# **Selective IgA Deficiency**

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#### Abstract

Introduction Immunoglobulin A (IgA) deficiency is the most common primary immunodeficiency defined as decreased serum level of IgA in the presence of normal levels of other immunoglobulin isotypes. Most individuals with IgA deficiency are asymptomatic and identified coincidentally. However, some patients may present with recurrent infections of the respiratory and gastrointestinal tracts, allergic disorders, and autoimmune manifestations. IgA and Its Functions Although IgA is the most abundant antibody isotype produced in the body, its functions are not clearly understood. Subclass IgA1 in monomeric form is mainly found in the blood circulation, whereas subclass IgA2 in dimeric form is the dominant immunoglobulin in mucosal secretions. Secretory IgA appears to have prime importance in immune exclusion of pathogenic microorganisms and maintenance of intestinal homeostasis. Despite this critical role, there may be some compensatory mechanisms that would prevent disease manifestations in some IgA-deficient individuals.

Pathogenesis In IgA deficiency, a maturation defect in B cells to produce IgA is commonly observed. Alterations in transmembrane activator and calcium modulator and cyclophilin ligand interactor gene appear to act as diseasemodifying mutations in both IgA deficiency and common variable immunodeficiency, two diseases which probably lie in the same spectrum. Certain major histocompatibility complex haplotypes have been associated with susceptibility to IgA deficiency.

Conclusion The genetic basis of IgA deficiency remains to be clarified. Better understanding of the production and function of IgA is essential in elucidating the disease mechanism in IgA deficiency.

**Keywords** IgA · function · immunodeficiency · pathogenesis

#### Introduction

Immunoglobulin (Ig) A deficiency (OMIM 137100) is defined as decreased or absent level of serum IgA in the presence of normal serum levels of IgG and IgM in a patient older than 4 years of age, in whom other causes of hypogammaglobulinemia have been excluded [1, 2]. In general, serum IgA level of less than 7 mg/dL (0.07 g/L) is considered as selective IgA deficiency since this concentration is the lowest detectable limit established by most of the laboratories. When serum IgA level is higher than 7 mg/dL but two standard deviations below normal for age, the condition may be referred to as partial IgA deficiency, which is quite common. The threshold of 4 years of age is used to avoid premature diagnosis of IgA deficiency which may be transient in younger children due to delayed ontogeny of IgA system after birth.

In here, selective IgA deficiency, with an emphasis on functions of IgA and disease pathogenesis, along with its epidemiology, clinical manifestations, disease associations, treatment approaches, and prognosis will be reviewed.

## Immunoglobulin A and Its Functions

First described in serum in 1953, IgA is the most abundant antibody isotype produced in the body [3-7]. It is the second dominant isotype in the blood circulation following IgG. It can be found in both monomeric and polymeric forms.

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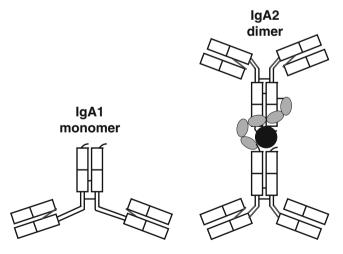
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Circulating IgA is in monomeric form, whereas secretory IgA, in the mucosal secretions of respiratory, intestinal, and genitourinary systems, is dimeric [4, 6, 7]. The monomeric structure of serum IgA has two heavy chains, each consisting of one variable and three constant regions, and two light chains, each of which is made up of one variable and one constant region. In humans, there are two subclasses of IgA: IgA1 and IgA2, constant heavy chains of which are encoded by two separate  $\alpha1$  and  $\alpha2$  genes on chromosome  $\alpha14$  [5–7]. The main structural difference between them is that IgA2 has a shorter hinge region which may render this isotype more resistant to bacterial proteases in the lumen of gastrointestinal or respiratory systems [8].

The function of serum IgA in the systemic immune response has not been clearly understood. Monomeric IgA in the circulation does not fix the classical pathway of the complement. However, it may have a role in activation of phagocytic system by means of the FcR $\alpha$  receptors [5, 7, 9]. It has been proposed that serum IgA binds to FcR $\alpha$  receptor on the monocytes and granulocytes; thereby, immune complexes formed by foreign antigens and IgA are cleared from the circulation by the phagocytic system without activating the complement system and without causing inflammation [10, 11]. Serum IgA may also have a role in controlling the immune system through inhibition of neutrophil chemotaxis by binding to other inhibitory proteins such as  $\alpha$ -1-antitrypsin and forming complexes [5, 7].

