

Forum Review Article

Redox Modifier Genes and Pathways in Amyotrophic Lateral Sclerosis

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Abstract

Enhanced redox-stress caused by neuroinflammation, mitochondria, and NADPH oxidases has been hypothesized to play critical roles in disease progression of amyotrophic lateral sclerosis (ALS). However, distinguishing whether the redox-stress observed in ALS is due to a primary defect in cellular reactive oxygen species metabolism/catabolism, or is a secondary consequence of neuroinflammation, has been difficult and the issue remains a matter of debate. Emerging evidence suggests that defects in genes that regulate NADPH oxidases may account for at least some forms of ALS. NADPH oxidases are key signaling complexes that influence cellular responses to growth factors and cytokines. In this context, NADPH oxidase-derived reactive oxygen species exert spatial control over the redox-dependent activation of certain pro-inflammatory receptors. Understanding the biology of how NADPH oxidases control cell signaling may help to clarify how genetic determinants of ALS lead to dysregulated pro-inflammatory signaling. This review provides a framework for understanding endosomal signaling through NADPH oxidases and potential mechanisms whereby gene defects in various forms of ALS may influence this cellular process and lead to motor neuron degeneration. Lastly, this review discusses past and current efforts to treat ALS using antioxidant therapies, as well as the limitations and advantages of each of these approaches. *Antioxid. Redox Signal.* 11, 1569–1586.

Introduction

AMYOTROPHIC LATERAL SCLEROSIS (ALS) IS A PROGRESSIVE and lethal degenerative disorder of upper and lower motor neurons in the brain and spinal cord. ALS disease onset is insidious and progresses linearly from time of diagnosis, with death occurring in most of the patients within 3–5 years. Most cases begin after age 40 and the mean age at onset is 58 years. About 50% of patients die within 3 years, and ~90% die within 5 years, while only a small proportion survive beyond 8 years. About 10% of ALS cases are clearly genetic or familial (FALS), with the rest being classified as sporadic (SALS).

Despite a great deal of academic, societal, and corporate interest in determining the root cause of the disease and developing adequate drugs and treatment regimens, to date only one drug—riluzole (Rilutek[™])—has been approved for the treatment of ALS (28). However, the mechanism of action

of riluzole is unclear and it has been estimated to prolong life by only ~3 months. Otherwise, treatments for ALS are largely palliative, and developing a new therapy for ALS remains a high priority. Thus, like many other progressive neurodegenerative diseases such as Parkinson's, Alzheimer's, and Huntington's disease, ALS represents a largely unmet medical need (28, 46, 125). Studies on rare familial forms of several of these diseases have provided insights into the molecular basis of their pathogenesis (20, 52, 109), and are beginning to reveal a general unifying set of factors in these slowly progressing diseases. These include protein aggregation, mitochondrial injury, inflammatory response, and nitrosative and oxidative stress.

ALS is a multigenic disease with a complex etiology (20, 46). Sporadic ALS cases may result from interactions between genetic and environmental components, each of which may contribute to the pathophysiology of the disease. Most, but not all, FALS exhibits an autosomal dominant pattern

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of inheritance, and several FALS-associated mutant genes have been identified (109). Most notable among these is the gene encoding superoxide dismutase type 1 (*SOD1*) (117). Recently, analysis of one such mutant gene, *SOD1*^{G93A}, in animal models has provided important insights into the mechanism underlying the dominant, or toxic gain-of-function, phenotype and its potential to lead to oxidative damage (55, 90). Similarly, research on a rare juvenile form of ALS caused by mutations in the *ALS2* gene has uncovered potential new paradigms for pathogenesis (26, 54, 144). Interestingly, as discussed in this review, the mechanisms involved in ALS pathogenesis in the context of *ALS2* and *SOD1* mutations overlap more extensively than previously thought.

Much of the difficulty in developing effective therapies for ALS reflects our relatively poor understanding of ALS pathogenesis (20, 46, 112). For instance, an extremely diverse set of biochemical processes has been implicated in the pathogenesis of ALS disease:

- glutamate-mediated excitotoxicity
- mitochondrial dysfunction
- neuroinflammation (microglial proliferation)
- apoptosis
- protein aggregation
- autophagy
- abnormal axonal transport
- oxidative stress

Several of these events—including glutamate excitotoxicity, mitochondrial dysfunction, and neuroinflammation—are discussed in detail in other reviews in this Forum issue.

One key hypothesis for the genesis of ALS is that motor neuron damage initially results as a consequence of oxidative stress (7). However, there is significant uncertainty as to whether the toxicity is mediated only by events that occur primarily in neurons, or if non-neuronal cells contribute as well. Recent evidence suggests that the neurodegeneration is non-cell autonomous (*i.e.*, not driven by events solely within the neuron), and that both neuronal cells and non-neuronal microglia are involved in ALS pathogenesis (13, 29, 86), even though degeneration is manifested only in the neurons. This review attempts to place in context potential pathologic mechanisms and disease-causing genes that may regulate oxidative stress in ALS.

Oxidative damage is known to increase with age, and the main risk factors for ALS—those that are consistently identified in epidemiology studies—are increasing age, male gender, and smoking (5, 10, 38). Moreover, biochemical markers of oxidative injury are abnormally high in postmortem samples from ALS patients (126). Oxidative stress results when the cell's ability to generate reactive oxygen species (ROS) exceeds its antioxidant capacity; when ROS or other radicals are not detoxified by antioxidants, cellular macromolecules (phospholipids, DNA, proteins) become targets for oxidative modifications that may compromise their function (46). In addition, other mechanisms that have been proposed to play a role in ALS disease, such as excitotoxicity and defective axonal transport, may be consequences of oxidative stress (21, 46). Excitatory neurotransmission and the metabolism of these neurotransmitters are sources of oxidative stress in cells such as motor neurons, which receive high levels of excitatory input. Mechanisms that lead to glutamate excitotoxicity include the disruption of intracellular calcium homeostasis and in-

creased ROS production due to perturbed mitochondrial function and activation of calcium-dependent enzymes. The latter include neuronal nitric oxide synthase (nNOS), which catalyzes the production of nitric oxide (57). Both superoxide and nitric oxide are produced in abundant quantity by phagocytes, including microglia of the nervous system (21, 141). There is also evidence for increased oxidative damage in both sporadic ALS and a familial form that is associated with mutations in *SOD1* (42, 141). Although the cause of disease initiation and progression in human patients is not known, and it also remains unclear whether oxidative damage occurs as a secondary consequence of the initiating toxic events, sequelae involving both motor neurons and surrounding glia are likely to contribute to neuronal death. Thus, mitigating ROS overproduction would be expected to prevent or retard the progression of this neurodegenerative disease. However, in clinical trials with antioxidants, ALS patients have not demonstrated a survival benefit (105). Hence, it appears that our understanding of the mechanisms that underlie the disease processes in ALS is not yet sufficiently advanced for us to rationally design antioxidant therapies (32, 81).

Genes known to cause juvenile (*ALS2*) and familial (*SOD1*) forms of ALS, as well as those closely linked to sporadic forms, are known to influence the activity of the small GTPase Rac1, which regulates several NADPH oxidase (Nox) complexes. Additionally, *Nox1* and *Nox2* appear to influence disease progression in the *SOD1*^{G93A} mouse model, whereas *Nox4* appears to be closely linked to sporadic ALS. Together, these findings suggest the intriguing possibility that multiple distinct defects in the regulation of Rac/Nox pathways may lead to enhanced redox-stress in ALS. This review will discuss the field's current understanding of how NADPH oxidases can control cell signaling and the relevance of these pathways to the pathoprogession and treatment of ALS.

NADPH Oxidases Influence Disease Progression in a *SOD1*^{G93A} Transgenic Mouse Model of Familial ALS

Redox-stress is thought to be an important component of disease progression in ALS (7, 109). For example, as was discovered more than 10 years ago, protein carbonyl and nitrotyrosine modifications (both markers of oxidative protein damage) occur at unusually high levels in the spinal cords of ALS patients (1, 2, 8, 122). What remains significantly less clear is whether this increased redox-stress is a primary defect or a secondary consequence of disease. Recently, two studies have shed some light on the involvement of NADPH oxidases in the disease progression observed in a familial mouse model of ALS that overexpresses a human *SOD1*^{G93A} transgene. NADPH oxidases are multi-subunit enzyme complexes that generate superoxide ($O_2^{\cdot-}$) by transferring an electron from NADPH to molecular oxygen. Seven known NADPH oxidase catalytic subunits exist (*Nox1*, *Nox2*^{gp91phox}, *Nox3*, *Nox4*, *Nox5*, *Duox1*, and *Duox2*) (60, 80). The most widely characterized of these is phagocytic gp91phox (also known as *Nox2*), which is also expressed in microglia (141) and a variety of other nonphagocytic cell types. Rac, a small GTPase, is an essential activator of *Nox2* and *Nox1*, along with several other subunits (p40phox, p47phox, p67phox, *NoxO1*, and *NoxA1*) that can act to promote *Nox* complex activation in a cell type-specific fashion (60, 80). Since NADPH oxidases (*Nox*) generate the $O_2^{\cdot-}$ substrate of the *SOD1* dismutation

reaction ($2O_2^{\cdot-} + 2H^+ \rightarrow H_2O_2 + O_2$) (94), this class of Nox enzymes has recently generated considerable interest in studies of ALS.

