

NIH Public Access

Author Manuscript

Biometrics. Author manuscript; available in PMC 2010 March 22.

Published in final edited form as:

Biometrics. 2007 June ; 63(2): 610–617. doi:10.1111/j.1541-0420.2006.00722.x.

On Models for Binomial Data with Random Numbers of Trials

W. Scott Comulada^{1,*} and Robert E. Weiss²

¹UCLA Center for Community Health, 10920 Wilshire Boulevard, Suite 350, Los Angeles, California 90024-6543, U.S.A

²Department of Biostatistics, UCLA School of Public Health, Los Angeles, California 90095-1772, U.S.A

Summary

A binomial outcome is a count *s* of the number of successes out of the total number of independent trials n = s + f, where *f* is a count of the failures. The *n* are random variables not fixed by design in many studies. Joint modeling of (s, f) can provide additional insight into the science and into the probability π of success that cannot be directly incorporated by the logistic regression model. Observations where n = 0 are excluded from the binomial analysis yet may be important to understanding how π is influenced by covariates. Correlation between *s* and *f* may exist and be of direct interest. We propose Bayesian multivariate Poisson models for the bivariate response (s, f), correlated through random effects. We extend our models to the analysis of longitudinal and multivariate longitudinal binomial outcomes. Our methodology was motivated by two disparate examples, one from teratology and one from an HIV tertiary intervention study.

Keywords

Logistic model; Longitudinal data; Multivariate discrete data; Poisson model; Random effects

1. Introduction

A logistic regression analysis with binomial outcome s_i for observation *i* conditions on the total number of trials $n_i = s_i + f_i$, where f_i is the number of failures. In many studies, n_i are not fixed by design but are random. In teratology studies on mice, researchers are interested in the proportion of fetuses with a given deformity (Paul, 1982; Machado et al., 2001). In public health studies on HIV transmission prevention, the proportion of protected sex acts, that is, sex acts using contraceptives, out of the number of total sex acts is a measure of the sexual risk behavior (Posner et al., 2001; Rotheram-Borus et al., 2001; Pinkerton, Chesson, and Layde, 2002; Foss et al., 2003; Rotheram-Borus et al., 2004). The total number of fetuses and sex acts, n_i , are random and not under the control of the investigator.

Choosing to model only successes or failures out of the total is no longer appropriate with random n_i . Teratology researchers expect the proportion of diabetic mice fetuses with a given deformity to increase as maternal mouse blood glucose levels increase. Analyzing the binomial outcome does not tell us if the increasing fraction of deformities is due to more deformities or fewer well-formed fetuses or a combination thereof. The effect of glucose levels on the number of viable fetuses may be uncertain, is also of interest, and is relevant to the proportion of viable fetuses.

^{*} scomulad@ucla.edu.

A high ratio of protected sex acts out of the total number of sex acts s_i/n_i and low number of unprotected sex acts f_i are both successful outcomes. High ratios s_i/n_i and low f_i might be expected to be correlated across the population. A person with $n_i = 0$ might, at a later observation, be anticipated to have high s_i/n_i , should they have $n_i > 0$ at that time. For example, low numbers of sex acts of any kind may be correlated with high fractions of protected acts. Both no sex acts and high fractions of protected acts are successes in HIV transmission prevention studies and the presence of no acts may indicate that a person is likely to have a high probability of protected acts, should any acts occur. This correlation is not included in the logistic regression of s_i successes out of $f_i + s_i$ trials, and the f_i are redundant in the logistic regression.

Several analytic techniques have been proposed to account for the random n_i . A logistic regression may include n_i as a covariate but does not allow s_i and f_i to be simultaneously modeled. Data points where $n_i = 0$ are still excluded, for example, observations are excluded for people who did not have sex. The multinomial-Poisson model (Terza and Wilson, 1990) models $n_i \sim \text{Poisson}(\lambda_i)$ and $s_i \mid n_i \sim \text{binomial}(n_i, \pi_i)$. Successes $s_i \sim \text{Poisson}(\lambda_i \pi_i)$ and failures $f_i \sim \text{Poisson} \{\lambda_i(1 - \pi_i)\}$ are modeled as independent given covariates. In our two examples, this independence seems a priori implausible. In the multinomial-Poisson model, observations with $n_i = 0$ contribute to the Poisson likelihood, but not to the binomial likelihood.

We recommend correlated bivariate Poisson models (Kocherlakota and Kocherlakota, 1992; Johnson, Kotz, and Balakrishnan, 1997) for the bivariate response (s_i, f_i) . We use random effects to model correlation between s_i and f_i . The bivariate Poisson model offers a number of advantages over analytic techniques that do not model s_i and f_i directly. The bivariate Poisson model may be easier to implement in standard statistical software packages than the multinomial-Poisson model; no transformation is needed to produce parameter estimates for outcomes s_i and f_i . The relationship between s_i and f_i is of interest and can be modeled. For models implemented in a Bayesian framework, regression coefficient and random effect priors are easier to formulate because they are on the same scale for both count responses, unlike for the multinomial-Poisson model.

