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Good Laboratory Practices Are Not Synonymous with Good Scientific Practices, Accurate Reporting, or Valid Data

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In her commentary, Tyl (2009) responded to our criticism (Myers et al. 2009) of her bisphenol A (BPA) research (Tyl et al. 2008), and she defended the reliance on Good Laboratory Practices (GLP) in animal studies concerning risks posed by chemicals. Her commentary, however, provides additional evidence that her research on BPA is flawed and that GLP can be unreliable.

The key evidence can be found in her treatment of the effect of BPA on prostate weight (Tyl et al. 2008). This effect is important because Tyl's data on adult prostate in mice has been used by the chemical industry—which has funded all of Tyl's research on BPA—and the Food and Drug Administration (FDA) to conclude that BPA has no effect at low doses. Indeed, Tyl argued that the weight of the evidence supports her findings that BPA is safe (all industry-funded studies report no low-dose effects of BPA). In contrast, > 200 studies in experimental animals, all funded by government agencies, have reported significant effects of BPA at low doses that are relevant to human and ecologic exposures (vom Saal et al. 2007).

We (Myers et al. 2009) concluded that prostate weights reported by Tyl et al. (2008) were abnormally high in control males, suggesting either that the dissections were done improperly, that control animals were exposed to a contaminating estrogen, or that their prostates were diseased. This would render the results invalid and therefore inappropriate to use in assessing BPA safety. It would also provide insights as to why, despite many other studies showing adverse effects of exposure to BPA at low doses (vom Saal et al. 2007), Tyl et al. (2008) detected none.

To counter this criticism, Tyl (2009) presented a table (her Table 2) of mouse prostate weights from other laboratories. The data she presented in fact show that no other laboratory measuring prostate weight in mice has reported mean weights as high as those reported by Tyl et al. (2008), except in old male mice with diseased prostates. Tyl's table cites data from research published by Heindel et al. (1995) previously conducted at her own institution, Research Triangle Institute, although she did not acknowledge this. The mean prostate weight reported by Heindel et al. (1995) for 16- to 17-week-old

CD-1 male mice was 48 mg, which is similar to most other findings, but contrasts sharply with Tyl's mean prostate weights of 74 mg for the F₁ males in her BPA study (these males were identified in Table 1 of Tyl's commentary as being examined at 18 weeks of age).

Table 2 of Tyl (2009) also includes data from a publication by Morrissey et al. (1988) showing a mean prostate weight of 58 mg in 23-week-old CD-1 mice. However, this study involved comparing data from two laboratories, and Tyl omitted from her table the data from the second laboratory that reported a mean prostate weight of 35 mg in 23-week-old CD-1 males. Morrissey et al. (1988) observed that the laboratory reporting the mean of 58 mg also had a higher standard deviation and lower statistical sensitivity than the laboratory reporting the 35 mg mean prostate weight. In studies in which prostate weight is high, such as that of Tyl et al. (2008), the findings are suspect in that the abnormally high prostate weight data show a poor relationship to other male reproductive organs (Morrissey et al. 1988). This strongly suggests that nonprostatic tissue has been included when prostate weights are abnormally high in the absence of disease.

Tyl's discussion of prostate weight effects also suggests that studies identified as GLP may not adhere to the strict record-keeping goals to which GLP aspires, undermining one of the arguments used for the value of GLP over research funding by the National Institutes of Health, which rarely follows the costly GLP guidelines. In the original publication, Tyl et al. (2008) reported that F₁ retained males were necropsied at approximately 14 weeks of age. In Table 1 of her commentary (Tyl 2009), Tyl stated that these males were 18 weeks of age at necropsy. However, in testimony before the FDA Science Board BPA Subcommittee hearing on 16 September 2008 (FDALive.com 2008), Tyl stated that these males were 24 weeks of age at necropsy as an explanation for their high prostate weights. Tyl assured the FDA panel that since "the difference in age influences growth rate and growth of organs, the comparison [of 12- and 24-week-old males] is specious, it is comparing apples and oranges." In fact, Tyl's data in Table 1 of her commentary (Tyl 2009) show no relationship between age and body weight. The inconsistencies in Tyl's FDA testimony, which could have had a significant impact on a regulatory decision concerning BPA, and the data concerning the age at tissue collection, prostate weights, and body weights presented in Table 1 of her

commentary are disturbing, and indicate that a thorough review of original data in Tyl et al. (2008) by scientific experts is warranted.