A recent study has demonstrated that *SOD1*^{G93A} ALS transgenic mice produce elevated levels of Nox2 and $O_2^{\cdot-}$ in spinal cord microglia (141). This same study demonstrated that *Nox2*-deficient *SOD1*^{G93A} congenic C57BL/6J mice live, on average, 13 days longer than *Nox2*-WT *SOD1*^{G93A} mice on the same background (increase from 122 to 135 days). Increased survival was accompanied by reduced ROS in the spinal cord (141). A subsequent study demonstrated that both *Nox1* and *Nox2* influence the survival of *SOD1*^{G93A} ALS transgenic mice on a B6SJL hybrid background (90). In this study, homozygous deletion of either *Nox1* or *Nox2* significantly delayed disease onset, and enhanced the survival of hemizygous *SOD1*^{G93A} ALS mice. Deletion of the *Nox2* gene had the greatest impact on survival (97 days difference; 132–229 days), and also led to a four-fold increase in the survival index (time to death after onset of disease). *Nox1* deficiency gave rise to a much smaller, but still significant, increase in survival (33 days difference; 129–162 days), but did not influence the survival index. Additional results from this study demonstrated reduced ROS production and microglial activation in the spinal cords, as well as increased rotarod performance and stride length, in *Nox2*-deficient *SOD1*^{G93A} ALS mice, consistent with changes in disease onset (90). Although these studies differed with respect to the magnitude of their findings, collectively they led to the conclusion that both *Nox1* and *Nox2* are NADPH oxidase isoforms that influence disease progression in the *SOD1*^{G93A} ALS mouse model, but that *Nox2* plays a more major role.

The reason for differences in outcomes between the above-described studies (90, 141) remains unclear, but likely has to do with background variation in the mice used to study *Nox2* involvement. Indeed, differences in genetic background have been shown to influence the survival of hemizygous *SOD1*^{G93A} transgenic mice (58, 139), with a 14–28 day increase in survival on the C57BL/6 congenic, as compared to the B6SJL hybrid, background. Currently, it is hypothesized that multiple SJL-derived modifier genes act in concert with *Nox2*-deficiency to significantly enhance the survival of *SOD1*^{G93A} mice, and pedigree analysis of survival from the F1 and F2 generations in the Marden *et al.* study is consistent with this hypothesis (90). The lack of a high degree of variability in survival among siblings with *Nox2*-deficient *SOD1*^{G93A} genotypes in the F1 and F2 generations suggests that a single modifier gene likely does not account for the increased survival associated with the *Nox2* mutant allele on the *SOD1*^{G93A} B6SJL mixed genetic background.

Despite the discovery that *Nox2* contributes to inflammatory components of ALS (90, 141), the mechanism(s) whereby mutations in *SOD1* lead to dysregulation of $O_2^{\cdot-}$ production by *Nox2* remains poorly understood. The two studies described above suggest that *Nox2* hyperactivation is likely the result of the enhanced inflammation and microglial activation that are typically associated with disease progression in ALS (109). In this context, one hypothesis to explain increased *Nox2* expression and enhanced redox-stress in ALS is shown in Fig. 1. In this working model, *Nox2* hyperactivation is not the primary event that leads to disease, but rather a consequence of some other cellular defect caused by a mutation in either *SOD1* or another gene(s) that leads to ALS (see other

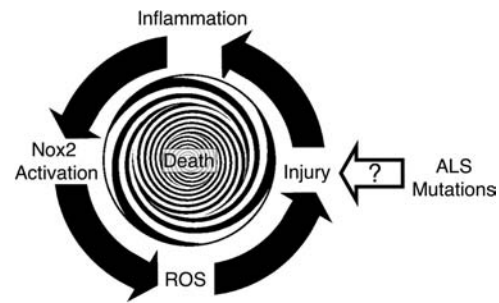


FIG. 1. Inflammation model for *Nox2* involvement in ALS disease progression. According to this model, primary cellular defects in ALS lead to cellular injury by unknown mechanisms. This injury leads to inflammation and *Nox2* activation in microglia. Activated microglia in turn produce ROS via *Nox2*, and this leads to further injury. This cycle repeats until death occurs.

Forum Review articles for elaboration). However, as disease progresses, inflammation and enhanced *Nox2* activation in microglia lead to redox-stress and neuronal death. Neuronal death, in turn, leads to more inflammation, recruitment of more microglia to the damaged tissues, and further *Nox2* activation. Thus, *Nox2*-mediated inflammation amplifies disease progression by increasing the redox-burden. This repetitive injury may lead to a spiraling decline in motor neuron function, and eventually to death. However, this model cannot explain how *Nox1* might also be involved in disease progression, since *Nox2* is thought to be the primary isoform that controls inflammatory responses in phagocytes and microglia.

Endosomal Signaling through Rac-Regulated NADPH Oxidases: Relevance to Pro-Inflammatory Signaling in ALS

Over the last two decades, superoxide and hydrogen peroxide produced by NADPH oxidases have been linked to a number of highly spatially- and temporally-regulated signaling mechanisms. In the context of the pathogenesis model presented in Fig. 1, understanding how NADPH oxidases control pro-inflammatory signaling pathways becomes extremely relevant to ALS. Two highly relevant pro-inflammatory cytokines linked to the progression of disease in ALS include interleukin-1-beta ($IL-1\beta$) and tumor necrosis factor alpha ($TNF\alpha$). Levels of these two pro-inflammatory cytokines are elevated in the blood and/or cerebrospinal fluid (CSF) of humans with ALS (33, 111). Elevation of $TNF\alpha$ protein and mRNA are also observed in spinal cords from *SOD1*^{G93A} (59) and *SOD1*^{G37R} (101) ALS mice, and this correlates with enhanced $NF\kappa B$ activation (101). $IL-1\beta$ is also induced in the spinal cords of ALS mice; however, gene disruption leading to the elimination of this cytokine does not delay disease onset (101). Similarly, *SOD1*^{G93A} and *SOD1*^{G37R} ALS mice bred onto the $TNF\alpha$ knockout background also do not exhibit increased survival (49). Hence, the current thinking is that it is the cumulative effect of multiple pro-inflammatory cytokines that influences the course of neuroinflammation and disease in ALS mice.

The activation of the pro-inflammatory transcription factor $NF\kappa B$ by $IL-1\beta$ and $TNF\alpha$ has been long recognized to be redox-dependent (68, 129). More recently, the redox-dependent processes that control $NF\kappa B$ activation by these cytokines have

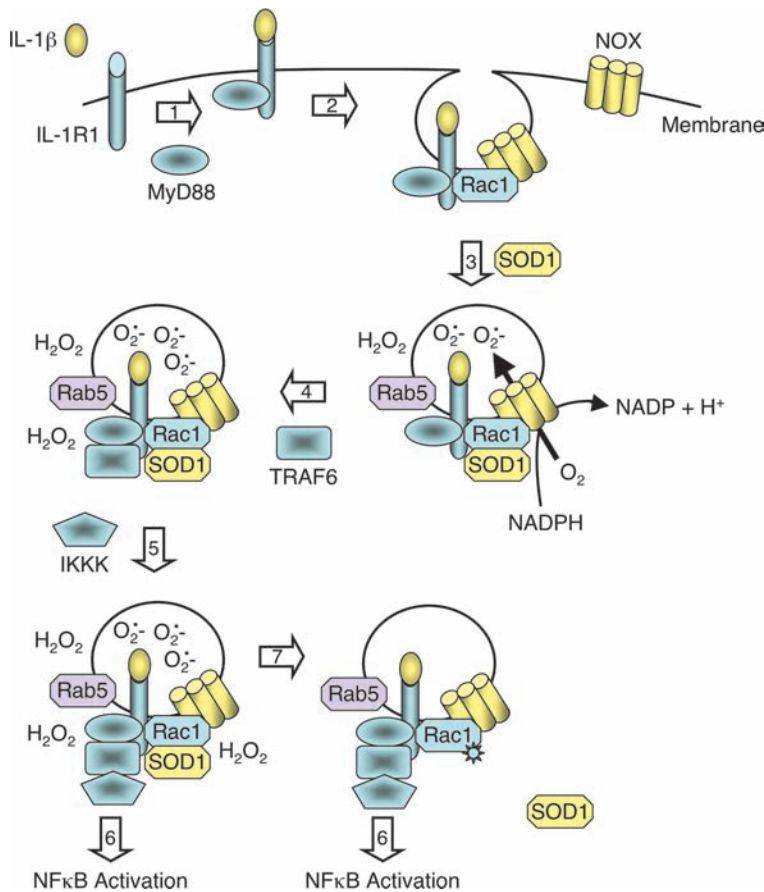


FIG. 2. IL-1 β mediates NF κ B activation through redoxosomal activation. Schematically drawn are steps involved in the redox-activation of NF κ B by IL-1 β (82, 97, 100). *Step 1:* IL-1 β binding to the IL-1 receptor (IL-1R1) facilitates recruitment of the effector MyD88 to the plasma membrane. *Step 2:* MyD88 binding to the ligand/receptor complex facilitates dynamin-dependent endocytosis and recruitment of Rac1 to the receptor. Rac1 is responsible for co-endocytosis of the Nox complex with the ligand-activated receptor. *Step 3:* Endocytosis of the IL-1R1/Nox complex leads to NADPH-dependent $O_2^{\bullet-}$ production in the endosomal lumen. In the case of Rab5 recruitment, is currently unclear whether it occurs at the endosome or the plasma membrane. However, superoxide production initiates in the Rab5-positive, early endosomal compartment. Additionally, SOD1 is recruited to the cytoplasmic face of the redoxosome, where it binds to Rac1 and stabilizes the GTP-bound active form of Rac1. *Step 4:* The localized production of ROS by newly formed redoxosomes facilitates H_2O_2 -mediated recruitment of TRAF6 to IL-1R1 on the redoxosome surface. Extraredoxosomal H_2O_2 may be generated either in the cytoplasm, following exit of $O_2^{\bullet-}$ from the lumen via chloride channels, or within the redoxosome lumen by spontaneous dismutation of $O_2^{\bullet-}$ within the lumen, followed by diffusion across the redoxosomal membrane. *Step 5:* The IL-1R1/MyD88/TRAF6 complex is now competent to recruit IKK kinases (IKKKs), and the recruited IKKKs can then phosphorylate cytoplasmically localized IKK complexes to activate NF κ B. *Step 6:* As ROS levels

outside the redoxosome rise, Rac1 is oxidized and SOD1 disengages. The uncoupling of SOD1 from Rac1 leads to: enhanced GTP hydrolysis by Rac1, inactivation of the Nox complex, and the termination of ROS production. This redox-dependent uncoupling of SOD1 from Rac1 appears to be defective in certain ALS-associated SOD1 mutants, and may lead to increases redox stress in the context of stimulating ligands that utilize redoxosomal pathways. Similar redoxosomal mechanisms appear to control TNF α -mediated activation of NF κ B (83, 97). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at www.liebertonline.com/ars).

been shown to also involve NADPH oxidases (specifically Nox1 and Nox2). In this context, endosomal activation of Nox1 and/or Nox2 has been shown to control redox-dependent activation of both the IL-1 β and TNF α receptors (82, 83, 97, 100). Although these receptors have unique effector pathways responsible for activating NF κ B, they share a similar dependence on redox-active signaling endosomes for their activation cascades. For these reasons, *redox*-active signaling endosomes have been recently named redoxosomes (103). The mechanisms of redoxosome activation may be particularly relevant to redox-signaling in ALS. In this section we review the features of redoxosome biology that are the foundation of later discussion, which places this subject in the context of new paradigms for thinking about disease mechanisms relevant to ALS.

