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T-cell based immunotherapy in experimental autoimmune encephalomyelitis and multiple sclerosis

Kate O'Brien¹, Bruno Gran^{1,2}, and Abdolmohamad Rostami^{2,†}

¹Division of Clinical Neurology, University of Nottingham, UK

²Department of Neurology, Thomas Jefferson University, Philadelphia, PA 19107, USA

Abstract

One of the reasons multiple sclerosis (MS) has been considered a T-cell mediated autoimmune disease is that a similar experimental disease can be induced in certain rodents and primates by immunization with myelin antigens, leading to T-cell-mediated inflammatory demyelination in the CNS. In addition, most if not all pharmacological treatments available for MS are biologically active on T cells. In this article we review the principles of T-cell-based immunotherapies and the specific actions of current and novel treatments on T-cell functions, when these are known. For both licensed and innovative agents, we also discuss biological actions on other immune cell types. Finally, we offer a brief perspective on expected changes in the use of MS immunotherapies in the near future.

Keywords

disease-modifying treatment; experimental autoimmune encephalomyelitis; immune system; immunotherapy; multiple sclerosis; T cell

Multiple sclerosis (MS) is a chronic, inflammatory, demyelinating disease of the CNS [1,2] that usually begins in young adulthood. It is characterized by lesions within the CNS and demyelination is a key feature of these lesions. There is also evidence of axonal transection or axonal degeneration. Genetic and environmental factors confer susceptibility to MS, as shown by epidemiological studies. These factors include gender, race, exposure to infections (particularly during childhood and adolescence), and geographical variables such as latitude and sun exposure [2]. The clinical symptoms vary depending on the parts of the CNS affected – they include focal neurological deficits such as motor weakness or paralysis, sensory loss, loss or blurring of vision and cognitive deficits [1].

The etiology of MS remains unknown; however, data from MS lesions, genetic susceptibility, the use of immunotherapies and extensive studies on animal models of MS suggest that it is an immune-mediated disease. Autoreactive T cells are thought to initiate an autoimmune response directed against components of CNS myelin (Figure 1). The main targets of the autoimmune reactions are thought to be myelin basic protein (MBP), proteolipid protein (PLP) and myelin oligodendrocyte glycoprotein (MOG) [3]. There is also thought to be a role for

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^{*}Author for correspondence Tel: +1 215 955 8100 Fax: +1 215 955 1390 a.m.rostami@jefferson.edu.

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antibody-mediated responses to myelin antigens in disease pathogenesis. Although primary demyelination is considered a key feature of MS, extensive axonal pathology is also present. Postmortem analysis of brains of MS patients demonstrated axonal transection in both active and chronic lesions, the frequency of which was related to the degree of inflammation [4]. Furthermore, loss of parental cell bodies in the cortex has also been observed [5]. Experimental autoimmune encephalomyelitis (EAE), an animal model of MS induced by immunization with myelin components in adjuvant substances, shows comparable neuronal pathology, which renders this model suitable for both neuro- and immunopathological studies of autoimmune inflammatory demyelination.

In this article, we will review the known roles of T cells in initiating and orchestrating autoimmune responses in EAE and MS, and define those that have been targeted for immunotherapy (Table 1).

T-helper cells in MS & EAE

T-helper 1/T-helper 2

Until recent years, pathological processes in MS and EAE were thought to be initiated by myelin-reactive T helper (Th)1 cells. However, abundant evidence has implicated IL-17-producing Th17 cells as the main cells involved in the pathogenesis of autoimmune demyelination in EAE and MS (Figure 2) [6–10]. Originally it was thought that CD4⁺ effector T cells were divided into two lineages: Th1 and Th2. Th1 are induced by IL-12 and produce IFN- γ , while Th2 cells secrete IL-4, IL-5 and IL-13. Th1 cells can mediate proinflammatory or cell-mediated immune responses, whereas Th2 cells mainly promote certain types of humoral immunity [11]. The Th1/Th2 paradigm in the mouse was subsequently extended to human T cells [12].

IL-12 is a heterodimeric cytokine formed by a large (p40) subunit and a small (p35) subunit. When it was discovered that IL-23 shares the p40 subunit with IL-12 [13], many questions were raised as to which cytokine was the key cytokine involved in disease. IL-12(p40)-deficient mice had been shown to be resistant to EAE [14]. However, it became clear after the discovery of IL-23 [13] that the deletion of IL-12(p40) had eliminated the expression of IL-12(p40p35) but also IL-23(p40p19) [15]. We and others then demonstrated that IL-12 was redundant in EAE susceptibility, as IL-12(p35)-knockout mice were in fact susceptible to disease [6,8]. In 2003, Cua and colleagues directly showed that IL-23 rather than IL-12 is crucial for autoimmune inflammation by using IL-23-knockout mice. The p19 subunit of IL-23 was knocked out and these mice were resistant to EAE [16]. IL-12 was in fact found to suppress EAE when administered to mice during the early phase of EAE induction. The induction of IFN- γ mediated this effect [17]. IL-23 was found to induce the production of IL-17 from both effector and memory CD4⁺ T cells, whereas IL-12 had only marginal effects on IL-17 production [18].

Th17/regulatory T cells

Langrish and colleagues clearly defined the role of IL-17-producing Th cells, which are driven by IL-23, in an adoptive transfer model of EAE. They cultured PLP-primed CD4⁺ T cells *in vitro* in the presence of IL-23. These cells that produce IL-17 were passively transferred into recipient mice, which developed severe EAE [10]. Komiyana and colleagues found that IL-17deficient mice develop attenuated EAE [9]. After initial identification of the Th17 lineage, research has focused on Th17 regulation [19–25]. Th17 polarization has been shown to be supported by a number of cytokines such as IL-1 γ , IL-23, IL-18, IL-22 and TNF- α . In addition, a surprising role of TGF- β in the presence of IL-6 has been elucidated. TGF- β induces naive T cells to produce IL-21, which then acts as a positive autocrine loop to upregulate the Th17specific transcription factor RORyt [23,24]. Along with the inducers of Th17 polarization, a number of negative regulators have also been identified. IL-27, a new member of the IL-12/IL-23 family of heterodimeric cytokines, is a potent negative regulator of Th17 differentiation [25,26]. IL-2 is also a negative regulator, as this cytokine is important for the generation and survival of natural regulatory T cells (Tregs) expressing the transcription factor Foxp3 [27]. CD4⁺CD25^{hi}Foxp3⁺ Tregs represent a naturally occurring immunoregulatory population of T cells. In contrast to effector T cells, Tregs protect against tissue damage and inhibit autoimmune disease [28]. Therefore, autoimmune responses appear to be regulated by a balance between effector cells and Tregs.

Role of T cells in EAE & MS

The important role of T cells in autoimmune inflammatory demyelination was established based on several lines of evidence; inflammatory lesions in the CNS of patients with MS were found to contain both CD4⁺ and CD8⁺ T cells [29]; adoptive transfer of immune cells, in particular Lyt $1^+ 2^-$ cells (later defined as CD4⁺ Th cells), from mice with EAE could transfer disease to healthy recipients [30,31]; depletion of CD4⁺ Th cells from mice with actively induced EAE led to disease suppression [30,32]; most therapies used in MS are in fact immunotherapies directed at T cells [33]; and genes that are associated with MS susceptibility are involved in antigen presentation to T cells or recognition of cytokine signals by T cells [34–36].

It should be noted that there are differences in the importance of different subsets of T cells between MS and EAE. EAE is mediated by CD4⁺ T cells. However, although CD4⁺ T cells dominate the perivascular regions of the inflammatory focus in EAE induced by MBP and PLP, MS lesions contain more CD8⁺ than CD4⁺ T cells. CD4⁺ T cells in MS lesions have been shown to have either pathogenic or neuroprotective functions dependent on the cytokines that are produced. Babbe and colleagues demonstrated that active MS lesions have a predominance of CD8⁺ T cells [37]. Upon further investigation it was found that expansion of the CD8⁺ Tcell repertoire was more antigen-driven than the CD4⁺ T-cell repertoire. Over-representation of CD8⁺ T cells was observed in cerebrospinal fluid of MS patients, and T cells were found to be stable over several months. Analysis of the *TCR* V β gene expression pattern indicated that in some patients, CD8⁺ T cells were clonally expanded [38].

Owing to the clear key role of T cells in MS and EAE, there has been a vast amount of research in the past two decades in an effort to elucidate new methods for T-cell immunotherapy. Some of the following immunotherapies are currently used, some have not been successful, and others warrant further study.

Principles of T-cell-based immunotherapies

Induction of tolerance

T cells have the ability to react to a variety of antigens, both self and nonself. Therefore, there are many mechanisms that exist naturally to eliminate potentially selfreactive responses – this is known as natural tolerance. The main mechanism for eliminating potential auto-reactive T cells occurs in the thymus and is known as central tolerance. Some potentially autoreactive T cells escape central tolerance and, therefore, peripheral tolerance mechanisms also exist. Despite these mechanisms, some selfreactive T cells may `escape' and be present in the repertoire, and their activation may lead to autoimmune diseases.

Studies on therapeutic tolerance have attempted to induce and amplify potent physiological mechanisms of tolerance in order to eliminate or neutralize selfreactive T cells and prevent or treat autoimmune diseases.

Experimental autoimmune encephalomyelitis can be induced via active immunization with CNS tissue or myelin peptides, such as MBP, PLP and MOG in complete Freund's adjuvant, or via adoptive transfer of CD4⁺ T cells. Several animal models of EAE have been developed over the last few decades. They include murine models such as the C57BL/6 mouse, which exhibits a monophasic or a chronic, sustained form of EAE induced by immunization with MOG35–55 (an immunodominant peptide of MOG) in complete Freund's adjuvant, and the SJL/J mouse, a relapsing model of EAE induced by immunization with PLP139–151 (an immunodominant peptide of PLP).

Antigen-specific inhibition of EAE can be achieved by administration of tolerogenic forms of myelin antigens. Tolerogenic antigens can be administered intravenously (iv.), intranasally or orally. Different administration routes may lead to different forms of tolerance induction [39]: clonal deletion – the elimination of antigen-specific cells by apoptosis; clonal anergy – the induction of functional hyporesponsiveness to antigens; and active suppression – the induction of an antigen-specific population of cells. This antigen-specific population negatively regulates the antigen-specific autoreactive cells. This type of suppression occurs in immune deviation where there is conversion from one Th phenotype (e.g., Th1) to another Th phenotype (e.g., Th2). The Th2 phenotype can then suppress disease by antagonising the Th1 phenotype, which is considered pathogenic [40].

