



Published in final edited form as:

AIDS. 2009 July 17; 23(11): 1449–1451. doi:10.1097/QAD.0b013e32832dbf91.

## Disruption of Env Tyrosine-Dependent Sorting Signal Does Not Affect Susceptibility of HIV-1 to Cytotoxic T Lymphocytes

Justin De La Cruz<sup>1</sup>, Ayub Ali<sup>2</sup>, Hwee L. Ng<sup>2</sup>, and Otto O. Yang<sup>\*,1,2,3</sup>

<sup>1</sup>Department of Microbiology, Immunology, and Molecular Genetics, David Geffen School of Medicine, University of California, Los Angeles, CA

<sup>2</sup>Department of Medicine, David Geffen School of Medicine, University of California, Los Angeles, CA

<sup>3</sup>UCLA AIDS Institute, University of California, Los Angeles, CA

### Keywords

HIV; HIV Envelope Protein gp41; AIDS Vaccines; Cytotoxic T Lymphocytes; SIV

Infections with live-attenuated viruses within SIV models of HIV-1 pathogenesis have offered the best evidence that a vaccine could generate protective immunity. Desrosiers and colleagues [1] first demonstrated this principle using *nef*-defective SIV, which established a chronic low-grade asymptomatic infection in macaques associated with protection against subsequent challenge with wild-type SIV. At least in part, this phenomenon appeared to be due to attenuation of viral replication.

More recently, Shacklett *et al* [2] and Hoxie and *et al* [3] have examined attenuated SIV infection via mutations in the transmembrane domain of gp41, which also subsequently protects from challenge by wild-type SIV. Shacklett *et al* found that SIV containing multiple (stop and point) mutations disrupting this domain reduced viral replicative capacity *in vitro*, likely due to effects on gp41 membrane trafficking in infected cells [4,5]. Hoxie and colleagues more specifically created mutations in the tyrosine-based sorting motif (Y712xx $\phi$ ) in the membrane-proximal cytoplasmic domain of gp41 [3]. Despite the role this motif in gp41 trafficking [4,5], the mutations appeared to have minimal impact on viral fitness, reflected by normal peak viremia during acute infection *in vivo* and growth kinetics *in vitro*. Thus the mechanism of attenuation and subsequent immune protection is unclear, but probably not due to markedly reduced viral replication capacity. Interestingly, *in vivo* CD8 depletion experiments have suggested that CD8<sup>+</sup> T lymphocytes (CTLs) may contribute to the protective immunity [6].

To assess whether analogous mutations in HIV-1 might affect viral susceptibility to CTLs, mutations in the Y712xx $\phi$  motif were constructed in HIV-1 NL4-3 [7] by point mutagenesis (QuikChange, Stratagene). These included EnvY712I, EnvY712S and Env $\Delta$ GY mutations in the cytoplasmic domain of gp41 (Figure 1A), engineered into the whole genome context of NL4-3.1 containing the clade B consensus sequence at Gag 77-85 (HXB2 a.a. numbering) [8]. These viruses were examined for their ability to replicate in T1 cells [9] (Figure 1B). The EnvY712I and EnvY712S mutants had growth kinetics similar to wild-type NL4-3 (EnvY712),

\*Corresponding author: Otto O. Yang, BSRB 163, 615 Charles E. Young Drive South, Los Angeles, CA, 90095, USA, oyang@mednet.ucla.edu, Phone: 310-794-9491, Fax: 310-825-3632.

similar to SIVmac239 [3]. In contrast, the Env $\Delta$ GY mutant was replication incompetent, suggesting that this mutation more severely impaired HIV-1 than SIV.

To assess the susceptibility of these mutants to CTLs, these viruses were tested in virus suppression assays [10,11] using HIV-1-specific CTL clones: S1-SL9-3.23T recognizing the HLA A\*02-restricted epitope SLYNTVATL in Gag p17 (a.a. 77-85) [12], S31-IV9-10.11T recognizing the HLA A\*02-restricted epitope ILKEPVHGV in RT (a.a. 309-317), and S58-PL10-10.8 recognizing the HLA A\*02-restricted epitope PLTFGWYCYKL in Nef (a.a. 136-145). The HIV-1-permissive target cells were A\*02-expressing T1 cells [10]. Comparisons of these viruses showed similar degrees of suppression of the mutant and wild-type viruses by the three CTL clones (Figure 1C and D). Similar results were noted in independent experiments using two other CTL clones recognizing other epitopes (not shown). Overall, these results suggested that directly increased susceptibility to CTL inhibition is not the mechanism of *in vivo* attenuation of infection with viruses containing these mutations.

