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A New Model of Cortical Stroke in the Rhesus Macaque

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Abstract

Primate models are essential tools for translational research in stroke, but are reportedly inconsistent in their ability to produce cortical infarcts of reproducible size. Here we report a new stroke model using a transorbital, reversible, two-vessel occlusion approach in male rhesus macaques that produces consistent and reproducible cortical infarcts. The right middle cerebral artery (MCA) (distal to the orbitofrontal branch), and both anterior cerebral arteries (ACA) were occluded with vascular clips. Bilateral occlusion of the ACA was critical for reducing collateral flow to the ipsilateral cortex. Reversible ischemia were induced for 45, 60, or 90 minutes (n=2/timepoint) and infarct volume and neurological outcome were evaluated. The infarcts were located predominantly in the cortex and increased in size with extended duration of ischemia determined by T_2 -weighted MRI. Infarct volume measured by 2,3,5-triphenyl tetrazolium chloride (TTC) and cresyl violet staining corroborated MRI results. Neurological deficit scores worsened gradually with longer occlusion times. A subset of animals (n=5) underwent 60 minutes of ischemia resulting in consistent infarct volumes primarily located to the cortex which correlated well with neurological deficit scores. This approach offers promise for evaluating therapeutic interventions in stroke.

Keywords

Cerebral ischemia; MRI; Neurological deficit; Non-human primate; Rhesus macaque; Stroke

INTRODUCTION

Animal modeling is essential for the preclinical evaluation of neuroprotective drugs for the treatment of stroke. Rodent studies have provided substantial understanding of the pathophysiology of stroke and efficacy of interventions to protect the brain against ischemic

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injury (Gidday *et al* 1999; Stenzel-Poore *et al* 2007; Tamura *et al* 1990). Additionally, the relative ease and efficiency of rodent models and their availability as inbred and genetically modified strains, have lead to comprehensive and successful stroke intervention studies, using an expanding array of molecular and pharmacologic agents (Huang *et al* 1994; Liu *et al* 2002). However, recent efforts to translate pre-clinical neuroprotective strategies from rodents to humans have been disappointing (Emerich 2000; Savitz 2007). There are numerous examples where promising neuroprotectants developed in rodent models have failed to reduce infarct size or improve functional outcome in humans. Moreover, some therapeutic trials in humans have actually lead to increased mortality (Davis *et al* 2000). Such discordance between rodent and human responses to these therapeutic approaches may be due to species-specific differences in response to brain ischemia (DeGraba and Pettigrew 2000; Grotta 1995). Such considerations have lead to the creation of STAIR guidelines (Stroke Therapy Academic Industry Round Table (Fisher M. Chair) 1999; Stroke Therapy Academic Industry Round Table (Fisher M. Chair) 1999; Stroke Therapy Academic Industry Round Table (Fisher M. Chair) 2005) and recent recommendations for preclinical testing of neuroprotective agents in non-human primates (Feuerstein *et al* 2008).

Non-human primate (NHP) models offer significant advantages for preclinical testing of candidate neuroprotective agents. Old-world monkeys such as macaques and baboons, have a close phylogenetic relationship to humans, which increases the likelihood that these animals share similar endogenous mechanisms of ischemic injury and neuroprotection (Schmitz *et al* 2005). Anatomically, most primate brains are much larger than rodent brains and possess gyrencephalic morphology and a gray/white matter ratio that is similar to that of humans. Critically, the anatomy of the cerebral vasculature in monkeys is analogous to humans, (Kapoor *et al* 2003) and thresholds for ischemia and infarction between monkeys and humans are very similar (Jones *et al* 1981). The complex behavior of these animals also allows for a better assessment of functional outcome of post-stroke recovery (Furuichi *et al* 2007; Mack *et al* 2003), which permits correlations of neurological function with infarct size and location (Mack *et al* 2003). Therefore, primate studies potentially offer an important translational bridge from rodents to humans by providing a pragmatic model in which to optimize therapeutic drugs and their range of effectiveness while minimizing toxicity.

Primate models of stroke are quite varied, but frequently involve some aspect of occlusion (aneurysm clips, thrombus, embolization, etc.) of the middle cerebral artery (MCAO). To access this vessel, craniotomy has been used (Hirouchi et al 2007), but this route is challenging, due to the very thick overlying temporalis muscle and also the required brain retraction, which may result in mechanical injury to the brain. Hence, a transorbital approach to access the proximal intra-cerebral vessels (Spetzler et al 1980) was employed in the current study. The results of MCAO, while promising (D'Arceuil et al 2006; Furuichi et al 2003; Kito et al 2001; Maeda et al 2005; Spetzler et al 1980; Young et al 1997), often produce infarcts that vary in location and size. Moreover, a long duration of ischemia is necessary to cause damage beyond the basal ganglia. Additional extended time of ischemia required for cortical involvement produces a mixture of cortical and subcortical stroke and can be associated with low survival. The resiliency of the monkey ipsilateral cortex to MCAO appears to be due to collateral flow to the region from the anterior cerebral arteries, as occlusion of both ACAs and the internal carotid artery (ICA) in the baboon does induce cortical damage. However, in the ACA/ICA baboon model the basal ganglia is also involved and heavily damaged; these animals frequently require extended post-operative care and may have significant morbidity and mortality (Huang et al 2000; Mack et al 2003). Moreover, reliance on occlusion of the ICA at the level of the anterior choroidal artery presents a technical challenge, due to lack of visualization of this artery and the proximity of the posterior communicating artery (PCA) which could result in persistent collateral flow through the PCA.