Intravenous tolerance

Tolerance induction in naive T cells: High doses of iv.-injected soluble antigen can induce T-cell anergy or apoptosis. Liblau and colleagues demonstrated tolerance induced by both thymic and peripheral apoptosis and also T-cell-receptor desensitization in mice injected with high doses of antigen [41]. Kearney and colleagues demonstrated that the route of administration of antigen could determine the level of tolerance achieved and showed that iv.-injected antigen was much more effective than subcutaneous injection [42]. Tolerance could further be improved by chemically cross-linking peptide antigen to splenocytes with 1-ethyl-3-carbodiamide, instead of using soluble antigen. This method was found to inhibit spinal cord homogenate-induced chronic relapsing EAE in SJL mice [43–45]. In another model of chronic relapsing EAE using Biozzi ABH mice [46], disease severity was also found to be reduced by iv. injection of myelin antigen. However, if the treatment is not initiated until late stage of disease, relapses are inhibited but disease progression is not prevented [47].

The effect of time of iv. administration of peptide to induce tolerance was also studied in Lewis rats. MBP was administered at different times both before and after active immunization of rats to induce EAE. The treatment of rats with iv. MBP before immunization was most successful 14 days prior to immunization, whereas after immunization the most effective iv. administration was at onset of disease. Disease was suppressed in a dose-dependent manner at these time points. In adoptive transfer experiments, it was found that recipient mice must be treated with iv. MBP after, but not before, the transfer of MBP-reactive EAE effector cells [48]. The tolerization of Lewis rats after iv. MBP administration was found to be controlled at least partly by a T-cell control mechanism in the spleen and the role of Tregs was speculated [49].

More recently, we have focused on the role of antigen-presenting cells (APCs) in iv. tolerance. We studied the interaction between APC and CD4⁺ T cells in iv. tolerance. Surprisingly, we found that APCs that had been purified from MOG iv.-treated mice were immunostimulatory [50]. Following this we examined the function of distinct subsets of APCs with particular focus on dendritic cells (DCs). We found that the population of CD11b⁺CD11c⁺ DCs are expanded in the spleen and CNS after iv. injection of MOG peptide to induce tolerance. These DCs exhibit immunoregulatory activity by increasing IL-10 and TGF- β , and inducing the generation of CD4⁺CD25⁺ Tregs [51]. Recently, we have assessed whether Th17 cells are involved in the

tolerance induced in mice after iv. MOG administration. We found a decrease in JAK/ STAT-1,4, ERK1/2 and NF- κ Bp65 phosphorylation in iv.-tolerized mice. A reduction in IL-17 was also observed. We transferred CD11b⁺ splenocytes of tolerized mice into actively induced EAE mice and found that these splenocytes delayed disease onset and reduced severity of disease [52].

Tolerance induction in memory autoreactive T cells: T cells that escape natural tolerance may become selfreactive memory T cells if they become activated. Such selfreactive T cells play major roles in the development of autoimmune diseases and because of this, much effort has been placed on designing strategies to tolerize these cells. As memory T cells do not require costimulation to the same extent as naive T cells they are more difficult to tolerize [53]. Verbeek and colleagues studied the effectiveness of iv. injection of soluble antigen in eliminating memory T-cell response against $\alpha\beta$ -crystallin, which is a candidate autoantigen in MS. They found a long-lasting specific tolerance induced by iv. administration [54].

Intranasal tolerance—The second route for inducing tolerance is intranasal administration. Bai and colleagues found that intranasal administration of MBP prevents relapses in the DA rat model of EAE where the rats were immunized with complete spinal cord homogenate. Prevention of relapses was associated with activation of regulatory cells expressing IL-4 and TGF- β [55]. Burkhart and colleagues studied intranasal administration of MBP in a mouse model of EAE. They found that intranasal tolerance was dependent on IL-10. They demonstrated this by using an IL-10-neutralizing antibody *in vivo* that restored susceptibility to EAE [56].

Oral tolerance—The third route for inducing tolerance is orally. Three different mechanisms of tolerance can be induced orally depending on the amount of oral antigen administered. Active suppression is favored by low doses, whereas clonal anergy or deletion is favored by higher doses of oral antigen. In the 1980s, the oral administration of antigen was found to suppress EAE [57,58] and, therefore, trials were initiated in patients. Weiner and colleagues found that the treatment of relapsing-remitting (RR) patients with bovine myelin containing MBP and PLP reduced the frequency of MBP-reactive T cells compared to controls. There was a tendency for the treated group to have less exacerbations and this was particularly evident in the subgroup of DR2-males [59]. It was later uncovered that oral antigen induces Th2 and Th3 (TGF- β -producing) T cells, CD4⁺CD25⁻ and latency-associated peptide⁺ (LAP) T cells [60]. This group then found that oral administration of CD3-specific monoclonal antibody suppresses EAE. The suppression was observed before the induction of disease and also at the peak of disease, and was associated with the induction of such CD4⁺CD25⁻LAP⁺ Tregs. These cells were found to be dependent on TGF- β for their suppressive function both *in vitro* and *in vitro* [61].

Use of altered peptide ligands

The trimolecular complex of T-cell antigen recognition involves an immunodominant antigenic peptide bound to a MHC (human histocompatibility antigens [HLA] in humans) molecule recognized by a T-cell receptor (TCR). An altered peptide ligand (APL) is the MHC-binding antigenic peptide modified by substitution of one or more amino acids to change the MHC or TCR binding characteristics. This can induce tolerance to the original peptide through several mechanisms, including partial activation of T cells [62,63] and antagonism at the T-cell activation level [64].

Brocke and colleagues used an altered peptide ligand of MBP to treat EAE. They induced EAE with a T-cell clone that is specific for MBP p87–99. They tolerized the clone *in vivo* with an analog of MBP p87–99 (phenylalanine at residue 96 was replaced by alanine) and found that

The generation of TCR transgenic (Tg) mice specific for MBP provided a system of extreme importance for evaluating the mechanisms of disease development and regulation in EAE. Goverman and colleagues developed mice that were Tg for either TCR- α or TCR- β chains of a T-cell clone specific for MBP [66]. T-cell responses to peptides were observed in double-Tg mice and some mice developed spontaneous EAE [66,67]. Further studies by Lafaille and colleagues demonstrated that spontaneous EAE could be significantly enhanced by crossing the TCR- $\alpha\beta$ Tg mice with recombination-activating gene-deficient mice that cannot rearrange endogenous TCR genes [68]. These results paved the way for studies using TCR in protection against EAE as they suggested that endogenous rearrangement using APLs may induce a population of Tregs that could prevent EAE. Offner and colleagues studied the immune regulation directed at TCR determinants using single TCR Tg mice expressing a TCR- β chain specific for MBP. They found that vaccination with the same target TCR protein amplifies TCR-specific regulation and protects against EAE [69].

one was sufficient to prevent disease [65].

`Humanized mice' that were Tg for HLA class II (human) molecules were also generated. These mice allowed for the testing of T-cell determinants that were vital in disease. Vandenbark and colleagues generated a HLD-DR2⁺ Tg mouse that lacked all MHC (murine) class II genes. These mice suffered from severe EAE after immunization with MOG35–55 peptide. They used these mice to test the efficacy of a novel recombinant TCR ligand in inducing tolerance. They found that this peptide could induce sustained tolerance in MOG-peptide induced EAE [70]. More recently, the human MBP84–102 epitope has been genetically linked to the HLA-DR2 by two heterodimeric receptors where a TCR-ζ signaling domain is either present or absent. The aim of this study was to use chimeric receptors to target therapeutic T cells against selfreactive T cells. The MBP-specific T cells were attacked by the ζ-modified cytotoxic T lymphocyte (CTL) and this CTL was found to block EAE [71].

The studies of Tg mice expanded to different regions of HLA-DR in addition to DR2. Ito and colleagues developed HLA-DR4-IE and HLA-DRB1*0401-IE β chimeric Tg mice and found these mice to be susceptible to EAE [72]. Muraro and colleagues studied HLA-DRB-restricted MBP111–129 as MBP111–129 is the immunodominant epitope in humans carrying HLA-DRB1*0401 [73]. They used HLA-DRB1*0401 Tg mice to study the pathogenic potential of HLA-DRB1*0401-restricted MBP111–129 and found that this was indeed pathogenic [73, 74]. Glatiramer acetate (GA) is a therapy used in MS that binds HLA tightly and can displace MBP [75] (see `Immunotherapies used in MS' below).

The use of TCR peptides was not only being studied in EAE, but also for the treatment of MS [76]. The hypothesis behind these experiments was as follows: as both α - and β -TCR chains are expressed in abundance, these unassembled TCR chains, or peptides thereof, could be presented on the surface of T cells in association with self-MHC molecules. This would provide MHC-restricted targets that could combine specifically with the TCR of Tregs (which were thought to be Th2 cells). Vaccination of a peptide highly similar to an autoreactive TCR fragment would enhance the regulatory Th2 priming and recognition that would control the Th1-mediated inflammation. This procedure was tested in patients and was found to be well tolerated and also showed modest signs of clinical improvement [77].

However, there were some major concerns raised about the use of APLs for immunotherapy, as a study by Bielekova and colleagues demonstrated that APLs could actually exacerbate MS. They used an MBP APL in a Phase II clinical trial and found that three patients developed

exacerbations, two of which were shown to be linked to the APL [78]. This study highlighted the level of caution needed when transferring a therapy that has been successful in animal models into humans.

Anti-cytokine monoclonal antibodies Anti-IL-12

IL-12 is a Th1 cytokine that was previously considered to be the main cytokine involved in disease pathogenesis in EAE and possibly in MS. As a result of this, much effort was placed into designing ways to decrease IL-12 production. Leonard and colleagues induced relapsing EAE in SJL/J mice by adoptive transfer of PLP peptide-specific T cells and found that administration of sheep-polyclonal anti-IL-12 antibody prevented the transient mild course of demyelinating neurological disease [79]. The treatment had to be prolonged for a dramatic reduction in disease severity to be observed. Constantinescu and colleagues also studied the effect of anti-IL-12 in relapsing EAE and found that when the antibody is administered after recovery from initial attack, spontaneous relapses are prevented [80]. Ichikawa and colleagues studied the effect of anti-IL-12 antibody in chronic relapsing EAE induced with MOG peptide. They observed suppression of disease clinically and histologically. Spleen cells of mice treated with anti-IL-12 antibody had a reduced proliferative response to MOG peptide, and IFN- γ production was also decreased [81].

The effect of the IL-12/23p40-neutralizing monoclonal antibody (ustekinumab) was recently tested in RRMS patients to assess the drug's safety and efficacy. This Phase II trial included 249 patients with RRMS. Patients were followed for 37 weeks and ustekinumab was generally well tolerated. However, there was no significant reduction in disease after ustekinumab treatment as assessed by the number of gadolinium-enhancing T1-weighted lesions [82].

Anti-IL-23—IL-23 has been found to be crucial in driving a pathogenic T-cell response, and results in the expansion and maintenance of Th17 cells. Therefore, it was vital that the effects of neutralizing IL-23 be examined. As IL-23 shares the p40 subunit with IL-12, an antibody specific to the p19 subunit of IL-23 was required. Chen and colleagues developed anti-IL-23p19 and tested the efficacy in EAE. Treatment with the antibody resulted in reduced IL-17, IFN-γ, IL-10, IL-6 and TNF in the CNS. Epitope spreading of PLP was prevented and this resulted in prevention of disease relapse [7].

