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Pathogenesis of *Escherichia coli* O157:H7 strain 86-24 following oral infection of BALB/c mice with an intact commensal flora

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Abstract

Escherichia coli O157:H7 is a food-borne pathogen that can cause hemorrhagic colitis and, occasionally, hemolytic uremic syndrome, a sequela of infection that can result in renal failure and death. Here we sought to model the pathogenesis of orally-administered *E. coli* O157:H7 in BALB/c mice with an intact intestinal flora. First, we defined the optimal dose that permitted sustained fecal shedding of *E. coli* O157:H7 over 7 days ($\sim 10^9$ colony-forming units). Next, we monitored the load of *E. coli* O157:H7 in intestinal sections over time and observed that the cecum was consistently the tissue with the highest *E. coli* O157:H7 recovery. We then followed the expression of two key *E. coli* O157:H7 virulence factors, the adhesin intimin and Shiga toxin type 2, and detected both proteins early in infection when bacterial burdens were highest. Additionally, we noted that during infection, animals lost weight and $\sim 30\%$ died. Moribund animals also exhibited elevated levels of blood urea nitrogen, and, on necropsy, showed evidence of renal tubular damage. We conclude that conventional mice inoculated orally with high doses of *E. coli* O157:H7 can be used to model both intestinal colonization and subsequent development of certain extraintestinal manifestations of *E. coli* O157:H7 disease.

Keywords

Escherichia coli O157:H7 mouse model; Shiga toxin; intimin

1. Introduction

E. coli O157:H7 is a food-borne pathogen that results in an estimated annual incidence of 73,000 cases of diarrheal illness each year in the United States [1]. Hemolytic uremic syndrome, or HUS, is the most serious sequela of *E. coli* O157:H7 infection that occurs on average in 4% of those infected [2]. However, the recent 2006 spinach-associated *E. coli* O157:H7 outbreak in the United States had a higher rate of HUS (16%) that was perhaps attributable to the emergence of a more virulent clade of *E. coli* O157:H7 [3,4]. HUS consists of a triad of

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symptoms that include microangiopathic hemolytic anemia, thrombocytopenia, and acute renal failure [5]. Although most people recover from HUS, late complications and even death can result [6].

The sequence of events by which *E. coli* O157:H7 establishes infection and causes disease are imperfectly understood. It is known that *E. coli* O157:H7 can cause illness in a person who has ingested as few as 100 organisms [7]. Therefore, at least a portion of that inoculum must be capable of surviving the acidic conditions of the stomach to colonize the intestine (reviewed in [8]). Intimate adherence of *E. coli* O157:H7 to the intestinal mucosa is considered to require expression of the bacterial outer membrane protein intimin and other factors secreted through the type III secretion system (TTSS) (reviewed in [9]).

The development of HUS is associated with the production of Shiga toxin (Stx) by *E. coli* O157:H7 and other Stx-producing *E. coli* (STEC) [10–12]. This toxin is produced by the infecting bacteria in the gut, as evidenced by the presence of Stx in the feces of *E. coli* O157:H7-infected individuals [11–13]. *E. coli* O157:H7 can produce two types of Stxs, Stx1 and/or Stx2, as well as variants of these toxin types. Stxs are AB₅ toxins that bind to cells through the pentameric B subunits via a glycosphingolipid receptor known as globotriaosylceramide, or Gb₃. Stx is an N-glycosidase that cleaves a purine residue in ribosomal 28S RNA. This catalytic event leads to inhibition of protein synthesis in the target cell and, subsequently, cell death by apoptosis (reviewed in [14]). The primary targets for Stx *in vivo* are small vessel vascular endothelial cells that express Gb₃ [15–17], such as those within the renal glomerulus [17,18]. It is not known how Stx crosses the intestinal barrier to target small vessel endothelial cells in the lamina propria of the gut (thought to be a primary event in the manifestation of hemorrhagic colitis), nor the mechanism by which the toxin enters the blood stream and travels through the circulatory system to the Gb₃-rich small vessel endothelia in the kidney.

Mouse infection models that mirror various aspects of STEC pathogenesis or disease (such as intestinal colonization, renal impairment, central nervous system damage, and death) have been developed (reviewed in [19]). While these animal systems are helpful for assessing features of STEC disease, many require that mice undergo artificial manipulations such as prolonged dietary restriction to promote colonization, mitomycin C injection to facilitate Stx expression, or antibiotic treatment to reduce the normal flora that can inhibit the establishment of an exogenous infection [20–25]. Furthermore, in antibiotic-treated or axenic animal model systems the STEC strain is likely to have a pathogenic advantage as it is no longer competing with the entire normal flora microbiome for resources. On the other hand, in a mouse model in which the normal gut flora is suppressed, the infecting *E. coli* O157:H7 strain may not receive molecular signals made by commensal bacteria that lead, through a quorum-sensing regulatory network, to TTSS expression by O157 [26]. Thus, to better reflect the typical gastrointestinal environment to which *E. coli* O157:H7 is exposed following ingestion, we sought to develop an oral model of *E. coli* O157:H7 infection in mice with an intact commensal flora (ICF).

2. Results

2.1 Dose response studies

As a first step in the development of a mouse model of *E. coli* O157:H7 infection in BALB/c mice with an ICF, we conducted dose-response experiments to determine whether inoculum size correlated with the percent of animals colonized or the level of colonization over time. For this purpose, groups of 5–10 mice were intragastrically administered approximately 10⁵, 10⁶, 10⁷, 10⁸, or 10⁹ CFU of *E. coli* O157:H7 strain 86-24Nal^R and the degree of bacterial colonization measured as defined by the number of 86-24Nal^R bacteria present per gram [colony forming units (CFU)/g] feces.

The percent of animals colonized on day 1 following infection with strain 86-24NaI^R varied by challenge dose (Fig. 1A). At an inoculum of 10⁵ or 10⁶ CFU, 40% or 80% of the mice, respectively, had detectable bacterial counts within their feces on day 1 post-infection. All animals challenged with 10⁷ CFU or greater shed detectable bacteria on day 1 of infection. The relative proportion of mice that remained colonized after day 1 was also dose-dependent (Fig 1A). Groups of animals given ≤10⁷ CFU of *E. coli* O157:H7 strain 86-24NaI^R had fewer infected mice on day 2 than on day 1, with 60% or less of the mice in those groups infected throughout the remainder of the study. Of the animals that received 10⁸ CFU, 7 of 8 mice remained colonized for the first 5 days after infection (2 of the original 10 animals died during the study and, thus, were no longer included in this analysis). At the highest inoculum of 10⁹ CFU, mice were both consistently (% of mice colonized) and persistently (prolonged, high-level colonization) infected for the 7 days that followed challenge (2 of 5 mice, both highly colonized, died on day 4 post-infection). In summary, as the inoculum dose increased there was a statistically significant increase in the percent of mice that were colonized over time (p=0.005 in a trend analysis).

Next, we assessed the differences in overall levels of colonization among the inocula groups (Fig. 1B). We found that as the challenge dose went up, the number of bacteria shed into the feces increased (p=0.001). At a low inoculum (10⁵ CFU), animals shed *E. coli* O157:H7 with a geometric mean (GM) less than 10³ CFU/g feces over the course of the experiment. The recoverable bacteria from mice infected with 10⁶ or 10⁷ CFU, while somewhat higher, still remained relatively low (GM at or below 10⁴ CFU/g feces). In comparison, animals that received an inoculum of 10⁸ or 10⁹ CFU had more recoverable bacteria on the day after infection (GMs of 6.5 × 10⁵ and 1.9 × 10⁶ CFU/g feces, respectively). Mice that received 10⁸ CFU had a statistically higher colonization level than animals challenged with 10⁵ CFU, irrespective of time (p=0.001). At the largest inoculum of 10⁹ CFU, statistically higher levels of colonization were measured compared to doses of 10⁵, 10⁶, or 10⁷ CFU (p≤0.01), again irrespective of time. However, as the overall colonization levels declined during the study these variations by inoculum dose in colonization loads eventually disappeared (except at the highest inoculum when comparing CFU/g feces on day 7 post-infection, Fig. 1B).

We next evaluated the impact on colonization of administration of 10⁸ or 10⁹ CFU of strain 86-24NaI^R to the mice by intragastric administration versus pipette feeding (orally through a micro pipette tip). We observed a more consistent infection after intragastric administration at the highest inoculum of 10⁹ CFU (p=0.023, Fig. 1C). When assessing the colonization levels in the various groups (Fig. 1D), significantly higher colonization levels were achieved at higher inocula (p=0.008), as was noted previously (Fig. 1B), regardless of method used to introduce the bacteria. Additionally, as infection progressed, colonization levels generally declined in all groups (p<0.001). When the data were averaged across all days, there was no overall difference between the procedures used to infect the animals at either dose. However, there was an indication that providing the inoculum by gavage resulted in increased levels of colonization on certain days post-infection, and this was most pronounced on day 3 (p=0.004) (Fig. 1D). Nevertheless, the most consistent and persistent colonization levels in mice were achieved with an inoculum of 10⁹ CFU regardless of the route of bacterial challenge used. Therefore, for all subsequent experiments, we orally infected mice either intragastrically by gavage or by pipette feeding with approximately 10⁹ CFU of the microbe. The inoculum method applied was largely dictated by the question under study in a particular experiment.

2.2 Sites of 86-24NaI^R colonization in the mouse intestinal tract

To determine the site of *E. coli* O157:H7 colonization within the intestines of the ICF mice after challenge with 86-24NaI^R, animals were orally inoculated with about 10⁹ CFU of the bacterium by pipette feeding. We elected to use the pipette feeding method for this study

because it is more like the natural route of exposure to the organism, ingestion, than is gavage. At various times following infection, groups of animals (n=8–17) were sacrificed, and the numbers of 86-24Nal^R bacteria associated with the tissues of the small intestine, cecum, and large intestine, as well as in the luminal contents of those intestinal segments were determined.

