

## Molecular cloning of rat type 2C (IA) protein phosphatase mRNA

(cDNA/adenylate cyclase)

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**ABSTRACT** A full-length cDNA encoding rat type 2C (IA) protein phosphatase was isolated from a kidney cDNA library. The cDNA was identified by screening the library with oligonucleotides based on a partial amino acid sequence determined from purified rat liver phosphatase. This clone is 2.35 kilobase pairs long and has a single extended translation reading frame that predicts a 382-amino acid protein of 42,416 daltons. The deduced amino acid sequence contains segments corresponding to three peptides from rat liver type 2C protein phosphatase and two peptides from rabbit skeletal muscle type 2C phosphatase. Rat kidney type 2C protein phosphatase is distantly related to yeast adenylate cyclase but is not related to the catalytic subunits of two other protein phosphatases (types 1 and 2A).

Protein phosphorylation plays a key role in the regulation of cellular functions. Although most studies focus on protein kinases, the importance of protein phosphatases in the phosphorylation/dephosphorylation cycle has been well documented (1). In this context, we previously reported the purification and characterization of three types of cytosolic rat liver phosphatases, types IA, IB, and II (2-5). These enzymes were later independently identified and named types 2C, 2A<sub>1</sub>, and 2A<sub>2</sub>, respectively (6). Since this later nomenclature is more widely used, we have adopted it in this report. The type 2C enzyme ( $M_r$  42,000-48,000) is monomeric and requires  $Mg^{2+}$  for activity (4, 5). Types 2A<sub>1</sub> and 2A<sub>2</sub> are heterooligomeric and have two common subunits termed  $\alpha$  ( $M_r$  35,000) and  $\beta$  ( $M_r$  69,000); the  $\alpha$  subunit is the catalytic moiety (2, 3). Type 2A<sub>1</sub> has a third subunit,  $\gamma$  ( $M_r$  58,000). The functions of the  $\beta$  and  $\gamma$  subunits are unknown. Although the type 2C enzyme was originally thought to be specific for glycogen synthase (7, 8), it was later shown that this enzyme is active with a wide variety of substrates, at least *in vitro* (5, 6). Neither the role of this enzyme in dephosphorylation *in vivo* nor the mechanism of its regulation has been described to date. To further understand the biology of the type 2C phosphatase, we undertook the molecular cloning of its mRNA.††

### MATERIALS AND METHODS

**Isolation and Sequence Analysis of Peptides.** Type 2C protein phosphatase was purified from rat liver as described (4, 5). The purified protein (100  $\mu$ g) was digested with trypsin and the resulting peptides were purified by HPLC (9). The peptides were sequenced by automated Edman degradation (10).

**Oligodeoxynucleotide Synthesis.** Oligonucleotides encoding regions of amino acid sequence were designed using the

consensus rules of Lathe (11) and synthesized on a BioSearch 8600 synthesizer.

**Cloning.** The rat liver and kidney libraries were constructed and screened as described (12, 13). The *Eco*RI inserts were subcloned into pGEM-series plasmids (Promega Biotec) and DNA sequence was determined by the dideoxy chain-termination technique of Sanger *et al.* (14) after construction of a series of nested deletions. In some cases, the dGTP analogue 7-deaza-dGTP (Toyoba, Osaka, Japan) was used to reduce band compression in G+C-rich sequences (15).

### RESULTS

**Amino Acid Sequencing.** Attempts to analyze the amino terminus of the type 2C phosphatase failed, presumably because the amino terminus is blocked. Tryptic peptides were therefore generated and isolated by reverse-phase HPLC, and limited amino acid sequence was determined from several of these by automated Edman degradation. The most probable nucleotide sequences encoding two of these peptides (Op-1 and Op-2) were determined using the rules of Lathe (11) and their complements were synthesized. The amino acid sequences of the peptides and the sequences of the corresponding oligonucleotides (On-1 and On-2) are presented in Table 1.