Thus, current primate stroke models have not reliably produced large infarcts restricted to the cortex. Availability of such a model would mimic a subclass of thrombotic stroke in humans and offer a relatively homogenous therapeutic target for studies of neuroprotection. A model of cortical stroke would help address criticisms of the overly broad inclusion criteria in recent failed stroke trials, as stroke patients with either cortical or subcortical infarcts entered into these trials may have caused mixed results. This is illustrated in studies using the immunosuppressive agent FK506 in the NHP, which demonstrated a significant reduction in infarct volume from MCAO, solely in the cortex and not in the basal ganglia (Furuichi *et al* 2003; Furuichi *et al* 2007). With long durations (hours) of ischemia, the cortical lesions often follow initial subcortical damage which progresses over time, creating secondary effects such as edema that can complicate interpretation of the effects of interventions.

The goals of this study are to create a reproducible cortical stroke model in the rhesus macaque and implement tools to evaluate damage as well as functional neurological outcomes. Desirable features of this model are good survivability to evaluate therapeutic interventions and minimal post-operative care. Using a transorbital approach, reversible occlusion of the distal right middle cerebral and both anterior cerebral arteries produced reproducible infarcts that are located primarily in the cortex and which can serve as a relevant final screen for neuroprotective efficacy in the advancement of promising therapeutics for human clinical trials.

MATERIALS AND METHODS

Animals

Eleven adult, male rhesus macaques (*Macaca mulatta*), with an average age of 8.8 ± 0.6 years and an average body weight of 9.3 ± 0.5 kg, were selected for this study. Animals were singlyhoused indoors on a 12h:12h light/dark cycle, with lights-on from 0700 to 1900 hr, and at a constant temperature of 24° C. Lab diet was provided twice daily, supplemented with fresh fruit and vegetables and drinking water *ad libitum*. Two weeks prior to surgery animals were screened for general health, endemic diseases and neurological disorders, and blood was drawn for baseline evaluation of peripheral blood cell counts. All animal experiments were subject to approval and surveillance by the Institutional Animal Care and Use Committee at the Oregon National Primate Research Center (ONPRC).

Experimental Design

Two experiments were conducted. An *Ischemia Duration Study* was performed to establish the optimal duration of ischemia necessary to generate a unilateral cortical infarct. An *Infarct Reproducibility Study* was performed to examine the reproducibility of stroke size and location after a specified duration of occlusion. The *Ischemia Duration Study* employed 45, 60, and 90 minute (n=2/group) occlusions and survival times of seven days for the 45 and 60 minute groups and two days for the 90 minute group. The *Reproducibility Study* involved 5 animals given 60 minutes of ischemia followed by a survival time of two days. Outcome measures (described below) for evaluating infarct size and neurological changes were performed throughout the prescribed experimental recovery periods.

Two-vessel Occlusion Protocol

Surgical procedures were conducted by a single surgeon to minimize variation. Anesthesia was induced with Ketamine (10 mg/kg, intramuscular injection). Animals were then intubated and maintained under general anesthesia using 0.8-1.3% isoflurane vaporized in 100% oxygen. A blood sample was taken for a complete blood count and a venous line was placed for fluid replacement. An arterial line was established for blood pressure monitoring throughout surgery to maintain a mean arterial blood pressure of 60-80 mm Hg. End-tidal CO₂ and arterial blood

gases were continuously monitored to titrate ventilation to achieve a goal of $PaCO_2$ of 35-40 mm Hg.

Animals were positioned supine without a head holding device. The right orbital area was clipped free of hair and cleaned with betadine. The medial and lateral canthi were incised as well as the periorbitum underneath the evelids. The eye was exonerated and the optic nerve and its vessels were severed at the optic canal. The orbital roof was then drilled using a high powered pneumatic drill and bone removed from the sphenoid wing, clinoid process, and medially to the planum sphenoidale. Using microdissection techniques under an operating stereomicroscope (Leica, Bannockburn, IL, USA) the dura was opened in a U-shaped fashion with the base against the sphenoid wing and clinoid process. Removal of the medial border of the orbital roof and planum sphenoidale allowed direct access to the internal carotid artery (ICA), ACA, and MCA permitting reversible occlusion with aneurysm clips of the distal MCA and both ACAs (Figure 1). The MCA could be readily identified just anterior to the sphenoid wing. A titanium aneurysm mini-clip (Aesculap Inc., Center Valley, PA, USA) was placed on the MCA distal to the orbitofrontal artery, and a second aneurysm mini-clip was placed to occlude both ACAs proximal to the formation of a single pericallosal artery (Figure 1). Following the occlusion period, mini-clips were removed, the cavity was irrigated, and periorbital fascia was placed over the dural opening to protect the brain surface. A large flat piece of bone wax was used to cover the bony opening and the eyelid was sutured closed.

Magnetic Resonance Imaging (MRI)

All scans were performed on a Siemen's 3T Trio system, housed near the surgical suite at ONPRC. Because of the small filling capacity of the rhesus macaque head, an extremity (EX) coil was employed to achieve better image quality of the brain. For animals not already anesthetized, anesthesia was induced initially with Ketamine (10 mg/kg, intramuscular injection). The animals were then intubated and administered 1% isofluorane vaporized in 100% oxygen was administered for anesthesia maintenance. Animals were scanned in the supine position and in most instances, were scanned prior to surgery (baseline), immediately after surgery (acute) and two days post-surgery. In the *Ischemia Duration Study*, four animals had a longer, (seven day) period of recovery and were subjected to MR scans during ischemia and also on days two and seven of recovery. Animals were monitored for physiological signs during scans, including pulse-oximetry, end-tidal CO₂ and respiration rate.

