

APOC1 T45S polymorphism is associated with reduced obesity indices and lower plasma concentrations of leptin and apolipoprotein C-I in aboriginal Canadians

Piya Lahiry,* Henian Cao,* Matthew R. Ban,* Rebecca L. Pollex,* Mary Mamakeesick,[†] Bernard Zinman,[†] Stewart B. Harris,[§] Anthony J. G. Hanley,** Murray W. Huff,* Philip W. Connelly,^{††} and Robert A. Hegele^{1,*}

Robarts Research Institute,* Schulich School of Medicine and Dentistry, University of Western Ontario, London, ON, Canada; Sandy Lake Health and Diabetes Project,[†] Sandy Lake, ON; Thames Valley Family Practice Research Unit,[§] University of Western Ontario, London, ON, Canada; Department of Medicine and Nutritional Sciences,** University of Toronto, and Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, ON, Canada; and St. Michael's Hospital,^{††} Toronto, ON, Canada

Abstract Apolipoprotein (apo) C-I is a constituent of chylomicrons, very low density lipoprotein, and high density lipoprotein. The role of apo C-I in human metabolism is incompletely defined. We took advantage of a naturally occurring amino acid polymorphism that is present in aboriginal North Americans, namely apo C-I T45S. We assessed the hypothesis that metabolic traits, including obesity-related and lipoprotein-related traits, would differ between carriers and noncarriers of apo C-I T45S. A genotyping assay was developed for *APOC1* T45S and genotypes were determined in a sample of 410 Canadian Oji-Cree subjects. The allele frequency of the apo C-I S45 allele was ~8% in this sample. We observed the apo C-I S45 allele was significantly associated with 1) lower percent body fat ($P < 0.05$), 2) lower waist circumference ($P = 0.058$), 3) lower serum leptin levels ($P < 0.05$), and 4) lower plasma apo C-I levels ($P < 0.0001$), using a newly developed ELISA-based method. Taken together, these results suggest that at the whole human phenotype level, apo C-I is associated with the complex metabolic trait of obesity as well as with serum leptin levels.—Lahiry, P., H. Cao, M. R. Ban, R. L. Pollex, M. Mamakeesick, B. Zinman, S. B. Harris, A. J. G. Hanley, M. W. Huff, P. W. Connelly, and R. A. Hegele. *APOC1* T45S polymorphism is associated with reduced obesity indices and lower plasma concentrations of leptin and apolipoprotein C-I in aboriginal Canadians. *J. Lipid Res.* 2010. 51: 843–848.

Supplementary key words nonsynonymous variant • leptin • Canadian First Nation population • genetics • serum levels • hypertriglyceridemic waist

Complex quantitative traits such as obesity are influenced by both genetic and environmental factors. Moreover, evaluation of the genetic contribution identifies gene products and their interactions in biological pathways, aiding in overall understanding of these complex traits. We have previously identified significant associations between genomic variants and complex traits in the Oji-Cree, an isolated Canadian First Nations population (1, 2). The Oji-Cree are an ideal population to study association of genetic factors with complex traits because their background genetic and environmental variation is relatively low.

Apolipoprotein (apo) C-I is a protein constituent of chylomicrons, VLDL, and HDL (3). Apo C-I is a member of the human apo C family, which also includes apo C-II and apo C-III. In contrast to other extensively investigated apolipoproteins such as apo E, B, and AI, and even apo C-II and C-III, the physiological role of apo C-I is less well established. In vitro, apo C-I has been suggested to be positively involved in HDL metabolism through activation of LCAT (4), inhibition of HL (5, 6), and inhibition of cholesteryl ester (CE) transfer protein (CETP) activity (7). Using in vivo models of apo C-I-deficient and apo C-I-over-expressing mice, apo C-I has also been suggested to have a positive

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Abbreviations: apo, apolipoprotein; BMI, body-mass index; CE, cholesteryl ester; CETP, CE transfer protein; NCEP ATP, National Cholesterol Education Program Adult Treatment Panel; HTGW, hypertriglyceridemic waist; MetS, metabolic syndrome; TC, total cholesterol; TG, triglyceride.

¹To whom correspondence should be addressed.
e-mail: hegele@robarts.ca

relationship with LDL. Apo C-I has been observed to affect metabolism of apo B-containing lipoproteins by attenuating VLDL clearance by inhibiting LPL, directly (8) or indirectly (9), and by inhibiting liver-specific LDL receptor (LDLR) (10) and LDLR-related protein (LRP) (11), as well as the peripheral tissue-specific VLDL receptor (VLDLR) (12). Overall, apo C-I has been shown to increase the production of VLDL (13), triglyceride (TG) and cholesterol in mice (3, 8).