IgA, which is mostly in dimeric form, is the dominant immunoglobulin in luminal secretions comprising more than two thirds of total IgA production in the body [8, 12, 13]. Being more resistant to proteolytic activity of the bacteria, IgA2 is the main IgA subclass found in secretions, although



**Fig. 1** Models of a monomeric IgA (*left*) and a dimeric secretory IgA (*right*). The monomer contains IgA1 subclass, whereas the dimer comprises two IgGA2 subclass monomers. The *black circle* denotes the joining (J) chain. The *gray chain of five oval links* indicates the secretory piece. Note the shorter hinge region in IgA2 [8]. Modified from [22]

both IgA subclasses can form dimers by covalent interaction with a joining (J) chain attached to the terminal constant region on the Fc portion. The secretory IgA dimer contains a secretory component which is actually the secreted component of the polymeric immunoglobulin receptor located on the basolateral surface of the mucosal epithelial cell [13, 14] (Fig. 1). More than 95% of secretory IgA is produced locally. In the gastrointestinal system, organized Payer's patches or isolated lymphoid follicles as well as nonorganized lamina propria can be sites for local IgA production by T cell-dependent as well as T cell-independent mechanisms [15-17]. Intestinal epithelial cells, dendritic cells, and local stromal cells may be contributing to T cellindependent production of IgA locally by secreting thymic stromal lymphopoietin, interleukin (IL)-6, interleukin-10, tumor necrosis factor-alpha, transforming growth factor beta (TGF-β1), B-cell activating factor (BAFF), and a proliferation-inducing ligand (APRIL) [16].

Mucosal membranes in the body cover an approximate area of 200-400 m<sup>2</sup> harboring an estimate of 15,000-36,000 species and 1,800 genera of microbiota [18-21]. Thus, the total number of prokaryotic cells exceeds the total number of eukaryotic cells in the body. Bacteria endogenous to the intestinal tract, oral cavity, and respiratory and genital tracts are coated with secretory IgA. As a result, the epithelial adherence and penetration of bacteria are limited, and the bacteria are confined to the mucosal surfaces [20]. The IgA coating of bacteria is traditionally considered to be through adaptive immunity by Fab-mediated antigen-specific binding. Recently, it has been proposed that there is a link between the specific antibody-dependent protection and the innate glycan-mediated mucosal immunity by means of N- and O-glycans of secretory IgA. It is likely that glycan-mediated interactions in concert with Fab-mediated polyreactivity enforce protective functions of secretory IgA [19, 20, 22, 23]. Despite this critical role of secretory IgA, surprisingly, some individuals with selective IgA deficiency are asymptomatic. The diagnosis of IgA deficiency depends on the measurement of IgA concentration in serum. Secretory IgA level is not determined; therefore, it is possible that the individuals diagnosed with selective IgA deficiency may still have some IgA in the mucosal systems enough to provide some protective functions. In addition, in most IgA-deficient patients, seemingly as a compensatory mechanism, production of secretory IgM is increased [24, 25]. IgA and IgM have evolutionary, structural, and functional similarities: homologies between the primary constant chain domains and the tail pieces, presence of a J chain in both, formation of polymers, ability to bind the basolateral polymeric Ig receptor on mucosal epithelial cell, and thereby forming secretory immunoglobulin molecule that contains the epithelial secretory component. The glycan moieties are



also similar in IgA and IgM molecules [26–28]. However, there are also some IgA-deficient patients who do not exhibit compensatory increase in secretory IgM [29].

Most of the commensal bacteria are located in the gastrointestinal tract [15, 16]. Therefore, maintenance of intestinal homeostasis is of prime importance. This homeostasis is achieved by means of immune defense mechanisms, in which secretory IgA has a central role. Growing body of evidence suggests that protective role of secretory IgA in the gastrointestinal system is not only through immune exclusion of bacteria [16–20]. In mice, it has been shown that secretory IgA is also critical in regulating bacterial communities of the intestinal lumen. Excessive expansion of anaerobic bacteria in the entire proximal intestinal system due to lack of secretory IgA has been reported in activation-induced cytidine deaminase knockout mice [30]. Similar aberrant anaerobic expansion is also observed in other mice with IgA deficiency, e.g., RAG2 -/-, SCID mice [31, 32]. This type of change in the intestinal microbial ecology may cause activation of mucosal immune cells including intraepithelial lymphocytes, cells of the isolated lymphoid follicles, Peyer's patches, and mesenteric lymph nodes. In addition, this activation status may become systemic and involve lymphocytes of all germinal centers and lymphoid tissues. Secretory IgA may also play a role in creating a noninflammatory host-microbial relationship as a consequence of its inability to fix the complement and the lack of proinflammatory IgA receptors on the intestinal macrophages.