All animals received anatomical MRI scans, which included T_1 and T_2 -weighted, high resolution scans, and a diffusion-weighted scan. The T_1 scan was a MPRAGE protocol, with TR=2500 msec, TE=4.38 msec, number of averages=1 and the flip angle=12°. Full brain coverage was attained at a resolution of 0.5 mm isovoxel. The T_2 scan was a turbo spin-echo experiment, with TR=5280 msec, TE=57 msec, number of averages=4, an echo train length of 5, and a refocusing pulse flip angle of 120°. The entire brain was imaged with a 0.5 × 0.5 mm in-plane resolution and a slice thickness of 1 mm. For measurements of the directionallyaveraged water diffusion coefficient (the diffusion tensor trace/3), a turbo STEAM-based pulse sequence (Nolte *et al* 2000) was employed. With this strategy, image distortions typically encountered with standard echo-planar-imaging-based diffusion measurements within the nonhuman primate brain were avoided. The pulse sequence parameters were, TR=22000 msec, TE=64 msec, b=900 s/mm². Full brain coverage was achieved with an in-plane resolution of 1.8×1.8 mm and a slice thickness of 2 mm.

Four of the animals from the *Ischemia Duration Study* which examined the effects of a variable duration of ischemia (45 and 60 minute groups) were also scanned during the time the aneurysm clips were applied, in order to document perfusion in sub-regions of the brain. This information was obtained to establish the amount of intra-animal variation in the location of the infarct zone and the diminution of cerebral blood flow due to two-vessel occlusion. Dynamic

susceptibility contrast (DSC)-MRI, a bolus tracking perfusion MRI method, was employed during the actual ischemic period, to generate full brain maps of cerebral blood flow and volume. Ultrafast imaging requires an echoplanar imaging (EPI) sequence for DSC-MRI and collects a rapid series of T_2 * weighted images during administration of contrast (Prohance, 0.1 mM/kg, *i.v.*). Full brain coverage yielded 20 slices at 1.5×1.5 mm in-plane, 2 mm slice thickness, with TR=1500 msec and TE=20 msec. Parametric maps were generated for cerebral blood flow (CBF in ml blood/100g tissue/minute), cerebral blood volume (CBV in blood volume percentage of total tissue volume), and mean transit time (MTT in seconds) using the JIM software package (Xinapse Systems, Thorpe Waterville, United Kingdom), following the model-independent method described in Ostergaard L. *et al.*(Ostergaard 2005). The arterial input function (AIF) was determined from (~20) brain pixels identified to have AIF characteristics (large, early, rapid intensity changes) by a two stage automatic scanning and selection routine.

Neurological Assessment

Neurological assessment was performed on a daily basis by a single operator following stroke using a 100 point examination scale developed by Spetzler and associates (Spetzler *et al* 1980). This scale evaluates motor function, behavior (mental status), and cranial nerve deficit. Higher scores represent better functional outcomes. Motor function is graded from 1 to 75, according to severity of hemiparesis in the extremities and face. Hemiparesis in the extremities is scored as 10=severe hemiparesis, 25=mild hemiparesis, 55=favors normal side or 70=normal ability. Facial weakness is scored as 1 = one sided paralysis and 5=normal facial movement. Behavior and alertness is scored from 0 to 20, with 0=dead, 1=comatose, 5=aware but inactive, 15=aware but less active, and 20=normal. Visual field deficits are scored as 1 when present or 5 when absent.

Tissue Collection

Animals were sacrificed two days post-stroke (n=7) or seven days following stroke (n=4). Stroke subjects were sedated and then deeply anesthetized with 25 mg/kg sodium pentobarbitol. A blood sample was drawn for a complete blood count prior to exsanguination and perfusion with cold heparinized saline (2U/ml) via the ascending aorta. Brains were rapidly removed, placed in a rhesus brain matrix (ASI, Warren, MI, USA) and cut into 15 consecutive, 4 mm thick coronal slabs per brain. For visualization of the region of infarction in a subset of animals, sections were immediately placed in 1.5% 2,3,5-triphenyltetrazolium chloride (TTC, Sigma) in 0.9 % phosphate-buffered saline and stained for 15 min at 37°C. A further subset of animal brain sections was stained for 45 min with TTC for increased infarct definition. After staining, sections were electronically imaged for subsequent analysis of infarct size and processed further for histology.

Histology and Histochemistry

Tissue slabs were fixed for four to five days in neutral buffered 10% formalin containing increasing concentrations of sucrose (10% to 30%) for cryoprotection. To prevent freezing artifact, tissues were equilibrated at 4°C for 24 h in 10% glycerol, 2% dimethyl sulfoxide in 0.02 M potassium phosphate buffer (pH 7.4). This was followed by 3-4 days of immersion in 20% glycerol, 2% DMSO in 0.02 M potassium phosphate buffer (pH 7.4). Following fixation and cryoprotection the 4 mm slabs were frozen and then sectioned (30-50µm) on a sliding microtome. The sections were mounted onto slides and one 50µm section from each 4 mm slab was stained with cresyl violet for high magnification visualization of the infarcted area. These sections also served to determine the distribution of the injury between gray and white matter and the severity of the infarct within specific brain regions. In addition, representative slides

with 30µm sections were also processed for Fluoro-Jade B staining (Chemicon, Temecula, CA) as described by (Schmued and Hopkins 2000) to identify damaged neurons.