Because attenuated SIV infection has been the most robust example of protective vaccination in the SIV-macaque model [1,6,13–15], understanding how viral attenuation affects antiviral immunity is clearly an important goal. The observation that SIV mutated in the Y712xx $\phi$  motif yields infections with typical high peak viremia followed by chronic low viremia in macaques subsequently protected from wild-type virus challenge [2,3,6] suggests that viral replication is not markedly affected by the mutations, but that replication is suppressed during chronic infection after development of CTL responses, which are the major determinant of set-point viremia [16–18]. Furthermore, preliminary data suggests that the low set-point viremia of macaques infected with Y712xx $\phi$ -disrupted SIV may be related to the CTL response [6]. Thus, a simple explanation could be that disruption of this motif somehow renders SIV directly more susceptible to CTL.

However, our results suggest that this is not the case for HIV-1; disruption of the Y712xx $\phi$  motif did not appear to increase susceptibility of HIV-1 to CTLs directly. While our *in vitro* assay may not necessarily predict the interaction of virus and CTLs *in vivo*, it seems likely that the mechanism of attenuation is either not mediated by CTLs, or indirectly increases the antiviral activity of CTLs. Interestingly, it has been suggested that the HIV-1 Env may facilitate viral escape from CTLs in lymph nodes [19,20]. Although our data do not address this issue directly, they are compatible with a mechanism whereby reduced levels of Env could reduce viral escape from CTLs. Further work would be required to explore this possibility.

## Acknowledgments

Funded by NIH grant AI043203 (O.O.Y.)

## REFERENCES

1. Daniel MD, Kirchhoff F, Czajak SC, Sehgal PK, Desrosiers RC. Protective effects of a live attenuated SIV vaccine with a deletion in the nef gene. *Science* 1992;258:1938–1941. [PubMed: 1470917]
2. Shacklett BL, Weber CJ, Shaw KE, Keddie EM, Gardner MB, Sonigo P, Luciw PA. The intracytoplasmic domain of the Env transmembrane protein is a locus for attenuation of simian immunodeficiency virus SIVmac in rhesus macaques. *J Virol* 2000;74:5836–5844. [PubMed: 10846063]
3. Fultz PN, Vance PJ, Endres MJ, Tao B, Dvorin JD, Davis IC, et al. In vivo attenuation of simian immunodeficiency virus by disruption of a tyrosine-dependent sorting signal in the envelope glycoprotein cytoplasmic tail. *J Virol* 2001;75:278–291. [PubMed: 11119598]
4. Berlioz-Torrent C, Shacklett BL, Erdtmann L, Delamarre L, Bouchaert I, Sonigo P, et al. Interactions of the cytoplasmic domains of human and simian retroviral transmembrane proteins with components

of the clathrin adaptor complexes modulate intracellular and cell surface expression of envelope glycoprotein. *J Virol* 1999;73:1350–1361. [PubMed: 9882340]

