Characterization of CSP-1, a Novel Extended-Spectrum β -Lactamase Produced by a Clinical Isolate of *Capnocytophaga sputigena*^{∇}

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The Capnocytophaga sputigena isolate NOR, responsible for septicemia, was resistant to amoxicillin and narrow-spectrum cephalosporins. In a cloning experiment, a new gene, bla_{CSP-1} , was identified; this gene encodes a novel extended-spectrum β -lactamase (ESBL) that shares only 52% and 49% identifies with the CME-1 and VEB-1 β -lactamases, respectively. The G+C content of this gene, its genetic environment, the absence of conjugation transfer, and its detection in two reference strains suggested that it was an intrinsic resistance gene located on the chromosome.

Capnocytophaga sputigena is a Gram-negative rod of the oral flora of healthy humans (8) that is responsible for systemic infections in immunocompromised patients with granulocytopenia and oral ulcerations, especially in children (5, 6, 8, 11, 13). Most of the clinical isolates of *C. sputigena* express β -lactamases (7). Almost 50% of these enzymes were previously identified as the transposon-encoded CfxA2 or CfxA3 enzyme (7, 10, 20). The other β -lactamases, which confer a drug resistance phenotype consistent with that of extended-spectrum β -lactamases (ESBLs), remained unidentified (7). The aims of this study were to perform the molecular characterization of a novel ESBL produced by a *C. sputigena* clinical isolate and to investigate its distribution among reference strains.

The *C. sputigena* isolate NOR was recovered at the teaching hospital in Amiens, France, in November 2008 from a 6-yearold child suffering from lymphoblastic leukemia. Ten days after the initiation of vincristine, the patient developed severe mucositis accompanied by a high fever. Despite ceftazidime and amikacin treatment, the patient's temperature remained $\geq 38^{\circ}$ C. Two blood cultures and a superficial sample of the necrotic oral lesions allowed isolation of *C. sputigena* NOR, which was identified at the genus level by the API 32ANA system (bioMérieux, Marcy l'Etoile, France) and at the species level by 16S rRNA sequencing. Once the bacteria was determined, the treatment was changed to imipenem (50 mg/kg of body weight daily) for 15 days. The course of the infection was favorable.

The NOR isolate was susceptible to fluoroquinolones (4). Moreover, its pattern of resistance was consistent with ESBL production (Table 1). PCR screening of *cfxA*-type genes (7) and conjugation experiments using the azide-resistant *Escherichia coli* J53 as the recipient strain failed (18). Cloning was attempted as previously described (3), giving rise to the recombinant clone *E. coli* TOP10(pNOR1). The recombinant plas-

* Corresponding author. Mailing address: Service de Bactériologie, Centre Hospitalo-Universitaire d'Amiens, Hôpital Nord, Place Victor Pauchet, 80054 Amiens, France. Phone: 33-32-266-84-30. Fax: 33-3-22-66-84-35. E-mail: mammeri.hedi@chu-amiens.fr. mid was purified and sequenced on both strands. Nucleotide sequence analysis failed to detect a genetic mobile element, such as an insertion sequence (IS) or class 1 integron. A 918-bp open reading frame (ORF), which encoded a 305-amino-acid protein that showed, in a BLASTx search, the highest similarity scores with other class A β -lactamases, was identified. This novel β -lactamase, which was named CSP-1 (for <u>Capnocytophaga sputigena</u> 1), had the following identities with the indicated β -lactamases: 52% identity with CME-1 (17), 49% with VEB-1 (15), 48% with TLA-1 (19), 42% with PER-1 (12), 41% with CblA (20), 40% with PER-2 (2), 38% with CepA (16), and 32% with CfxA (14).