When the 86-24Nal^R counts of the organ tissues were compared, we found that the bacterial load was the highest in the cecum, followed by the large intestine, while the small intestine had the fewest recoverable *E. coli* O157:H7 (Fig. 2A). The number of *E. coli* O157:H7 bacteria harvested from each of the tissues dropped from 6 hours to 24 hours, and then leveled off for the remainder of the study. The number of bacteria recovered from the luminal contents showed a similar pattern to that seen for the organs themselves, with the exception that the contents of the large intestines remained high throughout the study (Fig. 2B). We also observed that the large intestine luminal contents had significantly more recoverable *E. coli* O157:H7 than did the large intestinal tissue (compare Fig 2B line with open diamonds to Fig. 2A line with solid diamonds; p<0.001). In fact, the large intestine luminal contents had equivalent levels of *E. coli* O157:H7 to those found in the cecum, the cecal contents, or both, depending on the time-point assessed. As was seen for the small intestinal tissue, the small intestine luminal contents had significantly fewer recoverable *E. coli* O157:H7 bacteria throughout the study when compared with the cecal and large intestinal contents (p<0.001). Indeed, the small intestinal contents and tissue had the lowest number of 86-24Nal^R throughout the course of infection. Overall, we observed the highest GM CFU/g in the cecal tissue, cecal contents, and large intestinal contents, and these values were similar (compare Figs. 2A and 2B).

Because the cecal tissue had the highest *E. coli* O157:H7 counts of all the tissues examined, we concluded that the cecum is the site of optimal colonization by 86-24Nal^R. Therefore, we examined cecum-associated *E. coli* O157:H7 via immunofluorescence. Early in infection, we readily found fluorescent bacteria both within the cecal lumen (the majority of fluorescent organisms) and close, perhaps adherent, to the epithelial surface of the cecal tissue (Fig. 3A–F). At later time points during the course of infection, the numbers of O157-stainable bacteria decreased (not shown). As expected, *E. coli* O157:H7 was not detected in cecal sections from uninfected control mice (Fig. 3G–I).

2.3 Kinetics of intimin and Stx2 expression in vivo at intestinal sites

To further evaluate the pathogenesis of *E. coli* O157:H7 in mice with an ICF, we monitored the expression of intimin and Stx2 over time in animals fed $6-8 \times 10^8$ CFU of strain 86-24Nal^R. Three separate studies were conducted, and the pooled results are shown in Figure 4. For these studies, we did not evaluate small intestine or small intestinal contents because 86-24Nal^R levels were low at that location in the time course study (see Fig. 2).

Intimin was detected at all sites examined (cecum, cecal contents, large intestines, and large intestinal contents) from mice sacrificed at 3, 6, and 9 hours post-infection (Fig. 4A). A subset of tissue and tissue content samples were examined for absolute levels of intimin by comparison with values from a standard curve derived from control tissue spiked with different amounts of intimin (Fig. 4B). In this subset of specimens, the highest intimin values were detected within the cecum, cecal contents, and large intestinal contents. These latter findings likely reflect our earlier observations that the largest numbers of recoverable bacteria were found in these locations early in infection (see Fig. 2). Intimin was minimally detected at 24, 48, and 72 hours post-infection (Fig. 4A and B), most likely due to the declining numbers of recoverable CFU/g sample after 24 hours of infection (see Fig. 2).

The presence of Stx2 in the cecum, cecal contents, large intestines, and intestinal contents was first assessed by enzyme-linked immunosorbent assay (ELISA). Most of the samples positive for toxin were obtained during the first 9 hours of infection (Fig. 4C). We found the luminal

contents to have higher detectable toxin levels than the organ tissues when we measured absolute values versus a positive control in a subset of samples ($p \leq 0.002$) (Fig. 4D). Furthermore, we saw the highest detectable levels of toxin in the large intestinal contents ($p \leq 0.006$). In general, the locations where we found higher toxin levels corresponded to intestinal sites that had the highest numbers of recoverable organisms (compare Fig. 2 and Fig. 4C). To ensure that the positive results we obtained by ELISA were due to Stx2, we evaluated a subset of the samples for Stx2 by a neutralization assay (not shown). The findings in the neutralization assay and the ELISA were similar; toxin was most readily detectable early in infection and within the contents of the organs tested (the cecum and large intestine).

2.4 Systemic effects of infection

Over the course of longer infections, we noticed that a proportion of the animals that were fed 86-24Nal^R exhibited signs of illness (hunched posture, ruffled fur, lethargy, tremors, etc.) and succumbed to infection. To more closely examine the systemic effects of *E. coli* O157:H7 infection in mice with an ICF, groups of animals were infected by gavage with 10^9 CFU and monitored both for weight loss (indicative of illness) and death. We elected to use gavage for infection because we wanted to increase the likelihood that all animals would be colonized. All animals demonstrated a drop (about 1.5 grams) in weight from day -1 to day 0, a phenomenon we ascribed to overnight deprivation of food prior to infection (Fig. 5A). However, the mice returned to near pre-infection weights at day 1 of infection after they were permitted access to food and water *ad libitum*. In fact, even after *E. coli* O157:H7 inoculation, all animals gained weight between days 1 and 2. After day 2 of infection, there was a statistically significant divergence in weight of the control and infected animals. Indeed, the control mice continued to gain weight until day 3 when, as a group, their weight plateaued and remained steady at around 18 grams. Conversely, the experimental group started to lose weight after day 2 post-infection and weighed significantly less than did the control mice on days 3 to 5 post-infection ($p \leq 0.001$; Fig. 5A). Moreover, infected mice that demonstrated large amounts of weight loss succumbed to infection on days 5 and 6 post-*E. coli* O157:H7 challenge. In this experiment, 30% of the infected mice died, and, as expected, all control mice survived (Fig. 5B).

We then asked whether the mouse mortality seen after intragastric administration of *E. coli* O157:H7 could have resulted from bacteremia following gavage. To address this question, 20 mice were infected with 10^9 CFU of 86-24Nal^R intragastrically and monitored for mortality and the development of bacteremia. At 6 hours post-infection, blood was taken from each mouse and assessed for the presence of circulating bacteria. Five of the animals demonstrated the presence of bacteria in their blood. These same five animals died by 24 h post-infection. The remaining mice were divided into two groups: a group to check for the presence of bacteremia on days 2 and 4 post infection and a mortality group. The animals examined for bacteremia on day 2 post-infection demonstrated no detectable bacteria in their blood. Two animals from the bacteremia-assessment group were sacrificed on day 3 because they were moribund. These latter two animals had symptoms of disease that included neurological manifestations (ataxia, tremor, and convulsions), yet had no detectable bacteria in their blood. The final two animals from the bacteremia-assessment group were sacrificed on day 4 and were not bacteremic. In the mortality group, animals lost weight over time; furthermore, deaths (4/7 mice or 57% mortality) occurred on days 4 and 5 post-infection. All of the animals in the mortality group that succumbed to infection were free of circulating bacteria in their blood as determined post mortem. Thus, bacteremia was not seen, even in extremely moribund mice, after the first day of infection. Moreover, in a second study using a lower inoculum ($\sim 5 \times 10^8$ CFU 86-24Nal^R), no bacteremia was detected in any of the mice at 6 hours post-infection. In this latter study, the bacteremia-assessment group also had no detectable bacteria in the blood at day 2 or 4. The mortality group had a 20% death rate with no evidence of bacteremia

6 hours following gavage or post mortem. We concluded that at high challenge doses of 86-24NaI^R given by gavage, animals that succumb after day 2 post-infection are not dying as a consequence of bacteremia.

Since we observed deaths in mice infected by gavage with high doses of 86-24NaI^R in the absence of bacteremia, we speculated that this mortality was due to Stx2 produced by *E. coli* O157:H7 in the gut. Indeed, we have shown that streptomycin-treated mice that would normally die after oral STEC infection remain healthy if passively treated with neutralizing anti-Stx2 antibodies [25,27,28]. Here, we sought to demonstrate the central role of toxin in lethality after *E. coli* O157:H7 infection in an ICF model by assaying for the presence of Stx2 in the blood of animals gavaged with 10⁹ CFU of 86-24NaI^R. However, blood taken from animals that were infected with *E. coli* O157:H7 showed either no detectable toxin or sub-picogram levels of toxin (found in the blood of 2/15 mice). We next asked if it was possible to detect intravenously (IV) administered Stx2 in the blood of uninfected mice. We found that it was necessary to administer 100 – 1 000 LD₅₀s (100 – 1 000 ng) of Stx2 in order to detect toxin in blood taken from mice 2 hours post-intoxication. The amount of Stx2 detected in the blood from these highly intoxicated animals was 0 – 17 pg per 0.1 mL. Therefore, we surmised that the reason we did not detect Stx2 in the blood of more than 2 infected mice is most likely because the levels of toxin in the blood were below 17 pg/0.1 mL.

To further examine *E. coli* O157:H7 pathogenesis in mice with an ICF, animals were sacrificed on various days post-infection, blood was collected, and kidneys were surgically removed. Sera were analyzed for blood urea nitrogen (BUN) and creatinine levels, and whole blood samples were used to determine complete blood counts (CBCs, Table 1). When compared to the uninfected controls, *E. coli* O157:H7-infected mice showed significantly higher BUN ($p<0.01$) values and higher, but not statistically significant, creatinine levels ($p=0.07$) over the course of infection. The greatest elevations in these kidney function values were seen in animals sacrificed on days 4 and 5 (individual day data not shown). However, none of the key hematological features of HUS were apparent in the infected animals. Specifically, no differences in platelets, hemoglobin, or hematocrit values were noted between infected and control mice. Nevertheless, significant neutrophilia ($p=0.001$) was apparent in the blood samples of the experimental group compared to the controls (Table 1). In fact, as early as day 2 following infection, the blood of infected mice showed an increase in percentage of neutrophils that was evident throughout the experiment (days 2, 3, 4, and 5) with differences of up to 4-fold compared to controls (data not shown).

We then examined the histopathology of the kidneys from the *E. coli* O157:H7-infected and control animals whose blood samples were assayed as above. We found no evidence in any of the kidney sections from infected or control mice of lesions in the renal glomeruli (as assessed by H&E stain), or fibrin deposits in the glomerular capillaries or interstitial vessels [as evaluated by Mallory's phosphotungstic acid hematoxylin (PTAH) or a modified Carstairs' stain]. Additionally, we probed the kidney sections for the presence of Stx2. While we detected toxin in the kidney by immunofluorescence (IF) in one mouse that died following infection (not shown), that result was not duplicated in a second, moribund animal. That we found only minute amounts of circulating toxin in the blood also suggests that the quantity of Stx2 present for detection by IF in the kidneys is quite small. In contrast to the lack of glomerular changes in infected animals, histological evidence of injury to proximal and distal convoluted tubules was noted in kidneys from infected animals (Fig. 6A and B) but not in kidneys from control mice (Fig. 6C). Indeed, kidneys from several infected mice exhibited acute to subacute, multifocal, minimal to mild tubule degeneration, small amounts of epithelial necrosis, and varying degrees of regeneration.

