Molecular cloning and sequencing of chicken liver fatty acid synthase cDNA

(multienzyme complex/enzyme structure/protein sequence)

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ABSTRACT The complete amino acid sequence of chicken liver fatty acid synthase [acyl-CoA:malonyl-CoA C-acyltransferase (decarboxylating, oxoacyl- and enoyl-reducing, and thioester-hydrolyzing), EC 2.3.1.85] has been determined from the corresponding cDNA sequence. A 5.3-kilobase-pair (kbp) region of cDNA coding for chicken fatty acid synthase has been cloned and sequenced that is contiguous to the 2.3-kbp region previously sequenced [Yuan, Z., Liu, W. & Hammes, G. G. (1988) Proc. Natl. Acad. Sci. USA 85, 6328-6331]. The cDNA codes for the remaining 1677 amino acids of the previously unsequenced region of the protein. The amino acid sequence contains peptides known to be associated with the NADPH binding site of the enoylreductase active center, the acetyl/ malonyltransacylase active site, the "waiting" site containing cysteine, and a pyridoxal 5'-phosphate binding site. Locations of the NADPH binding site of the β -ketoacylreductase active site and of the dehydratase active site are proposed on the basis of protein sequence homologies to catalytic sites in other enzymes. The molecular weight of the complete polypeptide chain is 267,288. A linear functional map of the chicken fatty acid synthase derived from its primary sequence is presented.

Chicken fatty acid synthase [acyl-CoA:malonyl-CoA C-acyltransferase (decarboxylating, oxoacyl- and enoyl-reducing, and thioester-hydrolyzing), EC 2.3.1.85] is a multienzyme complex responsible for the synthesis of palmitic acid from acetyl-CoA and malonyl-CoA with NADPH as the reducing agent. The kinetics and stereochemistry of many of the six separate enzymatic activities of the complex have been described (1). The complex consists of two identical polypeptides of $M_r \approx 250,000$ (2). Limited tryptic digests of chicken liver fatty acid synthase lead to the identification of three major functional domains (2-4). Domain I, M_r 127,000, carries the acetyl- and malonyltransacylases as well as the β -ketoacylsynthase activities. Domain II, M_r 107,000, contains the dehydratase, β -ketoacylreductase, and enoylreductase activities as well as the acyl carrier protein. Domain III, M_r 33,000, carries the thioesterase active site. These molecular weights require a cDNA of about 7 kilobase pairs (kbp) to code for the entire polypeptide chain. The cDNA sequence of a 2.3-kbp 3'-terminal fragment of the fatty acid synthase coding region has been presented (5). The translated amino acid sequence contains peptides identified to be parts of the acyl carrier 4'-phosphopantetheine and thioesterase active sites. In this work the cDNA sequence of an additional 5.3 kbp, 5'-terminal to the previously described fragments, coding for domains I and II is presented.*

MATERIALS AND METHODS

Most of the materials and methods used in this work have been described (5-7). Escherichia coli strains used were JM101 and JM109 for M13 phage manipulation (8), Y1090 for λ gt11 phage library screening (9), and C600 and C600 *hfl* for work with λ gt10 phage (9).

cDNA libraries were prepared by Clontech from mRNA isolated from adult Leghorn rooster livers. The first-strand synthesis reaction for each library was primed with oligode-oxyribonucleotides synthesized by using sequence information from the 5' terminus of the clone obtained from the preceding library.

RESULTS AND DISCUSSION

Repeated screening of the first library, which was constructed in phage λ gt11 by priming the reverse transcription reaction with two oligodeoxyribonucleotides found 130 and 310 bp, respectively, from the 5' terminus of the previously determined fatty acid synthase cDNA sequence (5), yielded 75 clones, which were analyzed by *Eco*RI restriction mapping. Selected clones were analyzed by Southern blotting (10). All clones hybridizing the synthetic oligodeoxyribonucleotide used for library screening contained an *Eco*RI insert ≈800 bp long. This fragment was subcloned and sequenced by the strategy shown in Fig. 1. The sequence of the fragment (Fig. 2) was found to overlap the 5' end of the previously published chicken fatty acid synthase partial cDNA sequence (5).