Anti-IL-17—As mentioned above, recent data have shown that IL-17 is the main cytokine involved in EAE and not IL-12. Hofstetter and colleagues studied the pathogenic role of IL-17 in EAE and they also tested the effect of neutralization of IL-17 on disease severity. They neutralized IL-17 with IL-17-receptor-Fc-protein, which was found to slightly reduce clinical symptoms. Neutralization with an IL-17 monoclonal antibody was also tested and found to reduce disease severity, but not to the degree that was expected. It was hypothesized that other factors can compensate for the lack of IL-17. This view was confirmed by reduced EAE severity rather than complete resistance to disease in IL-17^{-/-} mice [83].

Translational pitfalls—One has to be extremely careful when transferring knowledge from animal models to humans. Therapies that are successful in animal models may be unsuccessful and possibly dangerous in humans. An example of this can be seen from the `cytokine storm' incident of 2006. TGN1412, which is an anti-CD28 monoclonal antibody, was administered to six healthy young male volunteers in Phase I of a clinical trial. All six volunteers developed a systemic inflammatory response and became critically ill, with renal failure, pulmonary infiltrates and lung injury. The volunteers were transferred to intensive care where they received dialysis and anti-IL-2 receptor antagonist antibody. Two of the patients required organ support for 8 and 16 days. Even though all patients survived, this episode highlights the need

for the extreme care to be taken when administering successful animal-model treatments to humans [84].

Therapies used in MS

The following therapies are used or under investigation for the treatment of MS. Many of the exact mechanisms of action are unknown; therefore, we are unsure if these are T-cell therapies. However, all exert some effect on T cells and are thus discussed here.

IFN-β

IFN- β is the most commonly used treatment for RRMS patients, although the mechanisms of action are still poorly understood. There are three commercially available IFN- β formulations, two forms of IFN-β1a (produced in eukaryotic cells, fully glycosylated, for subcutaneous or intramuscular injection) and one form of IFN-β1b (produced in prokaryotic cells, not glycosylated, for subcutaneous injection). IFN- β has been shown to be a Th1 and more recently a Th17 inducer, therefore it is a paradox that it can treat disease. It does, however, reduce Tcell activation by inhibiting the upregulation of HLA class II and thereby counteracting IFN- γ effects [85,86]. IFN- β is also thought to affect costimulatory molecule interaction such as CD40–CD40L [87] and also B7–CD28 [88]. Immune deviation can also be a result of IFN-β treatment where IL-10, a regulatory cytokine, is induced [89,90]. Another effect of IFN- β is on the blood-brain barrier. T-cell adhesion to the endothelium is thought to be interfered with, and also an increase in soluble vascular cell adhesion molecule (sVCAM) has been observed [91,92]. This sVCAM has been correlated with a reduction in disease severity as measured by gadolinium-enhancing lesions [91]. IFN- β s are biologically active on other immune-cell types, including APCs. The relevance of these actions to MS immunotherapy are being investigated [93,94].

Glatiramer acetate

Glatiramer acetate is a random copolymer of alanine, lysine, glutamic acid and tyrosine that is used in the treatment of RRMS. It was first studied in EAE and later was approved for use in MS patients. The effect of GA on T-cell clones was examined and it was found that there is preferential inhibition of self-reactive T-cell clones after treatment with GA. GA was crossrecognized by myelin basic protein-reactive T cells [95]. GA can bind to HLA class II molecules, including HLA-DR2, and thereby inhibit the activation of MBP-reactive T cells [75,96]. In human T-cell lines, it was found to induce proliferation without prior exposure to polymer; however, in MS patients that received daily injections of GA, this response was not present. GA was found to induce a Th2 rather than a Th1 response, which is usually present in MS patients [97]. GA not only has an effect on T cells but also has effects on APCs. GA was found to bind to MHC molecules on APCs and therefore competes with MBP binding to HLA [75]. GA has an inhibitory effect on monocyte reactivity. Activation of monocytes after TLR stimulation was considerably reduced after GA treatment [98]. GA activates type II monocytes that secrete IL-10 and TGF-B, and reduces the production of IL-12 and TNF. These type II monocytes were found to differentiate Th2 cells and Tregs [99]. In summary, the mechanism of action of GA is at least partly T-cell mediated, although other studies indicate that APCs are also involved. Neuroprotective mechanisms have also been proposed [100].

Natalizumab

The development of natalizumab arose from studies in EAE models where antibodies against α 4b1 integrin were found to prevent disease. This was associated with a prevention of leukocytes from accumulating in the CNS [101]. Natalizumab was later developed based on this principle, and it is an antibody against very late activating antigen. It blocks the endothelial interaction, thereby blocking entry of leukocytes into the CNS, and has been found to be a

successful treatment for MS. CSF leuckocyte counts of CD4⁺ and CD8⁺ T cells, CD19⁺ B cells and CD138⁺ plasma cells have all been found to be lower in MS patients treated with natalizumab compared with controls [102]. Natalizumab has also recently been approved for therapy in Crohn's disease. In spite of the very large number of MS patients treated worldwide, there is still some concern regarding safety, as rare cases of progressive multi-focal leukoencephalopathy (PML) have been reported. PML is a severe opportunistic infection of the brain that appears to arise owing to the lack of immune surveillance after natalizumab treatment, usually but not exclusively, when patients have also received additional immunotherapies [103].

Immunosuppressive agents

Immunosuppressive agents are drugs that inhibit or prevent activity of the immune system. However, because they dampen immune responses and are often nonspecific, there is concern over side effects such as being unable to resist infections. Azathioprine inhibits purine synthesis and cell proliferation, in particular that of leukocytes. It has been used as an oral treatment for MS where it has been found to reduce the relapse rate and be effective on MRI parameters of inflammation [104]. There was some concern over the risk of cancer after azathioprine treatment; however, the doses used to treat MS patients are quite low and overall safety data have been reassuring [105,106]. Mitoxantrone is a stronger immunosuppressive drug that inhibits Type II topoisomerase. It disrupts DNA synthesis and DNA in both healthy and cancer cells. As a Type II topoisomerase it inhibits the proliferation of T and B cells and macrophages [107]. Mitoxantrone is administered intravenously and is used for more aggressive disease (RR or secondary progressive with relapses) [108]. There are some concerns for myelosuppressive effects and cardiac toxicity and, therefore, there is a limit on the dose a patient can receive in a lifetime [109].

Cladribine

Cladribine is a lympholytic drug that is used in the treatment of chronic progressive MS patients and is generally well tolerated [110]. Cladribine targets lymphoctyes and can kill activated myelin-reactive T cells [111]. Cladribine also affects blast cells as well as mature cells. The efficacy of cladribine had been debated as it was found not to affect disability as measured by the expanded disability status score (EDSS) [112]. It was proposed for use in long-term patients but not as a first-line treatment of MS [113]. However, a recent trial for the use of cladribine in RR patients has shown that after a 2-year period, the EDSS of patients treated with cladribine remained stable and the need for steroid treatment was reduced. Cladribine administration was associated with lower lymphocyte counts [114]. Therefore, this therapy is showing more potential for the treatment of MS than originally anticipated.

Fingolimod

Fingolimod (FTY720) was developed as an immunomodulator for use in organ transplantation [115]. It was later tested for its efficacy in EAE. It was found to protect Lewis rats from EAE and this was associated with a reduction in Th1 cytokines in the CNS [116]. FTY720 antagonises sphingosine 1-phosphate receptor-1 on T cells and inhibits the S1P/SIP1- dependent lymphocyte migration from the lymph node. It is under investigation as an oral therapy for MS. FTY720 reduces the number of naive T cells and central memory T cells in the blood, but does not affect effector T cells [117]. FTY720 limits the efflux of immune cells from the lymph nodes, which reduces peripheral lymphocyte counts and the recirculation of lymphyocytes to the CNS [118î120]. A recent study of efficacy and safety has shown that FTY720 is well tolerated and reduces relapse rate and disease activity as measured by MRI [121].

Daclizumab

Daclizumab is a monoclonal antibody against CD25 (anti-IL2-R α). IL-2R α is upregulated on activated or abnormal T cells such as those in autoimmune diseases, and thus it was chosen as a target for therapy in MS [122]. Daclizumab has been well tolerated and has been found to be successful as both an add-on treatment to IFN- β and a standalone treatment [123,124]. The anticipated mode of action of daclizumab was that it would block the proliferation of activated autoreactive T cells; however, studies have found that this may not be the most important effect of treatment. Consistently, a striking expansion of a subset of natural killer (NK) cells has been observed. These NK cells are CD16–CD56^{bright} and have immunoregulatory as well as antiviral and antitumor properties. The expansion of these CD56^{bright} NK cells is correlated with a reduction in contrast-enhancing MRI lesions [125].

Alemtuzumab

Alemtuzumab is a monoclonal antibody against CD52, which is a cell-surface glycoprotein expressed on T and B cells, eosinophils and monocytes. It is expressed on mature lymphocytes, but not on lymphoid progenitor cells, which makes it a good candidate for therapy. It is used in the treatment of chronic lymphocytic leukaemia and has been found to impressively reduce relapses in RRMS patients and also to significantly reduce the rate of sustained neurological disability [126]. The mode of action of alemtuzumab is to deplete lymphocytes. CD4⁺ T cells are depleted for approximately 60 months, and monocytes and B cells are depleted for 3 months [127]. Because of this lymphocyte depletion there are concerns regarding the adverse effects. It is associated with a risk of secondary antibody-mediated diseases such as Graves disease [128]. A recent study has demonstrated that secondary autoimmunity after treatment with alemtuzumab occurs in patients that have genetically higher levels of IL-21 prior to treatment that leads to higher levels of T-cell apoptosis and cell cycling [129]. However, as the efficacy of alemtuzumab is impressive, there are currently two Phase III trials underway to determine the safety of the drug.

Teriflunomide

Teriflunomide is the active metabolite of leflunomide, a disease-modifying agent used in the treatment of rheumatoid arthritis. It is a pyrimidine-synthesis inhibitor with selective immunosuppressive and immunomodulatory properties [130]. It can inhibit T and B lymphocyte proliferation [131]. Teriflunomide has been successful in treating EAE and is now in Phase III clinical trials as an oral treatment for MS.

Retinoic acid

Retinoic acid (RA) has been implicated as a suppressive agent in EAE for many years. Massacesi and colleagues found that EAE can be prevented in the Lewis rat by two forms of RA [132]. It came to light that RA induces the production of TGF- β [133]. More light has now been shed on this topic since it was discovered that TGF- β can convert naive T cells into Tregs, which can suppress autoimmune diseases. Thus, if RA can induce TGF- β , this can then reduce disease severity by the induction of Tregs. It is very interesting to note that TGF- β in the presence of IL-6 can result in the differentiation of naive T cells into Th17 cells. The role of retinoic acid was studied with regards to which differentiation pathway TGF- β would lead T cells into. Mucida and colleagues found that retinoic acid regulates the TGF- β response and can drive the differentiation of Tregs and also inhibit the production of IL-17 [134].