3. Discussion and conclusions

3.1 Discussion

In this investigation, we developed a model of *E. coli* O157:H7 intestinal colonization and disease in BALB/c mice with an ICF. Although a large dose of *E. coli* O157:H7 was required in this murine model, high inocula of these organisms are also needed to achieve colonization and disease in monkeys [29] and rabbits [30]. Our data suggested that the predominate site of *E. coli* O157:H7 colonization was the cecum, although numerous challenge bacteria were also culturable from the cecal and large intestinal contents. Furthermore, mice challenged intragastrically with 10^9 *E. coli* O157:H7 often displayed such manifestations of disease as lethargy and weight loss, neutrophilia and increased levels of BUN. Moreover, about 30% of animals inoculated with *E. coli* O157:H7 by the intragastric route died. Finally, we observed evidence of tubular regeneration and some tubular damage in the kidneys of infected mice. We interpreted these findings to mean that damaged tubules were in the process of repair. Of note, tubular damage, in addition to signature glomerular lesions [17,31], have been seen in kidney sections from patients with HUS [32–34]. Furthermore, tubular necrosis and subsequent regeneration were described by Tesh and colleagues in mice that died after toxin administration [35].

The large numbers of *E. coli* O157:H7 strain 86-24Nal^R found in the cecum, cecal contents, and large intestinal contents of infected mice support the hypothesis that these organisms colonize the cecum and are then shed into the cecal contents before they pass into the large intestinal contents, where they are ultimately bound in fecal material and expelled. These results in aggregate indicate that the CFU/g feces formed from the contents of the large bowel most likely reflect the extent of organism replication in the cecum of the animal. This theory is consistent with the suggestion in the report by Nagano *et al.* that *E. coli* O157:H7 found in the large intestines and feces are shed from bacteria adherent in the cecum [36].

In addition to identifying the site of *E. coli* O157:H7 colonization in the ICF model, we detected intimin in the same areas as the bacteria early during *in vivo* infection but not readily thereafter. This pattern could have reflected down-regulation of intimin expression *in vivo*, as has been documented for the expression of intimin *in vitro* by a related organism, enteropathogenic *E. coli* (EPEC) [37]. However, because we also observed a significant decrease in bacterial numbers over this same time period, we could not ascertain whether *E. coli* O157:H7 intimin was down-regulated *in vivo* or if the reduced bacterial load meant that the intimin levels fell below the limit of detection for our ELISA.

As was seen with intimin, we observed the highest levels of Stx2 early in *E. coli* O157:H7 infection, and were only occasionally able to detect toxin in intestinal samples taken later during infection when total recoverable bacterial counts were low. Thus, our inability to detect toxin later in infection probably reflects the reduced bacterial numbers at that time in infection. Cornick and colleagues observed a similar phenomenon when they assessed colonization and toxin production within the small intestines of swine infected with an STEC [38]. They found the highest toxin titers when the greatest number of STEC were present in the ileum. In our study, the mice manifested systemic effects of disease, such as significant weight loss, despite the lower levels of toxin detected after 9 hours of infection. Additionally, mortality in infected animals did not occur until 3 to 6 days post-infection. Together these data suggest that a lag period occurs between the time of maximal toxin production in the gastrointestinal tract to the manifestation of clinical disease. Cornick and colleagues also found a lag period between toxin expression and disease manifestations in the pig [38]. During this lag period, toxin may transit from the site of infection to the bloodstream and then to the kidneys where it damages the cells within the tubules. That we and others have observed a delay between finding toxin in the

intestine and manifestations of systemic disease [23,38] suggests that therapeutic intervention may be possible during this interval (Melton-Celsa unpublished data, [39–41]).

Investigators from our laboratory previously reported that attainment of high-level STEC colonization and subsequent mortality in orally-infected mice necessitated the use of an antibiotic, i.e. streptomycin, to suppress the normal intestinal flora [25]. Subsequently, three other groups of investigators, in addition to ourselves, succeeded in the establishment of an STEC infection (using *E. coli* O157:H7) in mice in the absence of antibiotics [28,36,42,43]. However, prior to the study reported here, only Karpman and colleagues successfully administered *E. coli* O157:H7 to untreated mice with morbidity or death as a reported outcome [28]. These researchers, who did not monitor colonization, observed neurological and systemic illness in C3H/HeJ and C3H/HeN mice after gavage with strain 86-24 (or the non-toxin-producing *E. coli* O157:H7 strain 87-23) that included gastrointestinal damage, some loose stools, tubular and glomerular damage, and some fragmentation of blood cells (although the red cell fragmentation was not associated with Stx2 production in their study) [28]. The disparity in degree of systemic illness between the BALB/c mice used in our study and the C3H/HeN and C3H/HeJ mice used by Karpman and colleagues may reflect genetic differences in the mouse strains; or alternatively, a difference in the type or extent of their commensal gut flora that could impact the extent of colonization by the challenge *E. coli* O157:H7 strain. Inherent in the latter explanation is our presumption that mice colonized to a higher level with *E. coli* O157:H7 are more likely to display higher morbidity and perhaps mortality. Since both the Karpman and our studies utilized *E. coli* O157:H7 strain 86-24 (though our strain was NaI^R), the differences in the models should not reflect bacterial strain differences. Additionally, over half of the C3H/HeN and C3H/HeJ mice gavaged with strain 86-24 demonstrated positive blood cultures just prior to sacrifice. However, bacteremia is a feature not observed in humans with *E. coli* O157:H7 disease [44].

In our study that examined the impact of *E. coli* O157:H7 strain 86-24NaI^R administered by gavage on subsequent bacteremia in mice, we found positive blood cultures in a small portion of the mice after introducing 10⁹ CFU. Those mice with positive blood cultures for *E. coli* O157:H7 died within 24 hours of inoculation. However, none of the remaining animals in that same study were bacteremic, even when they appeared extremely moribund or died. Three lines of evidence support the hypothesis that *E. coli* O157:H7-infected mice that died 2 or more days after gavage with high doses of *E. coli* O157:H7 did so as a consequence of Stx2 delivered systemically from the gut. First, we found sub-picogram levels of Stx2 in the blood of some infected animals. Second, we detected Stx2 in the kidneys of one mouse (not shown). Third, we observed damage to the Gb3-rich [45] renal tubules, a pathology also seen after STEC infection in humans [32,34].

As noted above, two other groups of investigators reported establishment of *E. coli* O157:H7 infection in untreated mice but, unlike the original Karpman report or our study here, morbidity or mortality were not outcomes of infection. In 1998, Conlan and Perry showed that specific pathogen-free mice (CD-1, BALB/c, and C57BL/6) could be colonized for an average between 7 and 24 days following intragastric administration of greater than 10¹⁰ CFU of *E. coli* O157:H7 [42]. In a more recent study, Nagano and colleagues investigated *E. coli* O157:H7 colonization, in the absence of antibiotic treatment, in a number of inbred mouse strains that included BALB/c, C3H/HeN, C3H/HeJ, as well as out-bred ICR (also called CD-1) animals [36]. The mice were challenged with as much as 10¹¹ CFU introduced by gavage, and animals were given cimetidine, an H2 blocker, prior to infection to reduce the acidity of the stomach. Nevertheless, Nagano and colleagues were unable to detect *E. coli* O157:H7 shed in the feces from the majority of BALB/c mice with an intact commensal flora one week after infection but did find that ICR mice remained colonized in the absence of antibiotic treatment 7 days following challenge. While the reason we observed persistent colonization in ICF BALB/c mice when

Nagano *et al.* did not remain unclear, two possibilities may account for the varied findings. First, and likely most importantly, Nagano and colleagues used a different strain of *E. coli* O157:H7, GPU96MM (an Stx1- and Stx2-producing strain), than did we in this investigation. The *E. coli* O157:H7 strain used by Nagano *et al.* was associated with an outbreak in which patients had lower hospitalization and mortality rates than the illness linked to *E. coli* O157:H7 strain 86-24 used here [46,47]. Furthermore, our laboratory has previously shown that bacterial strain-specific differences can influence the colonization pattern seen in mice [25]. Second, the level (or type) of commensal flora in the gastrointestinal tracts of the BALB/c mice bred by the supplier used by Nagano *et al.* may have been different than the gut flora in the BALB/c mice that we obtained. Therefore, our current observation that a high initial inoculum of a virulent *E. coli* O157:H7 is necessary to attain elevated levels of the organism and persistent colonization of ICF BALB/c mice is most likely explained by competition for an intestinal niche between the in-coming *E. coli* O157:H7 strain and the already established resident bowel flora.

3.2 Conclusions

We established a model of *E. coli* O157:H7 oral infection in BALB/c mice with an intact commensal flora. Our data suggest that the following steps occur in the BALB/c ICF model of 86-24Nal^R infection: 1) the bacteria are ingested; 2) the bacteria colonize within the cecum and to a lesser extent, the large intestine; 3) Stx2 enters the blood stream from the site of *E. coli* O157:H7 colonization within the gastrointestinal tract; and, 4) toxin travels to the kidney, and either directly or indirectly causes renal damage and altered kidney function as seen in HUS. However, the absence of hemolytic anemia or thrombocytopenia indicates that not all the features of HUS are recapitulated in this ICF mouse infection model. Nevertheless, we believe that our small animal model will serve as a useful tool to assess various means to impact colonization levels or persistence of colonization and evaluate treatments that block or ameliorate the systemic, likely Stx2-mediated features of *E. coli* O157:H7 disease. We also speculate that this infection model can be readily applied to examine the relative colonizing capacity and virulence of different strains or clades of *E. coli* O157:H7.

