

## Efficiency of BRCAPRO and Myriad II mutation probability thresholds versus cancer history criteria alone for *BRCA1/2* mutation detection

J. J. T. van Harssel · C. E. P. van Roozendaal ·  
Y. Detisch · R. D. Brandão · A. D. C. Paulussen ·  
M. Zeegers · M. J. Blok · E. B. Gómez García

Published online: 1 December 2009  
© The Author(s) 2009. This article is published with open access at Springerlink.com

**Abstract** Considerable differences exist amongst countries in the mutation probability methods and thresholds used to select patients for *BRCA1/2* genetic screening. In order to assess the added value of mutation probability methods, we have retrospectively calculated the BRCAPRO and Myriad II probabilities in 306 probands who had previously been selected for DNA-analysis according to criteria based on familial history of cancer. DNA-analysis identified 52 mutations (16.9%) and 11 unclassified variants (UVs, 3.6%). Compared to cancer history, a

threshold  $\geq 10\%$  with BRCAPRO or with Myriad II excluded about 40% of the patients from analysis, including four with a mutation and probabilities  $<10\%$  with both programs. All four probands had a *BRCA2* mutation. BRCAPRO and Myriad II showed similar specificity at 10% threshold, overall BRCAPRO was more sensitive than Myriad II for the detection of mutations. Only two of the probands with an UV had probabilities  $>20\%$  with BRCAPRO and Myriad II. In summary, BRCAPRO and Myriad II are more efficient than cancer history alone to exclude patients without a mutation. BRCAPRO performs better for the detection of *BRCA1* mutations than of *BRCA2* mutations. The Myriad II scores provided no additional information than the BRCAPRO scores alone for the detection of patients with a mutation. The use of thresholds excluded from analysis the majority of patients carrying an UV.

**Keywords** *BRCA1* · *BRCA2* · BRCAPRO · Mutation · Myriad II · Probability model · Unclassified variants

---

J. J. T. van Harssel · C. E. P. van Roozendaal · Y. Detisch ·  
R. D. Brandão · A. D. C. Paulussen · M. J. Blok ·  
E. B. Gómez García (✉)  
Department of Clinical Genetics, University Medical Centre  
Maastricht, P.O. Box 5800, 6202 AZ Maastricht,  
The Netherlands  
e-mail: Encarna.Gomezgarcia@gen.unimaas.nl

*Present Address:*

J. J. T. van Harssel  
Department of Medical Genetics, University Medical Centre  
Utrecht, Utrecht, The Netherlands

R. D. Brandão · E. B. Gómez García  
Research Institute Growth & Development (GROW), University  
of Maastricht, Maastricht, The Netherlands

M. Zeegers  
Department of Complex Genetics, Nutrition and Toxicology  
Research Institute, Maastricht University, Maastricht,  
The Netherlands

M. Zeegers  
Unit of Genetic Epidemiology, Department of Public Health,  
Epidemiology and Biostatistics, School of Medicine, University  
of Birmingham, Birmingham, UK

### Abbreviations

AUC	Area under the curve
BBC	Bilateral breast cancer
BC	Breast cancer
<i>BRCA1/2</i>	Breast cancer 1 or 2 gene
DHPLC	Denaturing high performance liquid chromatography
MLPA	Multiplex ligation-dependent probe amplification
OC	Ovarian cancer
PTT	Protein truncation test
ROC curve	Receiver operating characteristics curve
UV	Unclassified variant

## Introduction

The two major susceptibility genes for breast cancer (BC) and ovarian cancer (OC), *BRCA1* (MIM: 113705) and *BRCA2* (MIM: 600185) were discovered in 1994 and 1995, respectively [1, 2]. Initially, the Protein Truncation Test (PTT) was the screening method widely used as a quick and inexpensive test to detect pathogenic truncating mutations in these genes. At a later stage, the screening was optimised using denaturing high performance liquid chromatography (DHPLC) [3], direct sequencing and Multiplex Ligation-dependent Probe Amplification (MLPA) [4], in which all types of mutations or deletions could be detected. A drawback of this optimised screening methodology is that, as compared to PTT, it is laborious and more expensive. Furthermore, inherited mutations in these two genes only account for a small fraction of the familial clustering of BC and OC in the total group of index patients [5–9]. Therefore, accurate selection criteria for patients eligible for DNA-analysis are mandatory.