Potential targets

Regulatory T cells

CD4⁺CD25⁺Foxp3⁺ Tregs occur naturally in the thymus and control effector T-cell responses in the periphery. TGF- β can act on naive T cells to induce Foxp3 and generate induced Tregs that suppress effector T-cell immune responses. However, in the presence of TGF- β and IL-6 there is a predominant generation of Th17 cells [19]. This suggests that not only is there an antagonistic relationship between Th17 and Tregs, there is also a dichotomy in their generation. During a steady state in the immune system in the absence of inflammation, TGF- β induces the differentiation of Foxp3⁺ Tregs. However, once inflammation is initiated, IL-6 is produced by the activated innate immune system and this suppresses the generation of TGF- β -induced Tregs and generates Th17 cells [135]. There is a Treg population present in the CNS during EAE. These Tregs are functional in suppressing MOG-specific responses of naive T cells, but they fail to inhibit antigen-specific effector T cells that have been isolated from the target organ at the acute disease phase [136]. Kohm and colleagues adoptively transferred Tregs into EAE mice and found suppression of disease [137]. Viglietta and colleagues found a significant decrease in function of Tregs from peripheral blood of MS patients compared with healthy controls [138]. However, recent studies have paradoxically shown that human Tregs produce IL-17, and these IL-17-producing Tregs can still retain their suppressive function [139]. Tregs offer therapeutic potential for MS, however further study is warranted before their therapeutic potential can be reached.

IL-21

IL-21 belongs to the IL-2 family of cytokines and is critical in the differentiation of Th17 cells. In the absence of IL-6, IL-21 has been found to cooperate with TGF- β to induce the differentiation of Th17 cells [23]. IL-6-induced expression of IL-21 leads to further IL-21 induction by an autocrine loop, which also induces the IL-23 receptor (IL-23R) in naive CD4⁺ T cells [140]. IL-23 function is at least partly dependent on IL-21, which renders Th cells susceptible to IL-23 signals. IL-21 effects are dependent on the transcription factor STAT3, which is required for the differentiation of Th17 cells *in vivo* [140,141]. IL-23 induces the orphan nuclear receptor ROR γ t in turn synergizes with STAT3 to promote the expression of IL-17 [142].

Interestingly, IL-21 also appears to mediate secondary autoimmunity in MS patients after treatment with alemtuzumab [129]. A monoclonal antibody against IL-21 could offer therapeutic potential in MS.

IL-22

IL-22 is a member of the IL-10 family of cytokines. However, unlike IL-10, it exhibits proinflammatory properties. IL-22 is secreted by Th17 cells and promotes antimicrobial defenses [21]. The IL-22R is a heterodimeric receptor complex that is composed of the IL-22R1 and IL-10R2 chains [143]. IL-22 does not itself directly regulate the function of CD4⁺ T cells as IL-22R is not expressed on these cells [144]. *In vitro*, the secretion of large quantities of IL-22 by Th17 cells requires IL-23, therefore it is thought that it is produced by terminally differentiated Th17 cells [145]. Becher and colleagues generated IL-22-knockout mice and found that they were fully susceptible to EAE, which indicates that IL-22 is not crucial to the development of EAE [146]. Nevertheless, further study is needed to elucidate its exact role in autoimmune diseases.

Negative regulation of Th17 differentiation: IL-27

IL-27 is a heterodimeric cytokine that is a member of the IL-12/IL-23 superfamily. IL-27 signals through WSX-1, and mice lacking this receptor develop increased pathological inflammation during Th1 and Th2 responses, indicating an anti-inflammatory role of IL-27. IL-27 has also been found to suppress Th17 responses [26]. This suppression is STAT1 dependent. Batten and colleagues demonstrated that mice lacking the IL-27 receptor are hypersensitive to actively induced EAE and show stronger Th17 responses [25]. For the purposes of immunotherapy it is critical to determine the phase of EAE at which IL-27 suppresses disease. We examined the effect of exogenous administration of IL-27 and found that over a 7-day period of subcutaneous osmotic pump administration, disease was suppressed, indicating a therapeutic potential of this cytokine. We also studied an adoptive transfer model of EAE and showed that Th17 effector responses are suppressed after IL-27 treatment, and the potential to transfer disease is reduced [147]. We and others later found that suppression of EAE by IL-27 was dependent on IL-10 [26,148,149]. The exact role of IL-27 in MS needs to be elucidated; however, given its role in EAE, it may be therapeutically relevant in MS.

Th9 cells

Until recent years, the lineages of Th cells were divided into Th1 and Th2. Th17 was the next addition and now it seems that there may be yet another Th lineage: Th9, independently described last year by both Veldhoen and colleagues and Dardalhon and colleagues [150, 151]. They demonstrated that TGF- β can divert Th2 cells to secrete IL-9. The differentiation of Th9 cells can inhibit Foxp3 expression, thereby suppressing the generation of Tregs [150]. IL-9-producing T cells also produce IL-10. However, these IL-10-producing cells have lost their immunosuppressive function and may even be involved in tissue inflammation. Therefore, the Th9 subset is a new potential target for autoimmune diseases, including EAE and MS.

Conclusion

In the past two decades, significant advancement has been made in the treatment of MS. However, therapies are still only partially effective. We believe that as our understanding of the pathophysiology of MS continues to develop this will pave the way for future drug development directed at these discoveries. We must note that extreme caution has to be taken when transferring a successful treatment in EAE into humans. This can be highlighted from previous failed clinical trials that were successful in EAE. An open-label trial in 16 patients was performed on a monoclonal antibody, anti-CD3. The anti-CD3 infusions were associated with significant toxicity where patients developed severe T-cell cytopenia and many adverse effects, including hypotension and nausea. An increase in IFN- γ and TNF- α levels was observed and, although there were no new MRI lesions, disease worsened [152]. Another example of failed translational immunotherapy can be seen in the trial of anti-CD4 monoclonal antibody. The Phase I trial results of this antibody were promising, as there was no major toxicity, and a long lasting reduction in CD4 T cells was observed [153,154]. However in the Phase II clinical trial there was no effect on lesions as measured by gadolinium-enhanced MRI, and EDSS was also not affected [155]. MS is a disease where many different components of the immune system interact and the importance of each component also depends on a number of variables including genetic and environmental factors. Therefore, using a therapy directed at one of these components obviously has its drawbacks. We are constantly gaining a greater insight into the pathogenesis of both MS and EAE and, therefore, we should be optimistic that new therapeutic opportunities will arise.

Future perspective

Considerable effort is going into the development of oral immunotherapies, an innovation that most MS patients would welcome. As treatment options increase, we anticipate that in the next 5–10 years it will be possible to increasingly tailor immunotherapies to the characteristics of individual patients. Variables such as age, gender, disease duration, severity as measured clinically but also radiologically, previous responses to other treatments, risk/benefit ratios and, of course, patient goals and preferences will be increasingly used to make therapeutic decisions. Administration of immunotherapies at earlier stages of disease, when they are more likely to be effective, is also expected to increase. There is also agreement among experts that combination therapy will be given more frequently, again taking into account the above mentioned factors and, importantly, specific interactions and synergy between different drugs. The fact that MS is more prevalent in affluent countries makes it likely that availability of treatments will be extended to larger percentages of patients affected by this debilitating disease.

Executive summary

- CD4⁺ T helper (Th) type 1 (Th1) and Th17 cells are thought to initiate and coordinate immune responses that lead to tissue damage in experimental autoimmune encephalitis and multiple sclerosis (MS). By contrast, regulatory CD4⁺ T cells, and to some extent Th2 cells, are considered anti-inflammatory and protective. However, the immunological scenario of immune-mediated CNS damage is more complex and the contribution of these and novel T-cell subsets continues to be clarified.
- CD8⁺ T cells participate in the pathogenesis of autoimmune inflammatory demyelination and are found in higher numbers than CD4⁺ T cells in MS lesions. In the spinal fluid of patients with MS, memory CD8⁺ T cells have been reported to be selectively enriched, suggesting a role for these cells in the pathogenesis of the disease.
- Induction of immune tolerance to myelin antigens such as proteolipid protein, myelin oligodendrocyte glycoprotein and myelin basic protein is an important strategy for MS immunotherapy. Tolerance can be induced via different routes, including intravenous, intranasal and oral administration of antigen, aiming at the deletion of autoreactive T-cell clones, the induction of clonal anergy (unresponsiveness to antigen), or the expansion of antigen-specific T cells secreting anti-inflammatory cytokines.
- Anti-cytokine antibodies offer the opportunity to neutralize proinflammatory cytokines. Theoretically they are an important tool, but in clinical trials these antibodies have not been particularly successful thus far. Monoclonal antibodies aimed at blocking the interaction of activated T cells with the endothelium (anti-VLA4) or at depleting CD4⁺ T cells as well as other immune cells (anti-CD52) appear much more effective in suppressing inflammatory activity in the CNS.
- The unexpected development of a cytokine storm and systemic inflammatory response syndrome in a group of healthy volunteers treated with an anti-CD28 antibody underscores the difficulty in predicting the biological effects of antibodies proven to be safe and effective in preclinical animal models. This is relevant to T-cell directed therapies (the above mentioned antibody was a humanized superagonist anti-CD28 monoclonal antibody that stimulates and expands T cells), but also to antibody therapies directed at other immune cell populations.

 Both established and innovative T-cell therapies used in MS very often have additional effects on other immune cell populations, including B cells and different types of antigen-presenting cells. In some cases these effects are known (e.g., anti-CD52 depletes CD4⁺ and CD8⁺ T cells, as well as monocytes and macrophages), whereas in others cases the effects are less clear or are under investigation.