4. Materials and methods

4.1 Bacterial strains and growth conditions

A nalidixic acid-resistant derivative of *E. coli* O157:H7 strain 86-24, referred to as 86-24Nal^R, was used for all experiments described herein (kindly provided by Dr. Arthur Donohue-Rolfe of Tufts University). The parental strain 86-24 produces Stx2 and was isolated in Washington State during a 1986 outbreak associated with contaminated beef products [47]. For all mouse infection studies, a sample of 86-24Nal^R was obtained from a freezer stock of the organism and inoculated onto a Luria Bertani (LB) agar plate supplemented with nalidixic acid (25 µg/mL). The plate was then incubated overnight (O/N) in air at 37°C. A single isolated colony was picked from the agar plate and inoculated into LB broth with nalidixic acid (25 µg/mL). The cultures were then grown O/N with aeration. The bacteria were harvested from the broth by centrifugation, and the pellets resuspended in 20% glucose-phosphate-buffered saline (PBS) to a 40 – 100× concentration.

4.2 Mouse infection studies

Six-week-old female BALB/c mice from Charles River Labs (Wilmington, MA) were used for all animal experiments. The mice were permitted food and water *ad libitum* except prior to *E. coli* O157:H7 infection. Food was taken from the animal cages the night before bacterial challenge. Water bottles were removed from the cages 2 hours prior to infection of the mice. Following infection, access to food and water was restored.

4.2.1 Experimental series #1—Dose-response studies were done to determine the optimal infectious dose of 86-24Nal^R required to permit sustained *E. coli* O157:H7 colonization of mice over 7 days. Groups of 5–10 animals were inoculated by intragastric administration, also referred to as gavage, with approximately 10⁵, 10⁶, 10⁷, 10⁸, or 10⁹ colony-forming-units (CFU) of strain 86-24Nal^R in 100 µL. In a second set of studies, animals were orally infected by pipette feeding 10⁸ or 10⁹ CFU of bacteria. Pipette feeding (where animals were allowed to ingest the bacterial suspension by lapping the liquid from the end of a micropipette tip) was done in two rounds of 50 µL each with 2 hours between feedings to facilitate ingestion of the complete dose. The extent of bacterial colonization was monitored daily for 7 days by quantitation of the *E. coli* O157:H7 shed into fecal pellets. For that purpose, animals were placed individually into clean, empty cages, allowed to defecate, and the feces collected and weighed. The fecal material was diluted 1:10 by weight into PBS then homogenized by mechanical disruption with a sterile wooden stick. Large debris was pelleted by brief centrifugation at 1 750 × g with an International Equipment Company clinical centrifuge (IEC Model CL, Damon IEC Division, Needam Hills, MA). The supernatants were diluted and plated onto Sorbitol MacConkey Agar (SMAC) plates supplemented with nalidixic acid (SMAC+Nal, 25 µg/mL) to determine CFU/g feces. Reported values represent the geometric mean (GM) of 5 or more animals per group.

4.2.2 Experimental series #2—Short-term infection studies were conducted to monitor levels of *E. coli* O157:H7 at various intestinal sites after challenge and to obtain organs for immunostaining studies. Animals were orally infected by pipette feeding of 50 µL containing 6–8 × 10⁸ CFU of strain 86-24Nal^R in three separate experiments. Control animals (generally 1 per time-point per experiment, with the exception that an additional control animal was required for the immunostaining experiments) were orally administered 20% glucose-PBS alone by pipette feeding. Groups of infected mice and control animals were anesthetized in isoflurane and euthanized by cervical dislocation at 3, 6, 9, 24, 48, and/or 72 hours after infection or buffer inoculation. Whole organs (cecum and kidneys) from at least one infected animal and one control animal at each time-point were removed and fixed in Formalde-Fresh (10% buffered formalin, Fisher Scientific, Pittsburgh, PA), then sectioned, placed on slides, and immunostained for detection of O157 (described in section 4.5 below). In addition, the cecum, large intestine, and a terminal section of the small intestines equal in length to that of the large intestine were surgically removed from the infected and control animals at each time point. The luminal contents were then removed from each organ by application of gentle pressure to the outside of the tissue to expel the internal material. Both the luminal contents and the organs were weighed and homogenized separately (Omni hand-held homogenizer) in 0.5 mL of PBS. The homogenates were then diluted (final dilution of 1:10 by weight) and samples were plated on SMAC+Nal plates for enumeration of CFU. An aliquot of each diluted homogenate was also treated with a protease inhibitor (complete, Mini, EDTA-free, Roche Applied Science, Indianapolis, IN) and stored at –20°C for subsequent analyses of intimin and Stx2 expression as detailed below in sections 4.3 and 4.4. Values for bacterial counts were reported as the combined GM from all experiments and represent a total of 8 or more (up to 17) animals per time-point analyzed.

4.2.3 Experimental series #3—For this multi-tiered investigation, mice were divided into 4 groups. Group A included infected experimental animals for blood work and tissue harvest (n=25). Group B contained uninfected control mice for blood work and tissue harvest (n=20). Group C consisted of infected experimental mice for morbidity studies (n=10); and, group D was comprised of uninfected controls for the morbidity study (n=10). *E. coli* O157:H7 counts in shed feces were tracked after intragastric administration of a concentrated culture of strain 86-24Nal^R in 20% glucose to groups A and C, or 20% glucose to groups B and D. Animal weights were measured as an indicator of morbidity in all groups. Based on colonization levels

and weight loss, five animals were selected on each of days 2, 3, 4, and 5 from the experimental group A, as well as five animals per day from the control group B. Blood was collected from these mice by exsanguination and placed into either microtainer tubes that contained potassium ethylenediaminetetraacetic acid (EDTA, BD diagnostics, Franklin Lakes, NJ) for CBC evaluation or into microcentrifuge tubes to process to obtain sera for kidney function analyses. For the latter purpose, blood in each microcentrifuge tube was first allowed to clot O/N at 4°C. The tubes were then subjected to two consecutive 10 minute centrifugation steps at 8 000 × g then ≥10 000 × g to separate the serum from the clotted red cells. Serum samples were stored at 4°C until they could be analyzed by the Uniformed Services University of the Health Sciences (USUHS) Diagnostic Services & Comparative Medicine Clinical Pathology laboratory (Bethesda, MD) for levels of blood urea nitrogen (BUN) and creatinine. Whole blood samples for determination of complete blood counts (CBC) were similarly stored and evaluated by the same facility.

The animals from which blood was obtained as above were then sacrificed by cervical dislocation. Their kidneys were removed at necropsy. These organs were then placed in the fixative Carnoy (60% ethanol, 10% glacial acetic acid, 30% chloroform) for 2 hours or Formalde-Fresh overnight, and then transferred to 70% ethanol for subsequent histopathological examination (described in section 4.5 below).

4.2.4 Experimental series #4—The possibility that deaths of mice infected with high doses of 86-24Nal^R by gavage may have occurred as a consequence of bacteremia induced by the procedure was evaluated. For those assessments two different infecting doses were used and two evaluation groups (blood evaluation and mortality assessment) were defined with 10 animals in each group. The mice were infected with ~2 × 10⁹ CFU (groups A and B) or ~5 × 10⁸ CFU (groups C and D) of 86-24Nal^R by gavage. Six hours post-inoculation blood was collected from the tail vein of all of the mice to test for bacteremia. On days 2 and 4 post-infection, five animals from the blood evaluation groups (B and D) were anesthetized in isoflurane and sacrificed by cervical dislocation. Blood was collected directly from the heart by cardiac puncture, and assessed for the presence of bacteria. On a few occasions, no blood was recovered by cardiac puncture and, in those instances, blood was taken from the thoracic cavity or the heart was cut open and the tissue smeared onto SMAC+Nal plates. The mortality groups (A and C) were followed for 10 days and upon death, blood and/or heart tissue was examined for bacteremia. If an animal appeared severely moribund, it was immediately sacrificed. Blood collected from each mouse (1–10 μL) was inoculated into LB broth that contained nalidixic acid. The broth was grown for 48 hours statically at 37°C and tested daily for the presence of bacteria by subculture onto SMAC+Nal plates. Additionally, the broth was visually assessed for turbidity. Blood (from the cardiac puncture or directly from the heart) was also plated directly on SMAC+Nal plates and grown overnight at 37°C.

4.2.5 Experimental series #5—Prior to infection, blood was obtained by tail bleed and collected into tubes containing heparin or lithium and heparin. Mice were infected by gavage with 1.9 × 10⁹ CFU of 86-24Nal^R. Colonization and weight were monitored. A group of mice (n=10) were followed for mortality. A separate group of mice (n=15) were sacrificed on days 4 and 5 post-infection with blood taken by retro-orbital eye bleed. The method used to test blood samples for toxin was Meridian's Premier EHEC enzyme-linked immunosorbent assay (ELISA) kit (as detailed in section 4.4 below). Blood cells were gently lysed by 3 rounds of freezing at -80°C and thawing at 37°C prior to use in the ELISA. This procedure was selected because we found in preliminary studies that blood, neat or diluted 1:50, interfered with interpretation of the Vero cell cytotoxicity assay, another test for Stx activity (described in section 4.4, below). The amount of Stx2 present in the blood was quantitated by comparison to a standard curve generated by adding purified Stx2 into whole mouse blood and gently lysing the blood/toxin mixture.

To determine the amount of Stx2 in the blood necessary for a positive result in the ELISA, groups of mice ($n = 2$) were administered purified Stx2 intravenously (IV). Prior to infection, blood was obtained by tail bleed and collected into tubes containing heparin or lithium and heparin. The groups then received 1, 10, 100, or 1 000 LD₅₀s of Stx2 (corresponding to 1 ng, 10 ng, 100 ng, or 1 µg of Stx2). Two hours post-injection, blood was collected from the mice by tail bleed. Lysed blood was tested for toxin using Meridian's Premier EHEC ELISA kit (as described above).

