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Engaging in paced mating, but neither exploratory, anti-anxiety, nor social behavior, increases 5α -reduced progestin concentrations in midbrain, hippocampus, striatum, and cortex

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Abstract

Sequential actions of 17β -estradiol (E₂) and progesterone (P₄) in the hypothalamus and the P₄ metabolite, 5α -pregnan- 3α -ol-20-one (3α , 5α -THP), in the midbrain ventral tegmental area (VTA) respectively mediate the initiation and intensity of lordosis of female rats and mayalso modulate anxiety and social behaviors, through actions in these, and/or other brain regions. Biosynthesis of E_2 , P_4 , and 3α , 5α -THP can also occur in brain, independent of peripheral gland secretion, in response to environmental/behavioral stimuli. The extent to which engaging in tasks related to reproductive behaviors and/or mating increased E₂ or progestin concentrations in brain was investigated. In Experiment 1, proestrous rats were randomly assigned to be tested in individual tasks, including the open field, elevated plus maze, partner preference, social interaction, or no test control, in conjunction with paced mating or no mating. Engaging in paced mating, but not other behaviors, significantly increased dihydroprogesterone (DHP) and 3α , 5α -THP levels in midbrain, hippocampus, striatum, and cortex. In Experiment 2, proestrous rats were tested in the combinations of the above tasks (open field and elevated plus maze, partner preference, and social interaction) with or without paced mating. As in Experiment 1, only engaging in paced mating increased DHP and 3α , 5α -THP concentrations in midbrain, hippocampus, striatum, and cortex. Thus, paced mating enhances concentrations of 5α -reduced progestins in brain areas associated with reproduction (midbrain), as well as exploration/anxiety (hippocampus and striatum) and social behavior (cortex).

Introduction

Increased production of the ovarian hormones, 17β -estradiol (E₂) and progesterone (P₄), which occurs during proestrus of rodents, modulate lordosis (the stereotypical posture that female rodents exhibit in response to male-typical stimuli in order for successful mating to occur). Systemic or intra-brain administration of E₂ and/or P₄ to the ventromedial hypothalamus (VMH) of ovari-ectomized rats reveals that the actions of these hormones are sufficient to initiate lordosis (Rubin & Barfield 1984, Pleim *et al.* 1993). However, P₄ also has effects in other brain regions, such as the ventral tegmental area (VTA) to influence the duration and intensity of lordosis (Frye 2001). In addition to divergent effects of P₄ in the VMH and VTA to alter sex behavior, there are also differences in P₄'s mechanisms of action

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in these regions. In the VMH, E_2 and P_4 initiate lordosis through classical actions at intracellular progestin receptors (PRs). However, in the VTA, P_4 modulates the duration and intensity of lordosis responses following metabolism to, and/or *de novo* synthesis of, 5α pregnan- 3α -ol-20-one (3α , 5α -THP) and its subsequent actions at GABA_A, dopamine-like type 1, and/or NMDA receptors and downstream signal transduction processes (Frye & Vongher 1999, Farye & Petralia 2003, Petralia & Frye 2003, Frye *et al.* 2004, Sumida *et al.* 2005). Using lordosis as a bioassay has revealed some of the mechanisms by which 3α , 5α -THP in the VTA facilitates mating; however, 3α , 5α -THP also has other important functional effects.

 $3\alpha,5\alpha$ -THP can influence exploratory, anti-anxiety, approach, social, and aggressive behaviors. Administration of P₄ or $3\alpha,5\alpha$ -THP to ovariectomized and/or adrenalectomized rats increases exploration, anxiolysis, approach, and social behavior to levels which are commensurate with that of proestrous rats, which have naturally elevated progestin levels (Bitran *et al.* 1999, Frye & Walf 2004, Jain *et al.* 2005, Shen *et al.* 2005) and blocking P₄'s metabolism to $3\alpha,5\alpha$ -THP attenuates these effects (Rhodes & Frye 2001, Frye & Walf 2002, Reddy *et al.* 2005, Walf *et al.* 2005). Among male rodents, $3\alpha,5\alpha$ -THP administration typically increases aggressive behavior, whereas in females $3\alpha,5\alpha$ -THP generally has a suppressive effect on agonistic behavior (DeBold & Miczek 1981, Fish *et al.* 2001, Pinna *et al.* 2005). These data suggest that $3\alpha,5\alpha$ -THP manipulations can enhance exploration, antianxiety, pro-social, and anti-aggressive behaviors, which may engender facilitation of reproduction.