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REFERENCES

- FDALive.com. 2008. Meeting of the Bisphenol A Subcommittee of the Science Board. Meeting Date: 9/16/08-9/16/08. Available: <http://www.fdalive.com/pastmeetings.cfm?committeekey=61> [accessed 21 October 2009].
- Heindel JJ, Chapin RE, George J, Gulati DK, Fail PA, Barnes LH, et al. 1995. Assessment of the reproductive toxicity of a complex mixture of 25 groundwater contaminants in mice and rats. *Fundam Appl Toxicol* 25(1):9-19.
- Morrissey RE, Lamb JC, Schwetz BA, Teague JL, Morris RW. 1988. Association of sperm, vaginal cytology, and reproductive organ weight data with results of continuous breeding reproduction studies in Swiss (CD-1) mice. *Fundam Appl Toxicol* 11(2):359-371.
- Myers JP, vom Saal FS, Akingbemi BT, Arizona K, Belcher S, Colborn T, et al. 2009. Why public health agencies cannot depend on good laboratory practices as a criterion for selecting data: the case of bisphenol A. *Environ Health Perspect* 117:309-315.
- Tyl RW. 2009. Basic exploratory research versus guideline-compliant studies used for hazard evaluation and risk assessment: bisphenol A as a case study. *Environ Health Perspect* 117:1644-1651.
- Tyl RW, Myers C, Marr M, Sloan CS, Castillo N, Veselica MM, et al. 2008. Two-generation reproductive toxicity study of dietary bisphenol A (BPA) in CD-1(r) (Swiss) mice. *Toxicol Sci* 104(2):362-384.
- vom Saal FS, Akingbemi BT, Belcher SM, Birnbaum LS, Crain DA, Eriksen M, et al. 2007. Chapel Hill bisphenol A expert panel consensus statement: integration of mechanisms, effects in animals and potential to impact human health at current levels of exposure. *Reprod Toxicol* 24(2):131-138.

Good Laboratory Practices: Tyl Responds

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I am responding to the comments by vom Saal and Myers on my commentary (Tyl 2009), which was in response to their commentary (Myers et al. 2009). Our dietary BPA mouse study was performed under Organisation for Economic Co-operation and

Development (OECD) Test Guideline 416 (OECD 2001) and Good Laboratory Practices (GLPs) (OECD 1998), with in-house quality control, and formal RTI quality assurance, and expert Developmental & Reproductive Toxicology (DART) panel oversight. We submitted all of our summary data online with our published mouse study (Tyl et al. 2008). I also discussed our BPA data, especially prostate weights (ventral and dorsolateral lobes separately) and ages of animals at scheduled necropsy, two areas of concern to vom Saal and Myers, in my commentary (Tyl 2009). We found no BPA effects on mouse prostate weights at any dietary dose, from 3 µg/kg/day to 600 mg/kg/day, whereas vom Saal and colleagues reported increased prostate weights at 2 and 20 µg/kg/day BPA, administered on gestational days 11–17 (Nagel et al. 1997). Vom Saal and Myers suggested that prostate weights were very large in our control mice (Tyl et al. 2008), in his view, likely due to prostatitis and/or poor dissection techniques resulting in extraneous tissue left on the prostates. We provided histopathologic confirmation of low (normal) rates of prostatitis, no increased incidences or severities from BPA, and no evidence of extraneous tissue from examination of the prostate paraffin block faces and histology slides. For animal ages at termination, we initially presented approximate ages of our animals at demise because we were not aware of their concerns at that time. In a letter to the editor of *Toxicological Sciences*, where the multigenerational BPA rat and mouse studies (Tyl et al. 2002, 2008) were published, I (Tyl 2009) explained in great detail the ages of our F₀, F₁, and F₂ animals at scheduled necropsy; the ages of the F₁ animals varied at most by 3 weeks in all groups, based on when the F₀ animals mated during the 2-week mating period, the need to have all F₁ offspring exposed for at least 8 weeks during the prebreeding period, and the need for all F₁s to be available for pairing in all groups at the same time. FDA auditors recently spent 11 days at RTI (30 March to

9 April 2009) inspecting our BPA rat (Tyl et al. 2002) and mouse (Tyl et al. 2008) multigenerational reproductive toxicity study data and records, with no study findings.