**cDNA Cloning.** The two oligonucleotides were labeled with phosphorus-32, mixed together, and used to screen  $\approx 10^6$  phage plaques from a rat liver cDNA library (12). Phage DNA was purified from 16 clones that yielded a signal; all of these hybridized to oligonucleotide On-2, whereas none hybridized to probe On-1, even under conditions of lowest stringency (data not shown). Cross-hybridization of the cDNAs reduced the 16 individual clones to four populations. The largest insert in each group was subcloned into the *Eco*RI site of pGEM-4Z and sequenced. One clone, pST-7, contained a 0.37-kilobase-pair (kbp) insert that encoded the peptide Op-2. This cDNA was used to probe RNA isolated from rat kidney and liver; a single-size RNA [estimated at 2.4 kilobases (kb)] from both tissues hybridized to this cDNA (data not shown). To obtain a full-length type 2C phosphatase cDNA,  $10^6$  clones from a rat kidney library (13) were probed with pST-7. This screening yielded an additional 5 clones, one of which, pST-11, had an insert of 2.35 kbp. DNA from clone pST-11 hybridized to a single 2.4-kb RNA species in rat liver and kidney (Fig. 1).

**Analysis of pST-11.** The 2.35-kbp insert was isolated by partial *Eco*RI digestion and subcloned into pGEM-7Zf(+) for sequence analysis. The sequence of 1.60 kbp of this cDNA is

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‡‡The sequence reported in this paper is being deposited in the EMBL/GenBank data base (accession no. J04503).

Table 1. Peptide and synthetic oligonucleotide sequences

Peptides		Oligonucleotides	
Name	Sequence	Name	Sequence (5' → 3')
Op-1	ALGDFDYK	On-1	CTTGTAGTTGAAGTTGCCAGGGC
Op-2	VEMEDAHTAV	On-2	CACAGCTGTGTGGCATTCTCCATCTCCAC
Op-3	TAVQVLIE	—	—
Op-4	GFELLD	—	—

Peptide sequences were determined from HPLC-purified tryptic fragments derived from 100  $\mu$ g of purified rat liver type 2C protein phosphatase and are given in standard one-letter code. The On-1 and On-2 oligonucleotide sequences are the complements of the most probable sequences [as determined by the rules of Lathe (11)] encoding the peptides Op-1 and Op-2, respectively.

presented in Fig. 2. This sequence contains a single long open translation reading frame of 1146 bp that encodes a protein of 382 amino acids. This reading frame is preceded by an 87-nucleotide G+C-rich region, and the putative translation initiation codon is in the context of the translation start consensus sequence (A at -3, G at +4, where the A in ATG is +1) (17). The open reading frame is terminated by a stop codon (TAA) at nucleotide 1294 and predicts a protein of 42,416 daltons. The sequences of both Op-1 and Op-2 are found within the conceptualized protein; furthermore, a peptide similar to Op-3 (six of eight amino acids) is also found. However, the sequence of Op-4 is not found within the protein sequence. Finally, the amino acid composition of the predicted protein is compatible with that determined for the purified rat liver phosphatase (data not shown).

**Sequence Comparisons.** The deduced amino acid sequence of the type 2C phosphatase was compared with other reported sequences by using the FASTP algorithm of Lipman and Pearson (18). This analysis demonstrated that the type 2C phosphatase is similar to the carboxyl-terminal region of yeast adenylate cyclase (19, 20); regions of similarity are shown in Fig. 3. The type 2C phosphate shares 22% identity with yeast adenylate cyclase (ref. 19 and Fig. 3); this similarity extends throughout 305 (of 382) amino acids of the type 2C phosphatase. To assess the significance of this similarity, the phosphatase and adenylate cyclase amino acid sequences were compared directly by using the RDF2 algorithm (21). In 200 shuffles of the adenylate cyclase sequence, the highest optimized similarity score was 84, while the similarity score of the type 2C phosphatase with adenylate cyclase was 140. The  $z$  value calculated from this RDF2 analysis is 11.7, indicating a high probability that the similarity between these two proteins represents an homology (21). Interestingly, the similar region overlaps the domain of yeast adenylate cyclase that reportedly (19, 20) contains its catalytic center. No other similarities were found among the sequences in two data bases (Genbank release no. 57 and the Protein Identification Resource release no. 17). Specifically, there is no significant similarity between protein phosphatase type 2C and the catalytic subunits of several other protein phosphatases (see below).