In addition to manipulations of 3α , 5α -THP having effects on mating and related behaviors, experiential factors can also influence production of 3α , 5α -THP in the brain. Rodents exposed to cold water-swim, foot shock, or ether have rapid and robust increases in biosynthesis of neurosteroids, such as 3α , 5α -THP, in the brain, which occur in the absence of peripheral sources of steroid hormones (Purdy *et al.* 1991, Erskine & Kornberg 1992, Drugan *et al.* 1993, Barbaccia *et al.* 1996). Mating can also increase 3α , 5α -THP biosynthesis. Mating of naturally receptive or hormone-primed, ovariectomized and/or adrenalectomized rodents increases 3α , 5α -THP concentrations in midbrain over that of nonmated rats (Frye & Bayon 1999, Frye & Vongher 1999, Frye 2001). As well, rats that can 'pace', or control their sexual contacts with a male, have enhanced luteal function, fertility, fecundity, serum P₄, and whole brain 3α , 5α -THP levels when compared with rats mated in a standard paradigm, which cannot control receipt and/or timing of their sexual contacts (Erskine *et al.* 1989, Frye & Erskine 1990, Coopersmith & Erskine 1994, Frye *et al.* 1996, 1998). However, the extent to which engaging in reproductive and related behaviors may alter progestin biosynthesis has not been well characterized.

Paced mating, when compared with standard mating, in a laboratory setting is more similar to naturalistic and semi-natural mating insofar as female rodents exert much more control over the mating process by leaving the male after a contact and remaining apart for several minutes (McClintock & Adler 1978, Erskine 1985, Frye & Erskine 1990, Xiao & Becker 1997, Frye *et al.* 1998, Zipse *et al.* 2000, Gardener & Clark 2001, Jenkins & Becker 2001, Gonzalez-Flores *et al.* 2004, Coria-Avila *et al.* 2005). Paced mating is characterized by greater expression of exploratory, approach, anti-anxiety, reproductive, and aggressive behaviors when compared with standard mating and altering 3α , 5α -THP levels can influence expression of these behaviors. Although there is some evidence that paced mating can increase 3α , 5α -THP production, where this occurs in the brain, and whether engaging in exploratory, approach, anti-anxiety, and/or social behavior alone can elicit these increases, is of interest. To investigate this, in Experiment 1, proestrous rats engaged in individual tasks used as indices of exploration (open field), anxiety (elevated plus maze), and pro-social (partner preference and social interaction) behaviors either alone, or in conjunction with,

paced mating and effects on progestin concentrations in areas involved in mediating these behaviors (midbrain, hippocampus, striatum, and cortex) were examined. In Experiment 2, proestrous rats engaged in exploratory/anxiety (open field and elevated plus maze) or social (partner preference and social interaction) behaviors with or without engaging in paced mating. If exploratory, anxiety, or social behaviors are sufficient to influence progestin production, then engaging in exploratory, anxiety, or social behaviors should increase progestin concentrations in brain areas involved in these processes (hippocampus, striatum, and cortex). Alternatively, if mating is necessary to increase progestin concentrations, then only rats that engage in paced mating would be expected to have increased progestin concentrations in the midbrain, an important brain region for reproductive behavior.

Materials and Methods

These methods were pre-approved by the Institutional Care and Use Committee at The University at Albany-SUNY.

Animals and housing

Adult, intact female Long–Evans rats (N = 211) were obtained from the breeding colony of the Laboratory Animal Care Facility in the Life Sciences Research Building at The University at Albany-SUNY (original stock Taconic Farms, Germantown, NY, USA). Rats were group-housed in a temperature- and humidity-controlled room on a reverse light cycle (lights off at 0800 h) with water and rat chow in their cages and allowed to feed *ad libitium*. All testing occurred between 0800 and 1300 h.

Determination of estrous cycle phase

Estrous cycle phase was determined by daily examination of vaginal epithelium (between 0700 and 0800 h), per previous methods (Long & Evans 1922, Frye *et al.* 2000). Rats were cycled through two normal estrous cycles (4–5 day cycle) prior to testing. All rats were tested on the evening of proestrus, which is characterized by the presence of many nucleated epithelial cells, when E_2 levels are declining, but progestin levels are high (Feder 1984, Frye & Bayon 1999).

Behavioral testing

Affective tasks utilized in the present studies are described below. Following testing in these tasks, some rats were allowed to engage in paced mating while others were not. Specific procedures for each experiment are detailed below.

Behavioral data were collected with the ANY-Maze data collection program (Stoelting Co., Wheat Dale, IL, USA) and by one of the two observers. There was a 97% concordance rate between data that was collected by ANY-Maze and that collected by the observer.

Open field—Behavior in the open field is an index of exploration, anxiety, and motor behavior (Blizard *et al.* 1975, Frye *et al.* 2000). The open field $(76 \times 57 \times 35 \text{ cm})$ has a 48-square grid floor $(6 \times 8 \text{ squares}, 9.5 \text{ cm/side})$: there is an overhead light illuminating the central squares (all but the 24 perimeter squares were considered central). Per previous methods, rats were placed in the open field and the path of their exploration was recorded for 5 min. The number of central, peripheral, and total entries was then calculated from these data as indices of anxiolysis, thigmotaxis, and motor behavior respectively.

Elevated plus maze—Behavior in the elevated plus-maze is also utilized to assess exploration, anxiety, and motor behavior (File 1993, Frye *et al.* 2000). The plus-maze was elevated 50 cm off the ground and consisted of four arms (49 cm in length and 10 cm in

width). Two arms were enclosed by walls 30 cm in height and the other two arms were exposed. As per the previous methods, rats were placed at the juncture of the open and closed arms and the number of entries into, and the amount of time spent on, the open and closed arms were recorded during a 5-min test. Time spent on the open arms is an index of anxiety and the total number of arm entries is a measure of motor activity.