5. Blot G, Janvier K, Le Panse S, Benarous R, Berlioz-Torrent C. Targeting of the Human Immunodeficiency Virus Type 1 Envelope to the trans-Golgi Network through Binding to TIP47 Is Required for Env Incorporation into Virions and Infectivity. *J. Virol* 2003;77:6931–6945. [PubMed: 12768012]
6. Hoxie J. Attenuated SIV Models and Protection from Pathogenic Heterologous Challenges. Conference on Retroviruses and Opportunistic Infections. 2008
7. Ali A, Jamieson BD, Yang OO. Half-genome human immunodeficiency virus type 1 constructs for rapid production of reporter viruses. *J Virol Methods* 2003;110:137–142. [PubMed: 12798240]
8. Yang OO, Sarkis PT, Ali A, Harlow JD, Brander C, Kalams SA, Walker BD. Determinant of HIV-1 mutational escape from cytotoxic T lymphocytes. *J Exp Med* 2003;197:1365–1375. [PubMed: 12743169]
9. Salter RD, Howell DN, Cresswell P. Genes regulating HLA class I antigen expression in T-B lymphoblast hybrids. *Immunogenetics* 1985;21:235–246. [PubMed: 3872841]
10. Yang OO, Kalams SA, Trocha A, Cao H, Luster A, Johnson RP, Walker BD. Suppression of human immunodeficiency virus type 1 replication by CD8+ cells: evidence for HLA class I-restricted triggering of cytolytic and noncytolytic mechanisms. *J Virol* 1997;71:3120–3128. [PubMed: 9060675]
11. Bennett MS, Ng HL, Ali A, Yang OO. Cross-clade detection of HIV-1-specific cytotoxic T lymphocytes does not reflect cross-clade antiviral activity. *J Infect Dis* 2008;197:390–397. [PubMed: 18184090]
12. Adnan S, Balamurugan A, Trocha A, Bennett MS, Ng HL, Ali A, et al. Nef interference with HIV-1-specific CTL antiviral activity is epitope specific. *Blood* 2006;108:3414–3419. [PubMed: 16882705]
13. Johnson RP, Desrosiers RC. Protective immunity induced by live attenuated simian immunodeficiency virus. *Curr Opin Immunol* 1998;10:436–443. [PubMed: 9722920]
14. Lohman BL, McChesney MB, Miller CJ, McGowan E, Joye SM, Van Rompay KK, et al. A partially attenuated simian immunodeficiency virus induces host immunity that correlates with resistance to pathogenic virus challenge. *J Virol* 1994;68:7021–7029. [PubMed: 7933084]
15. Wyand MS, Manson K, Montefiori DC, Lifson JD, Johnson RP, Desrosiers RC. Protection by live, attenuated simian immunodeficiency virus against heterologous challenge. *J Virol* 1999;73:8356–8363. [PubMed: 10482586]
16. Matano T, Shibata R, Siemon C, Connors M, Lane HC, Martin MA. Administration of an anti-CD8 monoclonal antibody interferes with the clearance of chimeric simian/human immunodeficiency virus during primary infections of rhesus macaques. *J Virol* 1998;72:164–169. [PubMed: 9420212]
17. Jin X, Bauer DE, Tuttleton SE, Lewin S, Gettie A, Blanchard J, et al. Dramatic rise in plasma viremia after CD8(+) T cell depletion in simian immunodeficiency virus-infected macaques. *J Exp Med* 1999;189:991–998. [PubMed: 10075982]
18. Schmitz JE, Kuroda MJ, Santra S, Sasseville VG, Simon MA, Lifton MA, et al. Control of viremia in simian immunodeficiency virus infection by CD8+ lymphocytes. *Science* 1999;283:857–860. [PubMed: 9933172]
19. Stevceva L, Yoon V, Anastasiades D, Poznansky MC. Immune responses to HIV Gp120 that facilitate viral escape. *Curr HIV Res* 2007;5:47–54. [PubMed: 17266556]
20. Stevceva L, Yoon V, Carville A, Pacheco B, Santosuosso M, Koriath-Schmitz B, et al. The efficacy of T cell-mediated immune responses is reduced by the envelope protein of the chimeric HIV-1/SIV-KB9 virus in vivo. *J Immunol* 2008;181:5510–5521. [PubMed: 18832708]

**A. Mutant Env Sequences**

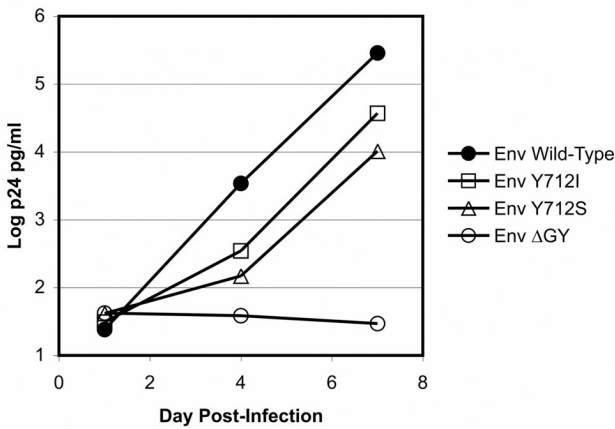
Env Wild Type 8355- GGATATTCACCATTA -8369  
 711- G Y S P L -715