Infarct Measurements

Images from T₂-weighted MRIs, TTC and cresyl violet stained sections were examined for the location of infarction, and the total affected area was measured using NIH ImageJ v1.38 software (Bethesda, MD, USA). For the transverse slices of T₂-weighted image, measurement started from the level of the gyrus rectus. The brainstem and cerebellum in the inferior slices of the MRI were included in the hemispheric measurements. T₂ signal at the location of the surgical cavity was not included as part of the infarction, as this was due to the residual effects (fluid collection near the orbit) of the surgery. Each of the techniques (MRI, cresyl violet, TTC) used to measure infarct volume analyzed comparable anatomical regions and sampled ~15 slices (4mm each). Measurements of infarct volume as a percentage of the ipsilateral hemisphere were made using the following formula: (area damaged/area of the ipsilateral hemisphere) x 100%. Total volume reflected calculations that compensated for slice thickness and intervening slices that were not directly measured. Basal ganglia involvement was specifically measured to compare the relative amount of damage versus the cortex.

Statistical analysis

Experimental effects were analyzed using one-way analysis of variance and in cases of significance Bonferroni's post-hoc test was used to determine pair-wise differences between treatment groups. Differences were considered statistically significant when p<0.05. Error calculations for the Infarct Reproducibility study are presented in standard error of the mean while calculations for the Duration of Ischemia study are presented in standard deviation as a result of small sample size. Correlations were prepared between pathologic and MR stroke sizes, as well as infarct size and neurologic score using the square of the Pearson Product Moment Correlation Coefficient to generate R² values. Power analyses were conducted using the online toolkit provided by DSS Research (www.dssresearch.com) with an alpha error of 5% and a beta error of 25%.

RESULTS

Intra-operative Monitoring and Hematology

The intra-operative vital signs and baseline hematological values for the five animals that underwent 60 minutes of ischemia in the *Infarct Reproducibility Study* are provided in Table 1. There was no significant variation in vital signs measurements from baseline, during ischemia and post-occlusion values. There was also no evidence of hemorrhage or operative complications on the immediate post-operative MRI. However, following ischemia there was a significant rise in the percentage of neutrophils and a significant reduction in lymphocytes. All animals survived for the pre-planned two or seven days of recovery, except one of the animals in the 90 minute group. Animals given 60 minutes of occlusion were capable of feeding, drinking and self-grooming behaviors although impaired due to the neurologic lesion. This outcome is desirable as it is consistent with similar neurologic assessments of humans who suffer cortical strokes.

MR imaging

Anatomical MRI scans of the animals revealed normal brain anatomy pre-ischemia, as well as during the time period immediately following two-vessel occlusion (data not shown). At two days after stroke, T_2 -weighted scans (Figure 2) closely matched the early lesion revealed by diffusion imaging, which retained a similar size and location at two days post-stroke. The

perfusion MR images (Figure 2A) revealed hypoperfusion that encompassed much of the ipsilateral hemisphere and some contralateral and midline structures immediately following the induction of stroke. MRI scans obtained during ischemia were taken to clearly delineate the regions of the brain that were hypoperfused. These data allow the calculation of the infarct lesion volume based on the fraction of the perfusion deficit region or area at risk rather than the fraction of hemispheric volume. For the three animals scanned in this manner, the infarct lesion was $37\pm2\%$ of the area at risk and the average perfusion deficit region was $29\pm3\%$ of the total brain volume. Generally included within the perfusion deficit region were smaller enhanced regions in the diffusion-weighted images (DWI), also obtained during the occlusion period, and similar regions of hypointensity in the associated apparent diffusion coefficient (ADC) maps created from them (Figure 2A). Hypoperfusion in the region of the cingulate cortex was observed, but this region was damaged bilaterally only after the more prolonged 90 minute duration of occlusion.

In the *Ischemia Duration Study*, the distribution of the infarct progressively enlarged as the duration of ischemia increased. After 60 minutes of two-vessel occlusion the infarct encompassed portions of the frontal, parietal, and insular motor cortex (Figure 2B), expanding with 90 minutes of ischemia to include midline structures, such as the cingulate cortex. Anatomical T_2 -weighted scans of animals with a longer survival time of seven days (data not shown) showed expansion of the anomalous T_2 -weighted image intensity beyond the original boundaries observed two days after ischemia. Despite the apparent diffusion of edema, the volume of infarct established using the T_2 MRI taken two days after stroke correlated well statistically (see below) with endpoint histology using TTC and cresyl violet (Figure 2C). Comprehensive cortical damage is evident throughout the ipsilateral hemisphere (Figure 2D).

Infarct Measurements: Correlation of MRI, TTC and Cresyl Violet Techniques

To establish the best methodology for measuring infarct size, TTC-stained slices were compared with cresyl violet-stained sections as well as with T₂-weighted MRI scans, obtained at day 2 or 7 post-ischemia from animals in the *Ischemia Duration Study*. Stroke volume measured from T₂-weighted MRIs had a high R² of 0.9866 (as a % of the ipsilateral hemisphere) and 0.9881 (actual volume) when correlated to TTC-based estimates (Figure 3A, B). The correlation between infarct volumes measured by T₂-weighted MRI and cresyl violet stained sections had a R² of 0.9633 (Figure 3C).