Partner preference—A modified version of the previously established partner preference task was utilized to assess preference for an intact male or a conspecific (Baum 1969, Frye *et al.* 2000). Experimental rats were placed in the center of an open field $(76 \times 57 \times 35 \text{ cm})$ that contained an ovari-ectomized stimulus female and an intact stimulus male in opposite corners, each of which was enclosed in Plexiglass compartments with small holes (1 cm in diameter) drilled in the bottom portion of the enclosure exposed to the center of the open field, so that experimental rats could receive visual and olfactory stimulation from stimulus rats in the absence of physical contact. The amount of time that experimental rats spent within a body's length of stimulus animals was recorded in a 5-min test. Increased time spent in close proximity to one stimulus rat versus another is an indication of a preference for that animal.

Social interaction—The social interaction task assessed exploratory and anxiety behavior associated with interacting with a novel conspecific (Frye *et al.* 2000, File & Seth 2003). Each member of a pair of rats (one experimental and one stimulus) was placed in opposite corners of an open field ($76 \times 57 \times 35$ cm). The total duration of time that experimental rats engaged an ovariectomized stimulus rat in crawling over and under partner, sniffing of partner, following with contact, genital investigation of partner, tumbling, boxing, and grooming was recorded during a 5-min test (Frye *et al.* 2000). An ovariectomized rat was utilized as the stimulus animal in order to avoid the possibility of vaginocervical stimulation of experimental rats, which might occur if a male rat had been used as the stimulus animal. Duration of time spent interacting with a conspecific is an index of anxiety behavior.

Paced mating—Paced mating was utilized over standard mating because of its greater ethological relevance and procedures were carried out as previously reported (McClintock & Adler 1978, Erskine 1985, Frye & Erskine 1990, Gans & Erskine 2003). Paced mating tests were conducted in a chamber $(37.5 \times 75 \times 30 \text{ cm})$, which was equally divided by a partition that had a small (5 cm in diameter) hole in the bottom center, to allow a female free access to both sides of the chamber, but which prevented the larger stimulus male from moving between sides. Females were placed in the side of the chamber opposite to the stimulus male. Rats were behaviorally tested for an entire ejaculatory series. Behaviors recorded were the frequency of mounts and intromissions that preceded an ejaculation. As well, the frequency (lordosis quotient = incidence of lordosis/number of mounts) and intensity (lordosis rating) of lordosis, quantified by rating of dorsiflexion on a scale of 0-3 (Hardy & DeBold 1972) was recorded. The percentage of proceptive (i.e., hopping, darting, ear wiggling, proceptivity quotient) and aggressive (i.e., vocalizations, defensive postures, aggression quotient) behaviors prior to contacts was also recorded. Pacing measures included the percentage of times the female left the compartment containing the male after receiving a particular copulatory stimuli (percentage exits after mounts, intromissions, and ejaculations) and latencies in seconds to return to the male compartment after these stimuli. The normal pattern of pacing behaviors for percentage exits and return latencies to be longer after more intensive stimulation (ejaculations > intro-missions > mounts) was observed in the present study.

Tissue collection

Immediately following testing, whole brains and trunk blood were collected for later measurement of E₂, P₄, DHP, and 3α , 5α -THP. Trunk blood was centrifuged at 3000 *g* for 10 min, serum was stored in eppendorfs at -80 °C. Brains were rapidly frozen on dry ice and stored at -80 °C for approximately 1 month prior to RIA.

Tissue preparation

Serum was thawed on ice and steroids extracted as described below. Immediately prior to measurement of steroids, brains were thawed and midbrain, hippocampus, striatum, and cortex were grossly dissected. The brain was initially positioned ventral side up for dissection. The anterior and posterior borders for gross dissection of midbrain were the optic chiasm and pontine regions respectively. The lateral borders were ~1.5 mm from midline and the dorsal aspect was just ventral to the cerebral aqueduct. For striatum, anterior/ posterior boundaries for dissection were the accumbens and optic chiasm, the lateral aspects were ~1.5 mm from midline, and the dorsal/ventral aspects were between the hypothalamus and thalamus and core and shell of the accumbens. For the dissection of hippocampus and cortex, brains were turned over. Intact bilateral hippocampus, including amygdala, was separated out from cortex. Bilateral anterior cortical tissues (~2 mm dorsal to olfactory bulbs) were dissected and used for cortex measurements. Following dissection, steroids were extracted from midbrain, hippocampus, striatum, and cortex as described below.

RIA for steroid hormones

 E_2 , P_4 , DHP, and 3α , 5α -THP concentrations were measured as described below, using previously reported methods (Frye *et al.* 1996, Choi & Dallman 1999, Frye & Bayon 1999).

Radioactive probes— E_2 (NET-317, 51.3 Ci/mmol), P_4 (NET-208: specific activity = 47.5 Ci/mmol), and $3\alpha,5\alpha$ -THP (used for DHP and $3\alpha,5\alpha$ -THP; NET-1047: specific activity = 65.0 Ci/mmol), were purchased from Perkin–Elmer (Boston, MA, USA).