Human genetic studies have had limited success in better elucidating the physiological role of apo C-I. This is due in part to the paucity of naturally-occurring human variants in apo C-I compared with numerous common and rare variants affecting the protein sequences of apo E, B, A-I, A-II, A-IV, A-V, C-II, and C-III (14). The human variants of these other apolipoproteins have often served to identify and specify key pathways and mechanisms for more intensive study (15). Common noncoding DNA variants of *APOC1* located in the promoter region were among the first to have been reported for genes affecting lipoprotein metabolism (16, 17) with somewhat variable associations with plasma lipoproteins (18–20) and neurological phenotypes (21–24). One naturally occurring structural variant of apo C-I, namely T45S, was correlated with the clinical trait of increased body mass index (BMI) in Native Americans and Mexican descendants (25). Thus far, this variant has not been associated with biochemical traits nor has this association been replicated in other ethnic populations.

In the process of systematically screening candidate genes in lipoprotein metabolism for new DNA sequence variants in Canadian subpopulations, we identified the nonsynonymous variant designated *APOC1* T45S in the Oji-Cree. We took advantage of this naturally occurring polymorphism to genotype Oji-Cree who were nondiabetic in order to detect associations with clinical and biochemical traits. We found that individuals with the *APOC1* T45S variant, when compared with other nondiabetic subjects, had lower indices for waist circumference, hypertriglyceridemic waist (HTGW) prevalence, percent body fat, as well as lower serum concentrations of leptin and apo C-I, the latter of which was measured using a newly developed quantitative assay.

METHODS

Study subjects

All subjects in the current study had formerly been participants in the original 1993–1995 Sandy Lake Health and Diabetes Project (26). The Oji-Cree community of Sandy Lake, Ontario, is located ~2000 km northwest of Toronto, in the subarctic boreal forest of central Canada. Seven hundred twenty-eight members of this community (72% of the total population) ≥ 10 years of age participated in the original survey. Detailed information on demographics, dietary habits, and physical fitness of the study participants has been previously reported (27–29). The studies were approved by the Sandy Lake First Nation Band Council and the University of Toronto Ethics Review Committee and signed informed consent was obtained from all participants.

Clinical characteristics and biochemical analysis

Body weight, height, waist circumference, and blood pressure were measured by standardized procedures (26). For waist circumference measurement, the natural waist was considered as the minimal circumference between the umbilicus and the xiphoid process. Hypertensive individuals were defined as those subjects having either blood pressure exceeding 130 mmHg (systolic) and/or 80 mmHg (diastolic), or those taking antihypertensive medications. Measurements of fasting blood analyses, including glucose, cholesterol, TGs, and leptin were performed as described elsewhere (26).

Genetic analysis of *APOC1* T45S

Leukocyte DNA was prepared as described elsewhere (30, 31) and was used for genotype analysis. Genotypes for *APOC1* codon 45 (T45S, exon 3) were determined using PCR. Exon 3 was amplified following established procedures (31) using primers 5' GGG AGG TAG CTG CAC ACA GT and 3' GGT GTG GGA AAT TTC AGA GG, followed by amplicon digestion using endonuclease *BsmAI* (New England Biolabs Inc., Ipswich, MA), as per manufacturer's recommendations. The digested fragments were electrophoresed in a 2% agarose gel. Using this restriction isotyping, the smaller (65 bp and 89 bp) and larger (154 bp) fragments represented the T45 and S45 variants, respectively.

HTGW case definition

According to Lemieux et al. (32), subjects have a HTGW if they present with both of the following: 1) abdominal obesity, defined by waist circumference ≥ 90 cm, and 2) hypertriglyceridemia, defined by plasma TG concentration ≥ 2 mmol/L. On the other hand, National Cholesterol Education Program Adult Treatment Panel (NCEP ATP) III guidelines for HTGW is gender-specific such that the waist circumference cut-off for males is >102 cm and >88 cm for females (33).

MetS case definition

According to the NCEP ATP III criteria (33), metabolic syndrome (MetS) was identified if a subject had ≥ 3 of: 1) increased waist circumference [>102 cm (>40 inches) for men, >88 cm (>35 inches) for women]; 2) elevated plasma triglycerides [≥ 1.69 mmol/L (≥ 150 mg/dl)]; 3) low plasma HDL cholesterol [<1.04 mmol/L (<40 mg/dl) for men, <1.29 mmol/L (<50 mg/dl) for women]; 4) increased blood pressure (≥ 130 mmHg systolic and/or ≥ 85 mmHg diastolic) or on antihypertensive drug treatment; and 5) impaired fasting glucose [≥ 6.1 mmol/L (≥ 110 mg/dl)].

Quantitation of serum apo C-I concentration

Apo C-I levels were quantified from frozen Oji-Cree serum samples by sandwich ELISA as described elsewhere (34). Briefly, polyclonal rabbit anti-human apo C-I antibody (Academy Biomedical Co., Houston, TX) was coated overnight at 7°C onto 96-well polystyrene plates (Nunc-Immuno MaxiSorp, NUNC, Rochester, NY) at a dilution of 1:200, followed by one wash with 300 μ L PBS per well. Each well was then blocked for 1 h at 26°C with 300 μ L PBS containing 0.05% Tween 20 (PBST) and 0.5% BSA (Sigma, St. Louis, MO) and followed by one wash with 300 μ L PBS. Subsequently, serum samples (dilution, 1:8000) and apo C-I standard (dilution, 1-60ng/ml; Biomedical Co., Houston, TX) diluted with PBST and 0.1% BSA and, respectively, were added to each well and incubated for 1 h at 450 rpm at 26°C. Wells were then washed three times with 300 μ L PBST and 0.5% BSA, followed by the addition of HRP-conjugated polyclonal goat anti-human apo C-I antibody (dilution, 1:8000; Academy Biomedical) diluted in PBST and 0.1% BSA. After two washes in PBST and one wash in PBS, color was developed by addition of