Various risk prediction algorithms and models have been developed to identify putative *BRCA1/2* mutation carriers. The first published model was that of the University of Pennsylvania (U-Penn), also called the Couch model [10]; followed by Shattuck-Eidens model or Myriad I [11]; Myriad II, an extension of the previous model [12]; the Myriad tables by Frank et al. [13]; the BRCAPRO model [14]; the Manchester scoring system of Evans et al. [15] and the recent BOADICEA model of Antoniou et al. [16].

In addition to the different types of models, also the thresholds used in these models to establish the indication to perform DNA-analysis, vary substantially among countries. In 1996, the American Society of Clinical Oncology recommended that consideration of *BRCA1/2* testing should be offered to patients with strong familial features, such as BC in the family or very early age at onset of the disease, c.q. corresponding to those patients whose prior probability of carrying a mutation exceeds 10% [17]. However, the updated American guidelines in 2003 did not recommend a numerical threshold [18]. Currently, the majority of Dutch Cancer Genetics Services apply a 10% pre-test probability as threshold to perform DNA analysis of the *BRCA1* and *BRCA2* genes [19], whereas in the UK most centres offer mutation analysis to families with a 20% or greater pre-test probability of carrying a mutation [20].

The goal of our study was to retrospectively analyze the added value of predictive mutation probability scores and optimal thresholds in terms of sensitivity and specificity of two currently used mutation probability models BRCAPRO and Myriad II, in a group of patients prior selected on the basis of family history of cancer.

**Table 1** Indications for DNA-analysis based on cancer history (classified by number of affected relatives)

Number of affected relatives		Indication for performing DNA-analysis
1	a	BC <35 years
	b	Bilateral BC; first time < 50 years
	c	Male BC
	d	OC <50 years
2	a	Two-first degree relatives with BC/OC, one case <50 years
	b	Two-first degree relatives with OC
	c	Two-second degree relatives with BC/OC, both <50 years
≥3		Three or more affected relatives in the family

BC Breast cancer, OC Ovarian cancer

## Materials and methods

### Patients and DNA-analysis

Our study includes 306 probands who had been selected for DNA-analysis based on established criteria about personal and family cancer history (shown in Table 1).

In the period 1996–2000 partial mutation screening of the *BRCA1* and *BRCA2* genes was performed using the protein truncation test (PTT). The PTT was used only for exon 11 of *BRCA1* and exons 10 and 11 of *BRCA2*. From 2000 on, complete genetic screening was performed using DHPLC [3], direct sequencing and for *BRCA1* also MLPA [4].

Clinical data of all the probands and their first and second degree relatives were retrieved from their medical records, which included cancer site: BC and/or OC, bilateral BC, BC in male relatives, age at diagnosis and number of affected relatives (data not shown). All probands were affected with breast cancer and/or ovarian cancer. There were no probands with known Jewish ancestry.

### Mutation probability methods

BRCAPRO [14] is a Mendelian model that incorporates mutated allele frequencies and cancer specific penetrances, in addition to the following clinical data about the probands and first and second degree relatives: number of women affected with BC; OC only; discrimination between paternal and maternal inheritance pattern; BC under age 50 and OC; bilateral BC; relative with both OC and BC; affected and unaffected individuals; Ashkenazi Jewish ancestry and male BC.

The Myriad II prevalence tables [12] are based on proband and family history accompanying results of *BRCA1/2* deleterious variant samples tested by the Myriad company.

The calculation takes into account if there are relatives who have been diagnosed with OC at any age, if relatives have been diagnosed with BC, grouped by those over and under age 50 years at diagnosis of BC, and the presence of male BC. It does not take into account bilateral BC and specific age at diagnosis. In addition, it only allows inclusion of a maximum of three relatives, including the patient; and it does not calculate *BRCA1* and *BRCA2* mutation probabilities separately like BRCAPRO. A more detailed description of both models has been published [12–14].

Carrier probabilities with BRCAPRO and Myriad II were calculated with the CancerGene software package from the University of Texas Southwestern Medical Center (CaGene version 4.0). Additional information about CancerGene is available at <http://www.utsouthwestern.edu>.

