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## Cardiac spectrins: Alternative splicing encodes functional diversity

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Normal cardiac physiology relies on the precise localization and function of ion channels, receptors, regulatory molecules, and structural proteins at distinct locations in the cell. Spectrins are actin-associated proteins that provide structural support to the cell membrane and play critical roles in compartmentalization of subcellular microdomains. Importantly, abnormalities in spectrin and spectrin-associated proteins (e.g. ankyrins, obscurin, protein 4.1) have been linked with disease including congenital and acquired arrhythmia syndromes and myopathy [1-10]. The unique structure of spectrin not only allows it to provide mechanical integrity to the cell membrane, but also organize macromolecular complexes at well-defined subcellular domains. Thus, increasing our understanding of spectrin function in the heart may illuminate exciting new pathways for altering cardiac excitability and function.

Spectrin is a flexible hetero-tetrameric molecule formed from anti-parallel heterodimers of  $\alpha$ - and  $\beta$ -subunits. Originally discovered in the erythrocyte, spectrins are now known to be widely expressed in mammalian tissues.  $\alpha$ -spectrin contains 22 functional domains comprised of 21 triple helical spectrin repeats responsible for binding associated proteins, and a C-terminal calmodulin-like domain (Figure 1).  $\beta$ -spectrin consists of an N-terminal actin-binding domain, 17 spectrin repeats, and a C-terminal domain containing a pleckstrin homology domain [11].  $\alpha$ -spectrin binds to  $\beta$ -spectrin at repeat 17 on  $\beta$ -spectrin and repeat one on  $\alpha$ -spectrin [12-15]. Importantly, ankyrins bind  $\beta$ -spectrin at repeat 15 [16]. Thus, each spectrin hetero-tetramer contains two binding sites for ankyrin allowing for construction of diverse macromolecular complexes.

To date, two  $\alpha$ - isoforms and five  $\beta$ -isoforms have been identified.  $\alpha$ I- and  $\beta$ I-spectrin are the predominant isoforms found in erythrocytes while other spectrin gene products are more broadly expressed. In the heart,  $\alpha$ II-spectrin is found with  $\beta$ II-spectrin at Z-line and sarcoplasmic reticulum (SR) membranes while both  $\alpha$ I- and  $\alpha$ II-spectrin are found with  $\beta$ I-spectrin at the plasma membrane [1]. Importantly, alternative splicing of both  $\alpha$ - and  $\beta$ -isoforms provides further functional diversity. Specifically, several splicing events have been described for  $\alpha$ II-spectrin resulting in small (6-20 residue) insertions in different spectrin

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**Disclosures** None

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repeats [17,18]. C-terminal regions of  $\beta$ I-,  $\beta$ II-, and  $\beta$ IV-spectrin are also alternatively spliced to produce “long” and “short” isoforms [11,19,20]. The “long”  $\beta$ -spectrin isoforms contain a pleckstrin homology (PH) domain, a roughly 100-residue domain with roles in ligand binding and intracellular signaling [11]. Greatly truncated forms of both  $\beta$ II- and  $\beta$ IV-spectrin are also generated through splicing events [19,21,22]. While the functional consequences for these splicing events remain unclear, they likely impact the localization and function of spectrin [20].

The importance of spectrin and associated proteins for normal human physiology is illustrated by the strong association between dysfunction in these proteins and human disease. Gene mutations in  $\alpha$ - and  $\beta$ - spectrin have been linked to hereditary spherocytosis and hemolytic anemia in humans and mice, although the most common cause of the disease are mutations in *ANK1*, the gene encoding the spectrin-associated protein ankyrin-R [23,24]. Spectrin mutations have also been identified as a cause of human spinocerebellar ataxia, a progressive neurological disorder characterized by loss of coordination. Specifically, several loss-of-function mutations in  $\beta$ III-spectrin have been linked to spinocerebellar ataxia type 5 [25]. Recently, a genome-wide association study identified linkage between variability in the human *ANK3* locus (encodes spectrin-associated ankyrin-G) and bipolar disorder [26]. Finally, dysfunction in ankyrins has been associated with potentially lethal cardiac arrhythmia syndromes. Human gene mutations in *ANK2* (encodes ankyrin-B) cause a complex arrhythmia syndrome characterized by sinus node dysfunction, atrial fibrillation, ventricular arrhythmias, and increased likelihood for sudden death [3,5,7,8,27]. Similarly, a human mutation that disrupts ankyrin-G/Nav1.5 interaction results in Brugada Syndrome, characterized by abnormal electrocardiograms and ventricular arrhythmia [4,9].