100 μ L of freshly prepared substrate solution containing 10 mg o-phenylenediamine dihydrochloride (Pierce, Rockford, IL), 20 ml sodium phosphate/citrate buffer, and 20 μ L of 30% H_2O_2 . The reaction was stopped after 20 min (450 rpm at 26°C) with 100 μ L of 2.0 M H_2SO_4 . Color development was measured at 490 nm using a microplate reader.

Statistical analysis

For this analysis, we studied a subset of Oji-Cree who were no closer than third-degree relatives to one another in order to control for confounding artifacts based on close kinship. SAS version 9.1 (SAS Institute, Cary, NC) was used for all statistical comparisons. Data are presented as mean \pm SD or as percentages for categorical variables. Logarithmic transformations (natural log) were used when data were not normally distributed. Transformed variables were used for parametric statistical analyses, but untransformed values were presented in tables. *P*-values were adjusted for age and sex when comparing differences in demographic and laboratory characteristics between those with and those without the S45 allele, using either the general linear model or the logistic model. Statistical significance was taken at a nominal *P* < 0.05 for all comparisons.

RESULTS

The baseline clinical and biochemical attributes of the 410 unrelated nondiabetic Oji-Cree subjects are shown in **Table 1**. This subset of the Oji-Cree had nearly even distribution of males and females. In addition to the traits indicated, their MetS status and their HTGW values were also recorded in Table 1.

In this sample of Oji-Cree, 348 had the *APOC1* T45/T45 genotype, 62 had the T45/S45 genotype, and none had

TABLE 1. Baseline clinical and biochemical traits of male and female nondiabetic Oji-Cree

Measurement	Males	Females
n	192	217
Age, y	26.9 \pm 13.2 ^a	23.7 \pm 11.5
Triglycerides, mmol/L	1.32 \pm 0.73	1.17 \pm 0.46
Total cholesterol, mmol/L	4.43 \pm 1.01	4.45 \pm 0.71
LDL cholesterol, mmol/L	2.58 \pm 0.87	2.34 \pm 0.60
HDL cholesterol, mmol/L	1.25 \pm 0.29	1.28 \pm 0.26
apo A-I, g/L	1.46 \pm 0.22	1.48 \pm 0.21
apo B, g/L	1.05 \pm 0.32	0.94 \pm 0.21
BMI, kg/m ²	24.7 \pm 4.93 ^a	25.8 \pm 5.80
Percent body fat, %	24.4 \pm 9.00 ^b	39.3 \pm 11.8
WHR—iliac crest	0.95 \pm 0.07 ^a	0.94 \pm 0.05
WHR—natural waist	0.91 \pm 0.07 ^a	0.86 \pm 0.05
Waist circumference, cm	89.9 \pm 14.0 ^a	86.5 \pm 13.2
Leptin, ng/ml	6.70 \pm 6.77 ^a	18.8 \pm 11.6
Apo C-I, μ g/ml	220.7 \pm 62.8 ^c	203.9 \pm 38.8 ^d
MetS, %	1.86 \pm 0.35	1.87 \pm 0.34
HTGW, %	1.84 \pm 0.36	1.96 \pm 0.19
HTGW—NCEP waist, %	1.91 \pm 0.28	1.96 \pm 0.19

BMI, body mass index; WHR, waist to hip ratio; MetS, diagnosis of metabolic syndrome using the National Cholesterol Education Program Adult Treatment Panel III criteria (34); HTGW, diagnosis of 'hypertriglyceridemic waist'; HTGW-NCEP waist, diagnosis of hypertriglyceridemic waist using cut values from the National Cholesterol Education Program Adult Treatment Panel III criteria (34).

^a n = 193.

^b n = 191.

^c n = 77.

^d n = 102.

the S45/S45 genotype (**Table 2**). Because there were no S45 homozygotes, T45 was treated as a dominant allele for the purpose of subsequent analyses. Overall, the *APOC1* T45S genotype frequencies in the Oji-Cree do not deviate significantly from predictions of the Hardy-Weinberg equation. In addition, the allele frequencies for T45 and S45 carriers are indicated in Table 2.

The clinical and biochemical attributes and significance of the nondiabetic Oji-Cree according to their *APOC1* genotypes are shown in **Table 3**. Significance for each biochemical or clinical trait was derived after adjustment for age and sex. There were no significant differences (*P* > 0.05) in any of the plasma lipid concentrations. In addition, there were no significant between-genotype differences in the prevalence of MetS or in either mean (BMI) or waist-to-hip-ratio. The lower values in waist circumference and NCEP-defined HTGW prevalence in S45 carriers, compared with T45 homozygotes, was close to being statistically significant with *P* = 0.070 (*P* = 0.066, using logarithmic adjustment) and *P* = 0.077, respectively. There were significant differences (*P* < 0.05) in percent body fat, serum leptin levels, and HTGW prevalence, with lower levels of these traits in the *APOC1* S45 carriers. In addition, we found that the *APOC1* S45 allele was associated with a significantly lower serum apo C-I concentration (*P* < 0.0001).