4.3 Detection of intimin by ELISA

A sandwich ELISA was developed to detect intimin in homogenates (see section 4.2.2) of the cecum, large intestine, or contents of those organs from 86-24NaI^R-infected mice. The capture antibody in the assay was a goat monospecific anti- *E. coli* O157:H7 intimin antibody that had been purified from polyclonal goat anti-intimin sera [48] by affinity chromatography over a column prepared by coupling full length, purified intimin to Pierce Ultralink Biosupport resin (Thermo Fisher Scientific, Rockford, IL) per the manufacturer's protocol. For the ELISA assay, wells of an Immulon 2HB (Thermo Scientific, Waltham, MA) high-binding microtiter plate were coated overnight at 4°C with 100 µL of a 1:50 dilution of affinity-purified monospecific goat anti-intimin antibody in a buffered solution of sodium bicarbonate/sodium carbonate. Next, 200 µL of 5% bovine serum albumin (BSA) in PBS-Tween 0.05% (PBST) was added to each well to block un-bound sites on the wells, and the plates were incubated at 4°C for at least 24 hours. The wells were washed 4–6 times with PBST and then homogenate samples (further diluted 1:2 from the original 1:10 dilution for a final dilution of 1:20) were added to the wells. The plates were incubated for 2 hours at 37°C, and the wells were then washed as above to remove any unbound sample. Intimin present within the homogenate and bound to the goat anti-intimin capture antibody was detected by incubation for 1 hour at room temperature with 100 µL of each of the following reagents in sequence with wash steps between each addition: a 1:5 000 dilution of rabbit anti-intimin polyclonal antibody [48] followed by a 1:5 000 dilution of horse radish peroxidase (HRP)-labeled donkey anti-rabbit IgG (H+L) antibody that was reported by the supplier (Jackson ImmunoResearch Laboratories, Inc., West Grove, PA) to display minimal cross reactivity with sera from other species of animals. Bound antibody was detected with a 3,3',5,5'-tetramethylbenzidine (TMB) peroxidase enzyme immunoassay (EIA) substrate kit (Biorad Laboratories, Hercules, CA) according to the manufacturer's protocol. The reaction was stopped with 100 µL of 1N sulfuric acid, and each well was read at an optical density (OD) of 450 nm on a spectrophotometric plate reader. An infected tissue or tissue content homogenate sample was considered to be positive for intimin if it had an optical density reading twice that of the average value for all the uninfected control tissue and tissue content homogenate samples. Quantitative values for intimin in samples from one of the three the experiments were derived from a standard curve constructed from ELISA values of dilutions of purified intimin spiked into control tissue homogenates. The limit of detection for the analysis was 360 ng/g tissue.

4.4 Stx assay procedures: ELISA, Vero cell cytotoxicity, and neutralization of activity assays

A commercially-available ELISA kit (Premier EHEC ELISA, Meridian diagnostics, Cincinnati, OH) was used to detect Stx2 in homogenates (see section 4.2.2) of the ceca, large intestines, or contents of those organs from 86-24NaI^R-infected mice. The ELISA kit was used according to the manufacturer's protocol with the following modifications: homogenates were diluted 1:2 in PBS (for a final dilution of 1:20) and then added directly to the ELISA. A sample was considered to be positive for Stx2 if it had an optical density reading twice the average value of the control tissues in aggregate. Stx2 levels in samples from one of the three experimental sets were determined based on a standard curve of purified toxin spiked into control tissue homogenates. In our hands, the limit of detection for quantitation with this assay was 6.4 ng toxin/g sample homogenate.

The presence of toxin in a subset of those samples was confirmed with a biological activity assay (Vero cell cytotoxicity and neutralization assay). To measure neutralizable toxin within the samples, a 1:100 final dilution of each homogenate was prepared in PBS and the diluted samples were then clarified at $8\,000 \times g$ for 10 minutes. The resultant supernatants were filtered first through a $0.8\ \mu\text{m}$ or $0.45\ \mu\text{m}$ syringe filter and then through a $0.2\ \mu\text{m}$ low-protein-binding syringe filter to remove bacteria and debris. Next, the filtrates were incubated in the presence or absence of a 1:5 000 (final concentration of 1:10 000) dilution of polyclonal rabbit anti-Stx2 antibody [49] for ~2 hours (at a 1:1 mixture of filtrate to antibody dilution or PBS) prior to the addition of 100 μL of the samples onto Vero cells that had been seeded into microtiter plates 24 hours previously. The overlaid Vero cell plates were incubated for 40–48 hours at 37°C in 5% CO_2 . The residual cells in the wells were then fixed in Formalde-fresh, stained with crystal violet, and read at 600 nm on a spectrophotometric plate reader.

4.5 Histopathology and staining of tissues

Kidneys and ceca were harvested from infected or control mice at necropsy, fixed (see section 4.2.2 and 4.2.3), and subsequently sent to HistoServ (Germantown, MD) for embedding in paraffin, sectioning, and placement on charged glass slides. Some sections were stained using hematoxylin and eosin (H&E) or a modified Carstairs's stain at HistoServ or stained using Mallory's phosphotungstic acid hematoxylin (PTAH) at the USUHS Laboratory of Animal Medicine (Bethesda, MD).

To locate *E. coli* O157:H7 bacteria bound to or near the surface of cecal tissue, slides with cecal sections were immunostained using an anti-O157 antibody. To reduce the background autofluorescence normally present in tissue, slides of sectioned tissues were first placed under fluorescent light for at least 48 hours [50]. Slides were then deparaffinized in HistoClear (National Diagnostics, Atlanta, GA) and rehydrated in a graded ethanol series. To increase antibody recognition, slides were treated with antigen retrieval buffer, pH10 [(5 \times AntigenPlus Buffer, pH10, EMD Biosciences, San Diego, CA) diluted to 1 \times , heated in a microwave for ~15 min with replenishment of the buffer as needed, cooled and dried]. Slides were blocked overnight in fetal bovine serum (FBS) supplemented with 0.1% Triton-X. Prior to the addition of primary antibody, the anti-O157 antibody was precleared against a laboratory strain of *E. coli* (DH5 α) and a powdered homogenate of normal mouse cecal tissue [51]. The cleared anti-O157 antibody was incubated on the tissue at a dilution of 1:500 in 3% bovine serum albumin (BSA) in PBS with 0.1% Triton-X for ~1h. The tissue sections were then washed and secondary goat anti-rabbit antibody conjugated to Alexa-fluor 488 (also pre-cleared against the cecal powder) was incubated on the tissue for ~1h at a dilution of 1:500 in 3% BSA in PBS with 0.1% Triton-X. Prior to microscopic observation, the slides were rinsed in PBS, mounted using Slowfade reagent, and a coverslip applied to each slide. Immunofluorescence of stained tissue sections was visualized with an Olympus BX60 microscope with a BX-FLA fluorescence attachment. Digital images of the fluorescence stains were obtained using a SPOT RT charge-coupled-device digital camera (Diagnostic Instruments, Inc., Sterling Heights, MI). Image J and DeconvolutionJ software (both developed by the National Institutes of Health) were used for processing the images and removing out-of-focus fluorescent signals. The images were overlaid and subjected to green false-coloration using Adobe Photoshop (Adobe Systems, Inc., San Jose, CA).

4.6 Statistical analyses

All statistical analyses were done through application of the SPSS v16 software (SPSS Inc., Chicago, IL). Specific analyses are described below.

Data for the dose response studies (section 4.2.1) were analyzed by both Kaplan-Meier analysis and repeated measures (RM) analysis of variance (ANOVA). The Kaplan-Meier analysis was

performed to ascertain differences in the amount of time mice were colonized by different doses of inoculum received. The RM ANOVA was used to assess statistical differences in levels of colonization by dose administered. The ANOVA was estimated by a linear mixed models approach that incorporated all available data including information from animals that had died before the end of the study. The model assumes equal correlation among repeated observations on the same animal. For the mixed model analysis, dose was considered the between subjects factor and day was set as the within subjects factor.

A repeated measures ANOVA was also used to test for statistical significance among the numbers of *E. coli* O157:H7 in various segments of the intestine over time (section 4.2.2) but, in this case, with time as a between subjects factor and organ as a within subjects factor. To more closely meet the assumptions of ANOVA, the dependent variable (CFU) was assessed on a log scale. After checking for significance by repeated measures ANOVA, we conducted a separate one-way ANOVA to compare bacterial counts at time-points among the organs. We also conducted repeated measures ANOVA for organs at each time-point and made separate pair-wise comparisons.

For the ascertainment of differences in expression of both intimin and toxin, a RM ANOVA was again used. To more closely meet the assumption for a normal distribution, the measurements for quantification of expression (the dependent variable) were assessed on a log scale. Measurements below the limit of detection were given an arbitrary value of 100 to allow the data to be included in the analysis. For the RM ANOVA, time was once again the between subjects factor and organ was again the within subjects factor. Pair-wise comparisons of the main effects for differences both by time and organ were also conducted using the least significant difference, a procedure that is equivalent to no adjustments for multiple comparisons.

Statistical differences in the mean weight between infected and control mice (section 4.2.3) were determined using a RM ANOVA. For the RM ANOVA, day was the between subjects factor and weight was the within subjects factor. Additionally, differences between infected and control mice (section 4.2.3) in terms of CBC analyses and serum chemistry results were assessed by performing independent samples t-tests on relevant numbers, such as BUN, creatinine, % neutrophils, % lymphocytes, hemoglobin, % hematocrit, and platelets.