Two oligodeoxyribonucleotides with sequences from the 5' end of the 800-bp sequence described above were used to prime first-strand cDNA synthesis in the construction of a second library. The mRNA was denatured with 2 mM methylmercuric hydroxide prior to cDNA synthesis to remove possible secondary structures that may have been responsible for consistent termination of the first-strand synthesis reaction 800 bp away from the priming site in the construction of the first library. The cDNA was subcloned into $\lambda gt10$ and amplified once. Screening with a synthetic oligodeoxyribonucleotide, which hybridizes to the noncoding strand of the fatty acid synthase cDNA in the 5' region of the fragment cloned from the first library, yielded >20 independent clones. The sizes of the inserts in $\lambda gt10$ estimated by EcoRI restriction mapping and electrophoresis in 0.7% agarose ranged from 0.9 to 3.1 kbp. Inserts with sizes over 2.6 kbp yielded two non- λ gt10 EcoRI fragments, of which one was 2.6 kbp. Southern analysis of the clones showed that the 2.6-kbp fragment hybridized to the oligodeoxyribonucleotide used to screen the library. This is consistent with the presence of an EcoRI restriction site 2.6 kbp to the 5' side of the library priming site in the fatty acid synthase coding region. Restriction mapping of the longest insert with *HindIII* and *Bgl II* restriction endonucleases yielded the map shown in Fig. 1. The fragments indicated on the map were subcloned into appropriate sites of M13mp19 in the orientations indicated. The Bgl II fragments were subcloned into the BamHI site. Sequencing was performed according to the scheme in Fig. 1, and the resulting sequence is shown in Fig. 2.

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^{*}The sequence reported in this paper has been deposited in the GenBank data base (accession no. M22987).



FIG. 1. Map of the chicken liver fatty acid synthase cDNA fragment. Clones used in sequencing are shown above the restriction map. Solid lines indicate clones in phage λ gt vectors. Lines with arrows indicating orientation represent clones in M13mp19. Dashed lines represent λ gt10 sequences. The sequencing strategy for the cDNA is shown below the restriction map.

dent clones. The longest insert in λ gt10 was 4.9 kbp long and yielded 2.6-kbp and 2.3-kbp non- λ gt10 fragments when digested with *Eco*RI. The 5'-terminal 2.3-kbp fragment was

Screening of the second library with an oligodeoxyribonucleotide found near the 5' terminus of the sequence obtained from the experiment described above, yielded 12 indepen-

10 20 30 40 50 60 70 80 90 AGAACCTGCT CAATGGGTG GAATGGTCA CAGAGGACGA TCGGAGGTGG AAGCCAGGGA TTTATGGACT GCCCAAAAGA AATGGAAAGC 100 110 120 130 140 151 160 175 TCAAGGACAT AAAAAATTC GATGCCTCT TCTTGGGTC CACCCCAAAC AAGCCTCATACA ATG GAT CCT CCA GTT CGC TTG TTG MET ABP PTO PTO VAI ATG LOU LOU $\begin{array}{c} 190 \\ 190 \\ 190 \\ 190 \\ 190 \\ 205 \\ 200 \\ 220 \\ 200 \\ 200 \\ 200 \\ 205 \\ 200 \\ 200 \\ 205 \\ 200 \\$ Glu Gin Gily Val Thr Phe ProSer Gily Glu Met Gil Gin Gin Gin Gin Gir Gor Ter CrG TAC AGA GAA TGT GGT ATC7908058108208357908058208358108158168176178176146176176186176178186176176186176176186176188198198107177176186176186176176188108108158808908959109259109259109258108808959709709859109251000636627727630636636636637727730100063662772763172786774772773073010006366277276327307301000104510601075730103010451060107510601075110011351150741103011051120113511501150115011501150741103011051120113511501150115011501150743148547148Asp ProAsp Pro744149</t Val Ser Asp Leu Leu Asn Ala Asp Giu Asn inr Pne Asp Asp inr val His Ala Pne val Giy Leu Ala Ala 1615 1630 1645 1660 1675 TA CAG ATT GCC CAA ATT GAT GTG GTA AAG GCT GCG GGT GTG CAA CCT GAT GGG ATT TTG GGC CAC TCA GTG GAA Ile Gin Ile Ala Gin Ile Asp Val Leu Lys Ala Ala Giy Leu Gin Pro Asp <u>Giy Ile Leu Giy His Ser Val Giy</u> 1690 1705 1720 1720 1735 1750 GAA CTA GCT TGT GGC TAT GCA GAT AAT TCC TTA AGT CAT GAA GAA GCT GTT CTT GCT GCT TAT TGG AGG GGC CGA <u>Giu Leu Ala Cys Giy Tyr Ala Asp Asn Ser Leu Ser His Giu Giu Ala Val Leu Ala Ala Tyr Trp Arg Giy Arg</u> 1780 1795 1810 1825 TGT GTG AAA GAG GCC AAA TTG CCC CCG GGA GGG ATG GCT GCT GGT CTG ACA TGG GAA GAG GCC Cys Val Lys Giu Ala Lys Leu Pro Pro Giy Giy Met Ala Ala Val Giy Leu Thr Trp Giu Giu Cys Lys Gin Arg