Within the mature protein sequence of CSP-1, seven boxes

TABLE 1. MICs of β -lactams for *C. sputigena* NOR and other *C. sputigena* and *E. coli* strains^{*a*}

	MIC (μ g/ml) of β -lactam for:					
β-Lactam	C. sputigena NOR	E. coli TOP10 (pNOR-1)	C. sputigena CRBIP 17.39	C. sputigena CIP 104301T	E. coli TOP10	
Amoxicillin	256	>512	0.25	0.25	2	
Amoxicillin-CLA ^b	0.25	4	0.25	0.25	2	
Amoxicillin-SUL ^b	0.25	8	0.25	0.25	2	
Ticarcillin	4	>512	0.25	0.25	1	
Ticarcillin-CLA ^b	0.25	4	0.25	0.25	1	
Piperacillin	0.5	16	0.25	0.25	1	
Piperacillin-TZB ^b	0.25	2	0.25	0.25	1	
Cephalothin	64	>512	0.125	0.25	4	
Cefoxitin	0.5	2	0.5	0.5	2	
Cefuroxime	64	256	0.5	0.5	0.5	
Cefotaxime	0.25	8	< 0.06	< 0.06	< 0.06	
Ceftazidime	4	512	< 0.06	< 0.06	< 0.06	
Ceftazidime-CLA ^b	< 0.06	0.5	< 0.06	< 0.06	< 0.06	
Aztreonam	0.125	16	0.125	0.125	0.06	
Cefepime	0.125	4	< 0.06	< 0.06	0.06	
Imipenem	0.06	0.06	0.06	0.06	0.06	

^{*a*} The MICs of β -lactams for *C. sputigena* clinical isolate NOR, two *C. sputigena* reference strains (CRBIP 17.39 and CIP 104301T), and the recombinant clone *E. coli* TOP10(pNOR-1) producing the CSP-1 enzyme are given.

 b CLA, clavulanic acid at 2 $\mu g/ml;$ SUL, sulbactam at 4 $\mu g/ml;$ TZB, tazobactam at 4 $\mu g/ml.$