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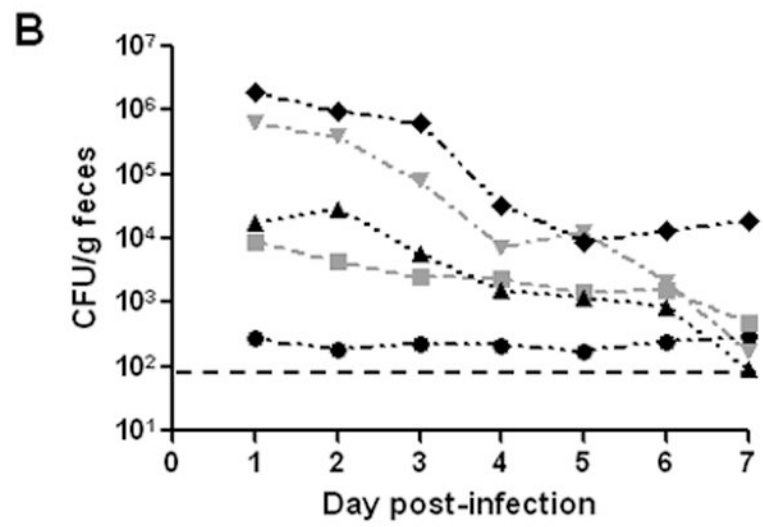
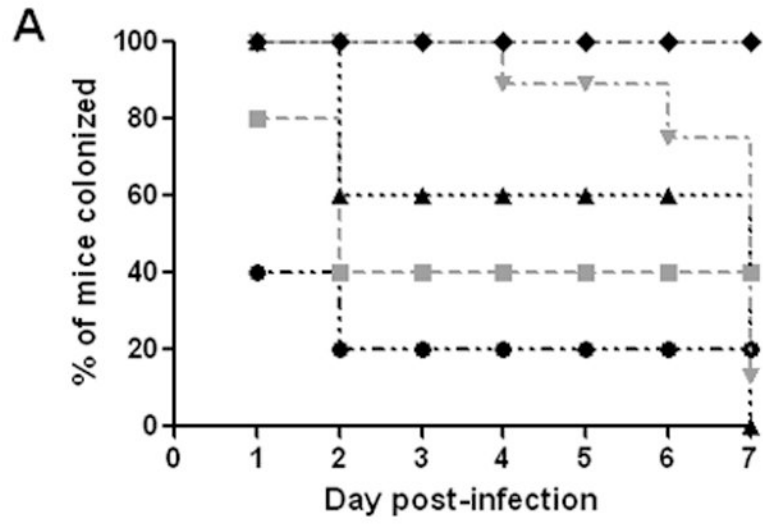
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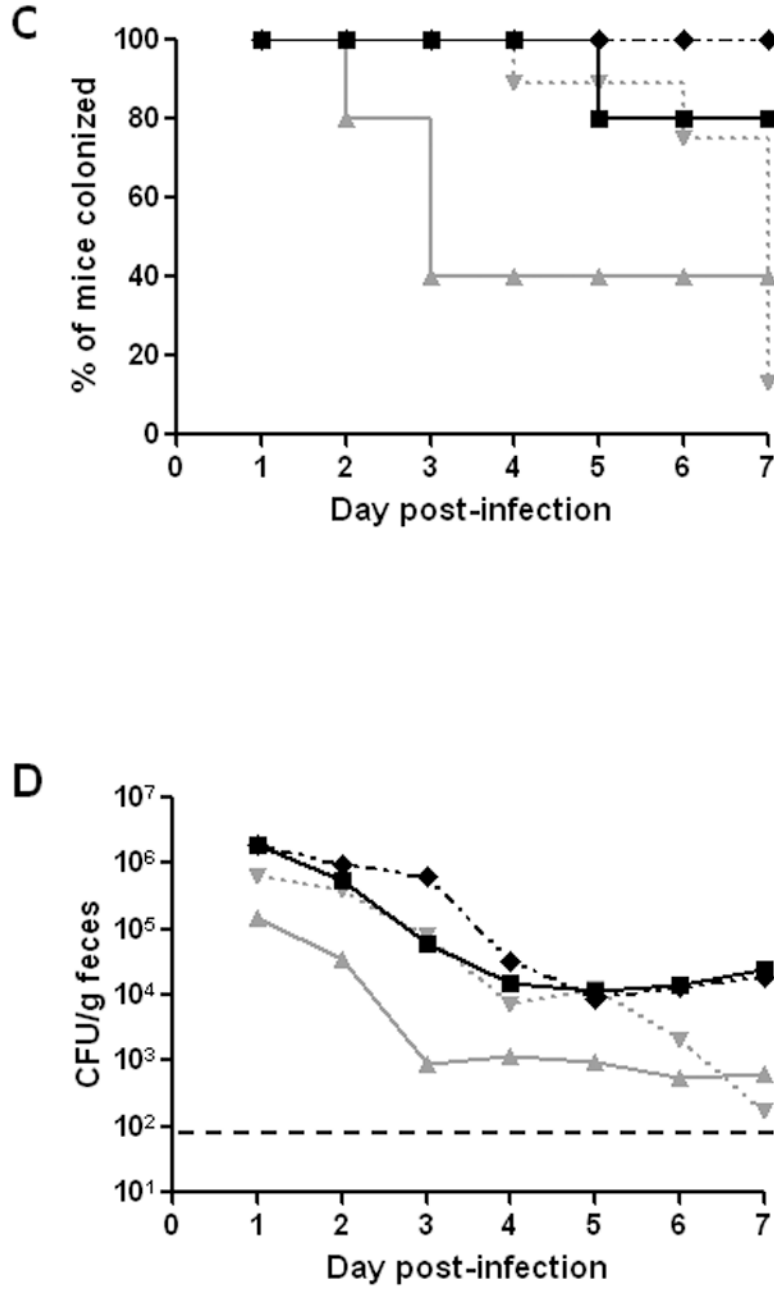


Fig. 1. Colonization of BALB/c mice by *E. coli* O157:H7 strain 86-24NaI^R. (A) Percent of infected mice colonized over time. Animals were intragastrically infected with 86-24NaI^R at inocula of 10⁵ (●), 10⁶ (■), 10⁷ (▲), 10⁸ (▼), and 10⁹ CFU (◆). Mice were considered colonized if there were recoverable 86-24NaI^R in the feces. (B) Colonization level of 86-24NaI^R overtime at various doses administered intragastrically (symbols as above). For panels C and D, animals were infected with 86-24NaI^R at doses of 10⁸ CFU or 10⁹ CFU either by intragastric administration (dashed lines with ▼ for 10⁸ CFU and ◆ for 10⁹ CFU) or pipette feeding (solid lines with ▲ for 10⁸ CFU and ■ for 10⁹ CFU). (C) Effect of administration route on percent of mice colonized. (D) Impact of administration route on colonization level. For panels A and

C, animals that died during the study were excluded from the study after the time of death; these included 2 mice on day 4 from the 10^9 dose (gavage) and one animal each on days 4 and 5 from the 10^8 dose (gavage). For panels B and D, each point represents the GM of the number of $^{86}\text{-}^{24}\text{NaI}^{\text{R}}$ shed into the feces from surviving, infected animals, and the limit of detection (indicated by the black dashed line) was 10^2 CFU/g feces.

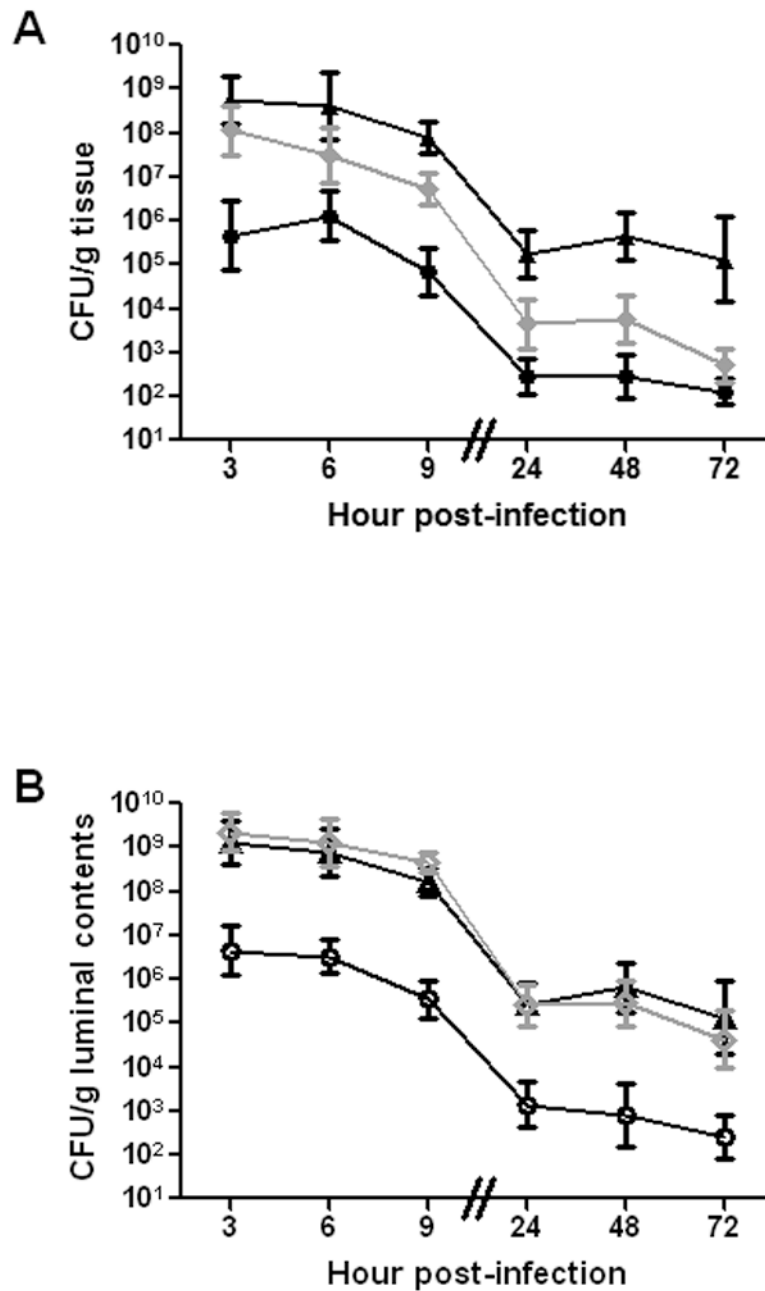


Fig. 2. Levels of *E. coli* O157:H7 over time at different sites in the gastrointestinal tract of BALB/c mice orally-infected with $6-8 \times 10^8$ CFU of strain 86-24NaI^R. The GM CFU/g tissue (A) or luminal contents (B) from groups of 8–17 mice each are shown for the small intestine (●), cecum (▲), and large intestine (◆) as well as the luminal contents of the small intestine (○), cecum (△), and large intestine (◇). Bars indicate the 95% confidence interval.