The bivariate Poisson model is easily extended to longitudinal multivariate binomial responses that are prevalent in public health studies on HIV transmission prevention. For example, the proportion of protected sex acts and proportion of sex partners to whom HIV-positive status is disclosed may be assessed (Rotheram-Borus et al., 2001). A negative correlation between protection and disclosure may be hypothesized; people may feel less of an obligation to disclose their HIV-positive status if protection is used. Models for multivariate binomial responses (Lefkopoulou, Moore, and Ryan, 1989; McCullagh and Nelder, 1989) and longitudinal bivariate discrete responses (Rochon, 1996) capture correlation between proportions but do not model correlation between successes and failures within or between binomial responses. We will show in our example on HIV transmission risk data that the correlation of successes and failures provides useful information.

Notation for cross-sectional and longitudinal multivariate binomial data with random n is introduced in Section 2. We consider several covariance models for the random effects of the successes and failures in Section 3 and discuss model selection using Bayes factors in Section 4. The logistic model priors and posteriors induced by the Poisson model are discussed in Section 5. The Poisson models are applied to univariate binomial data from a teratology study and longitudinal bivariate binomial data from an HIV transmission risk study in Section 6. Discussion follows in Section 7.

2. Model Notation

A random-intercept logistic model (Anderson and Aitkin, 1985; Beitler and Landis, 1985; Wong and Mason, 1985; Zeger and Karim, 1991; Agresti, 2002) with observations on experimental units i = 1, ..., N, taken at time point *j*, with longitudinal *K*-variate binomial data $s_{ijk} \sim$ Binomial (n_{ijk}, π_{ijk}), k = 1, ..., K, with probability π_{ijk} of success has linear predictor

$$\log \frac{\pi_{ijk}}{1 - \pi_{ijk}} = x'_{ij} \gamma_k + \delta_{ik},\tag{1}$$

where γ_k is a vector of regression coefficients for the *k* th response and covariate vector x_{ij} is assumed the same for all responses *k* for simplicity and random effect vector $\delta_i = (\delta_{i1}, ..., \delta_{iK})' \sim N_K(0, \tau)$. The model and notation applies to cross-sectional data by dropping subscript *j*. We first consider univariate longitudinal binomial data, K = 1 dropping the subscript *k*. We jointly model $z_{ij1} = s_{ij}$ and $z_{ij2} = f_{ij}$, our bivariate response, with a bivariate longitudinal Poisson model

$$z_{ijh} \sim \text{Poisson}(\lambda_{ijh}),$$
 (2a)

with means $\lambda_{ijh} = E(z_{ijh})$, h = 1, 2. For longitudinal multivariate binomial data, z_{ijh} corresponds to successes for h = 2k - 1 and failures for h = 2k from the *k* th binomial observation, $h = 1, \dots, H = 2 \times K$. Using a log link, the linear predictors are

$$\log(\lambda_{ijh}) = x'_{ii}\alpha_h + \beta_{ih},\tag{2b}$$

where again for simplicity the covariate vector x_{ij} is assumed the same for all responses h and α_h is a vector of coefficients for x_{ij} for the hth outcome. We use random effects β_{ih} to induce correlation between z_{ij1} and z_{ij2} . For $H \ge 2$, the vector β_i of random effects for each person $\beta_i = (\beta_{i1}, ..., \beta_{iH})' \sim N_H(0, \Sigma)$ with elements β_{ih} where Σ is an $H \times H$ covariance matrix.

3. Covariance Models

We discuss covariance models for *H*-variate cross-sectional Poisson random effects. The most general covariance model has $\Sigma_{uns} = (\Sigma_{ab})$, a, b = 1, ..., H, to be the unstructured covariance matrix. Define $\sigma_a^2 = \sum_{aa} \operatorname{and} \rho_{ab} = \sum_{ab} \sigma_a^{-1} \sigma_b^{-1}$. Special cases of Σ_{uns} correspond to interesting models.

For H = 2, we consider five covariance matrices Σ_q , q = 1, ..., 5, nested in Σ_{uns} . Uncorrelated random effects with $\rho_{12} = 0$, result in diagonal Σ_1 . The random effects account for overdispersion but do not induce correlation between z_{ij1} and z_{ij2} . Covariance matrices Σ_2 and

 Σ_3 are special cases of Σ_{uns} and Σ_1 , respectively, with diagonal variances equal, $\sigma_1^2 = \sigma_2^2$. In some data sets the success and failure counts may share one random effect, $\beta_{i1} = \pm \beta_{i2}$ and singular Σ_4 (+) or Σ_5 (-) result. For longitudinal models we consider multivariate random-intercept models. Chen et al. (2003) discusses additional covariance models.

4. Comparing Covariance Models

We fit multivariate Poisson models to binomial data in a Bayesian framework and employ Bayesian model checking techniques to compare different covariance model fits to the data. Bayes factors (Jeffreys, 1961; Kass and Raftery, 1995) are used to test random effect covariance models.