### Pathogenesis of IgA Deficiency

In IgA-deficient patients, the common finding is a maturation defect in B cells to produce IgA [6, 33]. The defect appears to involve the stem cells since IgA deficiency can be transferred by bone marrow transplantation [34]. The constant  $\alpha 1$  and  $\alpha 2$  genes are generally normal except for rarely described cases of heavy chain gene deletions involving various segments on chromosome 14 [35]. This may be the case in sporadic IgA-deficient patients who have associated IgG2, IgG4, and IgE deficiencies [36-38]. In IgA deficiency, B cells express IgA; however, they are of immature phenotype with the coexpression of IgM and IgD, and they cannot fully develop into IgA-secreting plasma cells [39, 40]. An intrinsic B cell defect, T helper cell dysfunction, and suppressor T cells have all been reported in IgA deficiency. Abnormalities in the cytokine network such as lack of IL-4, IL-6, IL-7, IL-10, TGF-β, and most recently IL-21 have also been proposed to play a role in IgA deficiency [6, 41-43]. It is interesting that IL-21 stimulation has been shown to induce class switch recombination to IgG and IgA and differentiation of IgA and IG secreting plasma cells with restoration of immunoglobulin

production ex vivo in patients with IgA deficiency and common variable immunodeficiency (CVID) [43].

There is a not well-defined genetic susceptibility in IgA deficiency. The pedigrees of IgA-deficient individuals show familial clustering with no distinct Mendelian inheritance pattern. Autosomal recessive, autosomal dominant, and sporadic transmission patterns have all been observed [44]. In view of the variation in the inheritance patterns and the lack of an identified primary genetic defect, it is likely that IgA deficiency represents a heterogeneous group of genetic abnormalities such as CVID. Mutations in transmembrane activator and calcium-modulator and cyclophilin ligand interactor (TACI, TNFRSF13B) have been found both in a subset of patients with IgA deficiency or CVID [45]. TACI, B-cell surface ligand for BAFF and APRIL, has a role in isotype switching in B cells. The same TACI mutation may be present in individuals with either IgA deficiency or CVID in the same family. However, it is controversial whether TACI mutations have a cause-effect relationship with IgA deficiency or CVID [46, 47]. Recently, a shared cytotoxic T lymphocyte-associated protein-4-inducible costimulator risk locus in IgA deficiency and CVID has been defined [48]. It is also known that IgA deficiency may progress to CVID, supporting the notion that IgA deficiency and CVID lie in the spectrum of the same disease [49, 50].

Associations between IgA deficiency and certain major histocompatibility complex (MHC) class I, II, and III haplotypes have been proposed [6, 51-54]. In IgA deficiency and type 1 diabetes mellitus, HLA-B8 frequency was found to be increased in earlier studies. HLA-B8 frequency was also higher in IgA deficiency and autoimmune disorders. However, these findings may be secondary to the presence of diabetes mellitus or autoimmune disorders rather than IgA deficiency itself. In another study in IgA-deficient individuals, HLA B8 allele was not related to history of autoimmunity; however, IgA-deficient patients who had HLA B8 were found to have significantly higher pneumococcal vaccination responses [55]. A recent study questioning the frequently implied higher risk for IgA deficiency with the HLA8 DR3 haplotype has shown that IgA deficiency is not associated with a distinct haplotype; rather, the risk is conferred by the common extended MHC haplotype HLA A1, B8, DR3, and DQ2 (the 8.1 haplotype) acting in a multiplicative manner [56]. An amino acid substitution at position 57 of the HLA-DQ beta chain gene has also been associated with susceptibility to IgA deficiency [57].