Bibliography

Papers of special note have been highlighted as:

- of interest
- ■■ of considerable interest
- Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. N. Engl. J. Med 2000;343(13):938–952. [PubMed: 11006371]
- 2. Sospedra M, Martin R. Immunology of multiple sclerosis. Annu. Rev. Immunol 2005;23:683–747. [PubMed: 15771584]
- 3. Grigoriadis N, Hadjigeorgiou GM. Virus-mediated autoimmunity in multiple Sclerosis. J. Autoimmune Dis 2006;3(1)
- Trapp BD, Peterson J, Ransohoff RM, et al. Axonal transection in the lesions of multiple sclerosis. N. Engl. J. Med 1998;338(5):278–285. [PubMed: 9445407]
- Peterson JW, Bo L, Mork S, Chang A, Trapp BD. Transected neurites, apoptotic neurons, and reduced inflammation in cortical multiple sclerosis lesions. Ann. Neurol 2001;50(3):389–400. [PubMed: 11558796]
- Becher B, Durell BG, Noelle RJ. Experimental autoimmune encephalitis and inflammation in the absence of interleukin-12. J. Clin. Invest 2002;110(4):493–497. [PubMed: 12189243]
- Chen Y, Langrish CL, McKenzie B, et al. Anti-IL-23 therapy inhibits multiple inflammatory pathways and ameliorates autoimmune encephalomyelitis. J. Clin. Invest 2006;116(5):1317–1326. [PubMed: 16670771]
- Gran B, Zhang G-X, Yu S, et al. IL-12p35-deficient mice are susceptible to experimental autoimmune encephalomyelitis: evidence for redundancy in the IL-12 system in the induction of central nervous system autoimmune demyelination. J. Immunol 2002;169(12):7104–7110. [PubMed: 12471147]
- Komiyama Y, Nakae S, Matsuki T, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. J. Immunol 2006;177(1):566–573. [PubMed: 16785554]
- Langrish CL, Chen Y, Blumenschein WM, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. J. Exp. Med 2005;201(2):233–240. [PubMed: 15657292]
- 11. Mosmann TR, Coffman RL. Th1 and Th2 cells: different patterns of lymphokine secretion lead to different functional properties. Annu. Rev. Immunol 1989;7:145–173. [PubMed: 2523712]
- Romagnani S, Parronchi P, D'Elios MM, et al. An update on human Th1 and Th2 cells. Int. Arch. Allergy Immunol 1997;113(1–3):153–156. [PubMed: 9130508]
- Oppmann B, Lesley R, Blom B, et al. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. Immunity 2000;13(5):715–725. [PubMed: 11114383]
- Segal BM, Shevach EM. IL-12 unmasks latent autoimmune disease in resistant mice. J. Exp. Med 1996;184(2):771–775. [PubMed: 8786337]
- 15. Segal BM, Dwyer BK, Shevach EM. An interleukin (IL)-10/IL-12 immunoregulatory circuit controls susceptibility to autoimmune disease. J. Exp. Med 1998;187(4):537–546. [PubMed: 9463404]
- 16. Cua DJ, Sherlock J, Chen Y, et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. Nature 2003;421(6924):744–748. [PubMed: 12610626]
- 17. Gran B, Chu N, Zhang GX, et al. Early administration of IL-12 suppresses EAE through induction of interferon-γ. J. Neuroimmunol 2004;156(1–2):123–131. [PubMed: 15465603]

- Aggarwal S, Ghilardi N, Xie M-H, de Sauvage FJ, Gurney AL. Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. J. Biol. Chem 2003;278 (3):1910–1914. [PubMed: 12417590]
- 19. Bettelli E, Carrier Y, Gao W, et al. Reciprocal developmental pathways for the generation of pathogenic effector Th17 and regulatory T cells. Nature. 2006
- Gutcher I, Urich E, Wolter K, Prinz M, Becher B. Interleukin 18-independent engagement of interleukin 18 receptor-α is required for autoimmune inflammation. Nat. Immunol 2006;7(9):946– 953. [PubMed: 16906165]
- 21. Liang SC, Tan XY, Luxenberg DP, et al. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. J. Exp. Med 2006;203(10):2271– 2279. [PubMed: 16982811]
- 22. Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stockinger B. TGF-b in the context of an inflammatory cytokine milieu supports *de novo* differentiation of IL-17-producing T cells. Immunity 2006;24(2):179–189. [PubMed: 16473830] ■■ The first report to show that TGF-β in the presence of IL-6 induces T-helper (Th)17 differentiation.
- 23. Korn T, Bettelli E, Gao W, et al. IL-21 initiates an alternative pathway to induce proinflammatory T (H)17 cells. Nature 2007;448(7152):484–487. [PubMed: 17581588] [23] and [24] show that IL-21 cooperates with TGF-β to induce Th17 differentiation.
- 24. Nurieva R, Yang XO, Martinez G, et al. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature 2007;448(7152):480–483. [PubMed: 17581589] [23] and [24] show that IL-21 cooperates with TGF-β to induce Th17 differentiation.
- Batten M, Li J, Yi S, et al. Interleukin-27 limits autoimmune encephalomyelitis by suppressing the development of interleukin 17-producing T cells. Nat. Immunol 2006;7(9):929–936. [PubMed: 16906167]
- 26. Stumhofer JS, Laurence A, Wilson EH, et al. Interleukin-27 negatively regulates the development of interleukin 17-producing T helper cells during chronic inflammation of the central nervous system. Nat. Immunol 2006;7(9):937–945. [PubMed: 16906166]
- 27. Laurence A, Tato CM, Davidson TS, et al. Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. Immunity 2007;26(3):371–381. [PubMed: 17363300]
- Sakaguchi S. Naturally arising CD4⁺ regulatory T cells for immunologic self-tolerance and negative control of immune responses. Ann. Rev. Immunol 2004;22(1):531–562. [PubMed: 15032588]
- Traugott U, Reinherz EL, Raine CS. Multiple sclerosis. Distribution of T cells, T-cell subsets and Iapositive macrophages in lesions of different ages. J. Neuroimmunol 1983;4(3):201–221. [PubMed: 6222066]
- 30. Brostoff SW, Mason DW. Experimental allergic encephalomyelitis: successful treatment *in vivo* with a monoclonal antibody that recognizes T helper cells. J. Immunol 1984;133(4):1938–1942. [PubMed: 6236260]
- Pettinelli CB, McFarlin DE. Adoptive transfer of experimental allergic encephalomyelitis in SJL/J mice after *in vitro* activation of lymph node cells by myelin basic protein: requirement for Lyt 1+ 2-T lymphocytes. J. Immunol 1981;127(4):1420–1423. [PubMed: 6168690]
- 32. Waldor MK, Sriram S, Hardy R, et al. Reversal of experimental allergic encephalomyelitis with monoclonal antibody to a T-cell subset marker. Science 1985;227(4685):415–417. [PubMed: 3155574]
- 33. Hemmer B, Hartung HP. Toward the development of rational therapies in multiple sclerosis: what is on the horizon? Ann. Neurol 2007;62(4):314–326. [PubMed: 17969020]
- 34. Compston A. The genetics of multiple sclerosis. J. Neurovirol 2000;6(Suppl 2):S5–S9. [PubMed: 10871776]
- Hafler DA, Compston A, Sawcer S, et al. Risk alleles for multiple sclerosis identified by a genomewide study. N. Engl. J. Med 2007;357(9):851–862. [PubMed: 17660530]
- 36. Haines JL, Ter-Minassian M, Bazyk A, et al. A complete genomic screen for multiple sclerosis underscores a role for the major histocompatability complex. The Multiple Sclerosis Genetics Group. Nat. Genet 1996;13(4):469–471. [PubMed: 8696344]

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- 37. Babbe H, Roers A, Waisman A, et al. Clonal expansions of CD8⁺ T cells dominate the T cell infiltrate in active multiple sclerosis lesions as shown by micromanipulation and single cell polymerase chain reaction. J. Exp. Med 2000;192(3):393–404. [PubMed: 10934227]
- 38. Jacobsen M, Cepok S, Quak E, et al. Oligoclonal expansion of memory CD8⁺ T cells in cerebrospinal fluid from multiple sclerosis patients. Brain 2002;125(3):538–550. [PubMed: 11872611]
- Miller A, Hafler DA, Weiner HL. Tolerance and suppressor mechanisms in experimental autoimmune encephalomyelitis: implications for immunotherapy of human autoimmune diseases. FASEB J 1991;5(11):2560–2566. [PubMed: 1868980]
- Racke MK, Bonomo A, Scott DE, et al. Cytokine-induced immune deviation as a therapy for inflammatory autoimmune disease. J. Exp. Med 1994;180(5):1961–1966. [PubMed: 7525845]
- Liblau RS, Tisch R, Shokat K, et al. Intravenous injection of soluble antigen induces thymic and peripheral T-cells apoptosis. Proc. Natl Acad. Sci. USA 1996;93(7):3031–3036. [PubMed: 8610163]
- 42. Kearney ER, Pape KA, Loh DY, Jenkins MK. Visualization of peptide-specific T cell immunity and peripheral tolerance induction. in vivo. Immunity 1994;1(4):327–339.
- 43. Miller A, Zhang ZJ, Sobel RA, al-Sabbagh A, Weiner HL. Suppression of experimental autoimmune encephalomyelitis by oral administration of myelin basic protein. VI. Suppression of adoptively transferred disease and differential effects of oral vs. intravenous tolerization. J. Neuroimmunol 1993;46(1–2):73–82. [PubMed: 7689596]
- Miller SD, McRae BL, Vanderlugt CL, et al. Evolution of the T-cell repertoire during the course of experimental immune-mediated demyelinating diseases. Immunol. Rev 1995;144:225–244. [PubMed: 7590815]
- 45. Tan JK, O'Neill HC. Maturation requirements for dendritic cells in T-cell stimulation leading to tolerance versus immunity. J. Leukoc. Biol 2005;78(2):319–324. [PubMed: 15809288]
- 46. Baker D, O'Neill JK, Gschmeissner SE, et al. Induction of chronic relapsing experimental allergic encephalomyelitis in Biozzi mice. J. Neuroimmunol 1990;28(3):261–270. [PubMed: 2373763]
- Pryce G, O'Neill JK, Croxford JL, et al. Autoimmune tolerance eliminates relapses but fails to halt progression in a model of multiple sclerosis. J. Neuroimmunol 2005;165(1–2):41–52. [PubMed: 15939483]
- Hilliard B, Ventura ES, Rostami A. Effect of timing of intravenous administration of myelin basic protein on the induction of tolerance in experimental allergic encephalomyelitis. Mult. Scler 1999;5 (1):2–9. [PubMed: 10096096]
- Hilliard BA, Kamoun M, Ventura E, Rostami A. Mechanisms of suppression of experimental autoimmune encephalomyelitis by intravenous administration of myelin basic protein: role of regulatory spleen cells. Exp. Mol. Pathol 2000;68(1):29–37. [PubMed: 10640452]
- Zhang GX, Yu S, Li Y, et al. A paradoxical role of APCs in the induction of intravenous tolerance in experimental autoimmune encephalomyelitis. J. Neuroimmunol 2005;161(1–2):101–112. [PubMed: 15748949]
- 51. Li H, Zhang GX, Chen Y, et al. CD11c+CD11b+ dendritic cells play an important role in intravenous tolerance and the suppression of experimental autoimmune encephalomyelitis. J. Immunol 2008;181 (4):2483–2493. [PubMed: 18684939]
- Jiang Z, Li H, Fitzgerald DC, Zhang GX, Rostami A. MOG(35-55) i.v suppresses experimental autoimmune encephalomyelitis partially through modulation of Th17 and JAK/STAT pathways. Eur. J. Immunol 2009;39(3):789–799. [PubMed: 19224632]
- London CA, Lodge MP, Abbas AK. Functional responses and costimulator dependence of memory CD4⁺ T cells. J. Immunol 2000;164(1):265–272. [PubMed: 10605020]
- 54. Verbeek R, van der Mark K, Wawrousek EF, Plomp AC, van Noort JM. Tolerization of an established αβ-crystallin-reactive T-cell response by intravenous antigen. Immunology 2007;121(3):416–426. [PubMed: 17386078]
- 55. Bai XF, Shi FD, Xiao BG, et al. Nasal administration of myelin basic protein prevents relapsing experimental autoimmune encephalomyelitis in DA rats by activating regulatory cells expressing IL-4 and TGF-β mRNA. J. Neuroimmunol 1997;80(1–2):65–75. [PubMed: 9413260]
- Burkhart C, Liu GY, Anderton SM, Metzler B, Wraith DC. Peptide-induced T cell regulation of experimental autoimmune encephalomyelitis: a role for IL-10. Int. Immunol 1999;11(10):1625– 1634. [PubMed: 10508180]