Extraction of steroids from serum— E_2 , P_4 , DHP, and 3α , 5α -THP were extracted from serum with ether following incubation with water and 800 c.p.m. of ³H steroid (Frye & Bayon 1999). After snap-freezing twice, test tubes containing steroid and ether were evaporated to dryness in a Savant. Dried down tubes were reconstituted with phosphate assay buffer to the original serum volume.

Extraction of steroids from brain tissues— E_2 , P_4 , DHP, and 3α , 5α -THP were extracted from brain tissues following homogenization with a glass/glass homogenizer in 50% MeOH and 1% acetic acid. Tissues were centrifuged at 3000 *g* and the supernatant was chromatographed on Sepak-cartridges equilibrated with 50% MeOH:1% acetic acid. Steroids were eluted with increasing concentrations of MeOH (50% MeOH followed by 100% MeOH). Solvents were removed using a speed drier. Samples were reconstituted in 300 µl assay buffer.

Set-up and incubation of RIAs—The range of the standard curves was 0–1000 pg for E_2 , and 0–8000 pg for P_4 , DHP, and 3α , 5α -THP. Standards were added to assay buffer followed by addition of the appropriate antibody (described below) and ³H steroid. Total assay volumes were 800 µl for E_2 and P_4 , 950 0µl for DHP, and 1250 µl for 3α , 5α -THP. All assays were incubated overnight at 4 °C, except for E_2 , which incubated at room temperature for 50 min respectively.

Antibodies—The E₂ antibody (E#244, Dr G D Niswender, Colorado State University, Fort Collins, CO, USA), which generally binds between 40 and 60% of $[{}^{3}H]E_{2}$, was used in a ratio of 1:40 000 dilution and bound 54% in the present study. The P₄ antibody (P#337 from Dr G D Niswender, Colorado State University) used in a ratio of 1:30 000 dilution typically binds between 30 and 50% of $[{}^{3}H]P_{4}$, and bound 48% in the present study. The DHP (X-947) and 3 α ,5 α -THP antibodies (#921412-5, purchased from Dr Robert Purdy, Veterans Medical Affairs, La Jolla, CA, USA) used in a ratio of 1:5000 dilution bind between 40 and 60% of $[{}^{3}H]3\alpha$,5 α -THP and bound 47% in the present study.

Termination of binding—Separation of bound and free steroid was accomplished by the rapid addition of dextran-coated charcoal. Following incubation with charcoal, samples were centrifuged at 3000 *g* and the supernatant was pipetted into a glass scintillation vial with 5 ml scintillation cocktail. Sample tube concentrations were calculated using the logit–log method of Rodbard & Hutt (1974), interpolation of the standards, and correction for recovery with Assay Zap. The inter- and intra-assay reliability coefficients were: E₂ 0.06 and 0.08, P₄ 0.10 and 0.11, DHP 0.10 and 0.09, and 3 α ,5 α -THP 0.10 and 0.11.

Procedure

Experiment 1—In order to determine whether individual exploratory or social tasks, with or without engaging in paced mating, can alter progestin concentrations in brain, rats were exposed to an individual task. Some rats were tested in the open field (n = 20), elevated plus maze (n = 20), partner preference (n = 20), or social interaction tasks (21), or not tested (n = 20). Other rats were tested in the open field (n = 22), elevated plus maze (n = 20), or social interaction tasks (n = 22), partner preference (n = 20), or not tested (n = 22), or not tested (n = 20), partner preference (n = 20), or social interaction tasks (n = 22), or not tested (n = 22) in conjunction with paced mating. Immediately following testing, trunk blood and whole brains were collected for later steroid hormone measurement. Two-way analyses of variance (ANOVAs) were used to examine effects of individual affective tasks (open field, elevated plus maze, partner preference, and social interaction) and paced mating (paced mating and no paced mating) on endocrine outcomes.

Experiment 2—To ascertain whether engaging in multiple exploratory or social tasks described above would have additive effects sufficient to alter progestin concentrations, behavioral tasks were grouped according to their exploratory/anxiety versus social aspects. The open field and elevated plus maze constituted the exploratory/anxiety tasks (exploratory group) and partner preference and social interaction were considered social tasks (social group). Rats that engaged in paced mating alone or were not behaviorally tested in any tasks served as controls.

Proestrous rats were tested in the exploratory tasks (open field and elevated plus maze) with (n = 10) or without (n = 10) paced mating, social tasks (partner preference and social interaction) with (n = 10) or without (n = 10) paced mating, or served as paced mated (n = 10) or non-tested controls (n = 10). Rats were tested sequentially through assigned tasks, without rest periods between tests. Immediately following testing (or in an analogous time frame for non-tested controls), tissues were collected for later measurement of hormones. Two-way ANOVAs were utilized to examine the effects of grouped affective tasks (open field and elevated plus maze, partner preference, and social interaction) and paced mating (paced mating and no paced mating) on endocrine outcomes.