Redoxosomes are signaling endosomes that generate $O_2^{\bullet-}$ in their lumen in response to stimuli such IL-1 β (82, 97) and TNF α (83, 97, 138) (Fig. 2). ROS generated by redoxosomes act as second messengers to facilitate the redox-dependent activation of the IL-1 β receptor (IL-1R) and TNF α receptor (TNFR1). An important feature of redoxosomal biology is its dependence on endocytosis of the ligand-activated receptor, together with NADPH oxidases from the plasma mem-

brane; inhibiting receptor endocytosis prevents the NADPH-dependent ROS production in redoxosomes that is required for activation of the proinflammatory transcription factor NF κ B (82, 83). In this context, Rac1 appears to play a critical role, as it is required for bringing both IL-1R and Nox2 from the plasma membrane into the endosome in a mammary epithelial cell line (82). Similar processes likely control TNFR1 recruitment into redoxosomes in this cell type (83). In smooth muscle cells, Nox1 appears to be capable of replacing Nox2 function in redoxosomes following both IL-1 β and TNF α stimulation (97), suggesting that Rac1-dependent NADPH oxidases (*i.e.*, Nox1 and Nox2) perform cell type-specific redoxosomal signaling. ROS generated by redoxosomes play an important role in the redox-activation of IL-1 and TNF receptor complexes at the endosome surface. It is hypothesized that the conversion of $O_2^{\bullet-}$ to H_2O_2 is required for redox-signaling, since the loading of endosomes with both purified SOD1 and catalase proteins is needed to inhibit redox-dependent activation of NF κ B by these receptors (82, 83).

The production of localized $O_2^{\bullet-}$ and H_2O_2 by redoxosomes is thought to restrict ROS production spatially to sites of receptor activation, and hence to control downstream signaling events. The redox-dependent event required for the activation

of both IL-1R and TNFR1 on redoxosomes appears to involve H₂O₂-mediated recruitment of TRAFs to these receptor complexes (Fig. 2). TRAF recruitment to each of these receptors (TRAF6 → IL-1R1 and TRAF2 → TNFR1) is a key event that is required for the formation of an active IKK kinase (IKKK) complex on these receptors, and this in turn facilitates phosphorylation and activation of the IKK complex. Activation of the IKK complex then leads to phosphorylation of IκB (inhibitor of NFκB) and subsequent mobilization of NFκB to the nucleus, where it can transcriptionally activate pro-inflammatory pathways. For a more comprehensive review on the effectors of both the IL-1R and TNFR1 pathways that lead to redoxosomal activation of NFκB, see the review by Oakley (103).

Why are redoxosomal pathways potentially important in ALS? The answer to this question stems from the finding that SOD1 actively recruits to both IL-1β- and TNFα-activated redoxosomes (Fig. 2) (82, 83, 100). Additionally, SOD1 appears to be functionally important for the activation of NFκB; SOD1-deficient fibroblasts fail to effectively activate NFκB following IL-1β stimulation (100). Furthermore, the expression of ALS mutant forms of SOD1 (L8Q, G93A, and G93C) in both glial and neuronal cell lines enhances NADPH-dependent O₂^{•-} production and Rac1 activation in endomembrane fractions (55). This enhanced redox-stress also led to decreased cell viability in both cell types. Given the link between Nox1/Nox2 and disease progression in SOD1^{G93A} transgenic mice (90, 141), these *in vitro* findings are of considerable interest, because they demonstrate that mutant SOD1 expression can lead to enhanced Nox-activation in the absence of inflammation. The placement of SOD1 on redoxosomes also suggested the potential for a direct functional involvement of SOD1 in regulating Nox-mediated signaling by redoxosomes.

The finding that SOD1 is recruited to the redoxosome surface might suggest that it functions to locally dismutate O₂^{•-} to H₂O₂, and in this manner facilitates redox-dependent receptor activation. However, unlike H₂O₂, which freely diffuses across membranes (25), O₂^{•-} is relatively impermeable to cellular membranes (88, 119). Hence, if indeed SOD1 dismutation activity plays a role in this process, a pathway that allows O₂^{•-} to pass through endosomal membranes must exist. Recently, studies in isolated endosomes and reconstituted endomembrane proteoliposomes have demonstrated that indeed O₂^{•-} can permeate redoxosomal membranes, and that it does so through an unknown chloride channel(s) (100). Cumulatively, these findings suggest that SOD1 is potentially important for redoxosomal ROS metabolism and signaling.

The SOD1/Rac1 Interaction Serves as a Redox-Sensor for Redoxosomal Nox Regulation: Functional Abnormalities in ALS SOD1 Mutants

The importance of SOD1 in regulating redoxosomal ROS was elucidated by the finding that SOD1 can directly associate with Rac1 (Fig. 2) (55). Immunoprecipitation studies have demonstrated that Rac1 is associated with SOD1 in multiple organs including the kidney, liver, and brain. Additionally, *in vitro* studies with purified His-tagged Rac1 deletion mutants and bovine SOD1 have demonstrated that these two proteins directly associate via a region of Rac1 contained between amino acids 35 and 70. This region of Rac1 spans several domains that are important for nucleotide binding (*i.e.*,

switch I, G2, switch II, and G3 domains) (140). Reconstitution assays using His-tagged Rac1 have demonstrated that the interaction between Rac1 and SOD1 depends on the redox-state and nucleotide-bound state of Rac1 (55). When Rac1 is in a reduced state and bound to GTP, it most efficiently binds to SOD1. Oxidation of Rac1 with as little as 50–100 pM of hydrogen peroxide prevents SOD1 association with Rac1. This process is reversible when Rac1 is reduced by the addition of 100–300 μM DTT. The association of SOD1 with Rac1 leads to an inhibition in the intrinsic GTPase activity of Rac1 (55), and hence would be expected to activate Rac1-regulated NADPH oxidases under reducing conditions. Indeed, the addition of SOD1 to isolated fibroblast endosomes *in vitro* leads to enhanced NADPH-dependent superoxide production by Nox2 (55); however, this enhancement is transient, since the accumulation of ROS leads to the dissociation of SOD1 from Rac1, and to inactivation of Rac1 through the hydrolysis of GTP to GDP. Thus, SOD1 appears to act as a redox-sensor that controls Rac1-mediated NADPH oxidase activation in the redoxosome (Fig. 3). Superoxides also appear to be capable of dissociating SOD1 from Rac1-GTP *in vitro*, leading to enhanced GTP hydrolysis by Rac1 (55). Although it is clear that the oxidation of Rac1 by very low levels of hydrogen peroxide plays an important role in dissociating SOD1 from Rac1, it remains to be determined if SOD1 locally provides this source of hydrogen peroxide for Rac1 oxidation on the redoxosome surface.

Of importance to ALS, studies have demonstrated that certain mutant forms of SOD1 (L8Q and G10V) have increased affinity for Rac1 (55). In this context, these SOD1 mutants more effectively stabilized the Rac1-GTP complex by

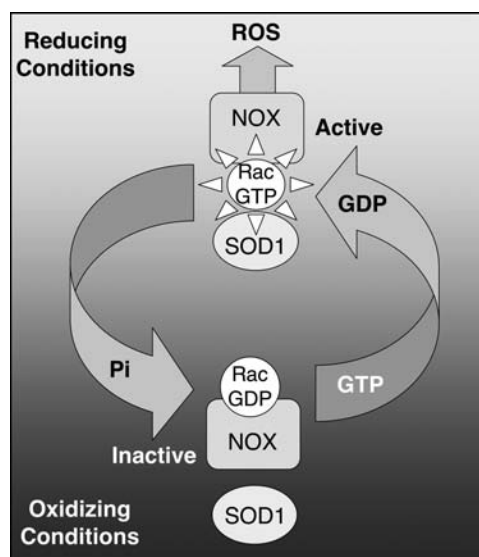


FIG. 3. Redox-dependent association of SOD1 with Rac1 serves as a redox-sensing mechanism that controls NADPH oxidase activation. Under reducing conditions, SOD1 associates with Rac1-GTP, and inhibits intrinsic and GAP-mediated GTP hydrolysis, maintaining Rac1 in an active state. This active form of Rac1 stimulates Nox activation and ROS production. When NADPH oxidase produces sufficient ROS to establish an oxidizing local environment, SOD1 disengages from Rac1 and Rac1-GTP is hydrolyzed to Rac1-GDP. This leads to shut down of the Nox complex.

decreasing intrinsic GTPase activity. Most interestingly, the association of mutant SOD1 with Rac1 was redox-insensitive, and the two proteins failed to dissociate in the presence of xanthine/xanthine oxidase derived $O_2^{\bullet-}$, a finding distinctly different from that of wild-type SOD1. These findings suggested that ALS-causing mutants of SOD1 can alter Rac1 activation, and hence may also influence the activity of Rac1-regulated Nox complexes. *In vivo* support for this hypothesis comes from findings in *SOD1^{G93A}* transgenic mice, in which both Rac1 activation and Nox-dependent ROS production are elevated in the spinal cord (55, 90, 141). However, conclusive evidence that an ALS-associated SOD1 mutant can alter redoxosomal Nox2 activation came from reconstitution studies in isolated endosomes from primary fibroblasts that had been isolated from wild-type and Nox2-deficient mice (55). In this study, the addition of purified *SOD1^{L8Q}* protein to isolated endosomes gave rise to prolonged NADPH-dependent $O_2^{\bullet-}$ production in comparison to that stimulated by the addition of *SOD1^{WT}* protein. However, Nox2-deficient endosomes exhibited significantly reduced NADPH-dependent $O_2^{\bullet-}$ production following the addition of *SOD1^{WT}* protein, suggesting that, at least in this cell type, SOD1 activated Nox2. These findings suggest that SOD1 can directly regulate endosomal Nox activation by acting as a redox-sensor for Rac1 activation and inactivation. Furthermore, redox-sensitive uncoupling of SOD1 from Rac1 appears to be dysfunctional in certain ALS-associated mutants of SOD1, and this defect leads to hyperactivation of Nox-derived $O_2^{\bullet-}$ by endomembranes. Such findings may help to explain the gain-of-function phenotype caused by familial ALS SOD1 mutations associated with enhanced redox-stress.