DISCUSSION

The principal findings of this study in nondiabetic Oji-Cree subjects were significant associations between the *APOC1* T45S polymorphism and variations in obesity indices, serum adipose-secreted hormone levels, and apo C-I levels. Specifically, in subjects carrying the *APOC1* S45 allele we found: 1) lower waist circumference, including lower frequency of HTGW, 2) lower percent body fat, 3) lower serum leptin concentrations, and 4) lower serum apo C-I concentrations, using an ELISA-based quantitative method. Thus, the nonsynonymous T45S variant in *APOC1* among the Canadian First Nations is associated with variations in adiposity, obesity, and lower serum apo C-I levels.

Human apo C-I, C-II, and C-III, are protein constituents of chylomicrons, VLDL, and HDL. Although much is known about the roles of the apo C-II and C-III in lipoprotein metabolism, less is known about the biological function(s) of apo C-I. The gene coding for human apo C-I, *APOC1*, is part of a 48-kb gene cluster on chromosome 19. The gene cluster also includes *APOC2*, *APOE*, and the pseudo-*APOC1*' gene (3, 35). The 4.7 kb *APOC1* gene is

TABLE 2. Genotype and allele frequencies in Oji Cree

	Genotype Frequencies ^a			Allele Frequencies	
	T45/T45	T45/S45	S45/S45	T45	S45
Male	166 (0.86)	27 (0.14)	0 (0)	0.93	0.07
Female	182 (0.84)	35 (0.16)	0 (0)	0.92	0.08
Total	348 (0.85)	62 (0.15)	0 (0)	0.92	0.08

^a percent in their respective populations given in parentheses.

TABLE 3. Clinical and biochemical traits of nondiabetic Oji-Cree according to *APOC1* T45S genotypes

Trait	TT n = 347	TS n = 62	P
Triglycerides, mmol/L	1.25 ± 0.61	1.15 ± 0.57	NS (0.15)
log triglycerides			NS (0.12)
Total cholesterol, mmol/L	4.28 ± 0.88	4.27 ± 0.84	NS (0.62)
log total cholesterol			NS (0.66)
LDL cholesterol, mmol/L	2.45 ± 0.75	2.46 ± 0.72	NS (0.82)
log LDL			NS (0.89)
HDL cholesterol, mmol/L	1.26 ± 0.27	1.29 ± 0.29	NS (0.56)
log HDL			NS (0.61)
BMI, kg/m ²	25.4 ± 5.5 ^a	24.7 ± 5.1	NS (0.15)
log BMI			NS (0.17)
Percent body fat, %	32.7 ± 12.9	30.0 ± 13.0 ^c	0.0081
log percent body fat			0.0131
WHR - iliac crest	0.94 ± 0.06 ^a	0.95 ± 0.06	NS (0.89)
WHR - natural waist	0.88 ± 0.07 ^a	0.89 ± 0.07	NS (0.68)
log WHR (natural)			NS (0.67)
Waist circumference, cm	88.5 ± 13.6 ^a	86.2 ± 13.8	NS (0.070)
log waist			NS (0.066)
Leptin, ng/ml	13.6 ± 11.9 ^a	10.7 ± 7.8	0.0065
log leptin			0.029
Apo C-I, µg/ml	221.7 ± 49.6 ^b	190.2 ± 47.7 ^d	<0.0001
MetS, %	14.1	9.09	NS (0.24)
HTGW, %	10.4	3.23	0.047
HTGW - NCEP waist, %	6.92	1.61	NS (0.077)

BMI, body mass index; WHR, waist to hip ratio; HTGW, diagnosis of hypertriglyceridemic waist; HTGW-NCEP waist, diagnosis of hypertriglyceridemic waist using cut values from the National Cholesterol Education Program Adult Treatment Panel III criteria (33); NS, not significant with nominal $P > 0.05$. Data are means ± SD. P -values are adjusted for age and sex. The P -values in bold indicates the difference observed for the trait is significant or close to significant.

^a n = 348.

^b n = 119.

^c n = 59.

^d n = 60.

primarily expressed in the liver, with lower amounts expressed in the lung, skin, testes, and spleen (35). The pseudo-*APOC1*' gene 7.5 kb downstream from *APOC1* has no detectable mRNA products in any tissue (35). *APOC1* gene expression is regulated by an array of elements found throughout the whole *APOE/C1/C2* gene cluster, such as the hepatic control region (36). The 6.6 kDa apo C-I protein is a polypeptide of 57 amino acid residues with residues 7 to 24 and 35 to 53 of apo C-I having importance in binding lipoproteins (37, 38).