We employ a Bayes factor B_{10} in favor of model M_1 against M_0 (Bernardo and Smith, 1994, p. 390; Raftery, 1995, p. 165) to compare models M_1 and M_0 . Let ω be a parameter or set of parameters from M_1 fixed at ω_0 in M_0 and let θ be the remaining model parameters. We assume prior densities under M_1 and M_0 satisfy $p(\theta \mid M_0) = p(\theta \mid \omega = \omega_0, M_1)$ and calculate B_{10} using the Savage–Dickey density ratio (Dickey, 1971; Verdinelli and Wasserman, 1995)

$$B_{10} = \frac{p(\omega)}{p(\omega|Y)} \bigg|_{\omega = \omega_0},\tag{3}$$

where $p(\omega)$ and $p(\omega \mid Y)$ are the prior and posterior densities of ω under M_1 evaluated at $\omega = \omega_0$ and Y is the observed data. Nonnested covariance models M_1 and M_2 are also compared using Bayes factors. If model M_2 can be compared to M_0 by B_{20} then M_2 can be compared to M_1 by a Bayes factor because $B_{21} = B_{20}/B_{10}$. Kernel density estimation as implemented in S-PLUS 6.0 (Venables and Ripley, 1999) was used to estimate $p(\omega)$ and $p(\omega \mid x)$.

For example, to test $M_0: \rho_{12} = 0$, draws are taken from the prior and posterior densities of the most general covariance matrix Σ_{uns} . Then $\beta_{uns,1}$ is calculated as the ratio of the prior density over the posterior density of correlation ρ_{12} estimated at $\rho_{12} = 0$ by kernel density estimate for both the prior and the posterior.

5. The Induced Logistic Model

5.1 Model Parameters

Although we model the counts, we maintain interest in the binomial probabilities π_{ik} . The logistic model parameters are functions of the Poisson model parameters. For *K*-variate binomial data, $\pi_{ik} = \lambda_{i(2k-1)}/{\{\lambda_{i(2k-1)} + \lambda_{i2k}\}}$. Substituting for λ_{ih} using equation (2b) gives

$$\pi_{ik} = \frac{1}{1 + \exp\left[-x_i' \left\{\alpha_{2k-1} - \alpha_{2k}\right\} - \left\{\beta_{i(2k-1)} - \beta_{i2k}\right\}\right]}.$$
(4)

The logistic coefficient vector is $\gamma_k = \alpha_{2k-1} - \alpha_{2k}$ and random effect is $\delta_{ik} = \beta_{i(2k-1)} - \beta_{i2k}$ with binomial random effect variance $\tau = \sigma_1^2 + \sigma_2^2 - 2\rho_{12}\sigma_1\sigma_2$ for K = 1 under $\Sigma_{\text{uns.}}$. For covariance matrix Σ_q , q = 1, 2, ..., 5, variance τ is $\sigma_1^2 + \sigma_2^2, 2\sigma_1^2(1 - \rho), 2\sigma_1^2, 0$, and $4\sigma_1^2$, respectively.

For K = 2, Σ_{uns} is a 4 × 4 matrix and $\tau = (\tau_{ab})$ is a 2 × 2 matrix with elements *a*, *b* = 1, 2, where $\tau_{11} = \sigma_1^2 + \sigma_2^2 - 2\sigma_{12}$, $\tau_{22} = \sigma_3^2 + \sigma_4^2 - 2\sigma_{34}$, and $\tau_{12} = \tau_{21} = \sigma_{13} + \sigma_{24} - \sigma_{14} - \sigma_{23}$. For $K \ge 2$ and other covariance models, covariance matrix properties may be utilized to determine τ .

5.2 Priors

We discuss priors for the *H*-variate Poisson model and the induced priors from the logistic regression coefficients. Priors for *p*-vector coefficient α_h and random effect vector β_i are set to

be $N_p(\mu_h, \eta I)$ and $N_H(0, \Sigma_{uns})$, respectively, where I is an identity matrix. The prior density for Σ_{uns} is inverse-Wishart, IW (*S*, *v*),

$$f(\sum_{\text{uns}}|S,\nu) = \left\{ 2^{\nu k/2} \pi^{k(k-1)/4} \prod_{i=1}^{k} \Gamma\left(\frac{\nu+1-i}{2}\right) \right\}^{-1} \\ \times |S|^{\nu/2} \left| \sum_{\text{uns}} \right|^{-(\nu+k+1)/2} \exp\left\{ -\frac{1}{2} \text{tr}\left(S \sum_{\text{uns}}^{-1}\right) \right\},$$

where tr (A) is the trace of a square matrix A, Γ is the gamma function, $S = (S_{ab})$, a, b = 1, ..., k, is a known symmetric, positive definite $k \times k$ scale matrix and degrees of freedom parameter v. We set S to be diagonal, implying a priori the random effects are equally likely to be positively or negatively correlated. Model Σ_1 has prior $P(\sigma_1^2, \sigma_2^2 | \rho_{12} = 0)$ with independent random effects and results in independent prior densities for $\sigma_1^2 \sim$ inverse-gamma(1.5, $S_{11}/2$) and $\sigma_2^2 \sim$ inverse-gamma(1.5, $S_{22}/2$).