The ability of H_2O_2 in pico-molar quantities to liberate SOD1 from Rac-GTP, and to allow for GTP hydrolysis to occur, suggests that the mechanism underlying *in vivo* Nox regulation may be exquisitely sensitive to small changes in cellular ROS. This mechanism may allow Rac1 to sense and regulate changes in cellular $O_2^{\bullet-}$ through SOD1 enzymatic conversion of $O_2^{\bullet-}$ to H_2O_2 . Such spatial regulation may be a key aspect of SOD1 function as a redox-sensor of inflammatory signals that utilize the redoxosome. Hence, we propose an alternative model for pathogenesis in certain familial forms of ALS involving SOD1 mutations (Fig. 4). According to this model, SOD1 mutations directly dysregulate redoxosomal Nox2 (and potentially also Nox1) activation, leading to heightened and prolonged ROS production in response to pro-inflammatory signals. In the context of $TNF\alpha$ and $IL-1\beta$ receptor activation, elevated endosomal Nox activation would be expected to enhance downstream pro-inflammatory signaling pathways such as $NF\kappa B$ (82, 83). Very recent support for the model shown in Fig. 4 has emerged from mixed cultures of human embryonic stem cell-derived astrocytes and motor neurons (89). This study demonstrated that expression of the *SOD1^{G37R}* ALS mutant protein in astrocytes leads to increased Nox2-dependent ROS production and hyperactivation of pro-inflammatory pathways that can selectively kill motor neurons in a non-cell-autonomous fashion.

Whether SOD1-mediated regulation of Nox2 is the event responsible for initiating disease progression in ALS, or only enhances ROS-mediated injury, remains to be determined. In this context, it is also important to point out that the regulation of ROS production by Rac1 is important for many cellular processes that are involved in signal transduction (129), actin

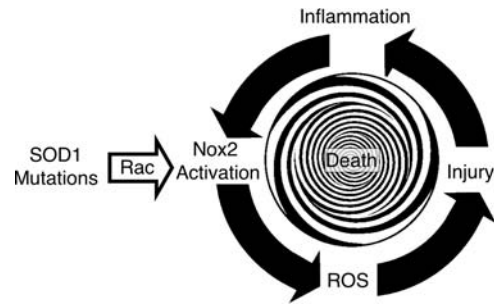


FIG. 4. Model for disease progression in familial forms of ALS that are caused by defects in SOD1-mediated regulation of the Rac/Nox2 complex. In this model, mutations in SOD1 leads to enhanced redox-insensitive binding to Rac-GTP that stabilizes the GTP-bound active form of Rac. This enhanced activation elevates Nox2 activity, increasing ROS production by microglia. This inciting event begins the cascade of injury and enhanced inflammation that in turn lead to microgliosis, and thus to more Nox2-expressing cells. The expanded population of Nox2-expressing microglia further increases ROS production via a dysregulated interaction between mutant-SOD1 and Rac, and this cycle repeats until death occurs.

cytoskeletal rearrangements (71, 87, 102), cell migration (143), cell proliferation (66, 99, 108, 143), and cell differentiation (113). Given that many of these pathways also involve activation and recruitment of Rac1 to specific membrane domains, it is also possible that SOD1/Rac1 interactions may influence other cellular aspects of disease progression in ALS.

ALS2 and SOD1 Inherited Forms of ALS Exhibit Defects in Rac1 GTPase Regulation: A Potential Unifying Theme for Molecular Pathophysiology in ALS

The identification of SOD1 as an effector of Rac1, and the finding that ALS mutations in SOD1 enhance Rac1 activity, becomes even more interesting when one considers that mutations in the *ALS2* gene lead to a juvenile form of inherited ALS (26, 54, 144). The *ALS2* gene encodes two alternatively spliced products that encode a long (1659 amino acid) and a short (396 amino acid) form of a protein called Alsin. The full-length Alsin protein has three independent guanine nucleotide exchange factor (GEF)-like domains. These include: an N-terminal regulator of chromatin condensation (RCC1)-like domain, which resembles the GEF for the Ran GTPase; a middle Dbl homology (DH)- and pleckstrin homology (PH)-like domain similar to the GEF for Rho GTPase; and a C-terminal vacuolar protein sorting 9 (VPS9)-like domain that is homologous to the Rab GTPase GEF (26, 54). The exact mechanism by which mutations in *ALS2* lead to ALS-like disease remains unknown, and this is confounded by the fact that motor neuron degeneration is not observed in *ALS2* knockout mice (23). Interestingly, however, *ALS2* knockout mice do have increased susceptibility to paraquat-induced neuronal oxidative stress (23), lymphopenia and hematopoiesis abnormalities (40), and endosome trafficking defects (34).

Several molecular findings suggest that Alsin may be an important determinant of endosomal dynamics, and hence relevant to SOD1-mediated redoxosomal signaling defects in ALS. Although a link between Alsin activity and the regula-

tion of endosomal NADPH oxidases remains highly speculative, we review aspects of Alsin biology that are potentially important to this line of thinking. Alsin was originally thought to serve as a GEF for both Rab5 and Rac1 GTPases, and hence is considered a candidate activator of each of these proteins (106, 131). Although the GEF activity of Alsin with respect to Rab5 has been reproduced by several research laboratories, a relatively recent study suggests that Alsin functions as an effector, rather than a GEF, of Rac1 (76). This finding is of considerable interest since SOD1 also acts as an effector for Rac1 (rather than as a traditional GEF or GTPase-activating protein (GAP)) (Fig. 3) (55). Both the Rab5-GEF and Rac1-effector functions of Alsin appear to influence endocytic mechanisms and endosomal dynamics; activated Rac1 appears to recruit Alsin to the plasma membrane at sites of Rab5-dependent macropinocytosis, and Alsin appears to also promote the fusion of macropinosomes with EEA1/Rab5-positive early endosomes (Fig. 5A) (76).

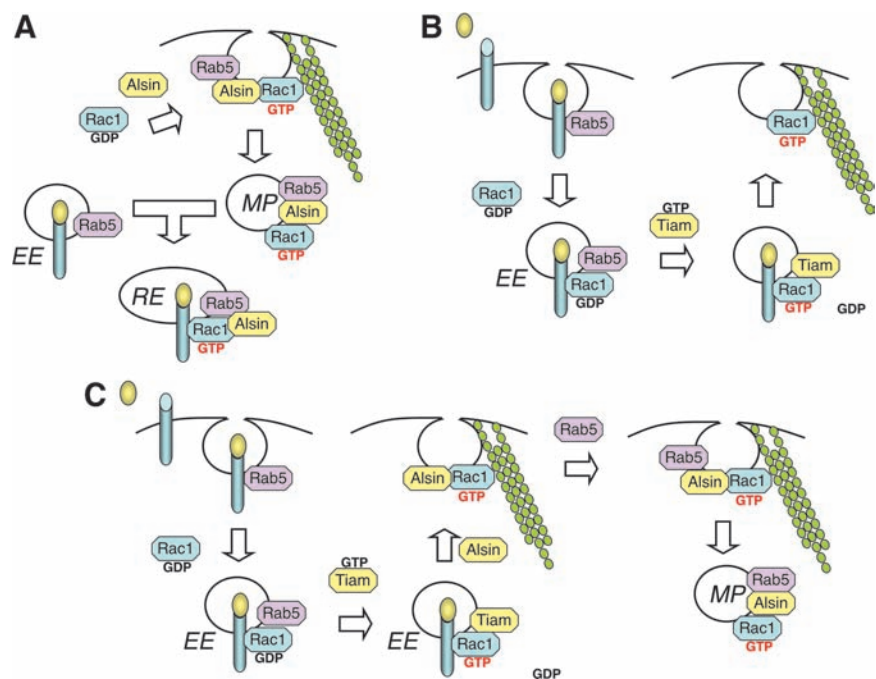
These functions of Alsin in endosome formation and trafficking are intriguing when compared to the biology of receptor-mediated redoxosome formation in the context of IL-1 β and TNF α signaling pathways. For example, IL-1 β -activated redoxosomes have been shown to initiate Nox2- and/or Nox1-dependent O₂^{•-} production in the early endosomal compartment, as determined by affinity isolation of Rab5 endosomes and the cellular co-localization of early endosomal antigen (EEA1) with O₂^{•-}-sensitive fluorochromes (82, 97) (Fig. 2). Furthermore, the recruitment of Nox2 from the plasma membrane into IL-1R1-stimulated redoxosomes is dependent on Rac1

(82). Lastly, SOD1 is recruited to Rab5-positive IL-1 β /IL-1R-containing redoxosomes (100), where it binds to Rac1 and regulates Nox2 activity in a redox-dependent fashion (55) (Fig. 3). However, in contrast to Rac1/Alsin-dependent macropinocytosis, which is most likely dynamin-independent (14, 31, 76), endocytosis of ligand-activated IL-1R1 is dependent on dynamin rather than on Rac1 (82). Interestingly, Alsin has been shown to selectively bind ALS mutants of SOD1 via its DH/PH domain (69), a finding reminiscent of enhanced binding of ALS SOD1 mutants to Rac1 (55).