Little is known about naturally occurring mutations in the *APOC1* gene contributing to lipid-related abnormalities in humans. Dumon et al. (39) reported a patient with chylomicronemia who had a naturally occurring mutation in *APOC1* resulting in a deficiency of apo C-I. However, this patient concurrently suffered from apo C-II deficiency due to an *APOC2* defect, which was more likely to have been the cause of the chylomicronemia (39). Mass spectrometry studies elucidated the first structural nonsynonymous variant of apo C-I T45S that was observed to have increased N-terminal truncation and increased in vitro distribution to the VLDL fraction in Native Americans and Mexicans (25). From mass spectrometry identification of the apo C-I T45S polymorphism in 228 Native Americans and in five sibling pairs of Mexican ancestry, S45 was associated with elevated BMI (25). The authors commented that more comprehensive studies of the *APOC1*

T45S polymorphism in independent samples were necessary (25).

Our results in a Canadian First Nation population indicate that the apo C-I T45S variant has the opposite effect, so that S45 carriers had lower percent body fat and BMI. This 'flip-flop' association has been previously reported across different ethnic groups because differences in the genetic background and/or environment cause heterogeneous effects due to the same variant (40). Impact of multiple loci may also affect association, causing the flip-flop phenomenon (40). In addition, our study was carried out in 424 individuals, which was a larger study than that reported in the initial study of Native American subjects.

In vitro studies using, for instance, ligand blotting assays, artificial bilayer vesicles, and cultured fibroblasts have shown that apo C-I has an inhibitory and/or stimulatory effect on many receptors and enzymes involved in lipoprotein metabolism, suggesting a complex role for apo C-I in human disease (3, 10, 11, 41). One such example is the potential HDL-raising effect that apo C-I has by regulating the activity of HL, CETP, as well as LCAT, a protein known to esterify cholesterol in HDL such that it increases HDL levels and particle size (4).


New insights into the metabolic properties of apo Cs have been provided by the technologies of gene targeting through human transgenic mice and knockout mice models. Human *APOC1*-transgenic animals (on a wild-type or *APOE*-deficient background) have been used to elucidate several roles in lipoprotein metabolism, such as inhibition of LPL (8, 13), inhibition of apo E-mediated lipid clearance via LRP (12), and blocking the binding of VLDL to the VLDLR (12). These roles have been associated with hyperlipidemia and increased atherosclerotic lesion development (9). In addition to hyperlipidemia, *APOC1*-transgenic animals exhibit several abnormalities, consisting of elevated plasma free fatty acid levels, inflammatory skin disorders, atrophic sebaceous glands, and subcutaneous adipose tissue (42), which suggests an additional role for apo C-I in epidermal lipid synthesis as well as adipose tissue formation. Because transgenic mice over-expressing *Apoc1* develop hyperlipidemia (43), a hypolipidemic phenotype was expected in *Apoc1* null mice. However, *Apoc1* null mice on a high-fat and high-cholesterol diet developed hypercholesterolemia due to impaired in vivo hepatic uptake of VLDL (44). In addition, a recent study demonstrated that *Apoc1* null mice had decreased serum HDL concentration (43). In contrast, human *APOC1* transgenic mice have increased HDL concentrations due to inhibition of scavenger receptor class B type 1, a protein receptor that facilitates the flux of HDL through the plasma membrane (45).

Overall, in vitro, in vivo, and clinical evidence demonstrates that apo C-I inhibits the uptake of TG-rich lipoproteins via hepatic receptors and as a consequence, the presence of apo C-I on the lipoprotein particle may prolong their residence time in the circulation and subsequently facilitate their conversion to LDL (3). Apo C-I also functions to activate LCAT, which exhibits two activities in normal plasma: α -LCAT activity leads to mature-HDL production whereas β -LCAT activity has a positive effect on

LDL concentrations (46). In addition, apo C-I has a role in adipose biology as observed in human transgenic mice (42).

Therefore, our study suggests that the S45 variant in the Canadian First Nations results in lower concentrations of apo C-I and leptin, which may lead to lower percent body fat and waist circumference. However, a previous study has suggested the S45 variant to have greater N-terminal truncation and preferential distribution to the VLDL fraction compared with the *APOC1* T45 variant (47); increased concentration of apo C-I was recently observed to be associated with decreased visceral fat in men with MetS (48).

Based on the studies thus far, apo C-I seems to promote HDL as well as LDL production due to its effects on CETP and on remnant lipoprotein clearance and LCAT activity. A prospective study of a genotype-determined concentration will be required to prove this, as circulating levels will be affected by increasing or decreasing lipoprotein concentrations. Further biochemical studies are required to uncover the effects of the T45S polymorphism on the structure and distribution of apo C-I in Canadian First Nations and other ethnicities using mass spectrometry and plasma distribution of apolipoproteins in order to understand the clinical consequences of this variant. In addition, long-term prospective studies might help to clarify the long-term effect of this structural variant on the propensity for obesity and diabetes. Overall, such studies using geographically isolated populations help further elucidate the physiological role of this relatively poorly defined protein in complex metabolic disorders.