In the logistic model, the induced priors for γ_k and δ_i are $N_p(\mu_{2k-1} - \mu_{k/2}, 2\eta I)$ and $N_K(0, \tau)$, respectively. We choose invertible $H \times H$ matrix *C* so τ is the upper left $K \times K$ submatrix of

$$C \sum_{\text{uns}} C' = \left\{ (C')^{-1} \sum_{\text{uns}}^{-1} C^{-1} \right\}^{-1} \sim \text{IW}(CSC', v).$$
(5)

By properties of the IW distribution τ is also distributed inverse-Wishart (Press, 1982, p. 118). For H = 2,

$$C = \left(\begin{array}{cc} 1 & -1\\ 1 & c_{22} \end{array}\right),\tag{6}$$

where $c_{22} \neq -1$ is an arbitrary real number. Then

$$\tau \sim \text{inverse-gamma}\left(\frac{v-1}{2}, \frac{V}{2}\right),$$
(7)

where $V = S_{11} + S_{22}$. For H = 4, $C = (r_1, r_2, r_3, r_4)'$ with row vectors $r_1 = (1, -1, 0, 0)$, $r_2 = (0, 0, 1, -1)$, and arbitrary row vectors r_3 and r_4 , linearly independent of r_1 and r_2 . Then

$$\tau \sim \mathrm{IW}(W, \nu),\tag{8}$$

where $W = (W_{ab})$, a, b = 1, 2, has elements $W_{11} = S_{11} + S_{22}$, $W_{22} = S_{33} + S_{44}$, and $W_{12} = W_{21} = 0$.

6. Examples

We consider teratology data containing a cross-sectional univariate binomial outcome and data from the Healthy Living Project (HLP), a public health study on HIV transmission behaviors containing a longitudinal bivariate binomial outcome. Multivariate Poisson models with

random effects covariance matrix Σ_{uns} are fit to the data. We analyze data sets containing all observations, referred to as complete data sets, and data sets excluding observations whose success and failure counts sum to zero, referred to as nonzero subsets, for closer comparison with logistic models.

Analyses are conducted in WinBUGS version 1.41 software (Spiegelhalter, Thomas, and Best, 2000) and based on a 10³ iteration burn-in followed by 10⁶ iterations. Inspection of time series and lagged autocorrelation plots showed the number of iterations to be satisfactory. Posterior means (M) and standard deviations (SDs) of the parameters are reported in Tables 1 and 2.

We plot posterior densities for probability π_x given covariate vector x as approximated by the S-PLUS 6.0 kernel density estimator. Comparisons are made between π_x estimated from logistic regression models and indirectly estimated from bivariate Poisson regression models

as discussed in Section 5. We use a = 1, ..., A draws $\alpha_h^{(a)}$ and $\gamma^{(a)}$ from the joint posterior of the parameter estimates, with A = 1000; every hundredth iteration out of the total 10^6 iterations was used. The $\delta^{(a)}$ are sampled from density $N(0, \tau^{(a)})$. The estimates are

$$\widehat{\pi}_{x} = \frac{1}{A} \sum_{a=1}^{A} \left[1 + \exp\left\{ -x'\gamma^{(a)} - \delta^{(a)} \right\} \right]^{-1}.$$
(9)

6.1 Teratology Data

The teratology data was originally reported on in Machado et al. (2001). It was collected to measure the effects of diabetes in 68 pregnant female C57BL/6J mutant mice on malformations in their litters. Nonzero subset analyses exclude six females with no live births. The binomial outcome is the fraction of live births with exencephaly. We split the binomial n_i into a bivariate count outcome, the number of viable fetuses with and without exencephaly from each litter. Diabetes was chemically induced in the female parents prior to pregnancy and assessed by blood glucose levels in milligrams per deciliter. Some parental mice contained a splotch allele thought to confer added susceptibility to malformations in their offspring. Covariates from the original analysis were used: an intercept; maternal blood glucose level (Glucose level); and a three-category splotch allele indicator, categorized as whether the father had the allele (Father), the mother had the allele (Mother), or neither parent had the allele. The experimental design excluded both parents having the allele. If both parents had the allele a fetus could not form. The linear predictor is parameterized as

$$\log(\lambda_{ih}) = \alpha_{h1} + \alpha_{h2} \text{Glucose level}_i + \alpha_{h3} \text{Father}_i + \alpha_4 \text{Mother}_i + \beta_{ih}.$$
(10)

6.1.1 Data priors—Priors on α_1 and α_2 are set to be N_4 (0, 10I), proper but basically uninformative. The prior on Σ is IW(*S*, *v*), with degrees of freedom, v = 4 and H = 2. We specify a diagonal scale matrix *S* so the expectation $E(\Sigma) = (v - H - 1)^{-1} S = S$ is a reasonable estimate of the random effect variances.

The mean litter size for the C57BL/6J mouse is listed as 7 on the Laboratory Animal Science Center section of the Boston University Medical campus web site, http://www.bumc.bu.edu. We use this to develop a prior for the random effects variances. We start with a guess that half of the litter is malformed so our prior point estimate of the mean litter size of malformed and nonmalformed fetuses is 3.5. We take 0.001 and 7 as a lower and upper 0.005 and 0.995

probability bounds on the mean for an individual litter. We want the 0.005 and 0.995 percentiles of the random effects distribution on the log scale to give us end points of $\log(0.001) = -6.9$ and $\log(7) = 1.17$. Six SDs span 99% of a normal density. The SD of the random effects is approximately $\{1.9 - (-6.9)\}/6 \approx 1.5$. The random effects variance is $1.5^2 \approx 2$ and we set $S_{11} = S_{22} = 2$.

Priors on γ and δ_i are then N_4 (0, 20I) and $N(0, \tau)$, respectively, and the prior on τ is inverse-gamma(1.5, 1).