Cumulatively, these findings place four early endosomal proteins (SOD1, Alsin, Rab5, and Rac1) in intersecting endosomal compartments where both Alsin and SOD1 have been shown to play roles as effectors of Rac1 (55, 76). Interestingly, it was also recently demonstrated that Rab5-mediated endocytosis is required for Rac1 activation in the endosomal compartment following mitogenic stimulation, and that this process plays a role in localizing activated Rac1 to the plasma membrane where it facilitates the formation of actin-based migratory protrusions (107) (Fig. 5B). In this context, Tiam1 (a Rac1-GEF) activates Rac1 in the early endosomal compartment prior to its association with actin at the plasma membrane. Given that actin-based rearrangements are also important for Alsin/Rac1-dependent macropinocytosis (76) (Fig. 5A), it is possible that this Tiam-mediated activation of Rac1 also plays a role in macropinocytic mechanisms involving Alsin (Fig. 5C). Hence, Rac1/SOD1/Alsin/Rab5 signaling networks may act in concert to regulate cell signaling at the endosomal level, and dysfunction of these pathways in ALS

FIG. 5. Models for Alsin function in endosomal dynamics.

(A) According to a model proposed by Kunita *et al.*, Alsin plays an important role in macropinocytosis following its recruitment by Rac1 to plasma membrane ruffles at sites of organized actin filaments (76). In this context, Alsin may stabilize Rac1-GTP and hence act as a Rac1-effector. In addition, the Rab5-GEF function of Alsin appears to control maturation of the macropinosome (MP) and later fusion of macropinosome with the early endosomal compartment (EE). This process may be important for the movement of endosomal components into the recycling endosome (RE). (B) In a model proposed by Palamidessi *et al.*, Rab5-mediated endocytosis into the early endosomal compartment is required for Rac1 activation by Tiam1, a Rac1-GEF that exchanges GDP for GTP. Rac1-GTP is then recruited to the plasma membrane where it facilitates the formation of actin-based migratory protrusions (107). (C) In a combined model that takes into account findings from both the Kunita and Palamidessi studies, Tiam1 activates Rac1 prior to recruitment of Alsin to the plasma membrane. As in model B, this process may be initiated by a ligand stimulus that leads to the formation of early endosomes. Once Rab5/Alsin/Rac1-GTP complexes are formed at the plasma membrane, spatially-restricted association with organized actin filaments leads to the formation of macropinosomes and downstream endosomal dynamics controlled by this compartment. The potential involvement of Tiam1 in this process is particularly interesting, since its homolog, Tiam2, has been associated with sporadic ALS (37). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at www.liebertonline.com/ars).



(through mutations in either Alsin or SOD1) may lead to disease. As discussed later in this review, the potential involvement of TIAM proteins in Alsin-mediated endosomal dynamics is of special interest since the *TIAM2* gene was identified as associated with sporadic forms of ALS (37).

Given that both Alsin and SOD1 are effectors of Rac1, it is tempting to speculate that defects in either protein could influence redox-signaling at the endosomal level. This would be consistent with two sets of findings. First, Alsin overexpression can mitigate neurotoxicity caused by the expression of a mutant form of SOD1 (69), suggesting that Alsin counterbalances a gain of function in SOD1. In glial and neuronal cell lines, the expression of SOD1 mutants leads to enhanced ROS production in the endomembrane compartment, and ultimately to cell death (55). Inhibiting ROS production under these conditions—by applying the Nox2 inhibitor apocynin—protects glial cells from this SOD1 mutant toxicity (55). Whether Alsin's association with mutant SOD1 and/or Rac1 also mitigates hyperactivation of Nox2 in the endosomal compartment remains to be determined. Second, *ALS2* knockout mice demonstrate an increased susceptibility to paraquat-induced neuronal oxidative stress (23) as well as defects in endosome trafficking (34). Hence, Alsin appears to protect cells from redox-stress while also influencing endosomal dynamics. Cumulatively, these findings

suggest the possibility that Alsin may protect cells from redox-stress by associating with mutant SOD1 and/or Rac1 on Nox-active endosomes.

Two models that could potentially explain how Alsin and SOD1 might influence Nox activation in the endosomal compartment are shown in Fig. 6. According to the first model, Alsin might play a role in the processing of receptor-activated redoxosomes to an inactive state (Fig. 6A). In this model, fusion of macropinosomes with receptor-activated redoxosomes leads to inactivation of the Nox complex by recycling or degradation of its components in the endocytic pathway. Loss of either Alsin or SOD1 function would in this case dysregulate the pathway by reducing the rate of fusion between these compartments, due to altered Rac1 or Rab5 activation. In contrast, Alsin overexpression might accelerate the fusion process, leading to a more rapid disposal of redoxosomes. According to the second model, Alsin may play an unknown role in receptor-mediated endocytosis and the regulation of Rab5, Rac1, and/or SOD1 in the redoxosome (Fig. 6B). In this context, Alsin binding to Rac1 and/or SOD1 could influence activation of the Nox complex in redoxosomes by facilitating conformational changes in the Rac1/SOD1 complex. In this model, Alsin could be part of a redox-sensor required to shut down the Nox complex, potentially by competing for SOD1 binding to Rac1 on the redoxosome.

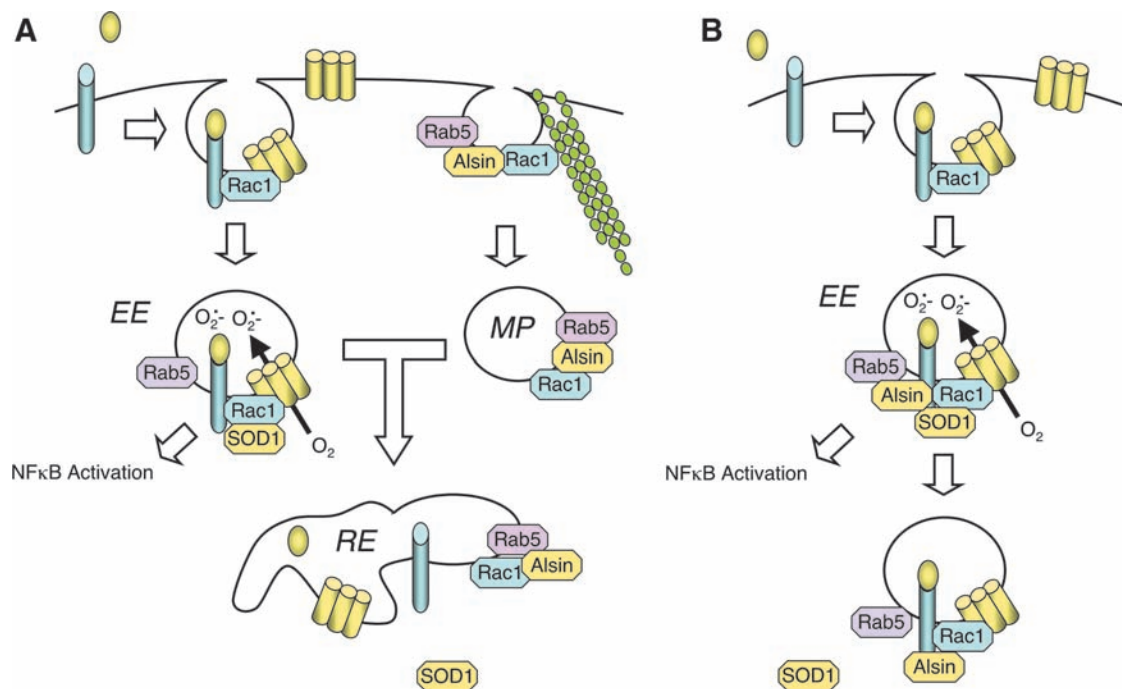


FIG. 6. Two models that position Alsin as a protective modulator of redox-signaling at the endosomal level. (A) This model proposes that Alsin facilitates endosomal processing of redoxosomes. Accordingly, ligand-triggered redoxosome formation mobilizes Nox to the early endosomal (EE) compartment, where Rac1/SOD1 complex formation leads to Nox activation and ROS production (55, 82). Macropinosomes (MP) form through the previously described process involving Alsin, Rac1, and Rab5 (76), and this compartment may be important for directing redoxosomes to the recycling endosomal (RE) compartment. This Alsin/Rab5-facilitated fusion event could lead to the processing of redoxosomal components and inactivation of the Nox complex. (B) In a second model, ligand stimulation leads to direct recruitment of Alsin to the redoxosome, through its binding to a component(s) of the Rab5, Rac1, SOD1 complex. This large molecular complex may facilitate the redox-dependent uncoupling of SOD1 from Rac1 to shut down the Nox complex following signal transmission. Such a scenario might involve competitive binding of Alsin to Rac1 in a manner that excludes SOD1 from the complex (as shown), or changes to the complex that enhance Rac1-GTP hydrolysis. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at www.liebertonline.com/ars).

Although these models are currently both highly speculative and other scenarios could be envisioned, both are consistent with Alsin protecting against redox-stress.