FIG. 2. (Figure continues on the next page.)

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 TGT CCT CCA AAC GTG GTA CCA GCA TGT CAC AAC TCT GAG GAT ACT GTT TCG GGG CCT CT GAT TCG GGG
 GCT CT GCA AAC GTG GTA CCA GCA TGT CAC AAC TCT GAG GAT ACT GTT TCG GGG CCT CT GAT TCG GGG
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 Cys Pro Pro Asn Val Val Pro Ala Cys His Asn Ser Glu Asp Thr Val Thr Val Ser Gly Pro Leu Asp Ser Val 1915
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 TCT GAG TTT GTA ACC AAA CTG AG AGA GAT GGT GTT GCA ACG GGC GAG GCC GAG GTT GCA TTT CAT Ser Glu Phe Val Thr Lys Leu Lys Lys Asp Gly Val Phe Ala Lys Glu Val Arg
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 GTG GAG ATT GCT CCA CAT GCT CTC TTA CAG GCT ATC TTG AGG AGA ATL TTG AAC LA CLT TG AC ACL TG ALT ATL CLA CCT

 Yal Glu Ile Ala Pro His Ala Leu Leu Gln Ala Ile Leu Arg Arg Thr Leu Lys Pro Thr Cys Thr Ile Leu Pro

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 CTG ATG AAG AAG GAC CAC AAA AAT AAC TTG GAG TTC TTC CTA ACG GACA ACT GGA AAG ATT CAT TTA ACT GGG ATA
 Leu Met Lys Lys Asp His Lys Asn Asn Leu Glu Phe Phe Leu Thr Glu Thr Gly Lys Ile His Leu Thr Gly Ile
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 AAT GTT CTT GGA AAT AAC TG TTC CCA CCT GTG GGA ATA CCT GTG GGA ACA CCT CTC ATT TCT CCA TAT
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 AAT GTT CTT GGA AAT AAC TTG TTC CCA CCT GTG GGA TAC CCT GTG GGA ACA CCT CTC ATT TCT CCA TTT TCT CCA T Asn Val Leu Gly Asn Asn Leu Phe Pro Pro Val Glu Tyr Pro Val Pro Val Gly Thr Pro Leu Ile Ser Pro 2440 2455 2470 2485 2
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 Arg Yal Leu Tyr Pro Ala Thr Gly Tyr Leu Val Leu Ala Trp Arg Thr Leu Ala Arg Ser Leu Gly Met Val Met
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 GAA CAA ACA GCT GTT ATG TTT GAA GAA GTT ACA ATC CAT CAG GCA ACT ATC CTT CCC AAA AAG GGA TCA ACA CAG Glu Gln Thr Ala Val Met Phe Glu Glu Val Thr Ile His Gln Ala Thr Ile Leu Pro Lys Lys Gly Ser Thr Gln 2740 2755 2770 2785 2800