### Statistical analysis

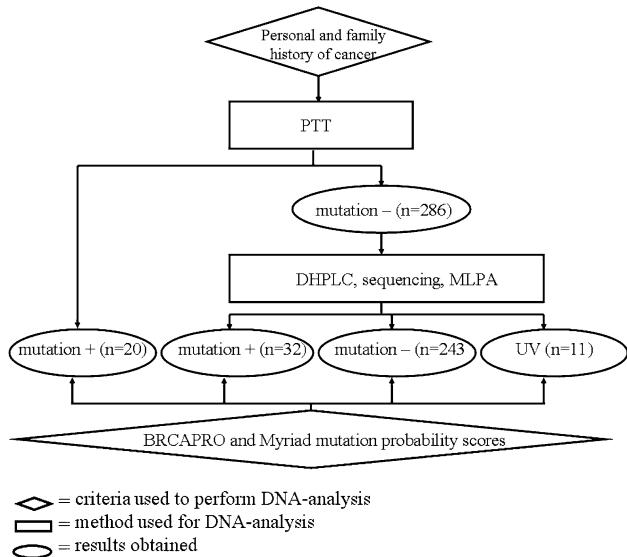
We calculated the carrier probabilities using the BRCAPRO and Myriad II models and compared the clinical characteristics between the group of probands with and those without a pathogenic mutation. Carriers of UVs were reported separately and not included in either the mutation positive or mutation negative groups for the purpose of the analysis of clinical features. Genetic alterations reported as polymorphisms were scored as mutation negatives. All data were analyzed using SPSS statistical software version 15.0 for Windows. The sensitivity, specificity, positive and negative predictive values, and positive and negative likelihood ratios were calculated for both BRCAPRO and Myriad II, and each of them for 10 and 20% thresholds.

Receiver operating characteristic (ROC) curves were drawn and the areas under the curve (AUC) were calculated in the group of 295 probands with either a mutation or a wild-type genotype. Calculations were made for BRCAPRO and for Myriad II and within each of them at 10 and 20% thresholds.

## Results

### DNA-analysis

Our study included a total of 306 probands who were eligible for *BRCA1* and *BRCA2* mutation detection, based on established personal and family cancer history criteria, as shown in Table 1. With the PTT we found 20 pathogenic mutations (6.8%). In the remaining 286 probands, complete *BRCA1* and *BRCA2* gene sequencing showed 43 additional genetic variations (14%). Thirty-two of them (10.4%) could be classified as a pathogenic mutation (*BRCA1* n = 13, *BRCA2* n = 19). The remaining 11 variants (3.6%) were classified as unclassified variants (UVs,



**Fig. 1** Chronological sequence of events in the study population

*BRCA1* n = 7, *BRCA2* n = 4). Figure 1 summarizes the chronological sequence of diagnostic methods and selection criteria applied in the studied population. An individual description of the genetic variants identified is presented in Table 2. Instructions for the nomenclature for the sequence variations can be seen at: <http://www.genomic.unimelb.edu.au/mdi/mutnomen/recs.html>.

Classification of the genetic variants (mutations or UVs) was done according to the Breast Cancer Information Core (BIC) database: [<http://research.nhgri.nih.gov/bic/>].

### Mutation probabilities

#### Pathogenic mutations

The mean carrier probabilities with BRCAPRO and Myriad II for the groups with a mutation and without a mutation are shown in Table 3. The probability scores are also shown for the subgroup of patients with mutations detected with PTT and for those with mutations detected by complete analysis. The carrier probabilities of the probands with a mutation (n = 53) were significantly higher than those without a mutation, in both models (BRCAPRO:  $P < 0.0001$ , Myriad II:  $P < 0.0001$ ). Apart from BRCAPRO and Myriad II, the occurrence of OC in the family ( $P = 0.001$ ) was the only clinical parameter which was significantly associated with the presence of a *BRCA1/2* mutation (data not shown).

The performance of the models, at 10 and 20% thresholds, is presented in Table 4 and Fig. 2 shows the corresponding ROC curves. Using 10% threshold with BRCAPRO 110 probands had a probability <10% of whom four had a mutation. With Myriad, nine out of 126 probands

