types of colitis. *J Clin Pathol* 1995; **48**: 749-753
- 12 Kumar NB, Nostrant TT, Appelman HD. The histopathologic spectrum of acute self-limited colitis (acute infectious-type colitis). Am J Surg Pathol 1982; 6: 523-529
- 13 Ayata G, Ithamukkala S, Sapp H, Shaz BH, Brien TP, Wang

- HH, Antonioli DA, Farraye FA, Odze RD. Prevalence and significance of inflammatory bowel disease-like morphologic features in collagenous and lymphocytic colitis. *Am J Surg Pathol* 2002; **26**: 1414-1423
- 14 West AB, Losada M. The pathology of diverticulosis coli. J Clin Gastroenterol 2004; 38: S11-S16
- Komorowski RA. Histologic spectrum of diversion colitis. Am J Surg Pathol 1990; 14: 548-554
- 16 Wehkamp J, Harder J, Weichenthal M, Mueller O, Herrlinger KR, Fellermann K, Schroeder JM, Stange EF. Inducible and constitutive beta-defensins are differentially expressed in Crohn's disease and ulcerative colitis. *Inflamm Bowel Dis* 2003; 9: 215-223
- 17 Wehkamp J, Harder J, Weichenthal M, Schwab M, Schäffeler E, Schlee M, Herrlinger KR, Stallmach A, Noack F, Fritz P, Schröder JM, Bevins CL, Fellermann K, Stange EF. NOD2 (CARD15) mutations in Crohn's disease are associated with diminished mucosal alpha-defensin expression. *Gut* 2004; 53: 1658-1664
- Wehkamp J, Salzman NH, Porter E, Nuding S, Weichenthal M, Petras RE, Shen B, Schaeffeler E, Schwab M, Linzmeier R, Feathers RW, Chu H, Lima H Jr, Fellermann K, Ganz T, Stange EF, Bevins CL. Reduced Paneth cell alpha-defensins in ileal Crohn's disease. *Proc Natl Acad Sci USA* 2005; 102: 18129-18134
- 19 Wehkamp J, Schmid M, Stange EF. Defensins and other antimicrobial peptides in inflammatory bowel disease. Curr Opin Gastroenterol 2007; 23: 370-378
- 20 Fahlgren A, Hammarstrom S, Danielsson A, Hammarstrom ML. beta-Defensin-3 and -4 in intestinal epithelial cells display increased mRNA expression in ulcerative colitis. Clin Exp Immunol 2004; 137: 379-385
- Vasquez N, Mangin I, Lepage P, Seksik P, Duong JP, Blum S, Schiffrin E, Suau A, Allez M, Vernier G, Tréton X, Doré J, Marteau P, Pochart P. Patchy distribution of mucosal lesions in ileal Crohn's disease is not linked to differences in the dominant mucosa-associated bacteria: a study using fluorescence in situ hybridization and temporal temperature gradient gel electrophoresis. *Inflamm Bowel Dis* 2007; 13: 684-692
- 22 Swidsinski A, Weber J, Loening-Baucke V, Hale LP, Lochs H. Spatial organization and composition of the mucosal flora in patients with inflammatory bowel disease. *J Clin Microbiol* 2005; 43: 3380-3389
- Braat H, McGuirk P, Ten Kate FJ, Huibregtse I, Dunne PJ, Hommes DW, Van Deventer SJ, Mills KH. Prevention of experimental colitis by parenteral administration of a pathogen-derived immunomodulatory molecule. *Gut* 2007; 56: 351-357
- 24 Fallon PG, Alcami A. Pathogen-derived immunomodulatory molecules: future immunotherapeutics? *Trends Immunol* 2006; 27: 470-476

S- Editor Tian L L- Editor Cant MR E- Editor Zheng XM