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	1	40	50
VEB-1MIKYYLRRAMNIIKICKPMS CME-2MEKNRKKQ CepAPER-1	 KKLTQMKKLTQMKKYFSFCLLGIFIFCFS SFSSLIKKELLYICIFCGALGCIQRKAQ MKKIILLFILTSQLVLAQ IVVLSIALVCIFILVFSLFHKSATKDSAN MNVIIKAVVTASTLLMVSFSSFETSAQ MNVITKCVFTASALLMLGLSSFVVSAQ FWGNIMKKHLVVIAFCVLFASASAFAAKG MSIQHFRVALIPFFAAFCLPVFAH MSIQHFRVALIPFFAAFCLPVFAH MRYIRLCIISLLATLPLAVHAS VKKSLRQFTLMATATVTLLLGSVPLYAQT MRFIHALLLAGIAHSAYASE IIFSGNLLTGCKTNLSDDYLPDDKITNNP MKFLLAFSLLIPSVVFASSSK	NPLEPLKT-EIRHIIKDKKATIGVA HTSILNDINAVTKDKKATVAVS IPPLTNVLTDSISQIVSACPGEIGVA SPLLKEQIESIVIGKKATVGVA SPLLKEQIESIVIGKKATVGVA TDSLKSSIEKYLKDKKAKVGVA TDSLKSSIEKYLKDKKAKVGVA QPLEQ-IKLSESQLSGRVGVI ADVQQKLAELERQSGGRLGVA NKGIDEIKDLETDFNGRIGVY KLTKTDLEKLEREKAAQIGVA	LILDGEDTLAVNNAEKY VLGIENDFQFSNANGNLKM VIVNNRDTVKVNNKSVY VWG-PDDLEPLLINPFEKF VWG-PDDLEPLLINPFEKF VWG-PDDLEPLLINPFEKF ULGIEDNFKLN-VNEKHHY ELDLNSGKILESFRPEEFF EMDLASGRTLTAWRADERF LINTADN-SQILYRADERF ALDTGSG-KSFSYKANERF IVDPQGE-VAGHRMAQFF IIADNGELSSHRGNEYF VLDTQNGEYWD-YNGNQRF
VEB-1PMMSVYKFHQALAVCDYLQ CME-2PMLSVFKFHIALAVLNQVD CfxAPMMSVFKVHQALALCNDFD CepAPMQSVFKLHLAMLVLHQVD PER-1PMQSVFKLHLAMLVLHQVD TLA-1PMQSTYKFHLALAVLDKLD TEM-1PMMSTFKVLLCGAVLSRVD SHV-1PMMSTFKVLLCGAVLSRVD CTX-M-1-AMCSTSKVMAVAAVLKKSE IMI-1PLCSSFKGFLAAAVLKGSQ GES-1AMCSTFKFPLAALVFERID BRO-1PVNSTIKAFIASHILLLVD	105 111 RGILDLEQNIFVKKSELLENTTWSPIRDKY KRHIPLSTSLYLDKRYFKPDTYSPLRDKY KGULTLDQKILIKKSDLLENTWSPLREKY NKGISLDTLVNINRDKLDPKTWSPMKAY QGKLDLNQTVIVNRAKVLQNTWAPIMKAY QGKLDLNQTVIVNRAKVLQNTWSPMKKDH KENISIDKKLFVKKSELLPNTWSPLRDKY AGQEQLGRRIHYSQNDLVEYSPVTEKH AGDEQLERKIHYRQQDLVDYSPVSEKH SEPENLLNQRVEIKKSDLVNYNPIAEKH DNQLNLNQIVNYNTRSLEFHSPITTKY SGTERGDRKLSYGPDMIVEWSPATERF KEKLDLNEKIIIKESDLIEYSPVCKY QGKVNPNSTVEIKKADLVTYSPVIEKQ box III box	PQGNLELPISELLAYTMQLSDNVAC (PGGNVELPISEIITYTVAQSDNNAC (SGPVISLTVRDLLRYTLTQSDNNAS (QGDEFSVPVQQLLQYSVSHSDNVAC (QGDEFVPVQQLLQYSVSHSDNVAC (QGDEFUAVQQLLQYSVSHSDNVAC (PGGNVDLSISEILKATVSRSDNNGC LLTDGMTVRELCSAAITMSDNTAA LVDGTMSLAELSAAALQYSDNVAM (KDNGMSLGDMAAAALQYSDNGAT 'LASGH-MTVLEAAQAAVQLSDNGAT 'LASGH-TTLEAAQAAVQLSDNGAT (VGQAITLDDACFATMTTSDNTAA	 DILLR-LIGGVDTVQKFIE DILFD-YIGGVNVVDEYIH DILLR-LIGGTKTVQKLMD NLMFK-DMVNVAQTDSFIA DLLFE-LVGGPAALHDYIQ DLLFE-LVGGPAALHDYIQ DLLFE-LVGGPAALHDYIQ DLLFF-FVGGTNKVHNFIS NLLLT-TIGGPKELTAFLH NLLLA-TVGGPAGLTAFLR NKLIS-HVGGPASVTAFAR NIILERYIGGPEGMTKFMR NLLLR-EIGGPAAMTQYFR NLLLR-EIGGPAAMTQYFK