Duration of Ischemia

The effect of duration of reversible two-vessel occlusion on infarct size was examined in three groups of animals (n=2/group) with occlusion times of 45, 60, or 90 minutes. As expected, increasing the duration of ischemia resulted in larger cortical infarct size (Table 2), with additional expansion into midline structures in hypoperfused regions defined by perfusion MRI. The associated neurological scores generally, but not absolutely, reflected increased damage with lower (worse) scores. The animals with 45 minute occlusions had a mean infarct volume of $1982.5 \pm 251 \text{ mm}^3$ which involved a mean of 19.0 ± 0.98 % of the ipsilateral hemisphere based on T₂-weighted MRI. There was no noticeable damage to the basal ganglia; the average neurological score at 48 hrs post-stroke was 34.5 ± 4.9 (Table 2).

Animals with 60-minute occlusions had a mean infarct volume of $2381 \pm 654 \text{ mm}^3$ which involved a mean infarct size of $22.1 \pm 4.6\%$ of the ipsilateral hemisphere based on T₂-weighted MRI. Damage to the basal ganglia continued to be very small and was not observed in all animals. The average neurological score at 48 hrs was 30.5 ± 0.7 (Table 2).

The 90 minute occlusion involved a greater portion of the cortex and resulted in the death of one of the animals prior to the day 2 post-stroke scan. Post-mortem T_1 , T_2 and diffusion-

weighted scans all revealed extensive ipsilateral damage that resulted in a slight midline shift of the brain (data not shown), similar to the other animal in this group. As assessed by T₂weighted MRI, the mean infarct volume was $4707 \pm 12.7 \text{ mm}^3$, which was $44.4 \pm 4.0\%$ of the ipsilateral hemisphere. Cresyl violet measurement of the mean infarct size was $34.4 \pm 7.8\%$ of the ipsilateral hemisphere. Unlike the shorter 45 or 60 minutes of ischemia, 90 minutes of stroke resulted in minor basal ganglia involvement of 4.1% of the ipsilateral hemisphere. The mean neurological score at 24 hours was 26 ± 1.4 . The surviving animal had a neurological score of 27 at 48 hrs post stroke (Table 2, animal #5).

Infarct Reproducibility

A duration of 60 minutes of ischemia resulted in moderate sized infarcts, good survivability and obvious neurological effects in the *Ischemia Duration Study*, thus we selected this duration of ischemia for the *Infarct Reproducibility Study* (Table 2, animals #7-11). T₂-weighted MRI scans and cresyl violet staining revealed a similar cortical pattern of damage as described above, with little (~1%) basal ganglia involvement. The amount of infarcted cortical gray matter measured from T₂-weighted MRI scans was 2834 ± 244 mm³, or $41.5 \pm 2.0\%$ of the ipsilateral cortex. This amount of damage was equal to $25.4 \pm 0.7\%$ of the ipsilateral hemisphere, very similar to cresyl violet ($24.1 \pm 0.9\%$) and TTC ($25.8 \pm 2.1\%$) measurements. Neurological deficits were moderate with a marked hemiparesis on the affected side and a total score at 48 hours post-stroke of 34.6 ± 1.8 . Neurological scoring and final infarct size revealed a consistent relationship (Figure 3D). A power analysis of the seven animals that received 60min of occlusion revealed that an N of 6 would be required to find a 15% or greater reduction in infarct size statistically significant.

Histology

Histopathologic evaluation of Fluoro-Jade B and cresyl violet stained specimens revealed that the portions of the frontal, parietal, somatosensory and the insular cortex were consistently injured in all animals subjected to 60 minutes of reversible two-vessel occlusion. Subcortical white matter injury was minor and proximal to cortical injury; no injury to deep white matter tracts was observed microscopically. The superior temporal gyrus and the basal ganglia were inconsistently injured. Representative Fluoro-Jade B and cresyl violet images showing neuronal injury in the ipsilateral cortex are provided in Figure 4. Animals with 60 minute occlusions had discontinuous patches of gray matter injury, while animals that underwent 90 minute occlusions had more confluent injury. In addition, 90 minute occlusions extended injury to midline structures in the ACA distribution, such as the cingulate and anterior frontal cortex, as described in T_2 -weighed MRI scans.

Extended Survival

Four animals from the *Ischemia Duration Study*, two from the 45 minute occlusion and two from the 60 minute occlusion, were scored neurologically each day for seven days following occlusion. This allowed evaluation of neurological deficits in the absence of the potentially confounding effects of analgesics commonly used immediately following surgery. All animals showed a steady improvement in behavior and motor function over seven days (Figure 5). The difference between animals receiving 45 minutes verses 60 minutes of occlusion demonstrates the potential ability to recognize the attenuation of neurological deficits in animals in a preclinical trial. The rate of recovery was attenuated for animals undergoing the longer stroke period of vascular occlusion. Also, there was sufficient room for improvement of the neurological scores suggesting that successful interventions would not be inhibited by a ceiling effect. This pattern of behavioral and motor recovery is consistent with observations from clinical stroke patients (Traversa *et al* 2000).