In this issue of *Journal of Molecular and Cellular Cardiology*, Ursitti and colleagues add to our understanding of the spectrin superfamily by identifying a novel splice variant of  $\alpha$ II-spectrin in heart [28]. This new variant, termed cardi+ (named for its predominant expression in heart), accounts for almost 30% of total  $\alpha$ II-spectrin in rat hearts at embryonic day 16, but only 6% after three weeks of age suggesting a possible role for  $\alpha$ II-spectrin cardi+ in development. The authors also identify two new  $\alpha$ II-spectrin isoforms,  $\Sigma$ 9 and  $\Sigma$ 10, containing the  $\alpha$ II-spectrin cardi+ splice variant. Interestingly, the cardi+ insertion decreases the binding affinity of  $\alpha$ II-spectrin for  $\beta$ -spectrin. Notably,  $\alpha$ II-spectrin cardi+ fusion proteins containing the two terminal spectrin repeats were highly insoluble compared with  $\alpha$ II-spectrin cardi-. As noted by the authors, these data suggest that the novel insertion may sequester the  $\alpha$ II-spectrin cardi+ population from  $\alpha$ II-spectrin cardi- polypeptides. However, the most significant finding from the study is the effect of the cardi+ sequence on cardiomyocyte cellular shape and organization. Specifically, neonatal cardiomyocytes positive for expression of GFP- $\alpha$ II-spectrin cardi+ display aggregates of the fusion protein throughout the cell. In contrast, GFP- $\alpha$ II-spectrin cardi- is diffusely spread throughout the myocyte cytoplasm. Moreover, GFP- $\alpha$ II-spectrin cardi+ expressing myocytes display pronounced processes with significant gaps between the membrane and the myofibrils. These processes are not observed in either control or GFP- $\alpha$ II-spectrin cardi- expressing myocytes.

In summary, the new findings of Ursitti, Bloch, and colleagues add important new understanding regarding the role of the spectrin-associated network in heart. However, these findings also illustrate how little we still understand about these critical molecules. For example, are specific spectrin isoforms restricted to unique excitable cell types (i.e. ventricular, atrial, sinus node)? Moreover, do C-terminal splicing events confer novel protein interaction sites or alter normal regulatory pathways (i.e. add/subtract phosphorylation sites)? Finally, do C-terminal splicing events alter the subcellular distribution and function of specific spectrin gene products? While much remains unknown about the function of different spectrin isoforms

in heart, this important new study adds to our growing appreciation for the diversity of spectrin polypeptides in excitable tissues.

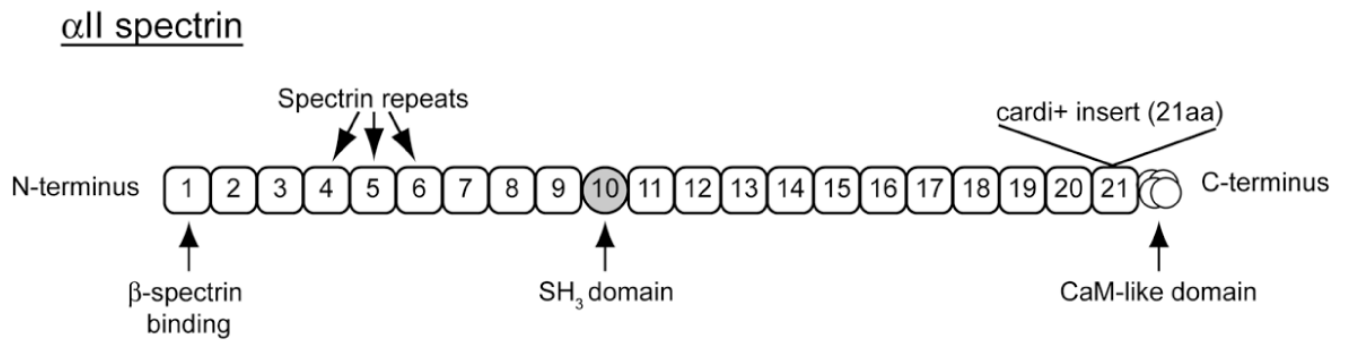
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## REFERENCES