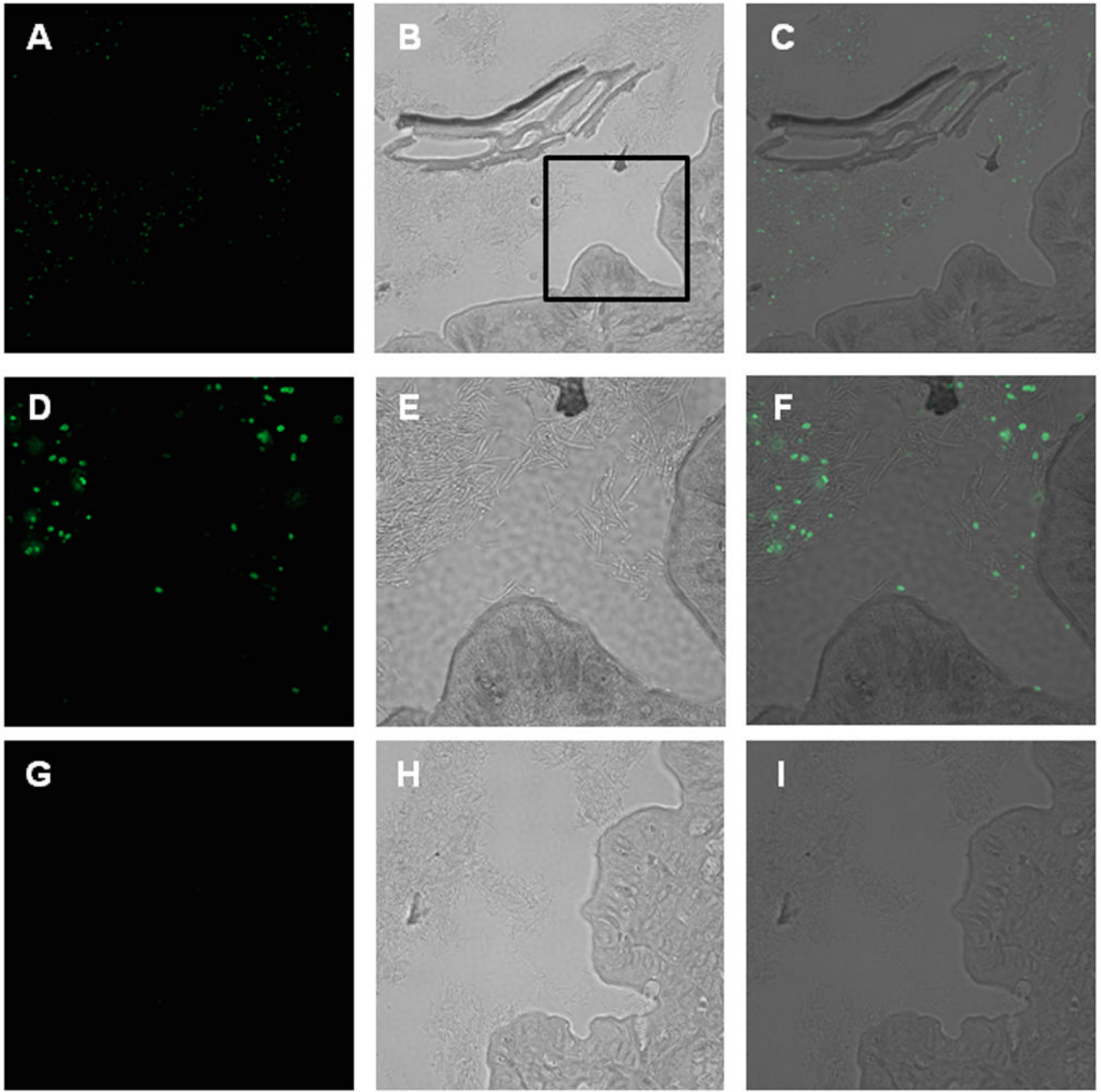
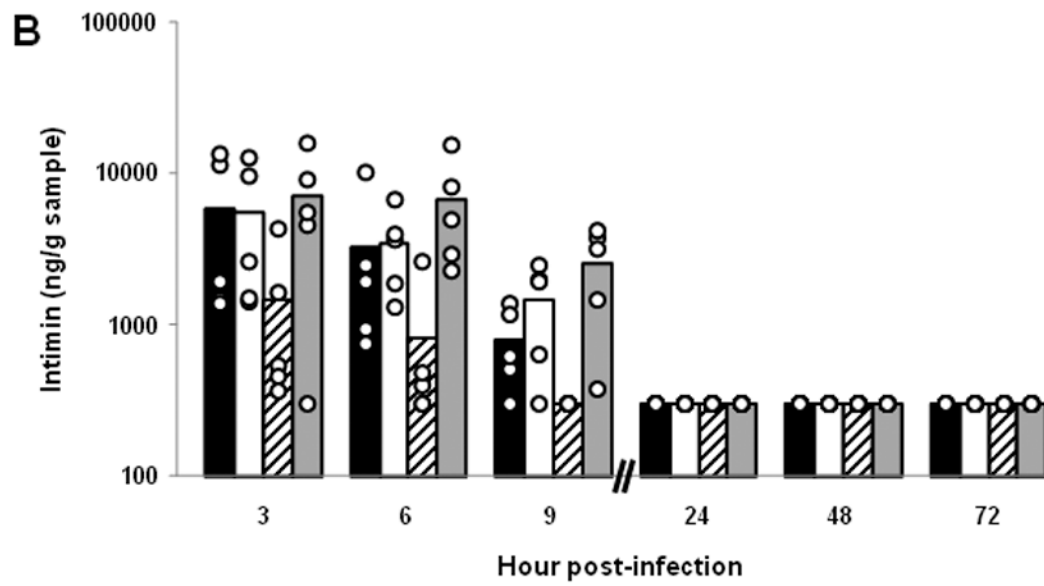
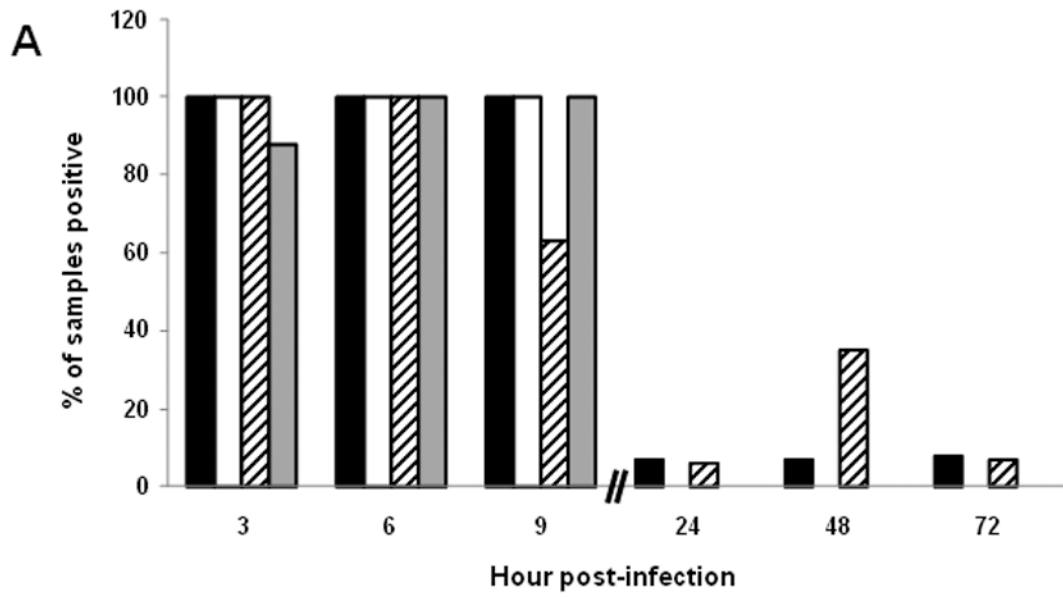


Fig. 3. Immunofluorescent *E. coli* O157:H7 in the cecum. Mice were infected with *E. coli* O157:H7 (A–F) or mock-infected (G–I) for 6 hours. Images display infected or control ceca stained for O157 (A, D, and G), the phase contrast image of the cecal section (B, E, and H), and the merged images (C, F, I). Panels D, E, and F are a higher magnification of the boxed area of panel B. For panels A–C and panels G–I the original image was obtained at a magnification of 40 \times . For panels D–F the original image was obtained at a magnification of 100 \times .