DISCUSSION

The ideal method for stroke modeling in the primate is uncertain. Although the MCAO offers significant advantages, previous NHP studies had limited success in producing reproducible cortical infarcts. MCAO in the baboon produced significant, but variably sized strokes located primarily in the basal ganglia (Del Zoppo *et al* 1986). A transorbital approach in baboons that used clips on the proximal MCA and the orbitofrontal branch of the MCA produced no infarction in six animals, and only a limited basal ganglia infarction in another six animals after 6 hours of occlusion (Young *et al* 1997). A similar approach in cynomolgus monkeys produced larger infarcts about 1000 mm³ in size after 3 hours of occlusion but infarcts were primarily localized to the basal ganglia (Takamatsu *et al* 2000). Ninety minute MCA only occlusion in the rhesus produced infarcts that were primarily located in the caudate and also varied in size (Murphy *et al* 2008). Thus, these approaches do not fully mimic the clinical situation of cortical stroke.

An alternative model of stroke uses thrombosis to create occlusion. However, one study of thromboembolic occlusion of the ICA in cynomolgus monkeys still results in a significantly variable infarct primarily located in the basal ganglia (Kito *et al* 2001). Thrombosis of the MCA, induced by activating intravascular rose bengal with photoirradiation (Maeda *et al* 2005) also produced significant variability in infarct size. PET performed during the ischemic induction showed cyclic flow reductions in the brain likely related to thrombus formation and embolization more distally. Therefore, we opted for the use of aneurysm clips for better control of placement, completeness and reversibility of occlusion.

To develop a model of cortical stroke in the rhesus macaque we initially performed a pilot study with different durations of reversible proximal MCA (single vessel) trunk occlusion. MCAO ranging from 3-6 hours resulted in progressively larger basal ganglia damage, but with little cortical involvement (data not shown). Four hours of occlusion of the orbitofrontal branch and the proximal trunk of the MCA also caused infarcts that were modest and localized to the basal ganglia (data not shown), suggesting sufficient collateral flow protected the cortex. Occlusion of both ACAs and one ICA in the baboon was reported to create a mixed stroke involving both the basal ganglia and cortex (Huang *et al* 2000), but with significant morbidity and mortality. The occlusion of the ICA at the level of the anterior choroidal artery can also be challenging, where lack of visualization and proximity of the posterior communicating artery could result in persistent collateral flow through the PCA. These data along with our experience suggesting a significant amount of collateral flow in the rhesus macaque, employing the 2-vessel occlusion approach to include the MCA (distal to the orbitofrontal and lenticulostriate arteries) and bilateral occlusion of ACAs would preserve the basal ganglia while establishing ischemia in the cortex.

There are apparent species differences on how ICA and/or MCA occlusion affects the evolution of stroke in humans versus macaques. The two proximal (A1) anterior cerebral arteries in the macaque merge to form the single (A2) pericallosal artery (Kapoor *et al* 2003), which supplies both hemispheres (Coceani *et al* 1966). While humans may also have cross-filling of the contralateral A2 via the ACA, collateral supply is variable and often limited by the small size of the ACA. Hence, strokes in humans by ICA occlusion or downstream ipsilateral MCA involvement alone can result in cortical lesions, while it is plausible that collateral flow in the macaque ACAs can provide significantly more cortical protection.

To maximize cortical infarction and minimize basal ganglia involvement and variability, we combined occlusion of the MCA (with preservation of blood flow in the orbitofrontal artery and lenticulostriate arteries) with the elimination of collateral circulation through the ACAs. We were able to demonstrate a close correlation of neurological deficits and infarct sizes

measured by T₂-weighted MRI, TTC and by cresyl violet staining in our model, similar to others (Huang *et al* 2000; Mack *et al* 2003). Damage to the cortex correlated with duration of ischemia and was directly related to neurological outcome. Although the bilateral ACA occlusion in this model is not directly translatable to human MCAO stroke, titrating the duration of the vascular occlusion, we avoided bilateral damage and contralateral edema.

In the peripheral blood, stroke resulted in an elevation in the neutrophil count and a significant reduction in the lymphocyte count analogous to findings in humans stroke, which may be a prognostic indicator (Clark *et al* 1996; Kasner *et al* 2001; Ross *et al* 2007). Murphy et al. also found neutrophil and macrophage infiltrates on the boarders of their lesions (Murphy *et al* 2008). These are stroke-induced changes that may point to interventions in immunological function that could modulate post-stroke effects.

The use of diffusion weighted MR images in our model allowed identification of an ischemic core region, indicating metabolically stressed areas at high risk of proceeding to infarction. Areas outside of these regions that are within the perfusion deficit region can be considered the ischemic penumbra, which in this model, as in clinical settings, is largely salvaged by reperfusion. The ischemic core however, which generally expands towards the boundaries of the perfusion deficit region with increased duration of ischemia, is the primary target region for neuroprotection. Generally, the pattern and scope of the DWI changes during ischemia were similar to T₂-weighted images of cortical edema at 48 hr post-stroke in these non-treated animals, but it is likely that a comparison with treated animals would reveal portions of the ischemic core that have been spared injury by antecedent therapy such as preconditioning. The timing of the collection of MR images specifically during this acute post-ischemic period may be particularly important. Other studies have observed that the affected volume, as seen by MRI (T₂-weighted, diffusion weighted, or FLAIR) peaks 1-3 days after reversible ischemia match the final infarct size, has been previously reported in monkeys (D'Arceuil et al 2006; Liu et al 2007) and rodents (Li et al 2000) (Sotak 2002). Thus, MR imaging in this timeframe can be an important tool for assessing the evolution and extent of infarct, as well as the effect of neuroprotection, as some tissue should be salvageable by successful treatment in the regions of hypoperfusion.