Env Y712I ---AT-----  
 - I - - - -

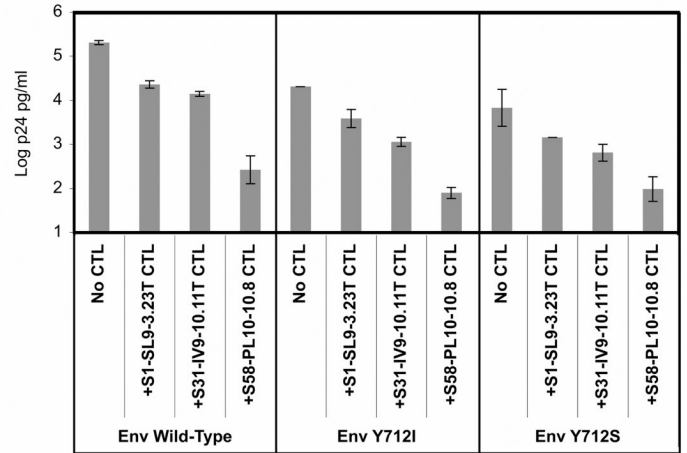
Env Y712S ---C-----  
 - S - - - -

Env ΔGY XXXXXX-----  
 X X - - - -

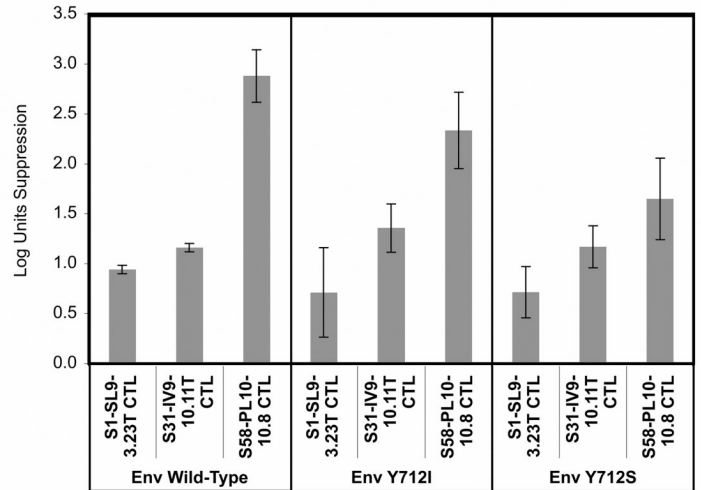
**B. Growth of Mutants**



**C. Mutant Replication With CTLs**



**D. Suppression of Mutants By CTLs**



**Figure 1. Susceptibility of Env Y712xxφ motif mutant viruses to HIV-1-specific CTL**

A. Nucleotide (NL4-3 residues 8355-8369) and amino acid (NL4-3 Env residues 711-715) sequences are given for the wild type NL4-3.1 virus and generated mutants. “-” indicates the same nucleotide or amino acid as wild type, and “X” indicates a deletion of a nucleotide or amino acid compared to wild type. B.  $1 \times 10^6$  HIV-1 permissive T1 cells were infected with the indicated mutant or wild type virus at an input of 1000pg p24 antigen and cultured in a 24-well tissue culture plate. Viral replication was measured by quantitative p24 ELISA (Perkin Elmer). These results are representative of 4 independent experiments. C. T1 cells were infected with 500 pg p24/ $10^6$  cells and cultured in triplicate in 96-well tissue culture plates with the indicated CTL clones ( $1.25 \times 10^4$  CTLs with  $5 \times 10^4$  target cells). Viral replication was measured by quantitative p24 ELISA (Perkin Elmer) on day 4 after infection. D. Inhibition was calculated by comparing replication in cells with or without CTLs. The plotted data indicate means of triplicates (error bars indicate 1 SD) for 1 experiment, and the results are representative of 2 independent experiments with these clones. Similar results were seen using a B\*57-restricted

CTL clone recognizing the epitope TSTLQEQIGW in Gag p24, and an A\*02-restricted CTL clone recognizing the epitope AIIRILQQL in Vpr in other experiments (data not shown).