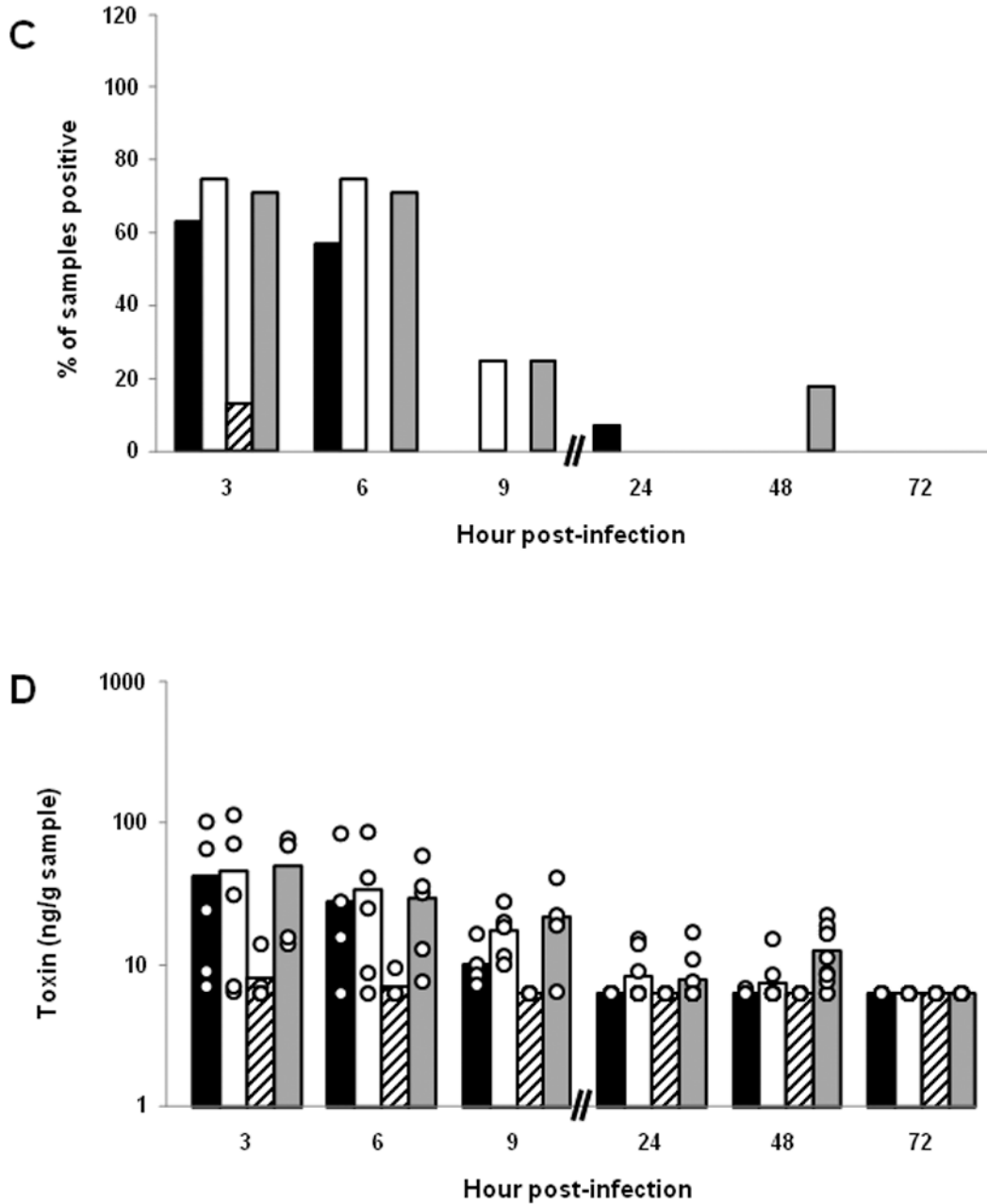


Fig. 4. Presence of intimin and Stx2 over time at different sites in the gastrointestinal tracts of BALB/c mice orally infected with *E. coli* O157:H7 strain 86-24NaI^R. (A) Percent of different intestinal samples positive for intimin over time. Each bar represents the mean of 8–17 samples tested. For panels A–D, the tissues are represented by black bars (cecum), white bars (cecal contents), hatched bars (large intestines), and gray bars (large intestinal contents). (B) Levels of intimin in different intestinal sites over time. Each bar represents the mean value of a subset of those samples presented in panel A (n=5–10). The limit of quantifiable intimin was 360 ng/g tissue or tissue contents. For panels B and D, each individual sample is represented by a ○. (C) Percent of different intestinal samples positive for Stx2 over time. Each bar represents the mean of 7–

16 samples tested. (D) Levels of Stx2 in different intestinal sites over time. Each bar represents the mean value of a subset of those samples presented in panel C (n=5–10). The limit of quantifiable Stx2 was 6.4 ng/g tissue or tissue contents.

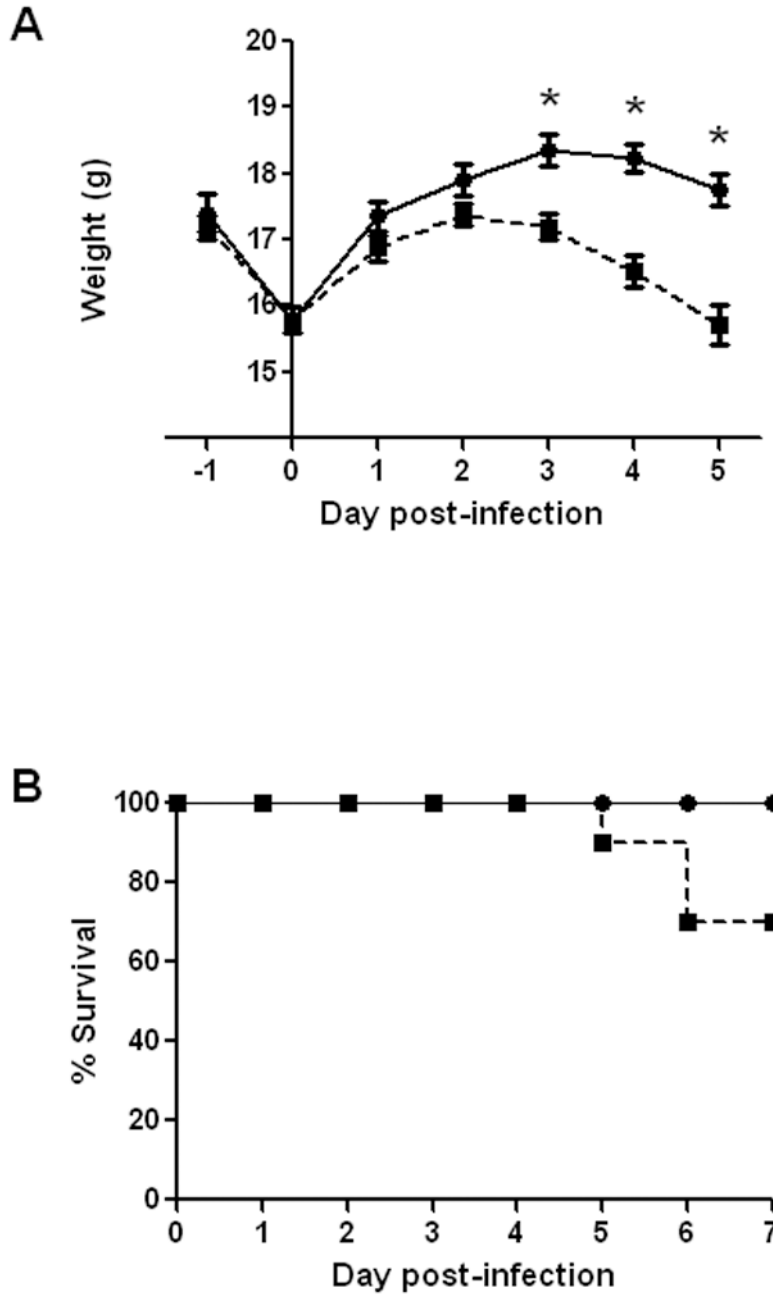


Fig. 5. Effect of intragastric infection of BALB/c mice with $\sim 10^9$ CFU of *E. coli* O157:H7 strain 86-24Nal^R on body weight and survival. Animals were starved overnight prior to intragastric inoculation with *E. coli* O157:H7 on day 0. (A) Weight was monitored for the control, mock-infected animals (●) and the 86-24Nal^R-infected, experimental group (■). Each point on the graph represents the mean weight of 10 (or less in the case of groups with mortality) animals per group. The bars depict \pm one standard error of the mean of weight. Asterisks indicate days that demonstrated a significant difference between weights of infected and control mice ($p \leq 0.001$). (B) Survival of animals infected with 86-24Nal^R or mock-infected. The percent of

surviving animals (10 mice per group) on each day post-infection inoculated with 86-24NaI^R (■) or mock-infected (●).

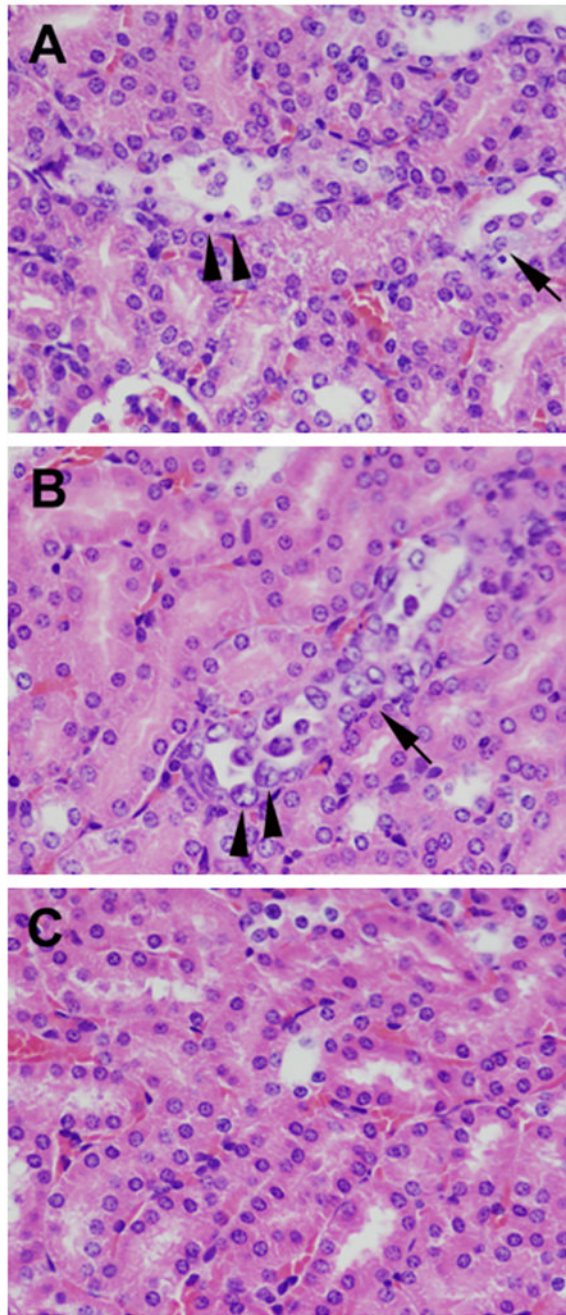


Figure 6.

Renal lesions in mice infected by gavage with *E. coli* O157:H7 strain 86-24Na^R. Panels A, B, and C display kidney sections stained with H&E and taken from infected (panels A and B) or control (panel C) mice. The original images were obtained at 40× magnification. (A) The mouse was necropsied on day 5 post-infection. Multiple dilated cortical tubules are lined by epithelial cells that are hyper eosinophilic, shrunken, angular, and pyknotic [indicative of necrosis (arrowheads)] or hypertrophic with vacuolated cytoplasm [indicative of degeneration (arrow)]. (B) The mouse was necropsied on day 5 post-infection. Cortical tubule epithelial cells have basophilic cytoplasm, are karyomegalic (arrowheads), and exhibit anisocytosis (presence in

the blood of erythrocytes with excessive variation in size) with mitotic figures (arrow), all of which is indicative of regeneration.

Table 1

Average^a values (and range) of serum chemistry and CBC analyses of mice infected with *E. coli* O157:H7 strain 86-24 Nal^R

Laboratory Test	Control	Infected	<i>p</i> -value
BUN^b (mg/dL)	17.8 (13,28)	32.4 (15,108)	<0.01
Creatinine (mg/dL)	0.26 (0.2,0.5)	0.34 (0.2,0.8)	0.073
% Neutrophils	14.8 (1.8,20.3)	39.0 (22.1,81.4)	0.001
% Lymphocytes	75.9 (61.4,84.8)	53.5 (13.6,74.4)	0.002
Hemoglobin (g/dL)	14.5 (12.4,16.3)	15.5 (12.4,17.2)	ND ^c
% Hematocrit	45.7 (39.4,51.2)	49.0 (39.5,56.6)	ND
Platelets (K/mL)	751 (335,1234)	881 (293,1189)	ND
WBC^d (K/mL)	4.79 (1.11,8.25)	5.04 (2.43,8.56)	ND

^a Averages represent data from all days (2,3,4,&5) combined

^b Blood Urea Nitrogen

^c No Difference

^d White Blood Cell