Statistical analyses

Two-way ANOVA were utilized to examine the effects of affective tasks (open field, elevated plus maze, partner preference, and social interaction) and paced mating on endocrine outcomes. Alpha level for statistical significance was P < 0.05. Where

appropriate, ANOVAs were followed by Fisher's PLSD *post hoc* tests to ascertain group differences. Power analyses were utilized to verify that all inferential statistics reported were valid with sufficient power. Six rats that were assigned to a pacing group in Experiment 1 did not exhibit normal pacing behavior and were excluded from further data analyses.

Results

Experiment 1: Effects of engaging in individual tasks

E₂ and **P**₄—There were no differences among groups in concentrations of E_2 or P₄ (Table 1) in any tissues examined. All levels were within the physiological range previously reported for proestrous rats (Frye & Bayon 1999).

DHP—Mating, but not test condition, increased DHP in midbrain (F(1,195) = 12.61, P = 0.0005), hippocampus (F(1,195) = 9.08, P = 0.003), striatum (F(1,195) = 15.40, P = 0.0001), and cortex (F(1,195) = 30.49, P = 0.0001, Fig. 1).

3 α ,**5** α -**THP**—Mating, but not test condition, also increased 3 α ,5 α -THP concentrations in midbrain (*F*(1,195) = 11.49, *P* = 0.0008), hippocampus (*F*(1,195) = 16.32, *P* = 0.0001), striatum (*F*(1,195) = 29.76, *P* = 0.0001), and cortex (*F*(1,195) = 35.62, *P* = 0.0001, Fig. 2).

Failure to engage in paced mating—Rats that failed to engage in paced mating performed similarly to other rats in affective behaviors (Table 2). Further, consistent with results discussed earlier, there were no differences in E_2 or P_4 in any brain area examined among rats that were exposed to the paced mating task, but did not exhibit normative pacing behavior, when compared with rats that were not exposed to paced mating (Table 3). In contrast to rats that actively engaged in paced mating, rats that were exposed to paced mating, but did not engage in pacing, did not demonstrate increases in DHP or 3α , 5α -THP concentrations in midbrain, hippocampus, striatum, or cortex, as was observed among rats that did engage in paced mating.

Experiment 2: Effects of engaging in multiple tasks

E₂ and **P**₄—Concentrations of E_2 and P_4 (Table 4) did not differ among groups in any tissues examined.

DHP—DHP levels were increased by mating, but not exploratory or social behaviors, in midbrain (F(2,54) = 34.84, P = 0.0001), hippocampus (F(2,54) = 46.59, P = 0.0001), striatum (F(2,54) = 17.21, P = 0.0001), and cortex (F(2,54) = 10.66, P = 0.001, Fig. 3).

3 α ,**5** α -**THP**—Mating, but not exploratory or social behaviors, increased levels of 3 α ,5 α -THP in midbrain (*F*(2,54) = 84.44, *P* = 0.0001), hippocampus (*F*(2,51) = 20.97, *P* = 0.0001), striatum (*F*(2,54) = 10.40, *P* = 0.002), and cortex (*F*(2,54) = 11.80, *P* = 0.001, Fig. 4).

Discussion

Results of the present study supported our alternative hypothesis that engaging in mating is necessary to produce increases in progestin concentrations. Rats that engaged in paced mating, irrespective of prior exposure to other tasks, either individually or in combination, had increased DHP and 3α , 5α -THP concentrations in midbrain, hippocampus, striatum, and cortex. There were no differences in E₂ or P₄ concentrations in any brain areas examined as a result of engaging in various exploratory (open field and elevated plus maze), social (partner preference and social interaction), or sexual behaviors. Together, these data suggest

that engaging in paced mating behavior produces increases in DHP and 3α , 5α -THP in areas traditionally (midbrain) and not typically (hippocampus, striatum, and cortex) associated with reproductive behavior.

The present results confirm and extend prior findings that mating can increase progestin levels. Prior investigations have demonstrated that mating increases levels of 3α , 5α -THP in brain. Mating increases central levels of 3α , 5α -THP of proestrous or ovariectomized and/or adrenalectomized, hormone-primed rats (Frye et al. 1996, Frye & Bayon 1999, Frye 2001). The results of the present study also demonstrate increased 3a,5a-THP concentrations in brain. Interestingly, we also saw increased levels of DHP among rats that were mated. Although our lab has focused primarily on effects of 3α , 5α -THP to modulate lordosis and other reproductively relevant behaviors, these data suggest that DHP may also play an important role to influence such behaviors. Whether increases in DHP were due to increased production of DHP, back-conversion of 3a,5a-THP to DHP, or a combination of these effects was not elucidated. Notably, DHP and 3α , 5α -THP have discrepant mechanisms of action. DHP has a very high affinity for intracellular PRs and is typically thought to act via more classical actions at PRs (Smith *et al.* 1974, Iswari *et al.* 1986). In contrast, 3α , 5α -THP, in physiological concentrations, does not have great avidity for intracellular PRs, but is a highly effective modulator of GABAA and NMDA receptors (Gee 1988, Cyr et al. 2001, Frye 2001). Hence, both actions may be occurring in these different brain regions. Although few intracellular PRs have been identified in the midbrain, hippocampus, striatum, or cortex and those that are present are not E₂-induced; it is possible that DHP has actions through these receptors to modulate exploratory, anxiety, and social behaviors. As well, $GABA_A$ receptors are present in each of the brain areas investigated (Hevers & Luddens 1998) and 3a,5a-THP can readily have actions through GABAA receptors (Gee 1988), which may underlie some of the effects observed in the present study. It is also possible that there is an integrated mechanism that involves actions at both intracellular PRs and GABAA receptors (Vasudevan et al. 2005, Frye et al. submitted). Therefore, the present data are important in that they may begin to elucidate the relative mechanisms by which progestins have their actions to modulate reproductively relevant behaviors beyond lordosis.