6.1.2 Results—Parameter estimates from random-intercept bivariate Poisson and univariate logistic models fit to the cross-sectional teratology data set are shown in Table 1. A higher proportion of malformed births with exencephaly is associated with increased blood glucose levels in the logistic model. The Poisson model shows the association with blood glucose is driven by a negative association between nonmalformed births is not seen. The association between the number of malformed births and blood glucose levels is close to being significant in the nonzero subset (M = 0.0022 and SD = 0.0012), showing that different conclusions might well be drawn from analyses on the complete data and on the nonzero subset.

For the nonzero subset, we estimate Bayes factors $\beta_{uns,1}$, comparing covariance structures Σ_{uns} to Σ_1 , to be 0.47 and similarly $\beta_{uns,2} = 0.60$. The first Bayes factor favors independent random effects, $\rho_{12} = 0$, and $\beta_{uns,2}$ favors equal random effect variances, $\sigma_1^2 = \sigma_2^2$.

6.2 HLP Data

We use data with n = 175 from the Choosing Life: Empowerment, Action, Results study (CLEAR; Rotheram-Borus et al., 2004) to create priors for models fit to HLP data (n = 936; Gore-Felton et al., 2005; Lightfoot et al., 2005). Nonzero subset analyses (n = 852) exclude observations for participants who were sexually inactive during the previous time period; 20%, 794 out of 3918, observations were excluded. The HLP study was modeled on the CLEAR study and bears many similarities. Both studies were designed to reduce HIV-transmission behaviors and improve the quality of life for persons living with HIV. Across studies, participants shared a similar socioeconomic status and were recruited from similar types of social service agencies in several metropolitan areas from 1999 to 2000 in the CLEAR study and from 2000 to 2002 in the HLP study. Sexual behavior, substance use, and quality-of-life measures were assessed at baseline, 3, 6, 9, and 15 months in the CLEAR study and at baseline and 5-month intervals up to 25 months in the HLP study. At baseline, participants in both studies were randomized to an immediate or delayed intervention condition.

There are two main differences across studies. The CLEAR sample is younger, ages 16 to 29 with a median age of 23, compared to the HLP sample, ages 19 to 67 with a median age of 40. Recruitment was based on different HIV-transmission behaviors. Current drug use and sexual activity were required to be recruited into the CLEAR and HLP studies, respectively. Study details may be found on the CHIPTS web site (http://chipts.ucla.edu/projects/chipts).

The bivariate binomial outcome, the proportion of sex partners to whom HIV-positive status is disclosed and the proportion of protected sex acts, is split into a multivariate Poisson outcome for the number of disclosed and undisclosed sex partners and number of protected and unprotected sex acts. We fit multivariate longitudinal Poisson random effects models. Relevant covariates based on analyses from prior studies include time, in units of 10 weeks (Wks); intervention arm (Intv), categorized as 1 = immediate or 0 = delayed; age at baseline (Age); and a four-category HIV risk indicator established by the Centers for Disease Control (CDC, 2001), categorized as men who are injecting drug users (IDU), non-IDU who are having sex with other men (MSM), non-IDU men not having sex with other men (HTM), and women.

Visual inspection of the HLP data suggested a piecewise regression allowing different trajectories for each intervention arm from baseline to 5 months and from 5 months to 25 months. Indicators for baseline (Base) and follow-up (Follow = 1 - Base) allow the trajectories to be modeled. Quadratic terms for time are included to model an observed curvilinear trend from 5 to 25 months. The linear predictor is parameterized as

$$\log(\lambda_{ijh}) = \alpha_{h1} \operatorname{Age}_{i} + \alpha_{h2} \operatorname{MSM}_{i} + \alpha_{h3} \operatorname{IDU}_{i} + \alpha_{h4} \operatorname{HTM}_{i} \\ + \alpha_{h5} \operatorname{Base}_{ij} + \alpha_{h6} \operatorname{Base} * \operatorname{Intv}_{ij} + \alpha_{h7} Follow_{ij} \\ + \alpha_{h8} Follow * \operatorname{Wks}_{ij} + \alpha_{h9} Follow * \operatorname{Wks}_{ij}^{2} \\ + \alpha_{h10} Follow * \operatorname{Intv}_{ij} + \alpha_{h11} Follow * \operatorname{Intv} * \operatorname{Wks}_{ij} \\ + \alpha_{h12} Follow * \operatorname{Intv} * \operatorname{Wks}_{ij}^{2} + \beta_{ih}.$$
(11)

We plot estimated mean counts λ_{jh} for outcome *h* at time point *j* in Figure 1. Outcomes are modeled on the logarithmic scale and transformed back to the original scale prior to plotting. The transformed expectation is of course not the expectation on the transformed scale for nonlinear transformations. A parametric version of the smearing estimate (Duan, 1983) is used

to calculate the mean on the transformed scale. We use a = 1, ..., A draws $\alpha_h^{(a)}$ and $\beta_{ih}^{(a)}$ from the joint posterior of the parameter estimates and random effects, with A = 100; every tenthousandth iteration out of the total 10^6 iterations was used. The estimate is

$$\widehat{\lambda}_{jh} = \frac{1}{AN} \sum_{a=1}^{A} \sum_{i=1}^{N} \exp\left\{x_{ij}^{\prime} \alpha_{h}^{(a)} + \beta_{ih}^{(a)}\right\}.$$
(12)

6.2.1 Data priors—Prior distribution parameter values for HLP data analyses were obtained from the CLEAR data set. Separate random-intercept Poisson regression models were fit to each outcome in SAS software, version 8.2, using the GLIMMIX macro (Wolfinger and O'Connell, 1993; Littell et al., 1996) using a pseudolikelihood approach. For the *h*th outcome and *p*th coefficient a $N(\mu_{hp}, \omega_{hp})$ prior was used with μ_{hp} and ω_{hp} set to the coefficient and coefficient variance estimate from the CLEAR data analyses.