**Table 2** Genetic variants identified

BRCA1			BRCA2		
Exon	Nucleotide change	Number of families	Exon	Nucleotide change	Number of families
<i>Pathogenic mutations found with PTT</i>					
11	c.2193_2197del5	5	11	c.5213_5216del4	1
11	c.2722G>T	4	11	c.5645C>G	1
11	c.1291_1292insT	1	11	c.5682C>G	1
11	c.2646_2648delTGC	1	11	c.6275_6576delTT	1
11	c.2685_2686delAA	1	11	c.6396_6397insA	1
11	c.2989_2990insAA	1	11	c.6486_6489del4	1
			11	c.6643delA	1
Total	6	13	Total	7	7
<i>Pathogenic mutations found with DHPLC, MLPA, and sequencing</i>					
1	del ex1a/b-2	1	3	c.115delG	1
2	c.68_69delAG	2	5	c.469_470delAA	2
5	c.191G>A	1	6	c.516 + 1G>T	1
8	c.514C>T	1	8	c.658_659delGT	2
17	c.4987- ?_2074 + ?del	1	10	c.1773-1776del4	1
18	c.5080G>T	1	10	c.1813insA	1
20	c.5277 + 1G>A	4	11	c.4314delC	1
22	c.5346G>A	1	15	c.7436_7437-2delAGAT	1
24	c.5503C>T	1	16	c.7734_7740del6ins9	2
			18	c.8058_8059delT	3
			19	c.8374_8384del11ins3	1
			23	c.9063_9078del16	1
			23	c.9117G>A	2
Total	9	13	Total	13	19
<i>Unclassified variants</i>					
11	c.692C>T	1	10	c.1385A>G	1
11	c.1258G>T	1	15	c.7469T>C	1
11	c.1486C>T	1	18	c.8111C>T	1
17	c.4987-3C>G	1	21	c.8723T>G	1
19(I)	c.5194-25A>C	1			
20	c.5216A>G	1			
23	c.5435C>G	1			
Total	7	7	Total	4	4

with a probability, <10% had a mutation. The four probands missed with BRCAPRO were also missed with Myriad. These four probands carried a *BRCA2* mutation, none of them had a family history of male BC, OC or bilateral BC. In one case the indication for DNA analysis was three BC cases in the family (indication 3, Table 1). The other three cases have a family history of two BC cases with one case below age 50 (indication 2a, Table 1).

Results using the 20% threshold, 7 out of 160 probands with BRCAPRO and 18 out of 249 probands with Myriad II had a mutation with a probability below the threshold. At both thresholds BRCAPRO is more sensitive than Myriad II, at 10% they both have similar specificity while at 20% is

Myriad II more specific (Table 4). As shown by the four AUC calculations from Fig. 2: (a) BRCAPRO performed better than Myriad II at both thresholds and (b) the AUC for the 20% thresholds were better than 10% thresholds.

#### Unclassified variants

The group of 11 UVs had median mutation probabilities with BRCAPRO of 13.6% (range 0.3–97.3%) and with Myriad II of 17.4% (range 7.3–37.6%). When compared with the group with a mutation, the UVs had lower probabilities than the group with a mutation: with BRCAPRO were statistically significant ( $P = 0.004$ ), with Myriad II

**Table 3** BRCAPRO and Myriad II scores of BRCA mutation positive versus negative populations

		Mutations with PTT N = 21	Mutations with complete analysis N = 32	Total mutations N = 53	No mutation N = 243	P values (*)
BRCAPRO	M	0.699	0.586	0.629	0.240	<0.0001
	±SD	(± 0.248)	(± 0.339)	(± 0.310)	(± 0.273)	
	Range	0.186–0.997	0.012–1.00	0.012–1.00	0.001–0.994	
Myriad II	M	0.293	0.202	0.237	0.119	<0.0001
	±SD	(± 0.124)	(± 0.126)	(± 0.131)	(± 0.102)	
	Range	0.122–0.465	0.019–0.421	0.019–0.465	0.007–0.603	

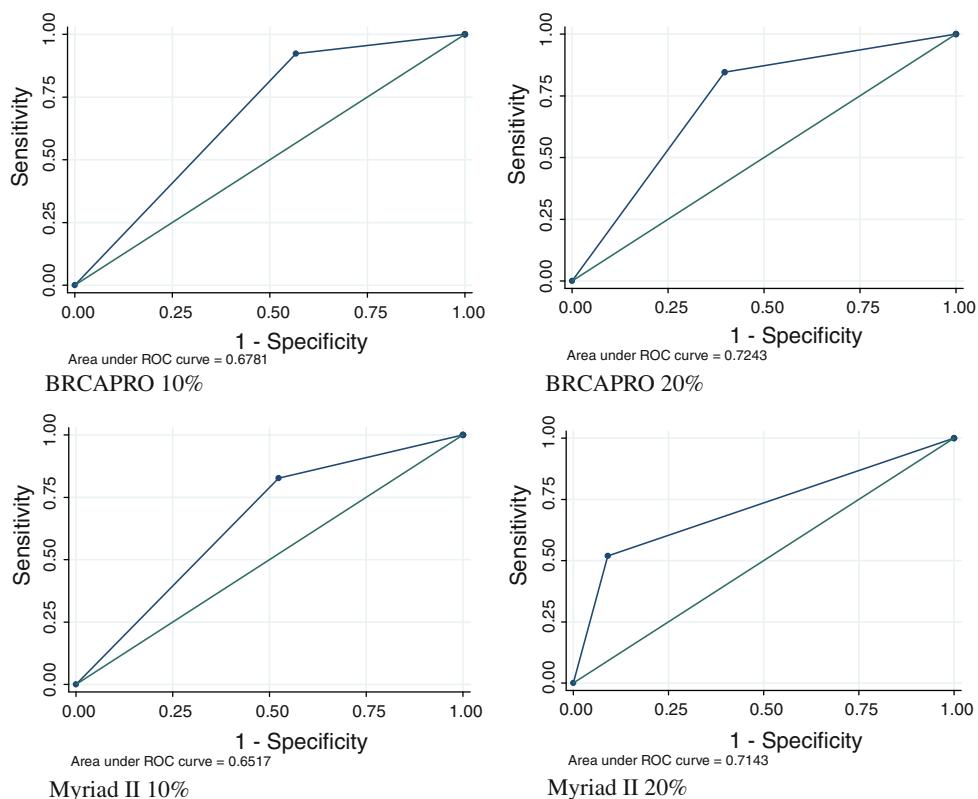
Results expressed as:  $M \pm SD$  mean ( $\pm$  standard deviation)