#### **Epidemiology**

Selective IgA deficiency is considered as the most common primary immunodeficiency. The worldwide incidence



varies depending on the ethnic background: 1:143 in the Arabian peninsula [58], 1:163 in Spain [59], 1:252 in Nigeria [60], 1:875 in England [61], and 1:965 in Brazil [62]. The incidence is lower among Asian populations, e.g., from 1:2,600 to 1:5,300 in China [63] and from 1:14,840 to 1:18,500 in Japan [64]. In general, IgA deficiency is more common in Caucasians. In the USA, the frequency is estimated to be from 1:223 to 1:1,000 in community studies and from 1:333 to 1:3,000 among healthy blood donors [6]. These numbers may in fact be higher because some individuals with IgA deficiency are asymptomatic, and there is no established routine screening program for IgA deficiency. The variation in incidence may also arise from the fact that the definition of selective IgA deficiency may differ in each study or registry.

#### **Clinical Manifestations**

There is a wide spectrum of clinical findings in IgA deficiency. Patients with IgA deficiency may be identified among blood bank donors, without any clinical findings [6]. In fact, 85–90% of IgA-deficient individuals are asymptomatic. This high percentage is interesting and still remains a puzzle to be solved since IgA is such a significant immunoglobulin in immune defense. Some patients with IgA deficiency have a tendency to develop recurrent sinopulmonary infections, gastrointestinal infections and disorders, allergies, autoimmune conditions, and malignancies.

Recurrent Sinopulmonary Infections Infections of the respiratory system are the most common findings in individuals with IgA deficiency [6, 55, 65]. These infections are mostly due to bacteria, e.g., Haemophilus influenzae and Streptococcus pneumoniae. Some patients may develop end organ damage such as bronchiectasis secondary to recurring or chronic infections [66]. Patients with associated antibody deficiency such as IgG2 subclass deficiency have a higher chance of having more severe infections and complications.

Gastrointestinal Infections/Disorders IgA-deficient individuals have a tendency to develop infections and disorders of the gastrointestinal tract [6, 44, 55]. Giardiasis, malabsorption, lactose intolerance, celiac disease, ulcerative colitis, nodular lymphoid hyperplasia, and malign proliferation are among the associated diseases. Since the protective barrier of the gastrointestinal system is impaired in IgA deficiency, protozoa such as Giardia lamblia can adhere to the epithelium, proliferate, and cause infection [67]. Malabsorption may ensue secondary to structural damage to the intestinal villi. Even in the absence of infection, some molecules may enter the subepidermal and submucosal tissue because of the

impaired mucosal clearance of macromolecules and proteins. This process may facilitate antibody production against certain antigens and intolerance to certain foods [68]. For instance, patients with IgA deficiency have a higher chance of developing celiac disease [69]. Patients with IgA deficiency are not expected to develop IgA isotype antibodies against gliadin, tissue transglutaminase, or endomysium; however, they may have IgG isotype antibodies against those antigens. Inflammatory bowel diseases, mostly ulcerative colitis, have also been reported in association with selective IgA deficiency [6, 55, 70, 71].

Allergic Disorders Allergic disorders appear to be common in patients with IgA deficiency [6, 55, 65]. The reported frequency of allergies varies according to the definitions of both IgA deficiency and allergy and also by the evaluation methods. In an earlier study, atopy was reported in 58% of pediatric and adult patients with IgA deficiency [72]. However, in a later study that surveyed 127 IgA-deficient patients between ages of 2 and 67 years, 13% of patients, a figure which was probably not higher than in the general population, was noted to have a history of allergy and asthma [55]. History of allergy was more common among younger patients (median age 10.5 years). In 126 Brazilian children and adolescents with IgA deficiency, 48% had respiratory allergies and atopic dermatitis [70]. In a recent prospective Swedish study in children, IgA-deficient patients were found to have an increased risk of pseudocroup at year 1 and parentally reported food hypersensitivity at year 4, both of which were possibly not IgE-mediated, as compared to children with normal serum levels of IgA [65]. In a more recent report, in which allergy status was determined more reliably by clinical presentation and skin prick testing using 14 common standard allergens, allergic manifestations including asthma, atopic dermatitis, allergic rhinitis/ conjunctivitis, urticaria, drug allergy, and food allergy were noted in 84% of patients (age range 4-32 years) with selective IgA deficiency [71]. In 40.5% of the patients, allergic manifestations were the presenting symptoms. It is thought that 25% of patients with IgA deficiency are identified during evaluation for allergic disorders [6].