 Glu Gln Thr Ala Val Met Phe Glu Glu Yal Thr Ile His Gln Ala Thr Ile Leu Pro Lys Lys Gly Ser Thr Gln
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 Leu Glu Val Arg Ile Met Pro Ala Ser His Ser Phe Glu Val Ser Gly Asn Gly Asn Leu Ala Val Ser Gly Lys
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 ATC TCC CTC CTA GAA AAC GAT GCT CTG AAG AAC TTT CAT AAC CAG CTG GCT GAC TTT CAG AGT CAA GCA AAC GTG
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 CAC GGA GTC AAA CTA GTT ATC CAT GGC CTA GAA ACC AAC GGG GCT GCT GCA GGA GGC TCC CCA CCA CAG AAA GGC
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 CAC GGA GTC AAA CTA GTT ATC CAT GGC CTA GAA ACC AAC GGG GCT GCT GCA GGA GCC CCA CCA CAG AAG GGC
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 His Gly Lau Glu Thr Asn Gly Ala Ala Ala Gly Ser Pro Pro Thr Gln Lys Gly
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 CTT CAG GAA AGT GTA GAA ACC GAT CCG CAT CTG GAA TG GC TH GTG HLEU HIS Ser Glu Leu Glu Gln Ile Val
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 ACT CAG GAA AGT GA CTC CCA GC GAT CCC CCA GAT CTG CAT CTA TA GC TTG CTA AT GGC TTG CAAG GTT GAA GAT GTG GAT CTC
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 ACT CAG GAG AAG ATG CAC CTC CAG GAC GAT CCC CTT CTC AAT GGC TTG CTG GAT TCT TCA GAG TTG AAG ACT TGC

 ACT CAG GAG AAG ATG CAC CTC CAG GAC GAT CCC CTT CTC AAT GGC TTG CTG GAT TCT TCA GAG TTG AAG ACT TGC

 Thr Gin Glu Lys Met His Leu Gin Asp Asp Pro Leu Leu Asn Gly Leu Leu Asp Ser Ser Glu Leu Lys Thr Cys 3640
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 CTG GAT GTG GCA AAG GAG AAC ACG ACC CAGT CAC AGG ATG AAG ATA GTG GAG GCT CTG GCA GGA AGT GGA CCT CTG Leu Asp Val Ala Lys Giu Asn Thr Thr Ser His Arg Met Lys Ile Val Glu Ala Leu Ala Gly Ser Gly Arg Leu 3715
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 TCT CTC GTG GTC CAA AGT ATT CTG AAT ACT CAG CCC CTG TTG CAG GCT CTG GCC ACT GAC TGC CCC TPhe Ser Arg Val Gln Ser Ile Leu Asn Thr GIn Pro Leu Leu Gln Leu Asp Tyr Ile Ala Thr Asp Cys Thr Pro 3790
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 CCC TCT GGA AAT CTG ACC AAT GCT GAC CTG GCA GTA TGC AAC TGT TCA ACA AGT GTT CTG GGG AAC ACA GCT GAA Pro Ser Gly Asn Leu Thr Asn Ala Asp Leu Ala Val Cys Asn <u>Cys Ser Thr Ser Val Leu Gly Asn Thr Ala Glu</u> 3940 3955 3970 3985 4000 ATT ATC TCT AAC TTA GCA GCT GCA GTG AAA GAA GGA GGG TTT GTT TTG CTG CAC ACC CTT CTT AAA GAG GAA
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 GIU GIU Leu Phe Ser Lys Ala Ser Leu Asn Leu Val Ala Met Lys Arg Ser Phe Phe Gly Ser Val Ile Phe Leu

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 TGT CGA CGG CAG TCC CCT GCC AAA GCA CCC ATT CTT CTG CCA GTA GAT GAC ACT CAT TAT AAG TGG GTT GAC TCC
 GCA GTC CCT GCC AAA GCA CCC ATT CTT CTG CCA GTA GAC ACT CAT TAT AAG TGG GTT GAC TCC
 GCA GTC CCT GCC AAA GCA CCC ATT CTT CTG CCA GTA GAC ACT CAT TAT AAG
 GGG GTT GAC GAC TACT CAT CAA GAC CAC TCT GTG TG ACT GCC ACC AAT TGT GGG AAC TCT GCA ATT

 TTA AAG GAG ATC TTG GCT GAC TCA TCA GAG CAG CCT CTG TGG TTG ACT GCC ACC AAT TGT GGG AAC TCT GCA ATT

 CCT TCA TCA ACT GTC CCA GCC ACT AGT CTT TCT TCC CTG GAG ATG CAG AAG ATT ATT GAG AGA GAT CTG GTG ATG

 Pro Ser Thr Yal Pro Ala Thr Ser Leu Ser Ser Leu Glu Met Gln Lys Ile Ile Glu Arg Asp Leu Yal Met