(\*) P values: total mutations versus no mutations: *t* test

**Table 4** Performance parameters of BRCAPRO and Myriad II

	Threshold 10%		Threshold 20%	
	BRCAPRO	Myriad II	BRCAPRO	Myriad II
Mutations/probands below threshold (*)	4/110	9/126	7/160	18/249
Sensitivity (95% CI)	0.875 (0.760–0.990)	0.719 (0.563–0.875)	0.781 (0.638–0.925)	0.438 (0.266–0.609)
Specificity (95% CI)	0.433 (0.372–0.494)	0.476 (0.415–0.538)	0.602 (0.542–0.663)	0.909 (0.874–0.945)
Positive predictive value (95% CI)	0.163 (0.108–0.218)	0.147 (0.092–0.203)	0.198 (0.129–0.268)	0.378 (0.222–0.535)
Negative predictive value (95% CI)	0.965 (0.931–0.999)	0.931 (0.887–0.974)	0.956 (0.925–0.988)	0.928 (0.896–0.960)

(\*) Number of probands with a mutation/total number of probands below the thresholds (i.e., <10% or <20%)

**Fig. 2** ROC curves for BRCAPRO and Myriad II, at 10 and 20% thresholds

**Table 5** Biochemical and epidemiological features of the unclassified variants with corresponding BRCAPRO and Myriad II scores

Exon	Nucleotide change	Effect on protein	Number of families	BRCAPRO (%)	Myriad II (%)	Polarity change	Conserved mammals/other	Times reported (BIC)	Co-segregation (literature)	Co-occurrence (literature)	Classification (literature)	References
<i>BRCA1</i>												
11	c.692C>T	p.T231M	1	59.9	17.8	Y	N/N	2	ND	N	Unknown	[31]
11	c.1258G>T	p.D420Y	1	6.5	10.5	Y	N/Y	3	ND	N	Unknown	[31]
11	c.1486C>T	p.R496C	1	0.3	3.4	Y	N/N	43	ND	Y	Neutral	[31, 32]
17	c.4987-3C>G	NA	96.9	37.6	—	—	—	0	ND	ND	ND	ND
20	c.5194-25A>C	NA	1	14.0	17.4	—	—	0	ND	ND	ND	ND
20	c.5216A>G	p.D1739G	1	97.3	32.5	Y	Y/Y	2	ND	ND	Cancer-associated	[30]
23	c.5435C>G	p.P1812R	1	4.6	7.5	Y	Y/Y	0	ND	ND	ND	ND
Total: 7			7									
<i>BRCA2</i>												
10	c.1385A>G	p.E462G	1	37.8	17.8	Y	Y/N	35	N	Y	Neutral	[29, 33, 34]
15	c.7469T>C	p.I2490T	1	13.6	19.9	Y	N/N	238	ND	ND	ND	ND
18	c.8111C>T	p.S2704F	1	11.5	7.3	Y	N/N	1	ND	ND	ND	ND
21	c.8723T>G	p.V2908G	1	1.2	9.8	N	Y/N	8	N	N	Neutral	[34]
Total: 4			4									

Alignments were based on following species and NCBI reference sequences: *BRCA1* Human (NP\_009225), Chimpanzee (NP\_001038958), Gorilla (AAT44835), Orang (AAT44834), Macaque (NP\_001108421), Dog (NP\_001013434), Rat (NP\_036646), Mouse (NP\_033894), Cow (NP\_848668), Opossum (NP\_001029141), Chicken (NP\_989500), Frog (AAI70141); *BRCA2* Human (CAA64484), Chimp (XP\_509619), Dog (BAB91245), Cat (NP\_001009858), Mouse (NP\_033895), Opossum (ABP48762), Chicken (NP\_989607). Y Yes, N No, ND Not done, NA Non Applicable

did not achieve statistical significance ( $P = 0.093$ ) (data not shown).