- Bitar DM, Whitacre CC. Suppression of experimental autoimmune encephalomyelitis by the oral administration of myelin basic protein. Cell Immunol 1988;112(2):364–370. [PubMed: 2451570]
- Higgins PJ, Weiner HL. Suppression of experimental autoimmune encephalomyelitis by oral administration of myelin basic protein and its fragments. J. Immunol 1988;140(2):440–445. [PubMed: 2447178]
- 59. Weiner HL, Mackin GA, Matsui M, et al. Double-blind pilot trial of oral tolerization with myelin antigens in multiple sclerosis. Science 1993;259(5099):1321–1324. [PubMed: 7680493]
- 60. Faria AM, Weiner HL. Oral tolerance. Immunol. Rev 2005;206:232-259. [PubMed: 16048553]
- 61. Ochi H, Abraham M, Ishikawa H, et al. Oral CD3-specific antibody suppresses autoimmune encephalomyelitis by inducing CD4⁺CD25⁻LAP⁺ T cells. Nat. Med 2006;12(6):627–635. [PubMed: 16715091] Immune tolerance was induced by administration of oral CD3-specific antibody. Tolerance correlated to the expansion of a CD4⁺CD25⁻LAP⁺ population of regulatory T cells.
- Evavold BD, Allen PM. Separation of IL-4 production from Th cell proliferation by an altered T cell receptor ligand. Science 1991;252(5010):1308–1310. [PubMed: 1833816]
- 63. Racioppi L, Ronchese F, Matis LA, Germain RN. Peptide-major histocompatibility complex class II complexes with mixed agonist/antagonist properties provide evidence for ligand-related differences in T cell receptor-dependent intracellular signaling. J. Exp. Med 1993;177(4):1047–1060. [PubMed: 8384651]
- 64. De Magistris MT, Alexander J, Coggeshall M, et al. Antigen analog-major histocompatibility complexes act as antagonists of the T cell receptor. Cell 1992;68(4):625–634. [PubMed: 1739971]
- 65. Brocke S, Gijbels K, Allegretta M, et al. Treatment of experimental encephalomyelitis with a peptide analogue of myelin basic protein. Nature 1996;379(6563):343–346. [PubMed: 8552189]
- 66. Goverman J, Woods A, Larson L, et al. Transgenic mice that express a myelin basic protein-specific T cell receptor develop spontaneous autoimmunity. Cell 1993;72(4):551–560. [PubMed: 7679952]
- 67. Brabb T, Goldrath AW, von Dassow P, et al. Triggers of autoimmune disease in a murine TCRtransgenic model for multiple sclerosis. J. Immunol 1997;159(1):497–507. [PubMed: 9200491]
- 68. Lafaille JJ, Nagashima K, Katsuki M, Tonegawa S. High incidence of spontaneous autoimmune encephalomyelitis in immunodeficient anti-myelin basic protein T cell receptor transgenic mice. Cell 1994;78(3):399–408. [PubMed: 7520367]
- Offner H, Adlard K, Bebo BF Jr, et al. Vaccination with BV8S2 protein amplifies TCR-specific regulation and protection against experimental autoimmune encephalomyelitis in TCR BV8S2 transgenic mice. J. Immunol 1998;161(5):2178–2186. [PubMed: 9725209]
- Vandenbark AA, Rich C, Mooney J, et al. Recombinant TCR ligand induces tolerance to myelin oligodendrocyte glycoprotein 35–55 peptide and reverses clinical and histological signs of chronic experimental autoimmune encephalomyelitis in HLA-DR2 transgenic mice. J. Immunol 2003;171 (1):127–133. [PubMed: 12816990]
- Moisini I, Nguyen P, Fugger L, Geiger TL. Redirecting therapeutic T cells against myelin-specific T lymphocytes using a humanized myelin basic protein-HLA-DR2-ζ chimeric receptor. J. Immunol 2008;180(5):3601–3611. [PubMed: 18292588]
- Ito K, Bian HJ, Molina M, et al. HLA-DR4-IE chimeric class II transgenic, murine class II-deficient mice are susceptible to experimental allergic encephalomyelitis. J. Exp. Med 1996;183(6):2635– 2644. [PubMed: 8676084]
- Muraro PA, Vergelli M, Kalbus M, et al. Immunodominance of a low-affinity major histocompatibility complex-binding myelin basic protein epitope (residues 111–129) in HLA-DR4 (B1*0401) subjects is associated with a restricted T cell receptor repertoire. J. Clin. Invest 1997;100 (2):339–349. [PubMed: 9218510]
- 74. Huh J, Yao K, Quigley L, et al. Limited repertoire of HLA-DRB1*0401-restricted MBP111-129specific T cells in HLA-DRB1*0401 Tg mice and their pathogenic potential. J. Neuroimmunol 2004;151(1–2):94–102. [PubMed: 15145608]
- 75. Fridkis-Hareli M, Teitelbaum D, Gurevich E, et al. Direct binding of myelin basic protein and synthetic copolymer 1 to class II major histocompatibility complex molecules on living antigenpresenting cells – specificity and promiscuity. Proc. Natl Acad. Sci. USA 1994;91(11):4872–4876. [PubMed: 7515181]

- Vandenbark AA, Chou YK, Whitham R, et al. Treatment of multiple sclerosis with T-cell receptor peptides: results of a double-blind pilot trial. Nat. Med 1996;2(10):1109–1115. [PubMed: 8837609]
- 77. Vandenbark AA, Morgan E, Bartholomew R, et al. TCR peptide therapy in human autoimmune diseases. Neurochem. Res 2001;26(6):713–730. [PubMed: 11519731]
- 78. Bielekova B, Goodwin B, Richert N, et al. Encephalitogenic potential of the myelin basic protein peptide (amino acids 83–99) in multiple sclerosis: results of a Phase II clinical trial with an altered peptide ligand. Nat. Med 2000;6(10):1167–1175. [PubMed: 11017150]
- Leonard JP, Waldburger KE, Goldman SJ. Prevention of experimental autoimmune encephalomyelitis by antibodies against interleukin-12. J. Exp. Med 1995;181(1):381–386. [PubMed: 7528773]
- Constantinescu CS, Wysocka M, Hilliard B, et al. Antibodies against IL-12 prevent superantigeninduced andspontaneous relapses of experimental autoimmune encephalomyelitis. J. Immunol 1998;161(9):5097–5104. [PubMed: 9794448]
- Ichikawa M, Koh CS, Inoue A, et al. Anti-IL-12 antibody prevents the development and progression of multiple sclerosis-like relapsing-remitting demyelinating disease in NOD mice induced with myelin oligodendrocyte glycoprotein peptide. J. Neuroimmunol 2000;102(1):56–66. [PubMed: 10626667]
- 82. Segal BM, Constantinescu CS, Raychaudhuri A, et al. Repeated subcutaneous injections of IL12/23 p40 neutralising antibody, ustekinumab, in patients with relapsing-remitting multiple sclerosis: a Phase II, double-blind, placebo-controlled, randomised, dose-ranging study. Lancet Neurol 2008;7 (9):796–804. [PubMed: 18703004] Neutralization of IL-12 and IL-23 was tested. The negative result may have been due to the inclusion of patients with advanced disease.
- Hofstetter HH, Ibrahim SM, Koczan D, et al. Therapeutic efficacy of IL-17 neutralization in murine experimental autoimmune encephalomyelitis. Cell Immunol 2005;237(2):123–130. [PubMed: 16386239]
- 84. Suntharalingam G, Perry MR, Ward S, et al. Cytokine storm in a Phase 1 trial of the anti-CD28 monoclonal antibody TGN1412. N. Engl. J. Med 2006;355(10):1018–1028. [PubMed: 16908486]
 Highlights the need for extreme caution in the translation of therapies that are considered safe and effective in animal models to use in human subjects.
- 85. Jiang H, Milo R, Swoveland P, et al. Interferon β-1b reduces interferon γ-induced antigen-presenting capacity of human glial and B cells. J. Neuroimmunol 1995;61(1):17–25. [PubMed: 7560008]
- 86. Panitch HS, Hirsch RL, Schindler J, Johnson KP. Treatment of multiple sclerosis with γ-interferon: exacerbations associated with activation of the immune system. Neurology 1987;37(7):1097–1102. [PubMed: 3110648]
- Teleshova N, Bao W, Kivisakk P, et al. Elevated CD40 ligand expressing blood T-cell levels in multiple sclerosis are reversed by interferon-β treatment. Scand. J. Immunol 2000;51(3):312–320. [PubMed: 10736102]
- Genc K, Dona DL, Reder AT. Increased CD80⁺ B cells in active multiple sclerosis and reversal by interferon β-1b therapy. J. Clin. Invest 1997;99(11):2664–2671. [PubMed: 9169496]
- 89. Dhib-Jalbut S. Mechanisms of interferon-β action in multiple sclerosis. Mult. Scler 1997;3(6):397–401. [PubMed: 9493642]
- 90. Rep MH, Schrijver HM, van Lopik T, et al. Interferon (IFN)-β treatment enhances CD95 and interleukin 10 expression but reduces interferon-γ producing T cells in MS patients. J. Neuroimmunol 1999;96(1):92–100. [PubMed: 10227428]
- 91. Calabresi PA, Tranquill LR, Dambrosia JM, et al. Increases in soluble VCAM-1 correlate with a decrease in MRI lesions in multiple sclerosis treated with interferon β-1b. Ann. Neurol 1997;41(5): 669–674. [PubMed: 9153530]
- 92. Dhib-Jalbut S, Jiang H, Williams GJ. The effect of interferon β-1b on lymphocyte-endothelial cell adhesion. J. Neuroimmunol 1996;71(1–2):215–222. [PubMed: 8982122]
- 93. Schreiner B, Mitsdoerffer M, Kieseier BC, et al. Interferon-β enhances monocyte and dendritic cell expression of B7-H1 (PD-L1), a strong inhibitor of autologous T-cell activation: relevance for the immune modulatory effect in multiple sclerosis. J. Neuroimmunol 2004;155(1–2):172–182. [PubMed: 15342209]