Genome Analysis of Sporadic ALS Uncovers Other Potential Nox/Rac Regulators

A recent study described a genome-wide search for single-nucleotide polymorphisms (SNPs) that associate with sporadic ALS (37). This search of over 750,000 SNPs in over 1,200 ALS patients and 1,500 normal subjects identified 10 genetic loci with a significant association with sporadic ALS in three independent population series, and 41 additional loci that were significant in two of the three series. Interestingly, six of the associated genes encode proteins known to either directly or indirectly influence Rac1 and/or NADPH oxidases activation (Fig. 7). This section places in context the known functions of these genes—in regulating Rac1, NADPH oxidases, and/or endosomal dynamics—that may influence redoxosome function. With speculative models, we will attempt to lay a foundation for further thinking about how diverse mutations in sporadic and inherited forms of ALS may have overlapping mechanisms of pathogenesis.

Nox4

The *Nox4* gene was identified as significantly linked to sporadic ALS in two of three series of patient studies (37). Given the finding that deletion of *Nox1* or *Nox2* genes in *SOD1*^{G93A} transgenic mice significantly prolongs survival (90, 141), the finding that the *Nox4* isoform of NADPH oxidase may also be linked to certain sporadic forms of ALS is quite intriguing. Unlike *Nox1* and *Nox2*, *Nox4* appears to be significantly regulated at the transcriptional level in response to disease processes and certain stimuli (9, 60). *Nox4* is expressed in neurons, and its mRNA increases following ischemia (133). The mechanisms that control *Nox4* activation appear to be quite different than those for *Nox1* and *Nox2*

activation (60). For example, Rac1 and other co-activator cytosolic subunits appear to be dispensable for *Nox4* activation in certain cell systems (9, 91). However, others have also demonstrated that Rac1 is required for *Nox4* activation by certain cell stimuli (48). It is interesting that mutant *SOD1* overexpression enhances *Nox*-dependent ROS in endomembranes from both glial and neuronal cell lines. However, only ROS production by glial cells is inhibited by apocynin (a known *Nox2* inhibitor that likely prevents p47phox recruitment to the *Nox2* complex) (55). Hence, it is possible that p47phox-independent NADPH oxidases (such as *Nox4* or *Nox1*) are responsible for neuronal ROS production stimulated by mutant *SOD1*.

TIAM2 (STEF)

The *TIAM2* gene was identified as significantly linked to sporadic ALS in two of three series of patient studies (37). *Tiam1* and *Tiam2* (also known as *STEF*) are two closely related Rac1 guanine nucleotide exchange factors (GEFs) (27, 53, 61, 95, 96). Both are expressed in the brain and other neuronal tissues, and activate Rac1 by exchanging bound GDP for GTP. *Tiam* GEFs are generally thought to regulate the actin cytoskeletal at the membrane through the activation of Rac1 (96), and have been shown to be involved in neuronal developmental, growth, and migration (70, 93, 146). *Vav1* is another Rac-specific GEF (22), and is generally thought to be responsible for activating Rac-dependent NADPH oxidases such as *Nox1* and *Nox2* (45, 98, 116, 137). Although *Tiam2* has not been reported to directly influence the activation NADPH oxidase, *Tiam2* association with the small GTPase *Rap1* can control activation of Rac1 (147). Interestingly, *Rap1* (also known as *Krev-1*), which associates with the phagocytic *Nox2* complex (39, 114), was recently demonstrated to positively influence *Nox2*-dependent superoxide production in neutrophils of *Rap1a* knockout mice (84). Hence, *Tiam2* may indirectly influence *Nox* activation through its control of *Rap1*/*Rac1*/2 dynamics at the membrane. The recent characterization of *Alsin* as a Rac1 effector that influences endocytosis from actin cytoskeletal contacts at the plasma membrane (76), provides yet another potential commonality between *ALS2* and *TIAM2*-associated sporadic forms of ALS (*i.e.*, both proteins regulate Rac1 and actin cytoskeletal dynamics). Additionally, *SOD1* has also been identified as a Rac1 effector that, when mutated in ALS, leads to excessive activation of both Rac1 and NADPH oxidase (55). Hence, *Tiam2*, *Alsin*, and *SOD1* are all Rac1 effectors/activators that are associated with both inherited and sporadic forms of ALS (Fig. 8).

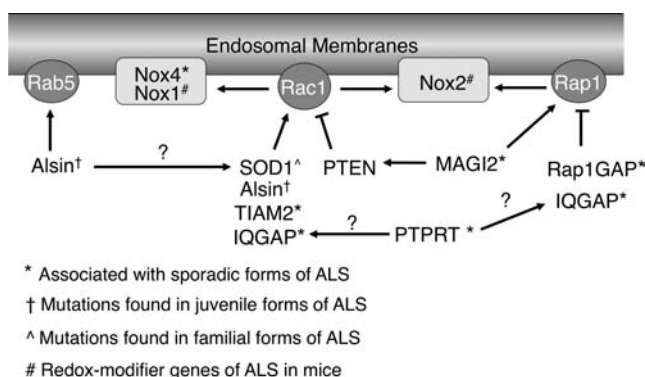


FIG. 7. Potential relationships between gene products that have been linked to ALS disease progression in mouse models, sporadic ALS, and various inherited forms of ALS. Arrows denote an activating relationship, and flat-headed lines denote an inhibitory relationship. Question marks denote a previously described relationship that would have unknown consequence in the current scheme (*i.e.*, *SOD1*/*Alsin* interaction), or a potential relationship based on a findings from a close homolog (*i.e.*, *PTPRT*/*PTPRM*). See text for details.

IQGAP2

The *IQGAP2* gene was identified as significantly linked to sporadic ALS in two of three series of patient studies (37). *IQGAP*-1, -2, and -3 are related calmodulin-binding IQ-motif proteins with significant homology to GAPs (17, 78). These proteins regulate actin cytoskeletal dynamics at the plasma membrane and cell migration, by controlling activation of the *Cdc42* and *Rac1* GTPases (17, 18, 92). Typically, guanine nucleotide exchange factors (GEFs) and GTPase-activating proteins (GAPs) are thought to either activate or inactivate small GTPases by exchanging GDP for GTP (GEFs), or by enhancing GTPase activity (GAPs), respectively (15). Although the name implies that *IQGAPs* might inhibit small GTPases through

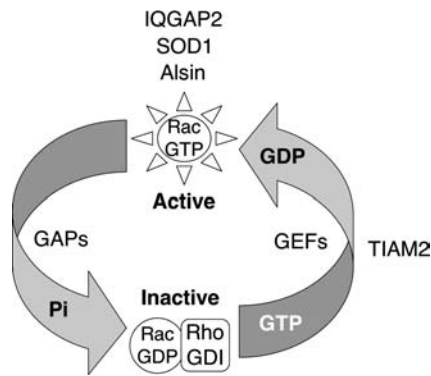


FIG. 8. Rac1 effectors and activators identified or associated with various forms of ALS in humans. Rac1 activity is controlled by nucleotide binding. The inactive form, Rac1-GDP, is held in the cytoplasm through its association with RhoGDI. Rac1 is activated by guanine nucleotide exchange factors (GEFs) and inactivated by GTPase activating proteins (GAPs). GEFs facilitate the exchange of GDP for GTP, while GAPs enhance GTP hydrolysis. IQGAP2, SOD1, and Alsin are known Rac1 effectors that bind to and stabilize the GTP-bound form of Rac1. These three molecules do not appear to act as either traditional Rac1-GEFs or GAPs (19, 55, 76). In contrast, TIAM2 serves as a GEF for Rac1 (27). *TIAM2* and *IQGAP2* genes were recently associated with sporadic ALS (37). SOD1 and Alsin have also been shown to influence the dynamics of Rac1 activation, to recruit to the endosomal compartment, and to also be mutated in certain forms of ALS (55, 76, 130, 131). See text for more detailed descriptions of these factors and their potential relationships to ALS.

GAP-associated activity, this is actually not the case. IQGAP1 associates with the GTP-bound forms of Cdc41 and Rac1 through a GAP-related domain (77), and suppresses intrinsic GTPase activity by stabilizing the GTP-bound form, which in turn increases the activity of both Cdc41 and Rac1 (17, 56). Unlike IQGAP1, IQGAP2 appears to associate equally with both GDP- and GTP-bound forms of Rac1 and Cdc42. However, similar to IQGAP1, IQGAP2 retains the ability to inhibit both RhoGAP-mediated and intrinsic GTP hydrolysis by Rac1 and Cdc42 (19, 120). Additionally, IQGAP3, which is highly expressed in the brain, has been shown to be required for the activation of Cdc42- and Rac1-dependent neurite outgrowth during neuronal morphogenesis (136). Hence, much like Alsin (76) and SOD1 (55), IQGAPs appear to function as effectors of Rac1. It is also interesting to note that IQGAP2 and SOD1 can both associate with GDP- and GTP-bound forms of Rac1 (19, 55, 120), although in the case of SOD1 there is a nucleotide preference that is dependent on the redox-state of Rac1 (55). IQGAP1 has been shown to increase ROS production by activating Nox2, and in this manner facilitates migratory wound healing in vascular endothelial cells (65). However, it is currently unclear if IQGAP2 also influences Nox activation. In summary, the studies by Dunckley *et al.* that have identified the *TIAM2* and *IQGAP2* genes as tightly linked to sporadic forms of ALS (37) add two additional Rac1 effectors to the existing list of Alsin and SOD1. Interestingly, *TIAM2*, *IQGAP2*, Alsin, and SOD1 all appear to influence the abundance/stability of the GTP-bound form of Rac1, and hence the mechanism by which defects in these proteins lead to ALS may overlap (Fig. 8).