As shown in Table 5, with Myriad II nine UVs and with BRCAPRO seven of the 11 UVs had mutation probabilities below 20%. The UVs: c.4987-3C>G (putative splice variant), and c.5216A>G in *BRCA1* were the only two UVs with probability values >20% both with BRCAPRO and Myriad II.

## Discussion

This study included 306 probands who had prior been selected for DNA-analysis of *BRCA1* and *BRCA2* mutations based on cancer history criteria. From 1995 until 2000, the PTT was the widely used quick, inexpensive detection technique, which at that time resulted in the detection of a mutation in 20 unrelated patients. At a later stage, complete sequencing analysis of the coding regions including intron-exon boundaries was performed, which resulted in the identification of an additional 43 genetic variations in individual probands of which 32 were pathogenic and 11 unclassified (UV). We then performed a retrospective study to estimate the mutation probabilities using BRCAPRO and Myriad II in all cases.

Concerning mutation detection rate, our results are in line with previous reports ([21] which show that complete *BRCA1/2* analysis substantially increases the number of variations detected as compared to PTT alone. However, it is less obvious which methods and which thresholds to use for selecting the patients for DNA-analysis. In this respect, the right balance between costs and mutation coverage needs to be determined: obviously, the ideal method has a high sensitivity, a high specificity and a high negative predictive value.

We found in our population study that BRCAPRO and Myriad II are more specific than cancer history, since they both excluded from analysis about 40% of the patients who did not have a mutation. Looking at sensitivity, BRCAPRO was more sensitive than Myriad II which is in line with previous publications by Berry et al. [18, 22]. Overall BRCAPRO performed better than Myriad II, as shown by their ROC curves.

Several groups have compared risk prediction models among each other [22–26]. Both Antoniou et al. [25] and Lindor et al. [26] compared performance of various carrier prediction algorithms, including BRCAPRO and Myriad II. In the study of Lindor et al. (26) no a priori selection criteria were required to perform DNA-testing. All models tested performed reasonably well. In the study of Antoniou et al. [25], with 1,934 families the largest published so far, families in whom DNA-testing had been performed were supplied by the clinical genetics centres and from research

studies. The indication criteria used to perform DNA-analysis in those settings are not specified. The authors show that BOADICEA provided the best discrimination of the five algorithms analyzed, though no significant differences were observed with this model and BRCAPRO. Both studies [25, 26] concluded that these prediction models under-predicted the numbers of carriers in the low estimated risk category. However, none of these studies have compared the added value of the use of quantitative methods, i.e., scores obtained with mutation probability detection methods to qualitative criteria based on cancer family history as indication for DNA analysis.

About the thresholds, the AUCs were better at 20% than at 10%, which also indicates the preference from an economic point. However, from a clinical point of view sensitivity is more important than specificity, which means that a choice has to be made between sparing 16% of the total of tests performed (i.e., 50 probands with mutation probabilities between 10 and 20% with BRCAPRO) at the price of missing three probands with a mutation.

Noteworthy is that the four pathogenic mutations that were missed with BRCAPRO, were also four of the nine which were missed with Myriad II using the 10% threshold and that all four are located in the *BRCA2* gene. This suggests that for detection of *BRCA1* mutations, BRCAPRO at a threshold of 10% is the method of choice: i.e., it has the same sensitivity as the cancer history criteria and reduces the number of tests by 40%. The fact that these probability models have a better performance for *BRCA1* than *BRCA2* has been reported previously by James et al. [23] and Bodmer et al. [24], and is related to the lower penetrance of the mutations in *BRCA2*. Indeed, as explained in the results section, none of the families of these patients included any of the most characteristic features of the *BRCA* families, such as OC, bilateral BC or male BC.