In the present study, the levels of DHP and $3\alpha,5\alpha$ -THP were increased not only in midbrain, as was expected, but also in hippocampus, striatum, and cortex. In a prior study, we examined the effects of non-paced mating on progestin concentrations and found matinginduced increases in $3\alpha,5\alpha$ -THP in midbrain, but not other areas (Frye & Bayon 1999). We also examined effects of specific stimuli associated with mating on $3\alpha,5\alpha$ -THP biosynthesis. Exposure to a mating arena, artificial vaginocervical stimulation (five-ten intromissions with glass eyedropper), or non-physical exposure to a male did not produce increases in 3α , 5α -THP concentrations in midbrain (Frye & Bayon 1999). This is congruent with the present data in that rats that were assigned *a priori* to a paced mating condition but failed to pace their mating contacts did not exhibit increases in DHP or $3\alpha,5\alpha$ -THP in any brain area examined. Together, these data suggest that aspect(s) of paced mating is important for neurosteroidogenesis; however, there have been no systematic studies directly comparing effects of standard versus paced mating on progestin biosynthesis. Ongoing investigations in our lab are examining this question.

The increased biosynthesis of DHP and $3\alpha,5\alpha$ -THP in hippocampus, striatum, and cortex, areas not typically associated with mating, may reflect $3\alpha,5\alpha$ -THP's neuromodulatory effects. $3\alpha,5\alpha$ -THP is produced *de novo* in brain in response to noxious stimuli to levels which can have agonist-like effects on GABA_A receptors and can dampen hypothalamic–pituitary–adrenal stress–responses (Purdy *et al.* 1991, Drugan *et al.* 1993, Barbaccia *et al.* 1996). As such, it has been proposed that $3\alpha,5\alpha$ -THP may increase parasympathetic activity and serve as an important mediator of homeostasis. The present data suggest that less

extreme, more normative, experiences, such as mating, can result in similar increases in progestins. The possible implications for this may be twofold. First, information about reproductive response is readily consolidated and this is adaptive in that it increases reproductive success (Domjan *et al.* 2004). We have found that other types of learning are enhanced by 3α , 5α -THP (Frye & Lacey 2000, Frye 2006*a*, Walf *et al.* 2006). Hence, 3α , 5α -THP may act to enhance consolidation of information important for reproductive behaviors. Secondly, 3α , 5α -THP's agonist-like actions at GABA_A receptors and anxiolytic effects may enable exploration, approach, and pro-social behaviors necessary for successful mating. However, the extent to which engaging in paced mating may have altered GABA_A receptor function was not addressed in the present study. These are intriguing data that suggest a role for actions of progestins in the VTA, hippocampus, striatum, and cortex to mediate anxiety, exploratory, and social behaviors. However, to more directly elucidate the role of these brain areas for effects of 3α , 5α -THP on the anxiety, exploratory, and social behaviors, it will be important to investigate the effects of manipulating 3α , 5α -THP directly in these areas.

In summary, the present data are an intriguing extension of prior reports examining effects of mating and related stimuli on progestin biosynthesis. In addition to the expected mating-induced increases in 3α , 5α -THP, we also saw increases in DHP. Further, rats that engaged in paced mating had increased levels of DHP and 3α , 5α -THP in midbrain, hippocampus, striatum, and cortex. Together, these data suggest that DHP and 3α , 5α -THP may be involved in reproductively relevant behaviors and that brain areas not traditionally associated with mating, the hippocampus, striatum, and cortex, may be important for these processes.

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Figure 1.

DHP concentrations (\pm S.E.M.) in midbrain (top left), hippocampus (top right), striatum (bottom left), and cortex (bottom right) of rats tested in the open field (OF, n = 20, n = 22), elevated plus maze (EPM, n = 20, n = 20), partner preference (SC, n = 20, n = 20), social interaction (SI, n = 21, n = 22), or no (NONE, n = 20, n = 20) affective task, alone (striped bars) or in conjunction with paced mating (closed bars). Asterisk indicates significantly different from respective no paced mating control group (P < 0.05).

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Figure 2.