PTPRT (PTPrho)

The protein tyrosine phosphatase receptor type-T (called *PTPRT* or *PTPrho*) gene was shown to be significantly linked to sporadic ALS in three of three series of patient studies (37). This gene encodes one of four related transmembrane proteins that fall into a family of receptor protein tyrosine phosphatases (RPTPs) (12). This family of proteins is characterized by the presence of a novel extracellular N-terminal MAM domain (meprin/A5/micro), which includes an immunoglobulin-like domain and four fibronectin type III (FN) repeats, and plays a role in homophilic cell adhesion (3, 4). *PTPRT* (*PTPrho*) is most closely related to *PTPRM* (*PTPmu*), another member of the family of RPTPs. Both of these proteins are highly expressed in the brain based on NCBI Unigene EST expression profiles. Interestingly, *PTPRM* directly interacts with IQGAP1 to promote neurite outgrowth, and peptide competition of IQGAP1 binding to Cdc42 and Rac1 blocks this *PTPRM*-mediated process (110). These findings are similar to those demonstrating that IQGAP3 is required for the induction of Cdc42- and Rac1-dependent neurite outgrowth during neuronal morphogenesis (136). Hence, *PTPRM* appears to activate Cdc42 and Rac1-dependent neurite outgrowth through IQGAP1. It is currently unclear if *PTPRT*, which has been linked to sporadic ALS (37), also possesses the ability to influence the activation of Cdc42 and Rac1. However, if *PTPRT* functions similarly to *PTPRM* in this context, *PTPRT* would join Alsin, SOD1, IQGAP2, and *Tiam2* as activators/ effectors of Rac1. More research in this area is obviously needed to support this hypothesis.

RAP1GAP (GARNL4)

The *RAP1GAP* gene (also known as *GARNL4*) was shown to be significantly linked to sporadic ALS in two of three series of patient studies (37). *Rap1GAP* (GTPase activating Rap/RanGAP domain-like 4) is a Rap1 GTPase activating protein (GAP) and hence inactivates the small GTPase Rap1 by promoting the hydrolysis of bound GTP (*i.e.*, Rap1-GTP is active and Rap1-GDP is inactive). Rap1 is a member of the Ras family of small GTPases and participates in numerous cell signaling events at the plasma membrane, where it controls integrin-mediated cell adhesion and the formation of cadherin-based cell-cell junctions (74). Interestingly, Rap1 physically associates with and activates VAV, *TIAM2*, and *TIAM1* (all GEFs for Rac1) (6, 43, 147). As discussed above, the *TIAM2* gene was also associated with sporadic ALS (37). It is currently thought that Rap1 may coordinate the recruitment of Rac1-GEFs to sites of cell-cell contact, and provide a link to the actin cytoskeleton (74) (Fig. 9). Interestingly, IQGAP also binds to both the GDP- and GTP-bound forms of Rap1, and appears to negatively regulate Rap1 activation, either by promoting Rap1 association with Rap1GAPs or by inhibiting its association with Rap1GEFs (67). The *IQGAP* gene was also associated with sporadic ALS in genetic studies (37). Activation of Rap1 is mediated by Rap1GEFs, including PDZ-GEF1, which is of particular interest since it associates with the MAGI scaffold proteins at β/α -catenin/cadherin-based cell-cell contacts to control Rap1 activation (74, 118) (Fig. 9). Overexpression of *Rap1GAP* also impairs Rap1-mediated VE-cadherin-dependent cell adhesion (118). In this context, MAGI controls the ability of PDZ-GEF1 to activate Rap1 following cell-cell contact, and hence counterbalances *Rap1GAP* activity. As discussed

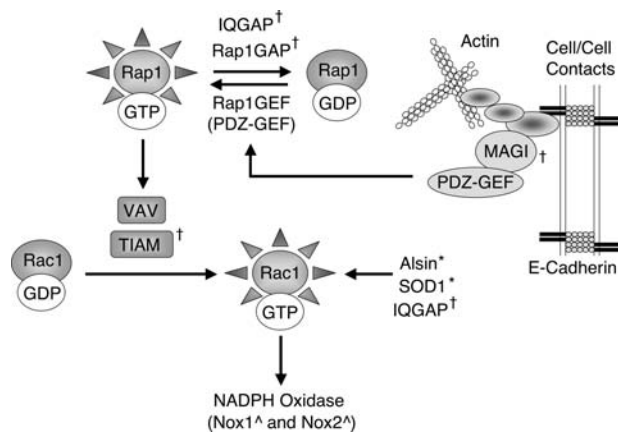


FIG. 9. Mechanistic relationships between actin cytoskeletal dynamics and the control of small GTPases Rap1 and Rac1. Shown is a schematic diagram outlining known relationships between genes associated with sporadic ALS (IQGAP, Rap1GAP, MAGI, and TIAM) (37), and their potential influence on Rac1 activation and function. Arrows denote an activating relationship between connected effectors. [†]denotes genes identified in genomic screens as strongly associated with sporadic ALS (37). *denotes genes known to be mutated in inherited forms of ALS. [^]denotes mouse redox-modifiers genes known to influence disease progression in the *SOD1*^{G93A} transgenic mouse model. See text for details.

below, *MAGI2* is yet another gene identified in genetic association studies of sporadic ALS (37).

Rap1GAP is also potentially of interest from the standpoint of redox-stress, since it was first shown to be highly expressed in neutrophils and to be associated with the NADPH oxidase complex (39, 114). For many years, Rap1 was thought to serve as the small GTPase required for Nox2 activation. Although Rac1/2 was later shown to have this role, recent studies have begun to indicate that Rap1 can also influence Nox2 activation. For example, Rap1a knockout mice have reduced respiratory bursts in isolated neutrophils (84), suggesting that Rap1 does indeed influence Nox2 activation (perhaps indirectly through the recruitment and activation of Vav, a Rac1-GEF). However, another report suggests that Rap1 suppresses Ras-mediated production of ROS in T-cells that is thought to be derived from NADPH oxidase (115).

Many of the emerging features of Rap1 paint a picture of protein/protein interactions and connected pathways suggestive of signaling networks for ALS-causing mutations and ALS-associated genes. At face value, many of these known or potential ALS-causing genes—such as *IQGAP*, *Rap1GAP*, *MAGI2*, *TIAM2*, *ALS2*, and *SOD1*—appear unrelated. However, as shown in Fig. 9, these genes and their protein products may indeed be connected—by cellular processes that regulate the cytoskeleton, Rac1, and/or Rac1-regulated NADPH oxidases. For example, mutations in Rap1Gap and IQGAP that inhibit their association with Rap1 might lead to higher levels of activated Rap1-GTP, which would in turn lead to enhanced Rac1 activation. Similarly, defects in MAGI2 could potentially influence PDZ-GEF activation, leading to reduced or enhanced Rap1 → Rac1 activation. Lastly, mutations in TIAM, IQGAP, Alsin, and/or SOD1 also have a proven ability, or the potential, to alter Rac1 activation. It remains to be determined whether the

connections across these pathways are a conserved feature of the pathophysiological mechanisms in ALS, or simply coincidence.

MAGI2

The *MAGI2* gene (also known as membrane-associated guanylate kinase inverted-2) was shown to be significantly linked to sporadic ALS in three of three series of patient studies (37). *MAGI1* and *MAGI2* are scaffold proteins that connect cadherin cell-cell contacts and actin cytoskeleton to signaling domains at the plasma membrane. As discussed above (Fig. 9), *MAGI* proteins are required for Rap1 activation initiated following cell–cell contact (74). Interestingly, *MAGI2* directly binds to and activates PTEN, a membrane-associated lipid phosphatase (64, 135, 142). The PTEN protein is also stabilized by interactions with *MAGI2* at the membrane (128), and negatively regulates Rac1 and Cdc42, and inhibits cell migration facilitated by these two small GTPases (85). In this context, Liliental *et al.* demonstrated enhanced Rac1 and Cdc4 activation in PTEN knockout cells that demonstrated a significantly increases migratory phenotype. This phenotype was also reversed by expression of dominant-negative forms of Rac1 and Cdc42. Hence, *MAGI2* may also have the ability to regulate Rac1 through its control of PTEN (Fig. 7). In this context, null mutations in *MAGI2* would be expected to lead to enhanced Rac1 activation by suppressing PTEN-mediated inhibition of Rac1.

Therapeutic Approaches to ALS

Clinical trials aimed at identifying drugs for the treatment of ALS have been conducted for over 50 years. Although over 20 drugs have been evaluated in clinical trials over the past 10 years, only riluzole (Rilutek) has emerged as a product (28, 81). Moreover, although the clinical trials for riluzole demonstrated a statistically enhanced survival of 60 days for patients being treated with the drug (11, 79), new therapeutic candidates are still being sought due to its limited efficacy. New sets of hypotheses have been generated based on the pathogenesis of ALS and on findings from genetic, pathological, and biochemical postmortem evaluations. Drugs candidates have been chosen for their potential to reduce excitotoxicity, apoptosis, motor neuron degeneration, bioenergetics, or oxidative damage (24, 62, 81, 132). As discussed throughout this review, oxidative stress has been implicated in the pathogenesis of ALS. Therefore, a relatively straightforward approach to development of therapies has been to evaluate anti-oxidants as a means of controlling this process. However, results from clinical trials for ALS that are based on anti-oxidant strategies have so far yielded unimpressive results (105), as highlighted below.

- Many natural biological agents such as vitamins C and E have antioxidant effects and are readily available; thus physicians commonly advise ALS patients to take these (105). However, although randomized clinical studies of high-dose vitamin E (5,000 mg) intake for 18 months in patients who were also taking riluzole found this regimen to be safe, they also indicated that the patients derived no survival benefit (44, 105).
- Creatine improves motor function in transgenic *SOD1*-based animal models of ALS, however, results in human patients have not been successful. In ALS transgenic

mice, oral administration of creatine has a positive impact on abnormal mitochondrial function (72, 73). However, in clinical studies in which creatine was supplemented in the diet, patients were found not to derive any benefit (51, 123, 124).

- Coenzyme Q10, a cofactor in the electron transfer chain, is also a scavenger of free radicals in lipid and mitochondrial membranes, and is thought to improve energy production by the mitochondrial membrane and the cell. Although the administration of high doses (up to 3,000 mg) of CoQ10 plus vitamin E (1,500 mg) to ALS patients was found to be generally safe, no clinical benefit was detected (41).
- Edaravone, a potent scavenger of free radicals that is approved in Japan for the treatment of ALS, was tested in 20 Japanese patients in a small double-blind study (145). The study showed only limited improvement in the ALSFRS-R score.