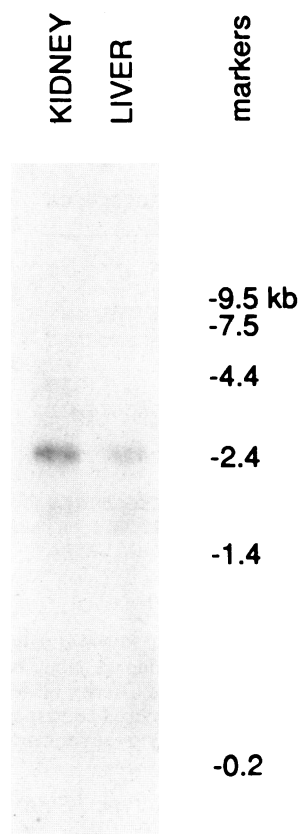


FIG. 1. Type 2C protein phosphatase RNA from rat liver and kidney. Two micrograms of poly(A)<sup>+</sup> RNA extracted from liver or kidney was electrophoresed through an agarose gel under denaturing conditions and transferred to a nylon membrane (Northern blot). The bound RNA was hybridized to 100 ng of an 800-bp 5' *Eco*RI fragment of cDNA pST-11 radiolabeled with phosphorus-32 to a specific activity of  $1.5 \times 10^8$  dpm/ $\mu$ g. Autoradiographic exposure time was 5 days in the presence of an image-intensifying screen. Methods were as described (12, 13).

## DISCUSSION

Our identification of cDNA clone pST-11 as encoding rat kidney type 2C protein phosphatase mRNA is based on the observation that two peptide sequences derived from the highly purified rat liver protein were identical to regions of the deduced protein, and a region similar to a third peptide sequence (75% identity) also was predicted. Furthermore, the size (382 amino acids, 42,416 daltons) of the predicted protein agrees well with the observed molecular weight ( $M_r$  42,000–48,000) from SDS/polyacrylamide gel electrophoresis (4, 5, 22, 23). The 2.35-kbp cDNA pST-11 is likely to be full-length because it hybridizes to a single, 2.4-kb poly(A)<sup>+</sup> RNA species from rat liver and kidney.

Cohen and colleagues recently reported the purification of two isozymes of rabbit skeletal muscle type 2C protein phosphatase (23) and determined the amino acid sequences of two peptides derived from each isozyme (16). Both peptide sequences of one isozyme (subtype 2C<sub>1</sub>) are accurately predicted by our cDNA pST-11 (Fig. 2). This finding lends further credence to our identification of this cDNA as encoding a type 2C protein phosphatase and shows unequivocally that our type IA and Cohen's type 2C phosphatases are the same protein. We note that one of the peptide sequences that we determined from rat liver type 2C phosphatase (peptide Op-4) was not found in the predicted amino acid sequence of cDNA pST-11, nor was a second pair of peptides identified by Cohen's group (isolated from isozyme 2C<sub>2</sub>) found in our cDNA-derived sequence. These missing pep-