Autoimmunity Autoimmune diseases are among the most important clinical manifestations in IgA deficiency [6, 55, 70]. Autoantibodies, such as antibodies against sulfatide, Jo-1, cardiolipin, phosphatidylserine, and collagen can be detected in IgA-deficient patients even if overt clinical disease is not the case [73]. It has been long known that several autoimmune disorders may occur in association with IgA deficiency. In a 2004 study, the second most common association with IgA deficiency after recurrent infections was autoimmunity (28%) [55]. Autoimmunity was more prevalent in adults (median age 29 years) and in



females (24 autoimmune conditions in females versus 14 in males in a total of 34 subjects). The most common autoimmune condition was idiopathic thrombocytopenic purpura followed by hemolytic anemia, juvenile rheumatoid arthritis, thyroiditis, systemic lupus erythematosus, and presence of various autoantibodies. In a younger population, autoimmune disorders, i.e., thyroid disease, arthropathy, celiac disease, anemia, and systemic lupus erythematosus, were detected in 19% of patients. This figure varies from 20% to 30% based on the age range of studied populations [71, 74]. In addition, autoimmune conditions were found to be higher among relatives of IgA-deficient patients. Of the first-degree relatives of IgA-deficient patients, 10% had autoimmunity compared to an estimate of 5% in the general population.

IgA-deficient patients may develop anti-IgA antibodies which have a potential to cause anaphylactic reactions upon transfusion of any blood product, such as red blood cells or platelets, which contains trace amounts of IgA [75, 76]. The anti-IgA antibody capable of causing a type I hypersensitivity reaction would be of IgE isotype. Therefore, testing for the IgE isotype antibodies against IgA would be meaningful in determining the possibility of a blood transfusion reaction in an IgA-deficient patient. However, this testing is not readily available in most of the laboratories. Instead, IgG anti-IgA antibodies may be measured as a screening tool.

*Malignancy* The association of IgA deficiency and malignancies have been reported in sporadic cases, particularly at older ages. Those are usually of lymphoid and gastrointestinal origins [55, 77].

#### **Laboratory Evaluation**

IgA deficiency should be a consideration in a patient with recurrent respiratory and gastrointestinal infections, allergies, and autoimmune disorders [1, 2]. Immunologic evaluation for IgA deficiency is also warranted in case of anaphylaxis secondary to a blood product transfusion, celiac disease, and a family history of IgA deficiency and/ or CVID. It would be prudent to take into account the medications that may cause decreased serum levels of IgA in the patient. Evaluation of a suspected IgA deficiency would generally include a complete blood count with differential, quantitative serum immunoglobulin levels, serum IgG subclasses, specific antibody response to protein and polysaccharide antigens, and lymphocyte subsets. In addition, pertinent laboratory testing for the associated conditions, e.g., recurrent infections, allergies, or celiac disease, should be performed. Celiac disease screening should include IgG isotype antibodies against gliadin and tissue transglutaminase since IgA isotype antibodies may not be detected because of the IgA deficiency.

#### Management

IgA-deficient patients who are diagnosed coincidentally and/ or who do not have any symptoms do not need any treatment. However, awareness and education are of prime importance, particularly to prevent a potential anaphylactic reaction secondary to blood transfusion. In this regard, patients with selective IgA deficiency should be recommended to wear a medical alert bracelet. In case of a blood transfusion requirement, the patient, ideally, should be screened for anti-IgA antibodies. The blood product should be prepared from an IgA-deficient individual, or saline-washed red blood cells should be the choice. All blood products should be given with caution, and the staff should be prepared to treat a potential anaphylactic reaction.

In IgA deficiency, the mainstay of treatment is the treatment of associated diseases. If the patient experiences recurrent infections, daily prophylactic antibiotics on a continuous or seasonal intermittent basis may be beneficial. In case of associated IgG subclass deficiency and/or specific antibody deficiency, immunoglobulin treatment via venous or subcutaneous route with a product that contains minimal IgA may be given. Standard treatment approach is entertained in case of an associated allergic disorder or autoimmune condition.

## **Prognosis**

The prognosis is good in patients with IgA deficiency if it is not associated with a significant disease. IgA deficiency in children may resolve over time. However, it is also known that IgA deficiency may progress into CVID, which has a less favorable outcome. Therefore, a patient with IgA deficiency, once identified, would deserve a regular follow-up of clinical and immunological findings.

In summary, although IgA deficiency is the most common primary immunodeficiency recognized for almost a half century, its genetic basis remains to be defined, and the cellular and molecular mechanisms involved in IgA physiology and functions as well as in disease pathogenesis need to be further elucidated. Molecular mechanisms of physiological and pathological actions are discussed in the accompanying review by Dr. Renato Monteiro.

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