In summary, although the precise function of apo C-I is unknown, the human genetic data presented herein suggest a possible role for the *APOC1* S45 variant in obesity and adipocyte regulation as demonstrated by the decrease in body fat, waist circumference, decreased prevalence of HTGW, as well as a decreased serum leptin levels. Moreover, this is the first demonstration of a direct relationship of *APOC1* genetic variation with the serum concentration of apo C-I. 

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REFERENCES

- Pollex, R. L., A. J. Hanley, B. Zinman, S. B. Harris, and R. A. Hegele. 2006. Clinical and genetic associations with hypertriglyceridemic waist in a Canadian aboriginal population. *Int. J. Obes.* **30**: 484–491.
- Pollex, R. L., M. Mamakeesick, B. Zinman, S. B. Harris, R. A. Hegele, and A. J. Hanley. 2007. Peroxisome proliferator-activated receptor gamma polymorphism Pro12Ala is associated with nephropathy in type 2 diabetes. *J. Diabetes Complications.* **21**: 166–171.
- Jong, M. C., M. H. Hofker, and L. M. Havekes. 1999. Role of ApoCs in lipoprotein metabolism: functional differences between ApoC1, ApoC2, and ApoC3. *Arterioscler. Thromb. Vasc. Biol.* **19**: 472–484.
- Soutar, A. K., C. W. Garner, H. N. Baker, J. T. Sparrow, R. L. Jackson, A. M. Gotto, and L. C. Smith. 1975. Effect of the human plasma apolipoproteins and phosphatidylcholine acyl donor on the activity of lecithin: cholesterol acyltransferase. *Biochemistry.* **14**: 3057–3064.
- Conde-Knape, K., A. Bensadoun, J. H. Sobel, J. S. Cohn, and N. S. Shachter. 2002. Overexpression of apoC-I in apoE-null mice: severe hypertriglyceridemia due to inhibition of hepatic lipase. *J. Lipid Res.* **43**: 2136–2145.
- Kinnunen, P. K., and C. Ehnolm. 1976. Effect of serum and C-apoproteins from very low density lipoproteins on human post-heparin plasma hepatic lipase. *FEBS Lett.* **65**: 354–357.
- Gautier, T., D. Masson, J. P. de Barros, A. Athias, P. Gambert, D. Aunis, M. H. Metz-Boutigue, and L. Lagrost. 2000. Human apolipoprotein C-I accounts for the ability of plasma high density lipoproteins to inhibit the cholesteryl ester transfer protein activity. *J. Biol. Chem.* **275**: 37504–37509.
- Berbee, J. F., C. C. van der Hoogt, D. Sundararaman, L. M. Havekes, and P. C. Rensen. 2005. Severe hypertriglyceridemia in human *APOC1* transgenic mice is caused by apoC-I-induced inhibition of LPL. *J. Lipid Res.* **46**: 297–306.
- Westerterp, M., M. Van Eck, W. de Haan, E. H. Offerman, T. J. Van Berkel, L. M. Havekes, and P. C. Rensen. 2007. Apolipoprotein CI aggravates atherosclerosis development in ApoE-knockout mice despite mediating cholesterol efflux from macrophages. *Atherosclerosis.* **195**: e9–e16.
- Schayek, E., and S. Eisenberg. 1991. Mechanisms of inhibition by apolipoprotein C of apolipoprotein E-dependent cellular metabolism of human triglyceride-rich lipoproteins through the low density lipoprotein receptor pathway. *J. Biol. Chem.* **266**: 18259–18267.
- Weisgraber, K. H., R. W. Mahley, R. C. Kowal, J. Herz, J. L. Goldstein, and M. S. Brown. 1990. Apolipoprotein C-I modulates the interaction of apolipoprotein E with beta-migrating very low density lipoproteins (beta-VLDL) and inhibits binding of beta-VLDL to low density lipoprotein receptor-related protein. *J. Biol. Chem.* **265**: 22453–22459.
- Jong, M. C., V. E. Dahlmans, P. J. van Gorp, K. W. van Dijk, M. L. Breuer, M. H. Hofker, and L. M. Havekes. 1996. In the absence of the low density lipoprotein receptor, human apolipoprotein C1 overexpression in transgenic mice inhibits the hepatic uptake of very low density lipoproteins via a receptor-associated protein-sensitive pathway. *J. Clin. Invest.* **98**: 2259–2267.
- Westerterp, M., W. de Haan, J. F. Berbee, L. M. Havekes, and P. C. Rensen. 2006. Endogenous apoC-I increases hyperlipidemia in apoE-knockout mice by stimulating VLDL production and inhibiting LPL. *J. Lipid Res.* **47**: 1203–1211.
- Lusis, A. J., A. M. Fogelman, and G. C. Fonarow. 2004. Genetic basis of atherosclerosis: part II: clinical implications. *Circulation.* **110**: 2066–2071.
- Hegele, R. A. 2009. Plasma lipoproteins: genetic influences and clinical implications. *Nat. Rev. Genet.* **10**: 109–121.
- Frossard, P. M., R. T. Coleman, M. J. Malloy, J. P. Kane, B. Levy-Wilson, and V. A. Appleby. 1987. Human apolipoprotein C1 (apoC1) gene locus: DraI dimorphic site. *Nucleic Acids Res.* **15**: 1884.
- Frossard, P. M., D. W. Lim, R. T. Coleman, H. Funke, G. Assmann, M. J. Malloy, and J. P. Kane. 1987. Human apolipoprotein C1 (ApoC1) gene locus: BglII dimorphic site. *Nucleic Acids Res.* **15**: 1344.
- Smit, M., E. van der Kooij-Meijns, L. P. Woudt, L. M. Havekes, and R. R. Frants. 1988. Exact localization of the familial dysbetalipoproteinemia associated HpaI restriction site in the promoter region of the *APOC1* gene. *Biochem. Biophys. Res. Commun.* **152**: 1282–1288.
- Hubacek, J. A., D. M. Waterworth, R. Poledne, J. Pitha, Z. Skodova, S. E. Humphries, and P. J. Talmud. 2001. Genetic determination of plasma lipids and insulin in the Czech population. *Clin. Biochem.* **34**: 113–118.
- Shachter, N. S., D. Rabinowitz, S. Stohl, K. Conde-Knape, J. S. Cohn, R. J. Deckelbaum, L. Berglund, and S. Shea. 2005. The common insertional polymorphism in the *APOC1* promoter is associated with serum apolipoprotein C-I levels in Hispanic children. *Atherosclerosis.* **179**: 387–393.
- Abildayeva, K., J. F. Berbee, A. Blokland, P. J. Jansen, F. J. Hoek, O. Meijer, D. Lutjohann, T. Gautier, T. Pillot, J. De Vente, et al. 2008. Human apolipoprotein C-I expression in mice impairs learning and memory functions. *J. Lipid Res.* **49**: 856–869.
- Li, H., S. Wetten, L. Li, P. L. St Jean, R. Upmanyu, L. Surh, D. Hosford, M. R. Barnes, J. D. Briley, M. Borrie, et al. 2008. Candidate single-nucleotide polymorphisms from a genomewide association study of Alzheimer disease. *Arch. Neurol.* **65**: 45–53.
- Tycko, B., J. H. Lee, A. Ciappa, A. Saxena, C. M. Li, L. Feng, A. Arriaga, Y. Stern, R. Lantigua, N. Shachter, et al. 2004. APOE and