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10          30          50          70
TGCCCGCCGCGCGTCCGCCACCGCCCGCCCGCCGACCGAGGGACCCGCCCGCGGCTGCTCCGGATCTAGAG
          90          110          130          150
GATCAAGTCATAATGGGAGCATTTTTAGACAAGCCAAAGATGGAAAAGCATAATGCCAGGGCCAGGGGAACCGG
          M G A F L D K P K M E K H N A Q G Q G N G
          170          190          210
TTGCGATACGGCCTAAGCAGCATGCAAGGTTGGCGAGTTGAAATGGAGGATGCACATACGGCTGTGATCGGTTTG
L R Y G L S S M Q G W R V E M E D A H T A V I G L
          230          250          270          290
CCAAGTGGACTTGAGACATGGTCATTCTTTGCTGTATATGATGGGCATGCTGGTTCTCAGGTTGCCAAATACTGC
P S G L E T W S F F A V Y D G H A G S Q V A K Y C
          310          330          350          370
TGTGAGCACTTGTTAGATCACATACCAATAACCAGGATTTAAAGGCTCTGCAGGAGCACCCCTCTGTGGAAAAC
C E H L L D H I T N N Q D F K G S A G A P S V E N
          390          410          430          450
GTAAAGAACGGAATCAGAACAGGTTTTCTGGAGATTGTAACACATGAGAGTTATGTCAGAGAAGAAACCGGT
V K N G I R T G F L E I D E H M R V M S E K K H G
          470          490          510
GCAGATAGAAGTGGGTCAACAGCTGTGGGTGCTTAATTTCTCCCAACATACTTATTTTCATTAAGTGTGGGGAC
A D R S G S T A V G V L I S P Q H T Y F I N C G D
          530          550          570          590
TCAAGAGGTTACTTTGTAGAAAATAGAAAAGTTCACCTTCTCACCAAGACCATAAACCAAGTAATCCTCTGGAA
S R G L L C R N R K V H F F T Q D H K P S N P L E
          610          630          650          670
AAAGAACGAATTCAGAATGCAGGGGGCTCTGTGATGATTCAGCGTGTCAATGGCTCTCTGGCTGTATCAAGAGCT
K E R I Q N A G G S V M I Q R V N G S L A V S R A
          690          710          730          750
CTTGGGGATTTTGATTACAAATGTGTCCATGGAAAAGTCCCACAGAGCAGCTTGTCTCGCCAGAGCCTGAAGTC
L G D F D Y K C V H G K G P T E Q L V S P E P E V
          770          790          810
CATGATATTGAAAGGCTGAAGAAGATGACCAGTTCATCTTTCATGTGATGGCATCTGGACGTCATGGGC
H D I E R S E E D D Q F I I L A C D G I W D V M G
          830          850          870          890
AACGAAGAGCTCTGTGACTTTGTGAGATCCAGACTTGAAGTCACTGATGACCTTGAGAAAGTTGCAATGAAGTA
N E E L C D F V R S R L E V T D D L E K V C N E V
          910          930          950          970
GTGCACACCTGTTTGTATAAGGGAAGTCGAGACAACATGATGTGATTTTGATCTGCTTTCCAAACGCGCTAAA
V D T C L Y K G S R D N M S V I L I C F P N A P K
          990          1010          1030          1050
GTCTCAGCAGAGCGGTGAAGAAGGAGCAGAGCTGGACAAGTACCTGGAGAACAGAGTAGAAGAAATCATAAAG
V S A E A V K K E A E L D K Y L E N R V E E I I K
          1070          1090          1110
AAGCAGGGGAAGCGCTCCCTGACTTAGTCCATGTGATGCGCACGTTAGCCAGTGAGAACATCCCCAGCCTCCCA
K Q G E G V P D L V H V M R T L A S E N I P S L P
          1130          1150          1170          1190
CCAGGGGTGAACTGGCAAGCAAGCGGAATGTAATGAAGCCGTTTACAATAGACTGAACCCCTTACAAAACGAC
P G G E L A S K R N V I E A V Y N R L N P Y K N D
          1210          1230          1250          1270
GACACTGATTCTGCGTCAACTGATGATATGGTAAAGCTGCTCATCTAGCCATGGTTTCACCTTTGCCTGGAAG
D T D S A S T D D M W
          1290          1310          1330          1350
AGAGAGAGAGATCAGCGCATCCTTGCTGAGCCTTTAACATCCATCACCGACTTAAGGAAGGGGTCGACATGGGT
          1370          1390          1410
GAGGGACTACAGCTTCAGCGATGGAACAGCCAGCCAGAATGGATTTTTTTTTTTTATTGTAATTTGAGACTTAT
          1430          1450          1470          1490
GTAAGCGTGATTTCAAACCGTAATTCATGTTGTAATCAGACTCCAGCAATTTTGTGTATGATTTGGTTTTT
          1510          1530          1550          1570
TGTAAGTGTAAATGTCCTTGTACAAAGTGCTCATATTTAATTATGAAGTCTTTAAAATCACTATCAATGATAA
          1590
AGGAAATTTGGCTTGTGTGTGGCACA

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FIG. 2. Nucleotide and predicted protein sequence of cDNA pST-11. The 2.35-kbp rat cDNA insert of pST-11 was removed from  $\lambda$ gt10 DNA by partial *EcoRI* digestion and subcloned into the same site in plasmid pGEM-7Zf(+). The nucleotide sequence along both strands of 1602 contiguous nucleotides was determined by the dideoxy chain-termination method (14) after construction of a series of nested deletions. The amino acids are given in one-letter code under the nucleotide character strings. The peptide sequences overscored correspond to peptides Op-2 (residues 34–43), Op-3 (residues 128–135), and Op-1 (residues 196–203). The underscored regions correspond to the sequences of peptides CB-1 (residues 116–145) and CB-2 (residues 335–370) (both from isozyme 2C<sub>1</sub>) described by another group (16).

tides are presumably parts of a type 2C phosphatase isozyme(s).