An additional advantage of the use of models with thresholds is the exclusion of many patients carrying an UV. Indeed, since the majority of the UVs that have been detected and reported in the literature are later reclassified as neutral variants [27], a specific method is also desirable which can selectively exclude those UVs which are not clinically relevant. We have previously reported that patients with a pathogenic mutation have significantly higher a priori probabilities with BRCAPRO and Myriad II than those with a UV [28], which also suggests that most of the UVs are neutral variants. In addition, BRCAPRO and Myriad II have been shown to be useful parameters also to predict whether a UV is deleterious when used in multi-parametric regression models [29].

Interestingly, from the two UVs with mutation probabilities above 20% with BRCAPRO and Myriad II, the p.D1739G variant in *BRCA1* has been classified as cancer-associated in the literature according to criteria, such as

polarity change, conservation amongst species and in silico models [30]. Conversely, for most of those UVs with a mutation probability below 20%, evidence from other sources also argue against them being deleterious. This is the case for: p.R496C in *BRCA1* [31, 32] and p.E462G [29, 33, 34] and p.V2908G [34] in *BRCA2*.

In conclusion, BRCA PRO and Myriad II probability scores allow exclusion of a large proportion of mutation negative probands for *BRCA1/2* diagnosis, as compared to using familial cancer history alone. When comparing the two models, BRCA PRO is more sensitive in detecting known pathogenic mutations than Myriad II. Myriad II scores provide no added value to BRCA PRO alone. BRCA PRO performs better for the identification of patients with a *BRCA1* than with a *BRCA2* mutation. The use of quantitative scores exclude from study a considerable number of patients carrying an UV. Information about the probability scores may be also valuable for the classification of UVs.

**Acknowledgments** Rita Brandão was supported by two Portuguese grants: Fundação Calouste Gulbenkian (79117), and Fundação para a Ciência e Tecnologia (SFRH/BD/32386/2006).