Two compounds with antioxidant properties remain in clinical studies. The manganese porphyrin AEOL 10150 is a potent catalytic antioxidant (16), and the injection of this compound into ALS mice when they became symptomatic markedly prolonged survival (30). Two Phase I studies testing the safety and feasibility of AEOL 10150 have been completed, and they indicate that the administration of this drug at doses up to 2 mg/kg/day was safe and well tolerated, with an excellent pharmacokinetic profile in ALS subjects (104, 132). A follow-on, phase I open-label study of AEOL 10150 was recently initiated in a patient diagnosed with progressive and debilitating ALS, as a compassionate-use, multiple-dose study; this trial is designed to evaluate the safety and efficacy of AEOL 10150 in an ALS patient over an extended period of time.

The second compound currently under study is KNS-760704, which provides protection from oxidative stress-related neurotoxic cascades and has neuroprotective properties (50). A double-blind, randomized, placebo-controlled study evaluating the safety and efficacy of KNS-760704 compared to placebo is ongoing. The study will be conducted in two parts. In Part 1, ~80 eligible patients will receive one of the following treatments for 12 weeks: placebo, low-dose, mid-dose, or high-dose KNS-760704. Participants who complete all 12 weeks of the Part 1 phase will be eligible for randomization in the Part 2 phase of the study. The duration of Part 2 of the study will be 28 weeks. Subjects of the Part 2 study will be divided randomly into one of two active treatment groups for 24 weeks (low-dose or high-dose KNS-760704), and will receive placebo for the remaining 4-week period (63).

Several therapies developed based on other current hypotheses for ALS pathogenesis have also been tested in double-blind, placebo-controlled clinical trials in ALS patients, but have failed (35, 81). Specifically, these trials have tested therapies based on anti-glutamate, antioxidant, neuroprotection, anti-inflammatory and anti-apoptotic strategies (35, 46, 81).

The antiexcitotoxic drugs are intended to have a therapeutic impact on the harmful effects of repetitive firing or on the elevation of intracellular calcium by calcium-permeable glutamate receptors (21, 46). Repetitive firing in response to excessive glutamate release is a well-known cause of neuronal cell death. Glutamate transporters normally provide rapid uptake of glutamate, and ALS patients often have increased

glutamate in the cerebrospinal fluid, due to reduced glutamate transport—most often because of loss of the excitatory amino acid transporter 2 (EAAT2) (46, 57). Therefore, drugs that can affect this process have been pursued as potential therapeutics. To date, however, riluzole is the only one of these drugs for which test results have been sufficiently compelling to lead to its approval for the treatment of ALS. Notably, the exact mode of action of riluzole, a benzothiazole compound, is unknown and may actually be related to inactivation of voltage-dependent sodium channels or interference with intracellular signaling events, rather than to inhibition of glutamate release (11, 36, 79).

Intuitively, and from data derived from rodent studies, it was reasonable to evaluate the activity of neurotrophic factors for their ability to rescue or protect motor neurons from degeneration. However, pharmacokinetics, bioavailability, dose-limiting toxicities, and antibody-based inactivation of the neurotrophic factor have all contributed to a lack of success to date in clinical trials that have evaluated such factors (132). Also, minocycline, a tetracycline antibiotic with neuroprotective and anti-inflammatory properties, appeared to be effective in delaying disease onset and survival in ALS transgenic mice (75, 134, 148, 149). However, in a Phase III clinical trial of ALS subjects, patients treated with this drug unexpectedly exhibited substantially more rapid disease progression than those treated with the placebo (47).

Other modalities such as protease inhibitors, stem cell therapy, and gene therapy are either being considered or tested in clinical trials (46). Also, novel approaches to shut down the production of mutant SOD1 are under development for the subset of patients with these mutations (127). However, effective drug treatments for patients have remained elusive despite advances in our understanding of ALS pathology. Although the ALS field has gained valuable experience in the design and conduct of clinical trials from the clinical studies completed to date, it is unclear why none of these therapies except riluzole, which has only a modest effect on survival, has had a positive impact in patients. Whether the reason the experimental treatment did not show efficacy in any of these cases was because the therapy was truly not effective, the biological targets were misguided, or the trial design was inadequate, is difficult to assess. For instance, in spite of a strong scientific rationale for developing an antioxidant-based therapy, antioxidants—including vitamins C, E, selegiline, selenium, methionine, and—no significant effect on the primary outcome measure was observed in a meta-analysis of all antioxidant trials combined (105). Suggested limitations of these trials include insufficient power (Vitamin E, creatine) and low dosage (creatine) (81).

Although some of the potential therapeutics described above were efficacious in animal models, such as the mutant *SOD1* transgenic mouse, these preclinical results obviously have been unreliable in predicting the results of human studies (35). A recently published study suggests that most of the published results on the mutant *SOD1* transgenic mouse models may actually reflect noise in survival distribution, and the authors recommend a stringent minimum study design (121). Whether the mutant *SOD1* transgenic mouse accurately represents human sporadic ALS is also uncertain.

Many of the antioxidant therapies tested thus far may not have been effective because they are not based on inhibiting a validated therapeutic target or a specific pathway. As discussed earlier in this review, an interaction between SOD1

and Rac1 serves to regulate ROS production by Nox2. Additionally, several other genes that are either known to cause ALS or are associated with the disease appear to converge on Rac1 regulation. As in the case of mutant SOD1, these mutations may lead to alterations in the regulation of NADPH oxidases and in the overproduction of ROS. These events may precipitate a pathogenic cascade that impairs mitochondrial function and inhibits EAAT2 (the main glial glutamate transporter protein), which results in the presence of an excess of glutamate and a consequent increase in intracellular calcium, and ultimately in enhanced oxidative stress (46). Microglial activation in patients with either sporadic ALS or familial ALS results in the release of neuroinflammatory proteins, amplifying oxidative stress that ultimately promote neuronal death via programmed cell death. Thus, inhibiting the key biological target Nox2 using a potent inhibitor should suppress the pathogenic cascade. The administration of apocynin, an inhibitor of Nox2 activity, has been shown to improve the survival of *SOD1*^{G93A} transgenic mice and also to protect a glial cell line from ROS-induced cell death caused by overexpression of *SOD1*^{L8Q} or *SOD1*^{G93A} (55). More recently, apocynin has also been shown to protect human motor neurons from non-cell-autonomous pro-inflammatory toxicity derived from co-cultured human astrocytes expressing *SOD1*^{G37R} (89). Thus, based on its mechanism of action, apocynin may be considered a disease-modifying therapy for ALS. Safety and toxicology studies are required to test its therapeutic activity in humans. Initial clinical studies of apocynin will include an evaluation of the dose-response relationship, and a demonstration of biological activity using relevant markers. Once these are obtained, efficacy studies will be considered.

Concluding Remarks and Future Directions

Despite a tremendous amount of research on ALS (over 10,800 publications to date), a unifying pathophysiologic mechanism for this devastating disease has eluded the field for decades. Additionally, apparent differences in the mechanism(s) that underlie disease pathology in the mouse models of ALS and human ALS patients have made it challenging to use rational design in developing therapies with clinical efficacy. One might argue that a unifying pathophysiologic mechanism does not exist because multiple independent pathways can lead to the motor neuron degeneration that is observed in ALS. This may indeed be the case. However, it is equally possible that ALS-causing genes and gene modifiers will fall into functionally-related groups—and that unifying pathways for disease causation will emerge from shared, overlapping, and interconnected cellular functions. The recent success in genome-wide genetic association studies in sporadic ALS patients has begun to support this concept, and has identified a wide array of candidate genes that may either cause ALS or influence the progression of this disease. Interestingly, many of these genes are known to influence endosomal and cytoskeletal dynamics, and the regulation of both Rac1 and NADPH oxidases. It is currently unclear which subsets of these genes directly dysregulate ROS production and/or the inflammatory responses observed in ALS. However, regulation of Rac1 appears to be central to both the inherited forms of ALS caused by defects in the *ALS2* and *SOD1* genes and a subset of genes that have been associated with sporadic ALS. Given the finding that ROS is elevated in ALS-

affected tissues, genes that affect ROS production, either directly or indirectly through inflammatory processes, may provide a unifying mechanism for disease causation and disease severity. The challenge over the next decade will be to place the abundance of genes that are associated with sporadic ALS into the biologic context of the simpler monogenetic causes of ALS. Such studies will hopefully lead to a clearer understanding of pathophysiologic mechanisms, and also to more effective methods with which to approach disease treatment.

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Abbreviations

ALS, amyotrophic lateral sclerosis; ALS2, amyotrophic lateral sclerosis 2 gene; Alsin, amyotrophic lateral sclerosis protein 2; CSF, cerebrospinal fluid; Duox, dual oxidase; EEA1, early endosomal antigen 1; FALS, familial amyotrophic lateral sclerosis; GDP, guanosine diphosphate; GFP, green fluorescent protein; GTP, guanosine triphosphate; H₂O₂, hydrogen peroxide; IL-1 β , interleukin-1 β ; IL-1R1, interleukin-1 receptor 1; IKK, I κ B kinase; IKKK, I κ B kinase kinase; MyD88, myeloid differentiation primary response gene; NADPH, nicotinamide-adenine dinucleotide phosphate (reduced); NF κ B, nuclear factor κ B; Nox, NADPH oxidase; Rac1, Ras related C3 botulinum toxin substrate 1; redoxosome, redox-active endosome; RhoGDI, Rho GDP-dissociation inhibitor; ROS, reactive oxygen species; SOD1, superoxide dismutase 1; SALS, sporadic amyotrophic lateral sclerosis; TNF α , tumor necrosis factor α ; TNFR1, tumor necrosis factor receptor 1; TRAF2, tumor necrosis factor receptor associated factor 2; TRAF6, tumor necrosis factor receptor-associated factor 6; TRADD, tumor necrosis factor receptor-associated death domain.

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