- APOC1 promoter polymorphisms and the risk of Alzheimer disease in African American and Caribbean Hispanic individuals. *Arch. Neurol.* **61**: 1434–1439.
24. Shi, J., S. Zhang, C. Ma, X. Liu, T. Li, M. Tang, H. Han, Y. Guo, J. Zhao, K. Zheng, et al. 2004. Association between apolipoprotein C1 HpaI polymorphism and sporadic Alzheimer's disease in Chinese. *Acta Neurol. Scand.* **109**: 140–145.
 25. Kasthuri, R. S., K. R. McMillan, C. Flood-Urdangarin, S. B. Harvey, J. T. Wilson-Grady, and G. L. Nelsestuen. 2007. Correlation of a T45S variant of apolipoprotein C1 with elevated BMI in persons of American Indian and Mexican ancestries. *Int. J. Obes.* **31**: 1334–1336.
 26. Hanley, A. J. G., S. B. Harris, A. Barnie, J. Gittelsohn, T. M. S. Wolever, A. Logan, and B. Zinman. 1995. The Sandy Lake Health and Diabetes Project: design, methods and lessons learned. *Chronic Dis. Can.* **16**: 149–156.
 27. Gittelsohn, J., T. M. Wolever, S. B. Harris, R. Harris-Giraldo, A. J. Hanley, and B. Zinman. 1998. Specific patterns of food consumption and preparation are associated with diabetes and obesity in a Native Canadian community. *J. Nutr.* **128**: 541–547.
 28. Harris, S. B., B. Zinman, A. Hanley, J. Gittelsohn, R. Hegele, P. W. Connelly, B. Shah, and J. E. Hux. 2002. The impact of diabetes on cardiovascular risk factors and outcomes in a native Canadian population. *Diabetes Res. Clin. Pract.* **55**: 165–173.
 29. Kriska, A. M., A. J. Hanley, S. B. Harris, and B. Zinman. 2001. Physical activity, physical fitness, and insulin and glucose concentrations in an isolated Native Canadian population experiencing rapid lifestyle change. *Diabetes Care.* **24**: 1787–1792.
 30. Hegele, R. A., A. J. Evans, L. Tu, G. Ip, J. H. Brunt, and P. W. Connelly. 1994. A gene-gender interaction affecting plasma lipoproteins in a genetic isolate. *Arterioscler. Thromb.* **14**: 671–678.
 31. Hegele, R. A., S. B. Harris, A. J. Hanley, F. Sun, P. W. Connelly, and B. Zinman. 1997. Angiotensinogen gene variation associated with variation in blood pressure in aboriginal Canadians. *Hypertension.* **29**: 1073–1077.
 32. Lemieux, I., A. Pascot, C. Couillard, B. Lamarche, A. Tchernof, N. Almeras, J. Bergeron, D. Gaudet, G. Tremblay, D. Prud'homme, et al. 2000. Hypertriglyceridemic waist: a marker of the atherogenic metabolic triad (hyperinsulinemia; hyperapoprotein B; small, dense LDL) in men? *Circulation.* **102**: 179–184.
 33. Executive Summary of The Third Report of The National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol In Adults (Adult Treatment Panel III). 2001. *J. Am. Med. Assoc.* **285**: 2486–2497.
 34. Bury, J., G. Michiels, and M. Rosseneu. 1986. Human apolipoprotein C-II quantitation by sandwich enzyme-linked immunosorbent assay. *J. Clin. Chem. Clin. Biochem.* **24**: 457–463.
 35. Lauer, S. J., D. Walker, N. A. Elshourbagy, C. A. Reardon, B. Levy-Wilson, and J. M. Taylor. 1988. Two copies of the human apolipoprotein C-I gene are linked closely to the apolipoprotein E gene. *J. Biol. Chem.* **263**: 7277–7286.