- [1]. Baines AJ, Pinder JC. The spectrin-associated cytoskeleton in mammalian heart. *Front Biosci* 2005;10:3020–33. [PubMed: 15970557]
- [2]. Arimura T, Matsumoto Y, Okazaki O, Hayashi T, Takahashi M, Inagaki N, et al. Structural analysis of obscurin gene in hypertrophic cardiomyopathy. *Biochem Biophys Res Commun* Oct 19;2007 362(2):281–7. [PubMed: 17716621]
- [3]. Mohler PJ, Schott JJ, Gramolini AO, Dilly KW, Guatimosim S, duBell WH, et al. Ankyrin-B mutation causes type 4 long-QT cardiac arrhythmia and sudden cardiac death. *Nature* Feb 6;2003 421(6923): 634–9. [PubMed: 12571597]
- [4]. Mohler PJ, Rivolta I, Napolitano C, LeMaillet G, Lambert S, Priori SG, et al. Nav1.5 E1053K mutation causing Brugada syndrome blocks binding to ankyrin-G and expression of Nav1.5 on the surface of cardiomyocytes. *Proc Natl Acad Sci U S A* Dec 14;2004 101(50):17533–8. [PubMed: 15579534]
- [5]. Mohler PJ, Splawski I, Napolitano C, Bottelli G, Sharpe L, Timothy K, et al. A cardiac arrhythmia syndrome caused by loss of ankyrin-B function. *Proc Natl Acad Sci U S A* Jun 15;2004 101(24): 9137–42. [PubMed: 15178757]
- [6]. Hund TJ, Wright PJ, Dun W, Snyder JS, Boyden PA, Mohler PJ. Regulation of the ankyrin-B-based targeting pathway following myocardial infarction. *Cardiovasc Res* Mar 1;2009 81(4):742–9. [PubMed: 19074823]
- [7]. Le Scouarnec S, Bhasin N, Vieyres C, Hund TJ, Cunha SR, Koval O, et al. Dysfunction in ankyrin-B-dependent ion channel and transporter targeting causes human sinus node disease. *Proc Natl Acad Sci U S A* Oct 7;2008 105:15617–22. [PubMed: 18832177]
- [8]. Mohler PJ, Le Scouarnec S, Denjoy I, Lowe JS, Guicheney P, Caron L, et al. Defining the cellular phenotype of “ankyrin-B syndrome” variants: human ANK2 variants associated with clinical phenotypes display a spectrum of activities in cardiomyocytes. *Circulation* Jan 30;2007 115(4): 432–41. [PubMed: 17242276]
- [9]. Lowe JS, Palygin O, Bhasin N, Hund TJ, Boyden PA, Shibata E, et al. Voltage-gated Nav channel targeting in the heart requires an ankyrin-G dependent cellular pathway. *J Cell Biol* Jan 14;2008 180(1):173–86. [PubMed: 18180363]
- [10]. Stagg MA, Carter E, Sohrabi N, Siedlecka U, Soppa GK, Mead F, et al. Cytoskeletal Protein 4.1R Affects Repolarization and Regulates Calcium Handling in the Heart. *Circ Res*. Sep 11;2008
- [11]. Bennett V, Baines AJ. Spectrin and ankyrin-based pathways: metazoan inventions for integrating cells into tissues. *Physiol Rev* Jul;2001 81(3):1353–92. [PubMed: 11427698]
- [12]. Cherry L, Menhart N, Fung LW. Interactions of the alpha-spectrin N-terminal region with beta-spectrin. Implications for the spectrin tetramerization reaction. *J Biol Chem* Jan 22;1999 274(4): 2077–84. [PubMed: 9890967]
- [13]. Kennedy SP, Weed SA, Forget BG, Morrow JS. A partial structural repeat forms the heterodimer self-association site of all beta-spectrins. *J Biol Chem* Apr 15;1994 269(15):11400–8. [PubMed: 8157672]
- [14]. Kotula L, DeSilva TM, Speicher DW, Curtis PJ. Functional characterization of recombinant human red cell alpha-spectrin polypeptides containing the tetramer binding site. *J Biol Chem* Jul 15;1993 268(20):14788–93. [PubMed: 8325856]
- [15]. Bignone PA, Baines AJ. Spectrin alpha II and beta II isoforms interact with high affinity at the tetramerization site. *Biochem J* Sep 15;2003 374(Pt 3):613–24. [PubMed: 12820899]