- 94. Shapiro S, Galboiz Y, Lahat N, Kinarty A, Miller A. The `immunological-synapse' at its APC side in relapsing and secondary-progressive multiple sclerosis: modulation by interferon-β. J. Neuroimmunol 2003;144(1–2):116–124. [PubMed: 14597105]
- 95. Gran B, Tranquill LR, Chen M, et al. Mechanisms of immunomodulation by glatiramer acetate. Neurology 2000;55(11):1704–1714. [PubMed: 11113226]
- Neuhaus O, Farina C, Wekerle H, Hohlfeld R. Mechanisms of action of glatiramer acetate in multiple sclerosis. Neurology 2001;56(6):702–708. [PubMed: 11288751]
- Duda PW, Schmied MC, Cook SL, Krieger JI, Hafler DA. Glatiramer acetate (Copaxone®) induces degenerate, Th2-polarized immune responses in patients with multiple sclerosis. J. Clin. Invest 2000;105(7):967–976. [PubMed: 10749576]
- 98. Weber MS, Starck M, Wagenpfeil S, et al. Multiple sclerosis: glatiramer acetate inhibits monocyte reactivity *in vitro* and *in vivo*. Brain 2004;127(Pt 6):1370–1378. [PubMed: 15090474]
- 99. Weber MS, Prod'homme T, Youssef S, et al. Type II monocytes modulate T cell-mediated central nervous system autoimmune disease. Nat. Med 2007;13(8):935–943. [PubMed: 17676050]
- 100. Maier K, Kuhnert AV, Taheri N, et al. Effects of glatiramer acetate and interferon-β on neurodegeneration in a model of multiple sclerosis:a comparative study. Am. J. Pathol 2006;169 (4):1353–1364. [PubMed: 17003491]
- 101. Yednock TA, Cannon C, Fritz LC, et al. Prevention of experimental autoimmune encephalomyelitis by antibodies against α4β1 integrin. Nature 1992;356(6364):63–66. [PubMed: 1538783]
- 102. Stuve O, Marra CM, Jerome KR, et al. Immune surveillance in multiple sclerosis patients treated with natalizumab. Ann. Neurol 2006;59(5):743–747. [PubMed: 16634029]
- 103. Stuve O. The effects of natalizumab on the innate and adaptive immune system in the central nervous system. J. Neurol. Sci 2008;274(1–2):39–41. [PubMed: 18474372]
- 104. Massacesi L, Parigi A, Barilaro A, et al. Efficacy of azathioprine on multiple sclerosis new brain lesions evaluated using magnetic resonance imaging. Arch. Neurol 2005;62(12):1843–1847. [PubMed: 16344342]
- 105. Amato MP, Pracucci G, Ponziani G, et al. Long-term safety of azathioprine therapy in multiple sclerosis. Neurology 1993;43(4):831–833. [PubMed: 8469348]
- 106. Casetta I, Iuliano G, Filippini G. Azathioprine for multiple sclerosis. Cochrane Database Syst. Rev 2007;(4):CD003982. [PubMed: 17943809]
- 107. Fox EJ. Mechanism of action of mitoxantrone. Neurology 2004;63(12 Suppl 6):S15–S18. [PubMed: 15623664]
- 108. Martinelli Boneschi F, Rovaris M, Capra R, Comi G. Mitoxantrone for multiple sclerosis. Cochrane Database Syst. Rev 2005;(4):CD002127. [PubMed: 16235298]
- 109. Fox EJ. Management of worsening multiple sclerosis with mitoxantrone: a review. Clin. Ther 2006;28(4):461–474. [PubMed: 16750460]
- Sipe JC, Romine JS, Koziol JA, et al. Cladribine in treatment of chronic progressive multiple sclerosis. Lancet 1994;344(8914):9–13. [PubMed: 7912347]
- 111. Sipe JC, Romine JS, Koziol JA, et al. Development of cladribine treatment in multiple sclerosis. Mult. Scler 1996;1(6):343–347. [PubMed: 9345414]
- 112. Rice GP, Filippi M, Comi G. Cladribine and progressive MS: clinical and MRI outcomes of a multicenter controlled trial. Cladribine MRI Study Group. Neurology 2000;54(5):1145–1155. [PubMed: 10720289]
- Brousil JA, Roberts RJ, Schlein AL. Cladribine: an investigational immunomodulatory agent for multiple sclerosis. Ann. Pharmacother 2006;40(10):1814–1821. [PubMed: 16985095]
- 114. Stelmasiak Z, Solski J, Nowicki J, et al. Effect of parenteral cladribine on relapse rates in patients with relapsing forms of multiple sclerosis: results of a 2-year, double-blind, placebo-controlled, crossover study. Mult. Scler 2009;15(6):767–770. [PubMed: 19482866]
- Budde K, Schmouder RL, Brunkhorst R, et al. First human trial of FTY720, a novel immunomodulator, in stable renal transplant patients. J. Am. Soc. Nephrol 2002;13(4):1073–1083. [PubMed: 11912269]

- 116. Fujino M, Funeshima N, Kitazawa Y, et al. Amelioration of experimental autoimmune encephalomyelitis in Lewis rats by FTY720 treatment. J. Pharmacol. Exp. Ther 2003;305(1):70– 77. [PubMed: 12649354]
- 117. Mehling M, Brinkmann V, Antel J, et al. FTY720 therapy exerts differential effects on T cell subsets in multiple sclerosis. Neurology 2008;71(16):1261–1267. [PubMed: 18852441]
- Brinkmann V, Cyster JG, Hla T. FTY720: sphingosine 1-phosphate receptor-1 in the control of lymphocyte egress and endothelial barrier function. Am. J. Transplant 2004;4(7):1019–1025. [PubMed: 15196057]
- 119. Mandala S, Hajdu R, Bergstrom J, et al. Alteration of lymphocyte trafficking by sphingosine-1-phosphate receptor agonists. Science 2002;296(5566):346–349. [PubMed: 11923495]
- 120. Matloubian M, Lo CG, Cinamon G, et al. Lymphocyte egress from thymus and peripheral lymphoid organs is dependent on S1P receptor 1. Nature 2004;427(6972):355–360. [PubMed: 14737169]
- 121. O'Connor P, Comi G, Montalban X, et al. Oral fingolimod (FTY720) in multiple sclerosis: two-year results of a Phase II extension study. Neurology 2009;72(1):73–79. [PubMed: 19122034]
- 122. Waldmann TA. Anti-Tac (daclizumab, Zenapax®) in the treatment of leukemia, autoimmune diseases, and in the prevention of allograft rejection: a 25-year personal odyssey. J. Clin. Immunol 2007;27(1):1–18. [PubMed: 17216565]
- 123. Bielekova B, Richert N, Howard T, et al. Humanized anti-CD25 (daclizumab) inhibits disease activity in multiple sclerosis patients failing to respond to interferon β. Proc. Natl Acad. Sci. USA 2004;101(23):8705–8708. [PubMed: 15161974]
- 124. Rose JW, Watt HE, White AT, Carlson NG. Treatment of multiple sclerosis with an antiinterleukin-2 receptor monoclonal antibody. Ann. Neurol 2004;56(6):864–867. [PubMed: 15499632]
- 125. Bielekova B, Catalfamo M, Reichert-Scrivner S, et al. Regulatory CD56^{bright} natural killer cells mediate immunomodulatory effects of IL-2Ra-targeted therapy (daclizumab) in multiple sclerosis. Proc. Natl Acad. Sci. USA 2006;103(15):5941–5946. [PubMed: 16585503]
- 126 ■. Coles AJ, Compston DA, Selmaj KW, et al. Alemtuzumab vs. interferon β-1a in early multiple sclerosis. N. Engl. J. Med 2008;359(17):1786–1801. [PubMed: 18946064] Alemtuzumab is a monoclonal antibody against CD52. It is considered the most effective experimental therapy for patients with early relapsing–remitting multiple sclerosis. Careful monitoring is required for potential immune-mediated side effects.
- 127. Cox AL, Thompson SA, Jones JL, et al. Lymphocyte homeostasis following therapeutic lymphocyte depletion in multiple sclerosis. Eur. J. Immunol 2005;35(11):3332–3342. [PubMed: 16231285]
- 128. Coles AJ, Wing M, Smith S, et al. Pulsed monoclonal antibody treatment and autoimmune thyroid disease in multiple sclerosis. Lancet 1999;354(9191):1691–1695. [PubMed: 10568572]
- 129. Jones JL, Phuah CL, Cox AL, et al. IL-21 drives secondary autoimmunity in patients with multiple sclerosis, following therapeutic lymphocyte depletion with alemtuzumab (Campath-1H). J. Clin. Invest. 2009
- Herrmann ML, Schleyerbach R, Kirschbaum BJ. Leflunomide: an immunomodulatory drug for the treatment of rheumatoid arthritis and other autoimmune diseases. Immunopharmacology 2000;47 (2–3):273–289. [PubMed: 10878294]
- 131. Korn T, Magnus T, Toyka K, Jung S. Modulation of effector cell functions in experimental autoimmune encephalomyelitis by leflunomide – mechanisms independent of pyrimidine depletion. J. Leukoc. Biol 2004;76(5):950–960. [PubMed: 15328336]
- 132. Massacesi L, Abbamondi AL, Sarlo F, Amaducci L. The control of experimental allergic encephalomyelitis with retinoic acid. Further studies. Riv. Neurol 1987;57(3):166–169. [PubMed: 3499654]
- 133. Roberts AB, Sporn MB. Mechanistic interrelationships between two superfamilies: the steroid/ retinoid receptors and transforming growth factor-β. Cancer Surv 1992;14:205–220. [PubMed: 1330304]
- 134. Mucida D, Park Y, Kim G, et al. Reciprocal Th17 and regulatory T cell differentiation mediated by retinoic acid. Science 2007;317(5835):256–260. [PubMed: 17569825]
- Bettelli E, Oukka M, Kuchroo VK. Th-17 cells in the circle of immunity and autoimmunity. Nat. Immunol 2007;8(4):345–350. [PubMed: 17375096]