The prior on Σ was set to be IW (*S*, *v*), with *v* = 6. The diagonal scale matrix, *S*, was specified so the expectation $E(\Sigma) = (v - H - 1)^{-1} S = S$ had diagonal variance elements corresponding to random effect variance estimates from the CLEAR data analysis.

Inferred priors on γ_k and δ_{ik} are $N_p(\Theta_k, \Lambda_k)$ and $N_K(0, \tau)$, respectively, where Θ_k is a vector with elements $\Theta_{kp} = \mu_{2k-1,p} - \mu_{2k,p}$ and diagonal covariance matrix Λ_k has diagonal elements $\Lambda_{kpp} = \omega_{2k-1,pp} + \omega_{2k,pp}$. The prior on τ is inferred to be IW(*S*, ν), where diagonal scale matrix *S* has elements $S_{11} = 5.49$ and $S_{22} = 4.47$, calculated from CLEAR random effect variance estimates of 0.99, 1.53, 0.74, and 1.50 for the number of partners disclosed to, partners not disclosed to, protected acts, and unprotected acts, respectively.

6.2.2 Analysis results—We fit random-intercept multivariate Poisson and bivariate logistic models to the bivariate binomial outcome for the fraction of protected sex acts and disclosure. Parameter estimates are similar to parameter estimates from random-intercept bivariate Poisson and univariate logistic models fit to each binomial outcome separately. This is not surprising considering the low correlation of -0.085, 95% probability interval = (-0.17, 0.0068), between random effects in the bivariate binomial model, indicating binomial outcomes are nearly independent. We present parameter estimates for one of the binomial outcomes, the fraction of protected sex acts, in Table 2.

The proportion of protected sex acts increases over time more rapidly at first in the immediate condition, compared to the delayed condition. In Figure 1, the Poisson model shows the initial increase in the logistic model to be driven by an initial decrease in the number of unprotected acts in the immediate condition, compared to the delayed condition. The number of protected acts follow a similar trajectory across intervention conditions; inclusion of the zero counts eliminates the level shift in trajectories between intervention conditions at follow-up time points. Inclusion of the zero counts also reverses the direction of the correlation between random effects.

For the nonzero subset, we estimate Bayes factors $\beta_{\text{uns},1}$ to be 1.23, favoring independent random effects, $\rho_{12} = 0$, and $\beta_{\text{uns},2}$ to be 105.96, favoring unequal random effect variances, $\sigma_1^2 \neq \sigma_2^2$.

Approximate posterior densities for teratology and HLP outcome probabilities under the logistic and bivariate Poisson models are shown in Figures 2 and 3, respectively. The density shapes from the cross-sectional teratology data are quite similar, whereas the bivariate Poisson densities resulting from the longitudinal HLP data are much narrower compared to the logistic densities. In the HLP data, the bivariate Poisson model produces much smaller confidence intervals compared to the logistic model.

7. Discussion

Our analyses highlight the importance of capturing the complex nature of binomial data with random numbers of trials through the multivariate Poisson model. In both of our examples, the expansion of the binomial outcome to the bivariate Poisson model allowed us to examine the relationship between successes and failures and key covariates, giving a more detailed picture of the data. Biologists may be interested to examine whether teratogenic agents lead to different rates of malformed and normal births. Interventions targeting HIV-transmission behaviors may be very different, depending on whether the goal is to decrease unprotected acts or increase the proportion of protected acts.

More complex random effect models are possible. We could replace the random effects in equations (1) and (2b) with a more complicated random effects structure. For example, we could have a random intercept and slope instead of a random intercept, or more complex random effects. Alternatively, we could induce an autoregressive structure on the random effects over time, so that within-subject random effects were not constant over time (refer to Chen et al., 2003). Another possible covariance model for time-varying random effects is the product correlation model (Weiss, 2005, Chapter 13). For both the autoregressive and product correlation models, there would be additional Poisson variability in addition to the random effect variance.

Acknowledgments

The authors thank Michael Collins, Ph.D., for permission to use the teratology data set and Mary Jane Rotheram-Borus, Ph.D., for permission to use the CLEAR and HLP HIV transmission risk data sets in our analyses. The CLEAR data was collected with the support of NIDA grant DA07903 and the HLP data was collected with the support of NIMH grant U10MH057615. REW was partially supported by NIMH grant 1 R01 MH60213 and by the Center for HIV Identification, Prevention and Treatment Services (CHIPTS) within the Center for Community Health (P30MH-58107). We thank the referees and editors for helpful comments.

References

Agresti, A. Categorical Data Analysis. 2nd. New York: Wiley; 2002.