- [16]. Kennedy SP, Warren SL, Forget BG, Morrow JS. Ankyrin binds to the 15th repetitive unit of erythroid and nonerythroid beta-spectrin. *J Cell Biol* Oct;1991 115(1):267–77. [PubMed: 1833409]
- [17]. Cianci CD, Zhang Z, Pradhan D, Morrow JS. Brain and muscle express a unique alternative transcript of alphaII spectrin. *Biochemistry* Nov 30;1999 38(48):15721–30. [PubMed: 10625438]
- [18]. Moon RT, McMahon AP. Generation of diversity in nonerythroid spectrins. Multiple polypeptides are predicted by sequence analysis of cDNAs encompassing the coding region of human nonerythroid alpha-spectrin. *J Biol Chem* Mar 15;1990 265(8):4427–33. [PubMed: 2307671]
- [19]. Berghs S, Aggujaro D, Dirx R Jr, Maksimova E, Stabach P, Hermel JM, et al. betaIV spectrin, a new spectrin localized at axon initial segments and nodes of ranvier in the central and peripheral nervous system. *J Cell Biol* Nov 27;2000 151(5):985–1002. [PubMed: 11086001]
- [20]. Hayes NV, Scott C, Heerkens E, Ohanian V, Maggs AM, Pinder JC, et al. Identification of a novel C-terminal variant of beta II spectrin: two isoforms of beta II spectrin have distinct intracellular locations and activities. *J Cell Sci* Jun;2000 113(Pt 11):2023–34. [PubMed: 10806113]
- [21]. Mishra L, Cai T, Yu P, Monga SP, Mishra B. Elf3 encodes a novel 200-kD beta-spectrin: role in liver development. *Oncogene* Jan 14;1999 18(2):353–64. [PubMed: 9927192]
- [22]. Tse WT, Tang J, Jin O, Korsgren C, John KM, Kung AL, et al. A new spectrin, beta IV, has a major truncated isoform that associates with promyelocytic leukemia protein nuclear bodies and the nuclear matrix. *J Biol Chem* Jun 29;2001 276(26):23974–85. [PubMed: 11294830]
- [23]. Bennett V, Healy J. Organizing the fluid membrane bilayer: diseases linked to spectrin and ankyrin. *Trends Mol Med* Jan;2008 14(1):28–36. [PubMed: 18083066]
- [24]. Agre P, Casella JF, Zinkham WH, McMillan C, Bennett V. Partial deficiency of erythrocyte spectrin in hereditary spherocytosis. *Nature* Mar 28;Apr 28;1985 314(6009):380–3. [PubMed: 3982506]
- [25]. Ikeda Y, Dick KA, Weatherspoon MR, Gincel D, Armbrust KR, Dalton JC, et al. Spectrin mutations cause spinocerebellar ataxia type 5. *Nat Genet* Feb;2006 38(2):184–90. [PubMed: 16429157]
- [26]. Ferreira MA, O'Donovan MC, Meng YA, Jones IR, Ruderfer DM, Jones L, et al. Collaborative genome-wide association analysis supports a role for ANK3 and CACNA1C in bipolar disorder. *Nat Genet.* Aug 17;2008
- [27]. Mohler PJ, Hoffman JA, Davis JQ, Abdi KM, Kim CR, Jones SK, et al. Isoform specificity among ankyrins. An amphipathic alpha-helix in the divergent regulatory domain of ankyrin-B interacts with the molecular co-chaperone Hdj1/Hsp40. *J Biol Chem* Jun 11;2004 279(24):25798–804. [PubMed: 15075330]
- [28]. Zhang Y, Resneck WG, Lee PC, Randall WR, Bloch RJ, Ursitti JA. Characterization and expression of a heart-selective alternatively spliced variant of alphaII-spectrin, cardi+, during development in rat. *Journal of Molecular and Cellular Cardiology.* 2010 x: xxx-xxx.



**Figure 1. Structure of  $\alpha$ II-spectrin and location of cardi+ insert**

$\alpha$ II-spectrin is comprised of 21 triple helical spectrin repeats and a C-terminal calmodulin-like domain. Binding to  $\beta$ -spectrin occurs in repeat 1. Repeat 10 contains an SH3 domain commonly found in cell signaling proteins. The newly identified cardi+ splice variant results in a 21 amino acid insert in repeat 21 that affects binding affinity for  $\beta$ -spectrin and may regulate cardiomyocyte growth [28].