 α ,5 α -THP concentrations (± S.E.M.) in midbrain (top left), hippocampus (top right), striatum (bottom left), and cortex (bottom right) of rats tested in the open field (OF, n = 20, n = 22), elevated plus maze (EPM, n = 20, n = 20), partner preference (PP, n = 20, n = 20), social interaction (SI, n = 21, n = 22), or no (NONE, n = 20, n = 20) affective task, alone (striped bars) or in conjunction with paced mating (closed bars). Asterisk indicates significantly different from respective no paced mating control group (P < 0.05).

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Figure 3.

Levels of DHP (± S.E.M.) in midbrain (top left), hippocampus (top right), striatum (bottom left), and cortex (bottom right) of rats tested in exploratory or social behaviors, or controls, alone (open bars) or in conjunction with paced mating (closed bars). Asterisk indicates significantly different from respective no paced mating control group (P < 0.05). # indicates ($P \le 0.10$).

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Figure 4.

Levels of 3α , 5α -THP (± S.E.M.) in midbrain (top left), hippocampus (top right), striatum (bottom left), and cortex (bottom right) of rats tested in exploratory or social behaviors, or controls, alone (open bars) or in conjunction with paced mating (closed bars). Asterisk indicates significantly different from respective no paced mating control group (P < 0.05).

 E_2 and P_4 levels in serum, midbrain, hippocampus, striatum, and cortex of rats tested in the open field, elevated plus maze, partner preference, social interaction, or no task in conjunction with (yes) or without (no) paced mating.

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	Oper	ı field	Elevated]	plus maze	Partner p	reference	Social in	teraction	Ž	one
	No $(n = 20)$	Yes $(n = 22)$	No $(n = 20)$	Yes $(n = 20)$	Yes $(n = 20)$	Yes $(n = 20)$	No $(n = 21)$	Yes $(n = 22)$	No $(n = 20)$	Yes (n = 20)
E ₂										
Serum (pg/ml)	36.0 ± 4.4	34.7 ± 1.0	32.0 ± 1.9	31.9 ± 1.3	32.1 ± 1.3	32.7 ± 1.0	30.9 ± 1.3	32.3 ± 1.4	30.7 ± 1.4	32.2 ± 1.8
Midbrain (pg/mg)	2.6 ± 0.2	3.0 ± 0.6	2.4 ± 0.2	2.8 ± 0.4	2.9 ± 0.6	2.4 ± 0.2	2.7 ± 0.4	2.6 ± 0.5	2.5 ± 0.4	2.2 ± 0.2
Hippocampus (pg/mg)	2.4 ± 0.3	2.5 ± 0.4	2.6 ± 0.5	2.3 ± 0.6	2.7 ± 0.4	2.0 ± 0.3	2.8 ± 0.6	2.1 ± 0.5	2.7 ± 0.7	2.8 ± 0.7
Striatum (pg/mg)	2.2 ± 0.5	1.9 ± 0.5	1.9 ± 1.0	1.3 ± 0.3	2.0 ± 0.5	1.9 ± 0.3	1.3 ± 0.3	2.5 ± 0.5	2.2 ± 0.4	2.5 ± 0.7
Cortex (pg/mg)	1.3 ± 0.5	1.4 ± 0.4	1.3 ± 0.3	1.4 ± 0.4	1.7 ± 0.6	1.1 ± 0.3	1.2 ± 0.3	1.4 ± 0.3	1.5 ± 0.3	1.4 ± 0.4
P_4										
Plasma (ng/ml)	17.5 ± 1.9	15.9 ± 0.7	16.4 ± 2.0	15.8 ± 0.6	16.3 ± 1.1	15.7 ± 0.5	15.4 ± 0.7	16.5 ± 0.5	16.4 ± 0.6	17.0 ± 0.6
Midbrain (ng/mg)	1.9 ± 0.3	2.1 ± 0.3	2.3 ± 0.3	2.7 ± 0.5	1.9 ± 0.2	1.9 ± 0.2	1.8 ± 0.3	1.8 ± 0.3	2.0 ± 0.3	2.2 ± 0.5
Hippocampus (ng/mg)	1.2 ± 0.2	1.9 ± 0.2	2.1 ± 0.5	2.2 ± 0.3	2.1 ± 0.4	2.5 ± 0.5	2.3 ± 0.4	2.1 ± 0.4	2.1 ± 0.3	2.2 ± 0.4
Striatum (ng/mg)	2.1 ± 0.7	2.0 ± 0.5	2.0 ± 1.1	1.7 ± 0.4	2.2 ± 0.4	1.5 ± 0.3	1.0 ± 0.2	1.4 ± 0.4	0.8 ± 0.1	2.3 ± 0.5
Cortex (ng/mg)	1.1 ± 0.2	1.3 ± 0.3	1.0 ± 0.2	1.6 ± 0.3	1.2 ± 0.3	1.0 ± 0.2	1.5 ± 0.3	1.7 ± 0.4	1.2 ± 0.3	1.6 ± 0.3

Behavioral data of rats that were not exposed to paced mating (no paced mating) and rats that were exposed to paced mating that either engaged (paced mating) or did not engage in paced mating (did not pace, n = 6).