- Anderson DA, Aitkin M. Variance component models with binary response: Interviewer variability. Journal of the Royal Statistical Society 1985;47:203–210.B
- Beitler PJ, Landis JR. A mixed-effects model for categorical data. Biometrics 1985;41:991–1000. [PubMed: 3830263]
- Bernardo, JM.; Smith, AFM. Bayesian Theory. Chichester: Wiley; 1994.
- CDC. HIV/AIDS Surveillance Report 13. 2001.
- Chen MH, Ibrahim JG, Shao QM, Weiss RE. Prior elicitation for model selection and estimation in generalized linear mixed models. Journal of Statistical Planning and Inference 2003;111:57–76.
- Dickey J. The weighted likelihood ratio, linear hypotheses on normal location parameters. The Annals of Statistics 1971;42:204–223.
- Duan N. Smearing estimate: A nonparametric retransformation method. Journal of the American Statistical Association 1983;78:605–610.
- Foss AM, Vickerman PT, Heise L, Watts CH. Shifts in condom use following microbicide introduction: Should we be concerned? AIDS 2003;17:1227–1237. [PubMed: 12819525]
- Gore-Felton C, Rotheram-Borus MJ, Weinhardt LS, Kelly JA, Lightfoot M, Kirshenbaum SB, Johnson MO, Chesney MA, Catz SL, Ehrhardt AA, Remien RH, Morin SF, The NIMH Healthy Living Project Team. The Healthy Living Project: An individually tailored, multidimensional intervention for HIVinfected persons. AIDS Education and Prevention 2005;17:21–39. [PubMed: 15843115]
- Jeffreys, H. Theory of Probability. 3rd. Oxford: Oxford University Press; 1961.
- Johnson, N.; Kotz, S.; Balakrishnan, N. Discrete Multivariate Distributions. New York: Wiley; 1997.
- Kass RE, Raftery AE. Bayes factor and model uncertainty. Journal of the American Statistical Association 1995;90:773–795.
- Kocherlakota, S.; Kocherlakota, K. Bivariate Discrete Distributions. New York: Marcel Dekker; 1992.
- Lefkopoulou M, Moore D, Ryan L. The analysis of multiple correlated binary outcomes; applications to rodent teratology experiments. Journal of the American Statistical Association 1989;84:810–815.
- Lightfoot M, Rogers T, Goldstein R, Rotheram-Borus MJ, May S, Kirshenbaum S, Weinhardt L, Zadoretzky C, Kittel L, Johnson M, Gore-Felton C, Morin SF. Predictors of substance use frequency and reductions in seriousness of use among persons living with HIV. Drug and Alcohol Dependency 2005;77:129–138.
- Littell, RC.; Milliken, GA.; Stroup, WW.; Wolfinger, RD. SAS System for Mixed Models. Cary, North Carolina: SAS Institute; 1996.
- Machado AF, Zimmerman EF, Hovland DN Jr, Weiss RE, Collins MD. Diabetic embryopathy in C57BL/ 6J mice: Altered fetal sex ratio and impact of the splotch allele. Diabetes 2001;50:1193–1199. [PubMed: 11334426]
- McCullagh, P.; Nelder, JA. Generalized Linear Models. 2nd. London: Chapman and Hall; 1989.
- Paul SR. Analysis of proportions of affected foetuses in teratological experiments. Biometrics 1982;38:361–370. [PubMed: 7115867]
- Pinkerton SD, Chesson HW, Layde PM. Utility of behavioral changes as markers of sexually transmitted disease risk reduction in sexually transmitted disease/HIV prevention trials. Journal of Acquired Immune Deficiency Syndromes 2002;31:71–79. [PubMed: 12352153]
- Posner SF, Pulley L, Artz L, Cabral R, Macaluso M. Psychosocial factors associated with self-reported male condom use among women attending public health clinics. Sexually Transmitted Diseases 2001;28:387–393. [PubMed: 11460022]
- Press, JS. Applied Multivariate Analysis: Using Bayesian and Frequentist Methods of Inference. Melbourne: Krieger Publishing Co.; 1982.
- Raftery, AE. Hypothesis testing and model selection. In: Gilks, WR.; Richardson, S.; Spiegelhalter, DJ., editors. Markov Chain Monte Carlo in Practice. London: Chapman and Hall/CRC; 1995.
- Rochon J. Analyzing bivariate repeated measures for discrete and continuous outcome variables. Biometrics 1996;52:740–750. [PubMed: 8672710]
- Rotheram-Borus MJ, Lee MB, Murphy D, Futterman D, Duan N, Birnbaum J, Lightfoot M. Efficacy of a preventative intervention for youths living with HIV. American Journal of Public Health 2001;91:400–405. [PubMed: 11236404]