- 136. Korn T, Reddy J, Gao W, et al. Myelin-specific regulatory T cells accumulate in the CNS but fail to control autoimmune inflammation. Nat. Med 2007;13(4):423–431. [PubMed: 17384649]
- 137. Kohm AP, Carpentier PA, Anger HA, Miller SD. Cutting edge: CD4⁺CD25⁺ regulatory T cells suppress antigen-specific autoreactive immune responses and central nervous system inflammation during active experimental autoimmune encephalomyelitis. J. Immunol 2002;169(9):4712–4716. [PubMed: 12391178]
- 138. Viglietta V, Baecher-Allan C, Weiner HL, Hafler DA. Loss of functional suppression by CD4⁺CD25 ⁺ regulatory T cells in patients with multiple sclerosis. J. Exp. Med 2004;199(7):971–979. [PubMed: 15067033]
- 139. Beriou G, Costantino CM, Ashley CW, et al. IL-17 producing human peripheral regulatory T cells retain suppressive function. Blood 2009;113(18):4240–4249. [PubMed: 19171879]
- 140. Zhou L, Ivanov, II, Spolski R, et al. IL-6 programs Th-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat. Immunol 2007;8(9):967–974. [PubMed: 17581537]
- 141. Wei L, Laurence A, Elias KM, O'Shea JJ. IL-21 is produced by Th17 cells and drives IL-17 production in a STAT3-dependent manner. J. Biol. Chem 2007;282(48):34605–34610. [PubMed: 17884812]
- 142. Huber M, Brustle A, Reinhard K, et al. IRF4 is essential for IL-21-mediated induction, amplification, and stabilization of the Th17 phenotype. Proc. Natl Acad. Sci. USA 2008;105(52):20846–20851. [PubMed: 19088203]
- 143. Wolk K, Sabat R. Interleukin-22: a novel T- and NK-cell derived cytokine that regulates the biology of tissue cells. Cytokine Growth Factor Rev 2006;17(5):367–380. [PubMed: 17030002]
- 144. Chung Y, Yang X, Chang SH, et al. Expression and regulation of IL-22 in the IL-17-producing CD4 ⁺ T lymphocytes. Cell Res 2006;16(11):902–907. [PubMed: 17088898]
- 145. Zheng Y, Valdez PA, Danilenko DM, et al. Interleukin-22 mediates early host defense against attaching and effacing bacterial pathogens. Nat. Med 2008;14(3):282–289. [PubMed: 18264109]
- 146. Kreymborg K, Etzensperger R, Dumoutier L, et al. IL-22 is expressed by Th17 cells in an IL-23dependent fashion, but not required for the development of autoimmune encephalomyelitis. J. Immunol 2007;179(12):8098–8104. [PubMed: 18056351]
- 147. Fitzgerald DC, Ciric B, Touil T, et al. Suppressive effect of IL-27 on encephalitogenic Th17 cells and the effector phase of experimental autoimmune encephalomyelitis. J. Immunol 2007;179(5): 3268–3275. [PubMed: 17709543]
- 148. Awasthi A, Carrier Y, Peron JP, et al. A dominant function for interleukin 27 in generating interleukin 10-producing anti-inflammatory T cells. Nat. Immunol 2007;8(12):1380–1389. [PubMed: 17994022]
- 149 . Fitzgerald DC, Zhang GX, El-Behi M, et al. Suppression of autoimmune inflammation of the central nervous system by interleukin 10 secreted by interleukin 27-stimulated T cells. Nat. Immunol 2007;8(12):1372–1379. [PubMed: 17994023] IL-27 is a negative regulator of Th17. It suppresses autoimmune inflammation by secreting IL-10.
- 150 . Dardalhon V, Awasthi A, Kwon H, et al. IL-4 inhibits TGF-β-induced Foxp3⁺ T cells and, together with TGF-β, generates IL-9⁺ IL-10⁺ Foxp3⁻ effector T cells. Nat. Immunol 2008;9(12):1347–1355. [PubMed: 18997793] [150] and [151] introduced a new lineage of T helper cells designated as Th9. These effector cells can inhibit the generation of regulatory T cells by suppressing Foxp3.
- 151 ■. Veldhoen M, Uyttenhove C, van Snick J, et al. Transforming growth factor-β `reprograms' the differentiation of T helper 2 cells and promotes an interleukin 9-producing subset. Nat. Immunol 2008;9(12):1341–1346. [PubMed: 18931678] [150] and [151] introduced a new lineage of T helper cells designated as Th9. These effector cells can inhibit the generation of Treg cells by suppressing Foxp3.
- 152. Weinshenker BG, Bass B, Karlik S, Ebers GC, Rice GP. An open trial of OKT3 in patients with multiple sclerosis. Neurology 1991;41(7):1047–1052. [PubMed: 1906145]
- 153. Lindsey JW, Hodgkinson S, Mehta R, et al. Repeated treatment with chimeric anti-CD4 antibody in multiple sclerosis. Ann. Neurol 1994;36(2):183–189. [PubMed: 8053654]
- 154. Lindsey JW, Hodgkinson S, Mehta R, et al. Phase 1 clinical trial of chimeric monoclonal anti-CD4 antibody in multiple sclerosis. Neurology 1994;44(3 Pt 1):413–419. [PubMed: 8145907]

155. van Oosten BW, Lai M, Barkhof F, et al. A phase II trial of anti-CD4 antibodies in the treatment of multiple sclerosis. Mult. Scler 1996;1(6):339–342. [PubMed: 9345413]

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Figure 1. Pathogenesis of experimental autoimmune encephalomyelitis and multiple sclerosis

In the peripheral immune system, APCs (mainly DCs but also B cells and macrophages) produce IL-12, TGF- β and IL-6, and prime naive CD4⁺ T cells during activation by interaction of the T-cell receptor with cognate antigens presented by MHC class II complexes. Activated Th1 and Th17 cells cross the BBB into the CNS. Resident APCs interact with activated myelin-specific Th1 and Th17 cells, which become reactivated (Th2 cells can also be primed by APCs). Inflammatory cytokines and chemokines are produced and an inflammatory cascade is initiated. Infiltrating macrophages become activated and these, alongside antibodies, attack the myelin sheath on neurons.

APC: Antigen-presenting cell; BBB: Blood-brain barrier; NO: Nitric oxide; ROS: Reactive oxygen species; Th: T helper.



Figure 2. T-cell differentiation

Naive T cells differentiate into four different subsets of effector cells depending on the cytokines present during initial activation. First, IL-12 induces a Th1 differentiation program via activation of the transcription factors, STAT1, STAT4 and T-bet resulting in IFN- γ production. Second, IL-4 induces Th2 differentiation via the transcription factors, STAT6 and GATA3 resulting in the production of IL-4, IL-5 and IL-13 by Th2 effector cells. Third, in the presence of IL-6, TGF- β drives Th17 differentiation via activation of the transcription factors ROR γ t and STAT3. IL-23 also supports the Th17 lineage of effector T cells, which produce IL-17A and IL-17F. However, when naive T cells are exposed to TGF- β only (without IL-6), the transcription factor FoxP3 is activated, resulting in the differentiation of regulatory T cells that produce IL-10.

Th: T helper; Treg: Regulatory T cell; nTreg: Naturally occurring Treg.

Table 1

Current and potential T-cell immunotherapies for multiple sclerosis.

Treatments currently used	Mechanis	m/targets	Commen	nts
IFN-6: type I interferon with immunoregulatory and antiviral properties	Targets T cells, and blood-brai productior other unkr	cells, antigen-presenting cells of the n barrier; induces the n of IL-10; nown mechanisms	Most con in RRMS	nmonly used treatment
GA : random copolymer of alanine, lysine, glutamic acid and tyrosine	GA can bi molecules and thereb myelin bas protein-rea	nd to HLA class II including HLA DR2, y inhibits the activation of sic active T cells	Used as a	a treatment for RRMS
Anti-adhesion molecule antibodies: directed against very late antigen-4 and other adhesion molecules (e.g., natalizumab)	Blocks act entering th	ivated leukocytes from le CNS	Most pot modifyin concerns immunos	ent licensed disease- g treatment for RRMS; for suppression in rare cases
Immunosuppressants: used to generally suppress immune system reactions (e.g., azathioprine and mitoxantrone)	Azathiopri inhibitor, i proliferati leukocytes Mitoxantr immunosu inhibits ty	ine is a purine-synthesis inhibiting the on of cells, especially one is a stronger ppressive drug that pe II topisomerase	Azathiop oral treat Mitoxant intravence aggressiv with rela There is a effects, a	prine has been used as an ment for MS; rone is given ous and is used for more re disease (RR or SP pses); caution regarding side s the immune system
			is not fur potential	nctioning to its full
Transformation to stand from 3 and incored in a stand		Maahanim #awaata	is not fur potential	Comments
Treatments tested/under investigation Tolerizing agents: the administration of tolerogenic forms of myelin to induce antigen-specific inhibition of disease		Mechanism/targets Apoptosis of antigen spec cells, anergy of T cells to antigens induction of anti specific cells that negativ regulate autoimmune cell	is not fur potential cific T gen- ely s	Comments Three types of administration intravenous, intranasal and oral; potential for individually tailored antigen-specific treatment
Treatments tested/under investigation Tolerizing agents: the administration of tolerogenic forms of myelin to induce antigen-specific inhibition of disease Altered peptide ligands: MHC binding peptide modified by substitution of one of more amino acids to change the MHC or receptor binding characteristics	antigenic or T-cell	Mechanism/targets Apoptosis of antigen spec cells, anergy of T cells to antigens induction of anti specific cells that negativ, regulate autoimmune cell Tolerance can be induced mechanisms including pa activation of T cells and antagonism at the T-cell a level	is not fur potential cific T gen- ely s	Comments Three types of administration intravenous, intranasal and oral; potential for individually tailored antigen-specific treatment Need to target specific MHC/HLA types to reduce potential side effects, including exacerbation of disease
Treatments tested/under investigation Tolerizing agents: the administration of tolerogenic forms of myelin to induce antigen-specific inhibition of disease Altered peptide ligands: MHC binding peptide modified by substitution of one of more amino acids to change the MHC or receptor binding characteristics Cytokine-based therapies:	antigenic or T-cell	Mechanism/targets Apoptosis of antigen spec cells, anergy of T cells to antigens induction of anti specific cells that negative regulate autoimmune cell Tolerance can be induced mechanisms including pa activation of T cells and antagonism at the T-cell a level	is not fur potential cific T gen- ely s by rtial ctivation	Comments Three types of administration intravenous, intranasal and oral; potential for individually tailored antigen-specific treatment Need to target specific MHC/HLA types to MHC/HLA types to reduce potential side effects, including exacerbation of disease
Treatments tested/under investigation Tolerizing agents: the administration of tolerogenic forms of myelin to induce antigen-specific inhibition of disease Altered peptide ligands: MHC binding peptide modified by substitution of one of more amino acids to change the MHC or receptor binding characteristics Cytokine-based therapies: a) Anti-cytokine monoclonal antibodie Neutralization of cytokines relevant to di (e.g., anti-IL-12 [ustekinumab]), anti-IL-	antigenic or T-cell sisease 23 anti-	Mechanism/targets Apoptosis of antigen spec cells, anergy of T cells to antigens induction of anti specific cells that negativ regulate autoimmune cell Tolerance can be induced mechanisms including pa activation of T cells and antagonism at the T-cell a level Blocking of cytokines tha considered crucial in dise pathogenesis	is not fur potential cific T gen- ely s by rtial ctivation	Comments Three types of administration intravenous, intranasal and oral; potential for individually tailored antigen-specific treatment Need to target specific MHC/HLA types to reduce potential side effects, including exacerbation of disease Limited efficacy to date

Treatments tested/under investigation	Mechanism/targets	Comments
RA : the oxidized form of vitamin A; immunomodulation	RA induces the production of TGF- β that can convert naive T cells into Tregs. RA has also been found to inhibit IL-17 production	Good characterization of pharmacokinetics in human therapy
Tregs : naturally occurring Tregs can inhibit autoimmune diseases by suppressing the function of effector T cells	Methods to expand populations of Tregs could provide the rapeutic potential by suppressing the function of effector Th1 and Th17 cells	Cell-based therapy would be individualized
Immunoregulators: used to generally suppress immune system reactions (e.g., fingolimod)	Immunoregulators can reduce the number of Th cells in the CNS. Fingolimod reduces Th1 cytokines in CNS by antagonising sphingosine 1-phosphate receptor-1 on T cells, which in turn inhibits lymphocyte migration	There is caution regarding side effects, as the immune system is not functioning to its full potential

GA: Glatiramer acetate; MS: Multiple sclerosis; RA: Retinoic acid; RR: Relapsing-remitting; SP: Secondary progressive; Tregs: Regulatory T cells.