Thus, transient occlusion of the MCA and bilateral ACAs in the rhesus macaque produces reproducible, clinically relevant, cortical infarcts with limited variability between MRI and histological measures and results in excellent survival. In addition, neurological deficits correlate well with levels of damage and subcortical (basal ganglia) injury is minimized. Due to the fact that the deep white matter is largely intact, preservation of penumberal areas of cortex by a neuroprotectant is likely to be apparent clinically. With a moderate cortical infarct volume of ~25%, this model provides latitude for modulating infarct size to test the negative or positive effects of candidate therapeutics. Assessment of putative therapeutics in such a nonhuman primate stroke model may reduce the frequency of failed human clinical trials and offers the potential for preclinical screening of neuroprotective agents aimed at reducing cortical infarction in large vessel stroke in humans.

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Figure 1. Schematic diagram of surgical site and vasculature of the rhesus two-vessel occlusion model

A) In situ surgical view of the right orbit of the rhesus macaque illustrating the exposure of major cerebral arteries and positioning of aneurysm clips for occlusion of the right middle cerebral artery and bilateral anterior cerebral arteries. **B**) *Ex vivo* illustration of the Circle of Willis with one clip location on the middle cerebral artery distal to the orbito-frontal artery (lateral clip) and a second clip on both anterior cerebral arteries (medial clip) just before the vessels join to form a single peri-callosal artery.





A) Cerebral blood flow (CBF map), diffusion weighted images (DWI), ADC maps and T2weighted images are shown in three transverse slices taken from one animal. CBF, DWI and ADC were obtained during occlusion and the T2 scan was taken after 48 hrs. Hypoperfusion (dark blue) during 60 minutes of ischemia resulted in smaller areas of sub-regional cortical damage (DWI and ADC) as verified with the T2 scans. Regions of interest are encircled in red. **B**) The effect of varying the duration of ischemia on the extent of cortical damage. The region of hypoperfusion is shown at one level in a typical example of a CBF map, and encompasses much of the ipsilateral hemisphere and some bilateral, midline structures. The amount of damage (T2 signal) at 48 hrs post-ischemia increases with the duration of ischemia, encompassing more of the region that underwent hypoperfusion, including the bilateral cingulated gyrus. C) Comparison of infarct damage following 60 minute occlusion using T2weighted MRI (48 hrs post-ischemia), TTC, and cresyl violet stained sections. There was excellent agreement in the amount of damage as assessed by the three different techniques. D) Pseudo-colored distribution of damage is shown in transverse sections two days after 60 min of reversible ischemia, by T2-weighted MRI. Comprehensive cortical damage is evident throughout the brain on the side ipsilateral to the two-vessel occlusion.



Figure 3. Correlation of primary outcome measures

A-C) Animals were subjected to 45, 60 or 90 minutes (n=2/timepoint) of ischemia followed by two or seven days of recovery. **A)** Correlation of infarct measurement using T2-weighted MRI verses TTC (percent of hemisphere). **B)** T2 infarct volume (mm3) verses TTC infarct volume (mm3). **C)** T2-weighted MRI verses cresyl violet percent of hemisphere. **D)** Animals (n=5) received 60min of ischemia followed by two days of recovery. Immediately prior to sacrifice, animals underwent neurological assessment followed by MR scanning. Comparison of T2-weighted MRI lesion measurements and neurological score.



Figure 4. Histological evidence of moderate to severe cortical injury following 60 minute occlusion A. Low magnification montage of infarct (Fluoro-Jade B; green label), predominantly

restricted to the cortical ribbon. **B** and **D** show an area of fronto-parietal cortex with a patchy, discontinuous region of injury. In **B**, the bright, Fluoro-Jade B-labeled neurons have suffered ischemic damage, and unaffected, unstained neurons are darker than the background neuropil. A Nissl stained adjacent section (**D**) shows the residual intact neurons (left) vs. the right side of the figure, which shows loss of neurons and prominent capillaries. **C & E** show uninjured contralateral fronto-parietal cortex. No Fluoro-Jade B neurons are seen in **C** and normal cortical architecture and neuronal staining are observed in **E**.





Animals were evaluated daily for seven days following 45 (n=2) or 60 (n=2) minute occlusion. Evaluations consisted of neurological assessments as described in Methods. Analgesics were discontinued 24hrs following surgery and animals were scored daily.

Table 1 Physiological parameters and complete blood count (% differential) of animals in the Infarct Reproducibility Study

Stability of physiological parameters and white blood cell measurements were compared (n = 5; mean \pm sem).