- Rotheram-Borus MJ, Swendeman D, Comulada WS, Weiss RE, Lee M, Lightfoot M. Prevention for substance using HIV positive young people: Telephone and in-person delivery. Journal of Acquired Immune Deficiency Syndromes 2004;37:S68–S77. [PubMed: 15385902]
- Spiegelhalter, DJ.; Thomas, A.; Best, NG. WinBUGS Version 1.4 User Manual. MRC Biostatistics Unit; 2000.
- Terza JV, Wilson PW. Analyzing frequencies of several types of events: A mixed multinomial-poisson approach. The Review of Economics and Statistics 1990;72:108–115.
- Venables, WN.; Ripley, BD. Modern Applied Statistics with SPLUS. New York: Springer; 1999.
- Verdinelli I, Wasserman L. Computing Bayes factors using a generalization of the Savage-Dickey density ratio. Journal of the American Statistical Association 1995;90:614–618.
- Weiss, RE. Modeling Longitudinal Data. New York: Springer-Verlag; 2005.
- Wolfinger R, O'Connell M. Generalized linear mixed models: A pseudo-likelihood approach. Journal of Statistical Computation and Simulation 1993;48:233–243.
- Wong G, Mason W. The hierarchical logistic regression model for multilevel analysis. Journal of the American Statistical Association 1985;80:513–524.
- Zeger SL, Karim MR. Generalized linear models with random effects: A Gibbs sampling approach. Journal of the American Statistical Association 1991;86:79–86.



Figure 1.

Estimated mean outcomes over time by intervention condition from longitudinal bivariate Poisson model fit to HLP (a) complete data set and (b) nonzero subset.



Figure 2.

Approximate marginal posterior density of π , the probability of exencephaly, from bivariate Poisson (- -) and logistic (—) regressions fit to teratology nonzero subset. Densities are shown for 25th and 75th percentiles of sample blood glucose levels.



Figure 3.

Approximate marginal posterior density of $\pi_{.j}$, the probability of protected sex, from longitudinal bivariate Poisson (- -) and longitudinal logistic (—) regressions fit to HLP nonzero subset. Densities are shown for each intervention arm at three time points.

_
_
_
_
_
_
· U
~
-
~
_
_
_
_
\sim
0
_
_
_
<
\sim
^
L L
=
_
-
~
10
0
0
U
-
- ` .
_
U
<u> </u>

Table 1

Posterior means (M) and SDs from bivariate Poisson and univariate logistic models fit to teratology data. Reference splotch allele group has neither parent with splotch allele

Comulada and Weiss

		Bivar	iate Poisson model			Lo	gistic model	
	Complete data			Nonzero subset		Induced		Estimated
Part of model	M	SD	W	SD	¥	SD	W	SD
With exencephaly								
Intercept	-1.57	0.89	-2.15	0.96	-4.69	1.08	-5.20	1.21
Blood glucose level	0.0012	0.0011	0.0022	0.0012	0.0041	0.0014	0.0046	0.0016
Splotch allele group								
Father	0.33	0.43	0.41	0.43	0.39	0.48	0.53	0.52
Mother	0.56	0.51	0.53	0.50	0.73	0.58	0.82	0.62
Without exencephaly								
Intercept	2.78	0.46	2.53	0.45				
Glucose level	-0.0024	0.00064	-0.0019	0.00063				
Splotch allele group								
Father	-0.027	0.21	0.013	0.21				
Mother	-0.21	0.29	-0.20	0.28				
Random effect variances								
With exencephaly	0.64	0.31	0.53	0.25	0.81	0.34	0.87	0.48
Without exencephaly	0.24	0.082	0.20	0.063				
Correlation	0.040	0.25	-0.12	0.24				

_
_
~
_
_
<u> </u>
-
~
_
<u> </u>
-
<u> </u>
-
\mathbf{O}
<u> </u>
_
_
<
\sim
01
D
_
_
<u> </u>
CD
0
~
— .
0
<u> </u>

da and V	Weiss									
	Stimated	SD	0.36	0.15	0.36	0.026	0.0019	0.20	0.052	0.0039
Binomial model	H	M	-0.023	0.87	0.73	-0.15	0.014	0.053	0.23	-0.018
Univariate	ıduced	SD	0.26	0.12	0.098	0.018	0.0013	0.16	0.036	0.0027
	Ц									3

Table 2

Posterior means (M) and SDs of parameters from longitudinal bivariate Poisson and univariate logistic models fit to HLP data. Models adjust for age at baseline and risk group

Bivariate Poisson model

0.290.0043 -0.024-0.008 0.660.620.520.13 0.43 5.03Σ 0.000810.0010 0.0016 Nonzero subset 0.0021 0.014 0.0290.0770.011 0.10 0.022 0.12 0.0910.21 0.21 0.21 0.170.11 0.20ß -0.0480.0057 0.00540.0097 -0.13-0.11-0.25 -0.180.014 0.27 -0.211.602.16 0.452.22 1.741.862.74 Σ 0.00076 0.0020 0.0015 0.0097 0.010 0.0200.014 0.027 0.0980.140.250.250.11 0.12 0.290.12 0.280.21 S **Complete data** 0.000460.000400.0073 -0.061-0.0690.0033 0.033 0.011 0.064-0.11 2.22 1.692.54 0.37 0.31 3.38 2.25 1.82 Σ Random effect variances Follow * Intv * Wks² Follow * Intv * Wks² Follow * Intv * Wks Follow * Intv * Wks Unprotected acts $Follow * Wks^2$ $Follow * Wks^2$ Follow * Wks Follow * Intv Unprotected acts Follow * Wks Protected acts Follow * Intv Part of model Base * Intv Base * Intv Protected acts Follow Follow Base Base

Biometrics. Author manuscript; available in PMC 2010 March 22.

Page 16

0.41

6.28

0.038

-0.095

0.037

0.13

Correlation