VEB-1SLG-INDVSITTTEDEMHQ CME-2VNG-VKNFQIKYNEEEMHK CfxATLIPRSSFQIAYTEEEMSA CepASMG-IKETAVVANEAQMHA PER-1SMG-IKETAVVANEAQMHA TLA-1KLG-VKNISIKATEEEMHK TEM-1NMG-DHVTRLDRWEPELNE SHV-1QIG-DNVTRLDRWETELNE CTX-M-1-QLG-DETFRLDRTEPTLNT IMI-1SIG-DKDFRLDRWELDLNT GES-1KIG-DSVSRLDRKEPEMGD BRO-1EIG-ADMVLANNEPLLNR	-NGKSIYSNYTTANASSRLLQKFYNGEII -DMDDCYKNWTTPMEAANLLELFMTQDSM NDVKTLYANYTTASMVKTLKAFYKGMFI -DHNKAYSNYTSPLGAAMLMNRLFTEGLI -DDQVQYQNWTSMKGAAEILKKFEQKTQI -DDQVQYQNWTSMKGAAEILKKFEQKTQI -DDQVQYQNWTSMKGAAEILKKFEQKTQI -ADVQYQNWTSMKAAAQVLQKFEQKKQI -ANVQYTNWTTPDATVQLLKKFYKNEIL -AIPNDERDTTMPAAMATTLRKLLTGRLI -AIPGDERDTTSPRAMAQTLRNLTLGKAL -AIPGDERDTTSPRAMAQTLNLTLGKAI -NIPGDLRDTTTPIAMARTVAKVLYGGAL -NIPGDLRDTTTPIAMARTVAKVLYGGAL SHYGETSDTAKPIPYTKSLKALIVGNII -GKLGDLRDTTTPKAIASTLNKFLFGSAL	IKNEYTNFLKHIMIECETGKDRLPAP .SKRSTIFLMDIMTKTNTGMSKLEG- .DDEKQSFIKNTLKECKTGVDRIAAP .SETSQALLWKWMVETTTGPERLKG- .SETSQALLWKWMVETTTGPRLKG- .SKNSYDYLLNTMIETTTGPKRLKG- .TLASRQQLLDWMEADKVAGPLLRS- .GDSQRAQLVTWMKGNTTGAASIQA- .NEREKETYQTWLKGNTGAASIQA- .STSTHTIERWLIGNQTGDATLRA- .SNQSKEQLITWLINDKVADNLLRK-	LPESE-VKIGHKTGTSDKN -LLPK-VRMARKTGSSGKM LLDKEGVVIAHKTGSG-I -LLPAGTVVAHKTGTSG-I -LLPAGTIVAHKTGTSG-U -LLPDGTVVAHKTGSDTN -ALPAGWFIADKSGAG -VLPAGWFIADKTGAG -GLPASWVVGDKTGSG -GFPKDWVVGDKTGSC -GFPKNWVIGDKTGTC
VEB-1ERGEYIGINDIGFVILP-D CME-2KNGLTIAENDSGIVTLA-N CfxAENGVLAAHNDVAYICLP-N CepAKAGKTAATNDLGIILLP-D PER-1KAGKTAATNDLGIILLP-D TLA-1DKGITAATNDIGIILLP-N TEM-1E-RGSRGIIAALGPD-G SHV-1E-RGSRGIVALLGPN-N CTX-M-1D-YGTNDIAVIWPK-D IMI-1A-YGTANDYAVVWFK-N GES-1A-NGGRNDIGFFKAO-E BRO-1SESKNIIAVIWNE-N	DEYYTISVFVINSKENTSTNEKII GNRYVAVFVKDSKENMETNTKII GKHYAIAVFVKDSKESESQASQYV GRPLLVAVFVKDSAESSRTNEAII GRPLLVAVFVKDSAESSRTNEAII GRFLLVAVFVKDSAESSRTNEAII GKHFAIAVYVSDSEKSDERNRQI KAERIVVIYTTGSQATMDERNRQI KAERIVVIYLRDTPASMAERNQQI RAPLILVTYFTQPQPKAESRRDVL RAPLISVYTTAPKLSAKHEDKVI RD-YAVAVYTTAPKLSAVERDELV NKPYFISLFITQPHOGKSLDFKNQKDEIM QAPIIVSIYLAQTQASMERNDAI	SDISAAIYRYARNR AQVSKIVWDALNKKNKP /AHISAVVYSLLMQTSVKS AQVAQTAYQFELKKLSALSPN AQVAQTAYQFELKKLSALSPN AQVAQAAYQFELKKLSAVSPD AEICKSVWDYLVKDGK /AEIGASLIKHW /ASUGALIEHWQR /ASVGQVITQLILSTDK /AQIGKEIYPFL	