**Open Access** This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

## References

- Miki Y, Swensen J, Shattuck-Eidens D et al (1994) A strong candidate for the breast and ovarian cancer susceptibility gene *BRCA1*. *Science* 266(5182):66–71
- Wooster R, Bignell G, Lancaster J et al (1995) Identification of the breast cancer susceptibility gene *BRCA2*. *Nature* 378(6559):789–792
- Wagner T, Stoppa-Lyonnet D, Fleischmann E et al (1999) Denaturing high-performance liquid chromatography detects reliably *BRCA1* and *BRCA2* mutations. *Genomics* 62(3):369–376
- Hogervorst FB, Nederlof PM, Gille JJ et al (2003) Large genomic deletions and duplications in the *BRCA1* gene identified by a novel quantitative method. *Cancer Res* 63(7):1449–1453
- Antoniou A, Pharoah PD, Narod S et al (2003) Average risks of breast and ovarian cancer associated with *BRCA1* or *BRCA2* mutations detected in case series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet* 72(5):1117–1130
- King MC, Marks JH, Mandell JB (2003) Breast and ovarian cancer risks due to inherited mutations in *BRCA1* and *BRCA2*. *Science* 302(5645):643–646
- Easton DF, Ford D, Bishop DT (1995) Breast and ovarian cancer incidence in *BRCA1*-mutation carriers. Breast cancer linkage consortium. *Am J Hum Genet* 56(1):265–271
- Ford D, Easton DF, Stratton M et al (1998) Genetic heterogeneity and penetrance analysis of the *BRCA1* and *BRCA2* genes in breast cancer families. The breast cancer linkage consortium. *Am J Hum Genet* 62(3):676–689
- Thompson D, Easton D (2001) Variation in cancer risks, by mutation position, in *BRCA2* mutation carriers. *Am J Hum Genet* 68(2):410–419
- Couch FJ, DeShano ML, Blackwood MA et al (1997) *BRCA1* mutations in women attending clinics that evaluate the risk of breast cancer. *N Engl J Med* 336(20):1409–1415
- Shattuck-Eidens D, Olliphant A, McClure M et al (1997) *BRCA1* sequence analysis in women at high risk for susceptibility mutations. Risk factor analysis and implications for genetic testing. *JAMA* 278(15):1242–1250
- Frank TS, Manley SA, Olopade OI et al (1998) Sequence analysis of *BRCA1* and *BRCA2*: correlation of mutations with family history and ovarian cancer risk. *J Clin Oncol* 16(7):2417–2425
- Frank TS, Deffenbaugh AM, Reid JE et al (2002) Clinical characteristics of individuals with germline mutations in *BRCA1* and *BRCA2*: analysis of 10, 000 individuals. *J Clin Oncol* 20(6):1480–1490
- Parmigiani G, Berry D, Aguilar O (1998) Determining carrier probabilities for breast cancer-susceptibility genes *BRCA1* and *BRCA2*. *Am J Hum Genet* 62(1):145–158
- Evans DG, Eccles DM, Rahman N et al (2004) A new scoring system for the chances of identifying a *BRCA1/2* mutation outperforms existing models including BRCA PRO. *J Med Genet* 41(6):474–480
- Antoniou AC, Pharoah PP, Smith P, Easton DF (2004) The BOADICEA model of genetic susceptibility to breast and ovarian cancer. *Br J Cancer* 91(8):1580–1590
- Statement of the American Society of Clinical Oncology (1996) Genetic testing for cancer susceptibility. *J Clin Oncol* 14(5): 1730–1736 (discussion 1737–1740)
- American Society of Clinical Oncology policy statement update (2003) Genetic testing for cancer susceptibility. *J Clin Oncol* 21(12): 2397–2406
- Kwaliteitsinstituut voor de Gezondheidszorg CBO, Vereniging van Integrale Kankercentra. Richtlijn mammacarcinoom. Utrecht, 2007
- National Institute for health and clinical Excellence (NICE). (2006) Familial breast cancer—NICE clinical guideline 41. [www.nice.org.uk](http://www.nice.org.uk)
- Hogervorst FB, Nederlof PM, Gille JJ et al (2003) Large genomic deletions and duplications in the *BRCA1* gene identified by a novel quantitative method. *Cancer Res* 63(7):1449–1453
- Berry DA, Iversen ES Jr, Gudbjartsson DF et al (2002) BRCA PRO validation, sensitivity of genetic testing of *BRCA1/2*, and prevalence of other breast cancer susceptibility genes. *J Clin Oncol* 20(11):2701–2712
- James PA, Doherty R, Harris M et al (2006) Optimal selection of individuals for *BRCA* mutation testing: a comparison of available methods. *J Clin Oncol* 24(4):707–715
- Bodmer D, Ligtenberg MJ, van der Hout AH et al (2006) Optimal selection for *BRCA1* and *BRCA2* mutation testing using a combination of ‘easy to apply’ probability models. *Br J Cancer* 95(6):757–762
- Antoniou AC, Hardy R, Walker L et al (2008) Predicting the likelihood of carrying a *BRCA1* or *BRCA2* mutation: validation of BOADICEA, BRCA PRO, IBIS, Myriad II and the Manchester scoring system using data from UK genetics clinics. *J Med Genet* 45(7):425–431
- Lindor NM, Lindor RA, Apicella C et al (2007) Predicting *BRCA1* and *BRCA2* gene mutation carriers: comparison of LAMBDA, BRCA PRO, Myriad II, and modified Couch models. *Fam Cancer* 6(4):473–482
- Goldgar DE, Easton DF, Deffenbaugh AM et al (2004) Integrated evaluation of DNA sequence variants of unknown clinical significance: application to *BRCA1* and *BRCA2*. *Am J Hum Genet* 75(4):535–544

28. Gómez García EB, Ambergen T, Blok MJ, van den Wijngaard A (2005) Patients with an unclassified genetic variant in the BRCA1 or BRCA2 genes show different clinical features from those with a mutation. *J Clin Oncol* 23(10):2185–2190
29. Gómez García EB, Oosterwijk JC, Timmermans M et al (2009) A method to assess the clinical significance of unclassified variants (UVs) in the *BRCA1* and *BRCA2* genes based on cancer family history. *Breast Cancer Res* 11(1):1–12
30. Mirkovic N, Marti-Renom MA, Weber BL et al (2004) Structure-based assessment of missense mutations in human BRCA1: implications for breast and ovarian cancer predisposition. *Cancer Res* 64(11):3790–3797
31. Judkins T, Hendrickson BC, Deffenbaugh AM et al (2005) Application of embryonic lethal or other obvious phenotypes to characterize the clinical significance of genetic variants found in trans with known deleterious mutations. *Cancer Res* 65(21):10096–10103
32. Chenevix-Trench G, Healey S, Lakhani S et al (2006) Genetic and histopathologic evaluation of BRCA1 and BRCA2 DNA sequence variants of unknown clinical significance. *Cancer Res* 66(4):2019–2027
33. Wu K, Hinson SR, Ohashi A et al (2005) Functional evaluation and cancer risk assessment of BRCA2 unclassified variants. *Cancer Res* 65(2):417–426
34. Farrugia DJ, Agarwal MK, Pankratz VS et al (2008) Functional assays for classification of BRCA2 variants of uncertain significance. *Cancer Res* 68(9):3523–3531