Parameter	Befor	e occlusion	During occlusion	Post-occlusion
heart rate (b/min)	11	0.1 ± 5.5	124.7 ± 1.8	118.1 ± 9.0
temp (°C)	98	3.3 ± 0.5	99.7 ± 0.3	99.8 ± 0.4
SpO ₂	99	9.3 ± 0.5	98.7 ± 0.6	99.3 ± 0.4
EtCO ₂ (mmHg)	36	5.1 ± 1.6	36.5 ± 0.5	41.5 ± 4.8
systolic blood pressure	(mmHg) 60	0.2 ± 3.7	67.0 ± 1.4	68.8 ± 2.3
diastolic blood pressure	e (mmHg) 35	5.6 ± 1.2	38.7 ± 2.5	40.2 ± 2.6
% Differential	Before occlusion	During	g occlusion	Post-occlusion
neutrophils	48.6 ± 6.4		N/D	$69.0^{*} \pm 3.9$
lymphocytes	44.5 ± 5.6		N/D	$23.7^{*} \pm 3.9$
monocytes	4.0 ± 0.6		N/D	8.0 ± 1.7
eosinophils	2.0 ± 0.8		N/D	1.2 ± 0.2
basophils	0.6 ± 0.9		N/D	0.6 ± 0.1

N/D = not done.

p value < 0.05 compared to baseline samples.

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Table 2

Summary of primary outcome measurements

Ischemia Duration Study (rhesus #1-6) and Infarct Reproducibility Study (rhesus #7-11).

Study # Lethenia Survival (aiys) TTC T2 Cressi (nim) T2 MR1 Motor Behavior Fundion Behavior Total Total Motor Behavior Total Total Total Total Motor Behavior Total Constrained Behavior Motor Behavior Motor Behavior Altrines Source Altrines Altrine Altrine Altrines Altrines Altrines Altrines Altrines Altrines Altrine Altrines Altrine					hen	% ıpsılatera iisphere infaı	I rcted	% ipsulateral cortex infarcted	Infarct Volume	Ne (48h	urological Score ir post-occlusion)	
	Study	#	Ischemia (min)	Survival (days)	TTC	T2 MRI	Cresyl violet	T2 MRI	T2 MRI (mm ³)	Motor Function	Behavior and Alertness	Total Score
		1	45	7	8.6	18.3	21.53	U/N	1805	23	15	38
Instant 3 60 7 9.1 18.9 25.25 ND 1918 21 10 31 Instant 4 60 7 14.4 25.4 26.88 ND 2844 20 10 30 Instant 5 90 2 36.4* 47.3 39.9 ND 4716 15 10 30 6 90 2 $36.4*$ 41.6 28.9 ND 4716 15 27 27 7 6 90 2 $34.8*$ 41.6 28.9 ND 4716 15 27 7 6 90 2 $21.1*$ 23.3 21.1 42.5 2568 ND ND ND 8 60 2 $23.2*$ $21.3*$ $21.4*$ $25.6*$ 20° 16° 36° 9 60 2 $23.3*$ $23.5*$ $25.6*$ $23.6*$ 20° 16°		2	45	7	8.7	19.7	28.84	N/D	2160	19	12	31
Duration 4 60 7 144 25.4 26.88 N/D 2844 20 10 30 5 90 2 36.4* 47.3 39.9 N/D 4716 15 12 27 6 90 2 36.4* 41.6 28.9 N/D 4698 N/D 12 27 7 60 2 34.8* 41.6 28.9 N/D 4698 N/D 12 27 7 60 2 21.1* 23.3 21.1 42.5 258 23 14 1 8 60 2 23.1* 24.2 26.9 39.5 264 20 16 36 Reproducibility 9 60 2 33.5 264 20 16 36 10 60 2 31.7 25.7 24.3 369 18 31 11 60 2 33.1 26.9	Ischemia	ю	60	7	9.1	18.9	25.25	N/D	1918	21	10	31
	Duration	4	60	7	14.4	25.4	26.88	N/D	2844	20	10	30
		5	06	2	36.4 [*]	47.3	39.9	N/D	4716	15	12	27
7 60 2 21.1 23.3 21.1 42.5 2558 23 18 41 8 60 2 23.2* 24.2 26.9 39.5 266.4 20 16 36 Reproducibility 9 60 2 23.3.* 27.0 25.0 47.0 2980 20 13 33 10 60 2 31.7 25.7 24.3 43.6 3699 18 13 31 11 60 2 30.0 26.6 23.3 35.1 2269 20 12 32		9	06	2	34.8*	41.6	28.9	Q/N	4698	Q/N	N/D	N/D
8 60 2 23.2* 24.2 26.9 39.5 2664 20 16 36 Reproducibility 9 60 2 23.0* 27.0 25.0 47.0 2980 20 13 33 Inductibility 9 60 2 31.7 25.7 24.3 43.6 3699 18 13 31 10 60 2 30.0 26.6 23.3 35.1 2269 20 12 31		7	60	2	21.1*	23.3	21.1	42.5	2558	23	18	41
Infarct 0 60 2 23.0* 27.0 25.0 47.0 2980 20 13 33 Reproducibility 10 60 2 31.7 25.7 24.3 43.6 3699 18 13 31 11 60 2 30.0 26.6 23.3 35.1 2269 20 12 32		8	60	2	23.2^{*}	24.2	26.9	39.5	2664	20	16	36
10 60 2 31.7 25.7 24.3 43.6 3699 18 13 31 11 60 2 30.0 26.6 23.3 35.1 2269 20 12 32	Infarct Reproducibility	6	60	2	23.0^{*}	27.0	25.0	47.0	2980	20	13	33
11 60 2 30.0 26.6 23.3 35.1 2269 20 12 32		10	60	2	31.7	25.7	24.3	43.6	3699	18	13	31
		11	60	2	30.0	26.6	23.3	35.1	2269	20	12	32

J Cereb Blood Flow Metab. Author manuscript; available in PMC 2010 February 25.

* Measurements from brain slices stained with TTC for 45 min. All other slices stained with TTC for 15 min.