 36. Simonet, W. S., N. Bucay, S. J. Lauer, and J. M. Taylor. 1993. A far-downstream hepatocyte-specific control region directs expression of the linked human apolipoprotein E and C-I genes in transgenic mice. *J. Biol. Chem.* **268**: 8221–8229.
 37. Shulman, R. S., P. N. Herbert, K. Wehrly, and D. S. Fredrickson. 1975. The complete amino acid sequence of C-I (apoLp-Ser), an apolipoprotein from human very low density lipoproteins. *J. Biol. Chem.* **250**: 182–190.
 38. Rozek, A., G. W. Buchko, and R. J. Cushley. 1995. Conformation of two peptides corresponding to human apolipoprotein C-I residues 7–24 and 35–53 in the presence of sodium dodecyl sulfate by CD and NMR spectroscopy. *Biochemistry.* **34**: 7401–7408.
 39. Dumon, M. F., and M. Clerc. 1986. Preliminary report on a case of apolipoproteins CI and CII deficiency. *Clin. Chim. Acta.* **157**: 239–248.
 40. Lin, P. I., J. M. Vance, M. A. Pericak-Vance, and E. R. Martin. 2007. No gene is an island: the flip-flop phenomenon. *Am. J. Hum. Genet.* **80**: 531–538.
 41. Kowal, R. C., J. Herz, K. H. Weisgraber, R. W. Mahley, M. S. Brown, and J. L. Goldstein. 1990. Opposing effects of apolipoproteins E and C on lipoprotein binding to low density lipoprotein receptor-related protein. *J. Biol. Chem.* **265**: 10771–10779.
 42. Jong, M. C., M. J. Gijbels, V. E. Dahlmans, P. J. Gorp, S. J. Koopman, M. Ponc, M. H. Hofker, and L. M. Havekes. 1998. Hyperlipidemia and cutaneous abnormalities in transgenic mice overexpressing human apolipoprotein C1. *J. Clin. Invest.* **101**: 145–152.
 43. Shachter, N. S., T. Ebara, R. Ramakrishnan, G. Steiner, J. L. Breslow, H. N. Ginsberg, and J. D. Smith. 1996. Combined hyperlipidemia in transgenic mice overexpressing human apolipoprotein C1. *J. Clin. Invest.* **98**: 846–855.
 44. Jong, M. C., J. H. van Ree, V. E. Dahlmans, R. R. Frants, M. H. Hofker, and L. M. Havekes. 1997. Reduced very-low-density lipoprotein fractional catabolic rate in apolipoprotein C1-deficient mice. *Biochem. J.* **321**: 445–450.
 45. de Haan, W., R. Out, J. F. Berbee, C. C. van der Hoogt, K. W. van Dijk, T. J. van Berkel, J. A. Romijn, J. W. Jukema, L. M. Havekes, and P. C. Rensen. 2008. Apolipoprotein C1 inhibits scavenger receptor BI and increases plasma HDL levels in vivo. *Biochem. Biophys. Res. Commun.* **377**: 1294–1298.
 46. Zhao, Y., F. E. Thorngate, K. H. Weisgraber, D. L. Williams, and J. S. Parks. 2005. Apolipoprotein E is the major physiological activator of lecithin-cholesterol acyltransferase (LCAT) on apolipoprotein B lipoproteins. *Biochemistry.* **44**: 1013–1025.
 47. Wroblewski, M. S., J. T. Wilson-Grady, M. B. Martinez, R. S. Kasthuri, K. R. McMillan, C. Flood-Urdangarin, and G. L. Nelsestuen. 2006. A functional polymorphism of apolipoprotein C1 detected by mass spectrometry. *FEBS J.* **273**: 4707–4715.
 48. van der Ham, R. L., R. Alizadeh Dehnavi, J. F. Berbee, H. Putter, A. de Roos, J. A. Romijn, P. C. Rensen, and J. T. Tamsma. 2009. Plasma apolipoprotein C1 and CIII levels are associated with increased plasma triglyceride levels and decreased fat mass in men with the metabolic syndrome. *Diabetes Care.* **32**: 184–186.