It is clear that vom Saal and Myers apparently still do not understand or appreciate the discipline, power, importance, and usefulness of GLPs on study design, performance, documentation, and interpretation (which is why GLP-compliant studies are preferentially used in formal hazard identification and risk assessment).

The effects of dose levels, route, timing (life stages), and duration of BPA exposures on reported early and late effects, and whether there is a linkage between the low-dose early end points from short-term, small, basic exploratory studies and the outcomes from long-term guideline studies, need to be evaluated. Long-term, robust oral studies (Ashby et al. 1999; Cagen et al. 1999) and guideline-compliant oral multigenerational studies (Ema et al. 2001; Tyl et al. 2002, 2008), regardless of sponsorship, have not confirmed the low-dose effects reported in the basic studies, nor any long-term consequences anticipated from these reported early effects. Determination of whether there is hazard or risk of BPA to humans and wildlife from low, environmentally relevant doses by relevant exposure routes is based on available appropriate data. To date, governmental and other organizational hazard, risk, and weight-of-evidence assessments have concluded, based on the data, that there is no evidence of any adverse effects from oral BPA at low doses.

The author is employed by RTI International, a contract research organization.

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REFERENCES

- Ashby J, Tinwell H, Haseman J. 1999. Lack of effects for low dose levels of bisphenol A and diethylstilbestrol on the prostate gland of CF1 mice exposed *in utero*. *Regul Toxicol Pharmacol* 30:156–166.
- Cagen SZ, Waechter JM, Dimond SS, Breslin WJ, Butala JH, Jekat FW, et al. 1999. Normal reproductive organ development in CF-1 mice following prenatal exposure to bisphenol A. *Reprod Toxicol* 15:505–523.
- Ema M, Fujii S, Furukawa M, Kiguchi M, Ikka T, Harazono A. 2001. Rat two-generation reproductive toxicity study of bisphenol A. *Reprod Toxicol* 15:505–523.
- Myers JP, vom Saal FS, Akingbemi BT, Arizono K, Belcher S, Colborn T, et al. 2009. Why public health agencies cannot depend upon Good Laboratory Practices as a criterion for selecting data: the case of bisphenol A. *Environ Health Perspect* 117:309–315.
- Nagel SC, vom Saal FS, Thayer KA, Dhar MG, Boechler M, Welshons WV. 1997. Relative binding affinity-serum modified access (RBA-SMA) assay predicts the relative *in vivo* bioactivity of the xenoestrogens bisphenol A and octylphenol. *Environ Health Perspect* 105:70–76.
- OECD. 1998. OECD Series on Principles of Good Laboratory Practice and Compliance Monitoring. No. 1 OECD Principles of Good Laboratory Practice. Paris:Organisation for Economic and Co-operative Development.
- OECD (Organisation for Economic Cooperation and Development). 2001. Guideline for Testing of Chemicals. Proposal for Updating Guideline 416, Two-Generation Reproduction Toxicity Study. Paris:Organisation for Economic Co-operation and Development. Available: <http://www.oecd.org/dataoecd/18/13/1948466.pdf> [accessed 17 September 2009].
- Tyl RW. 2009a. Basic exploratory research versus guideline-compliant studies used for hazard evaluation and risk assessment: Bisphenol A as a case study. *Environ Health Perspect* 117:1644–1651.
- Tyl RW. 2009b. Letter to the Editor. *Toxicol Sci* 34(5):587–588.
- Tyl RW, Myers CB, Marr MC, Sloan CS, Castillo NP, Veselica MM, et al. 2008. Two-generation reproductive toxicity study of dietary bisphenol A (BPA) in CD-1 (Swiss) mice. *Toxicol Sci* 104(2):362–384.
- Tyl RW, Myers CB, Marr MC, Thomas BF, Keimowitz AR, Brine AR, et al. 2002. Three-generation reproductive toxicity study of dietary bisphenol A (BPA) in CD (Sprague-Dawley) rats. *Toxicol Sci* 68:121–146.