A surprising finding came from our comparison of the predicted type 2C phosphatase primary structure with those of other proteins, particularly other protein phosphatases. Type 2C protein phosphatase is unrelated to the catalytic subunit of bovine (24, 25), rabbit (26), porcine (27), or human (28) type 2A protein phosphatase as determined by an algorithm that detects local similarities (21). Furthermore, rabbit skeletal muscle type 1 protein phosphatase catalytic subunit, which is clearly related to type 2A protein phosphatase

(29), is also not related to type 2C protein phosphatase. Ingebritsen and Cohen (30) have classified the four serine/threonine protein phosphatases into two groups (type 1 and type 2) on the basis of activity against the  $\beta$  or  $\alpha$  subunit of phosphorylase kinase, respectively, and on differential sensitivity to two inhibitors. The type 2 protein phosphatases were further differentiated into types 2A, 2B, and 2C (our type 1A) on the basis of divalent cation requirements. The sequence relatedness of the type 1 and type 2A phosphatases and the lack of relatedness between these phosphatases and the type 2C phosphatase belies this classification scheme;

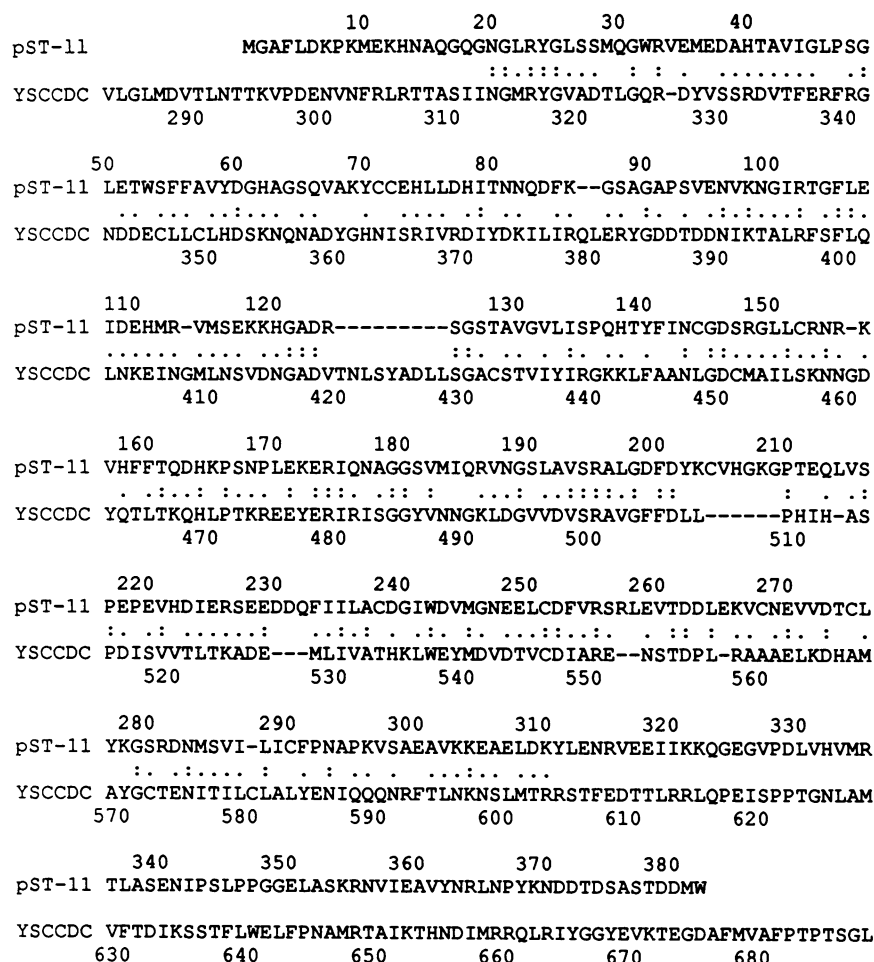


Fig. 3. Similarity between type 2C protein phosphatase (encoded by pST-11) and yeast adenylate cyclase (YSCCDCS), as detected with the FASTP algorithm of Lipman and Pearson (18). Two dots denote amino acid identity; a single dot denotes a conservative amino acid substitution. This analysis revealed 22% identity in a 305-amino acid overlap. The numbering of the yeast adenylate cyclase amino acids is that of Masson *et al.* (19).

perhaps a more meaningful classification would be based on sequence homologies and thus awaits the molecular description of type 2B phosphatase.

A global search of protein phosphatase type 2C against all known protein sequences by the FASTP algorithm (18) detected a similarity with yeast adenylate cyclase (22% identity in a 305-amino acid overlap, Fig. 3). The similarity score, percent identity, and the results of the RDF2 analysis all suggest that the type 2C protein phosphatase and yeast adenylate cyclase share a common ancestor. It is intriguing that the region of similarity overlaps a portion of the yeast adenylate cyclase catalytic center; perhaps this homology reflects the Mg<sup>2+</sup> requirement of these two enzymes.