	Central square entries	Open arm time	Time in proximity to male	Social interaction	Lordosis quotient	Lordosis rating	Percent exits
No paced mating	70 ± 5	34 ± 16	83 ± 9	72 + 5	NA	NA	NA
Paced mating	60 ± 7	46 ± 15	96 ± 9	85 + 6	57 ± 4	1.6 ± 0.1	26 ± 2^{d}
Did not pace	51 ± 10	36 ± 7	NA	NA	37 ± 8	2.0 ± 0.1	8 ± 1

 $^{\prime\prime}$ Significantly different from rats that engaged in paced mating.

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DHP and 3α , 5α -THP levels in midbrain, hippocampus, striatum, and cortex of rats that were not exposed to paced mating (non-paced controls) or were exposed to paced mating, but failed to exhibit normative pacing behaviors.

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	OF		EPN		None	
	Non-paced controls $(n = 20)$	Failed to pace $(n = 3)$	Non-paced controls $(n = 20)$	Failed to pace $(n = 2)$	Non-paced controls $(n = 20)$	Failed to pace $(n = 1)$
Midbrain DHP (ng/mg)	19.7 ± 3.6	15.2 ± 1.7	21.2 ± 3.5	17.9 ± 3.7	22.1 ± 4.2	19.5 ± 0
Midbrain 30,50-THP (ng/mg)	4.6 ± 1.3	3.3 ± 0.1	6.0 ± 1.4	4.9 ± 0.5	6.0 ± 2.1	5.9 ± 0
Hippocampus DHP (ng/mg)	21.9 ± 3.9	20.0 ± 2.5	27.3 ± 5.1	20.3 ± 2.7	17.7 ± 3.2	18.2 ± 0
Hippocampus $3\alpha, 5\alpha$ -THP (ng/mg)	14.3 ± 1.9	12.4 ± 3.3	13.9 ± 1.8	15.0 ± 1.5	13.2 ± 1.8	11.5 ± 0
Striatum DHP (ng/mg)	11.9 ± 3.3	10.2 ± 0.3	11.6 ± 2.4	11.5 ± 1.5	10.7 ± 2.6	9.6 ± 0
Striatum 3α , 5α -THP (ng/mg)	6.1 ± 0.9	4.8 ± 0.4	6.1 ± 0.8	5.8 ± 0.7	5.6 ± 0.9	4.5 ± 0
Cortex DHP (ng/mg)	19.7 ± 2.9	18.1 ± 11.5	23.1 ± 4.5	21.2 ± 1.8	20.0 ± 3.1	16.1 ± 0
Cortex 3a,5a-THP (ng/mg)	9.9 ± 1.5	8.8 ± 3.2	8.6 ± 1.7	6.3 ± 0.2	10.9 ± 1.1	9.5 ± 0

 E_2 and P_4 concentrations in serum, midbrain, hippocampus, striatum, and cortex of proestrous rats exposed to the open field (OF) and elevated plus maze (EPM) or partner preference (PP) and social interaction (SI) tasks, with or without paced mating, or non-tested controls.

	OF + EPM	OF + EPM + paced mating	PP + SI	PP + SI + paced mating	Paced mating	Non-tested controls
E ₂						
Serum (pg/ml)	31.9 ± 2.8	29.8 ± 3.4	29.3 ± 3.7	34.1 ± 2.8	33.7 ± 2.3	25.5 ± 4.1
Midbrain (pg/mg)	2.2 ± 0.1	2.2 ± 0.2	2.1 ± 0.1	2.0 ± 0.2	2.0 ± 0.2	2.1 ± 0.1
Hippocampus (pg/mg)	2.3 ± 0.5	2.4 ± 0.3	2.7 ± 0.6	2.5 ± 0.3	2.0 ± 0.1	1.8 ± 0.1
Striatum (pg/mg)	1.9 ± 0.7	2.0 ± 0.5	1.7 ± 0.6	1.7 ± 0.3	2.0 ± 0.7	1.5 ± 0.2
Cortex (pg/mg)	1.8 ± 0.2	1.5 ± 0.1	2.5 ± 0.6	2.1 ± 0.2	2.2 ± 0.7	1.8 ± 0.1
P_4						
Serum (ng/ml)	16.2 ± 1.3	19.8 ± 1.8	19.7 ± 1.1	20.5 ± 1.6	20.1 ± 1.5	17.5 ± 1.1
Midbrain (ng/mg)	2.1 ± 0.2	1.8 ± 0.3	1.9 ± 0.3	1.9 ± 0.4	1.9 ± 0.1	2.4 ± 0.3
Hippocampus (ng/mg)	1.6 ± 0.2	2.1 ± 0.2	1.5 ± 0.1	1.8 ± 0.2	1.7 ± 0.3	1.5 ± 0.1
Striatum (ng/mg)	1.5 ± 0.1	1.1 ± 0.2	1.5 ± 0.1	1.6 ± 0.1	1.7 ± 0.1	1.6 ± 0.1
Cortex (ng/mg)	1.8 ± 0.2	1.7 ± 0.2	1.4 ± 0.2	1.4 ± 0.1	1.8 ± 0.1	1.3 ± 0.1