FIG. 1. Multiple alignment of the amino acid sequences of the closely related class A β-lactamases with that of CSP-1. Boxes I through VI correspond to those described by Joris et al. (9). TLA-1, *E. coli* (GenBank accession no. AF148067); CME-1, *Chryseobacterium meningosepticum* (EMBL accession no. AJ006275); VEB-1, *E. coli* (EMBL accession no. O87489); PER-1, *Pseudomonas aeruginosa* (SwissProt accession no. P37321); PER-2, *Salmonella enterica* serovar Typhimurium (EMBL accession no. X93314); CblA, *Bacteroides uniformis* (SwissProt accession no. P30898); CepA, *Bacteroides fragilis* (GenBank accession no. L13472); CfxA, *Bacteroides vulgatis* (SwissProt accession no. P30899); TEM-1 (GenBank accession no. AAR25033); SHV-1 (GenBank accession no. AAR69828); CTX-M-1 (GenBank accession no. CAJ01342); IMI-1 (GenBank accession no. AAA93461); GES-1 (GenBank accession no. AAA93256); BRO-1 (GenBank accession no. AAA92126); CARB-1 (GenBank accession no. P16897).

characteristic of B-lactamases possessing a serine active site were found (Fig. 1) (9). CSP-1 had the four conserved elements for class A β-lactamases according to Ambler et al. and Joris et al. (1, 9): the Ser-X-X-Lys consensus active site serine residue at position 70, the SDN loop at position 130, the conserved Glu166, and the KTG sequence at position 234. Moreover, the CSP-1 enzyme shares the aspartic acid residue at position 136 and the insertion of four amino acids in the region from positions 238 to 242 that are the landmarks of the PER-1 lineage, which includes the PER and VEB β-lactamases (the genes encoding these enzymes are located on an integron). The peculiar Ω loop fold, which resulted from the Asp136 together with the inserted four residues, causes a significant increase in the size of the substrate binding pocket, leading to an extended hydrolysis spectrum as revealed by the crystallographic study of PER-1 (21).

DNA sequence analysis of the regions flanking the bla_{CSP-1} gene revealed an ORF located upstream encoding a putative peptidase E sharing 92% identity with that of *Capnocytophaga gingivalis*, and two ORFs located downstream, which encoded a putative HopPmaJ type III effector protein (virulent factor) sharing 60% identity with that of *Chryseobacterium gleum*, and a TonB-dependent receptor plug domain protein (porin family), respectively.

The G+C content of the bla_{CSP-1} gene is 34%, which is similar to the values for other sequenced *C. sputigena* genes already deposited in the EMBL sequence database (overall G+C content, 38%; GenBank accession no. NZ A BZV00000000).

To investigate the distribution of the bla_{CSP-1} gene in the *C.* sputigena species, a PCR screening was performed using whole-cell DNA from *C. sputigena* reference strains CIP 104301T and CRBIP 17.39 (Pasteur Institute, Paris, France), primers CSP-1-A (5'-CAGAGAATAGATTTGCGACC-3') and CSP-1-B (5'-TGTAAGCAATCACCTCATCG-3'), and a standard PCR protocol (denaturation for 10 min at 94°C; 35 cycles of 1 min at 94°C, 1 min at 55°C, and 2 min at 72°C; and a final extension step of 10 min at 72°C), which gave rise to two specific amplicons.

Isoelectric focusing (IEF) analysis of the β-lactamase content (3) from C. sputigena NOR gave a single band with a pI value of 6.4 that comigrated with the β-lactamase extract from *E. coli* TOP10(pNOR-1). The MICs of several β -lactams, which were determined as previously described (3, 6), are reported in Table 1. E. coli TOP10(pNOR-1) showed a resistance pattern that mirrored that of the parental strain, thus indicating that B-lactamase production constituted the main mechanism of resistance. E. coli TOP10(pNOR-1) was resistant to amoxicillin, ticarcillin, narrow-spectrum cephalosporins, ceftazidime, cefotaxime, and aztreonam (4). The addition of clavulanic acid, sulbactam, and tazobactam restored the susceptibility to penicillins. The clinical isolate and its recombinant clone were both susceptible to piperacillin, cefoxitin, and imipenem (Table 1). The two reference strains were susceptible to β -lactams, thus indicating that the level of expression of the bla_{CSP-1}-like genes may vary, which might be attributable to the promoter strength. Further experiments are required to investigate this discrepancy.

The detection of a β -lactamase activity in a clinical *C. sputigena* isolate that did not harbor the *cfxA*-type gene yielded the characterization of a novel ESBL. The *bla*_{CSP-1} gene is very likely an intrinsic resistance marker resident in the chromosome of *C. sputigena* as suggested by its genetic environment, the G+C content relatedness with the overall G+C content in *C. sputigena* (38%), the absence of conjugation transfer, and the detection of bla_{CSP-1} -like genes in the two reference strains.

Nucleotide sequence accession number. The nucleotide sequence of the complete *bla*_{CSP-1} gene from *C. sputigena* isolate NOR has been deposited in the GenBank nucleotide database under accession no. GQ217533.

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