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1. Ballou, L. M. & Fisher, E. H. (1986) in *The Enzymes*, ed. Boyer, P. D. (Academic, New York), Vol. 17, pp. 311-361.
2. Tamura, S., Kikuchi, H., Kikuchi, K., Hiraga, A. & Tsuiki, S. (1980) *Eur. J. Biochem.* **104**, 347-355.
3. Tamura, S. & Tsuiki, S. (1980) *Eur. J. Biochem.* **111**, 217-224.
4. Hiraga, A., Kikuchi, K., Tamura, S. & Tsuiki, S. (1981) *Eur. J. Biochem.* **119**, 503-510.

5. Tsuiki, S., Hiraga, A., Kikuchi, K. & Tamura, S. (1988) *Methods Enzymol.* **159**, 437-446.
6. Ingebritsen, T. S., Foulkes, J. A. & Cohen, P. (1983) *Eur. J. Biochem.* **132**, 263-274.
7. Kikuchi, K., Tamura, S., Hiraga, A. & Tsuiki, S. (1977) *Biochem. Biophys. Res. Commun.* **75**, 29-37.
8. Tamura, S., Kikuchi, K., Hiraga, A., Kikuchi, H., Hosokawa, M. & Tsuiki, S. (1978) *Biochim. Biophys. Acta* **524**, 349-356.
9. Hughes, G. J. & Wilson, K. J. (1983) in *Methods of Biochemical Analysis*, ed. Glick, D. (Wiley, New York), Vol. 29, pp. 59-135.
10. Hunkapiller, M. W., Hewick, R. M., Dreyer, W. J. & Hood, L. E. (1983) *Methods Enzymol.* **91**, 399-413.
11. Lathe, R. (1985) *J. Mol. Biol.* **183**, 1-12.
12. Lynch, K. R., Simnad, V. I., Ben-Ari, E. T. & Garrison, J. C. (1986) *Hypertension* **8**, 540-543.
13. Burnham, C. E., Hawelu-Johnson, C. L., Frank, B. M. & Lynch, K. R. (1987) *Proc. Natl. Acad. Sci. USA* **84**, 5605-5609.
14. Sanger, F., Nicklen, S. & Coulson, A. R. (1977) *Proc. Natl. Acad. Sci. USA* **74**, 5463-5467.
15. Mizusawa, S., Nishimura, S. & Seela, F. (1986) *Nucleic Acids Res.* **14**, 1319-1324.
16. McGowan, C. H., Campbell, D. G. & Cohen, P. (1987) *Biochem. Biophys. Acta* **930**, 279-282.
17. Kozak, M. (1984) *Nucleic Acids Res.* **12**, 857-872.
18. Lipman, D. J. & Pearson, W. R. (1985) *Science* **227**, 1435-1441.
19. Masson, P., Lenzen, G., Jacquemin, J. M. & Danchin, A. (1986) *Curr. Genet.* **10**, 343-352.
20. Kataoka, T., Broek, D. & Wigler, M. (1985) *Cell* **43**, 493-505.
21. Pearson, W. R. & Lipman, D. J. (1988) *Proc. Natl. Acad. Sci. USA* **85**, 2444-2448.

22. Pato, M. D. & Adelstein, R. S. (1983) *J. Biol. Chem.* **258**, 7055–7058.
23. McGowan, C. H. & Cohen, P. (1987) *Eur. J. Biochem.* **166**, 713–722.
24. Green, D. D., Yang, S.-I. & Mumby, M. C. (1987) *Proc. Natl. Acad. Sci. USA* **84**, 4880–4884.
25. da Cruz e Silva, O. B., Alemany, S., Campbell, D. G. & Cohen, P. T. W. (1987) *FEBS Lett.* **221**, 415–422.
26. da Cruz e Silva, O. B. & Cohen, P. T. W. (1987) *FEBS Lett.* **226**, 176–178.
27. Stone, S. R., Hofsteenge, J. & Hemmings, B. A. (1987) *Biochemistry* **26**, 7215–7220.
28. Arino, J., Woon, C. W., Brautigan, D. L., Miller, T. B. & Johnson, G. L. (1988) *Proc. Natl. Acad. Sci. USA* **85**, 4252–4256.
29. Berndt, N., Campbell, D. G., Caudwell, F. B., Cohen, P., da Cruz e Silva, E. F., da Cruz e Silva, O. B. & Cohen, P. T. W. (1987) *FEBS Lett.* **223**, 340–346.
30. Ingebritsen, T. S. & Cohen, P. (1983) *Science* **221**, 331–338.