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 AAT GTG TAT CGT GAT GGA AAG TGG GGT TCC TTC AGG CAT CTC CCA TTG CAG CAG CCT CAG GAG CTG ACG

 ASN VAL TYr Arg Asp Giy Lys Trp Giy Ser Phe Arg His Leu Pro Leu Gin Gin Ala Gin Pro Gin Glu Leu Thr

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 6AT GCT TAC GT GAT AGT GTG TGC GTT GAG GAG TCT CTC TCC CTT CGT TGG ATT GTT TCC CCC CTT CGG CAG CAG

 GAT TAT GCC TAC GTA AAT GTG TTG ACT GCT GGA GAT CTC TCT TCC CTT CGT GGA TT GTT TCC CCC CTT CGA CAG

 GLU TYr Ala TYr Val Asn Val Leu Thr Arg Gly Asp Leu Ser Ser Leu Arg Trp Ile Val Ser Pro Leu Arg His

FIG. 2. (Figure continues on the next page.)

FIG. 2. Sequence of the cDNA fragment coding for domains I and II of the chicken liver fatty acid synthase. The underlined DNA region overlaps the previously published sequence (5). Underlined amino acid sequences indicate protein sequences that have been obtained independently and are discussed in the text.

subcloned into M13mp19, and the unknown part was sequenced according to the scheme presented in Fig. 1. The resulting sequence is shown in Fig. 2.

The amino acid sequence encoded by nucleotides 5183-5254 of the sequence reported here matches exactly the 5'-terminal sequence of the fatty acid synthase cDNA fragment reported previously (5). The open reading frame of both sequences is continuous (Fig. 2). The cDNA fragment sequenced by Yuan *et al.* (5) codes for domain III (thioesterase) and the acyl carrier protein region of domain II of fatty acid synthase. The sequence reported here codes for domain I and the remaining part of domain II.

The amino acid sequence coded by nucleotides 5024–5056 corresponds exactly to the sequence of the enoylreductase active site tryptic fragment isolated by Chang and Hammes (11). The amino acid sequence coded by nucleotides 1928– 1954 matches exactly the sequence of the other pyridoxal 5'-phosphate-labeled tryptic fragment of fatty acid synthase (11). The latter fragment is located in domain I of the enzyme in agreement with peptide mapping (4).

The amino acid sequence coded by nucleotides 1652-1678 corresponds exactly to the essential serine site of acetyl/malonyltransacylase of chicken fatty acid synthase (12). This serine "loading" site is located in domain I of fatty acid synthase. The cysteine-containing "waiting" site peptide identified by iodoacetamide labeling (12) and located in domain I (13) is encoded by nucleotides 359-427.

Nucleotides 3185–3202 and 245–265 code for the amino acid sequences Cys-Leu-Asp-Ser-Leu-Lys and Val-Trp-Val-Gly-Ala-Ser-Gly, which match those of the fluorescent fragments isolated from the trypsin digest of chicken fatty acid synthase labeled with the nucleotide analog 2-[(4-bromo-2,3-dioxobu-tyl)thio]-1, N^6 -ethenoadenosine 2,5-diphosphate (S.-I. Chang and G.G.H., unpublished results). The match of the known sequences with the protein sequence deduced from the cDNA

Table 1. Homology between the amino acid sequences of the regions containing the enoylreductase and the proposed β -ketoacylreductase sites*

Active site	Amino acid sequence			
Enoylreductase	MGCRVFATVGSAEKR			
	XX xxxXXXx xxXX			
β-Ketoacylreductase	MGLLPAKGLATVVDCDKR			

*The sequences are aligned to give the best homology. No amino acids are omitted. The symbol – is used to denote a gap in a sequence introduced to improve alignment. Homologous residues are marked with X, and conservative substitutions are marked with x. indicates that the reading frame of the sequence is correct and that no omissions in nucleotides are present.

The chicken liver fatty acid synthase polypeptide contains two NADPH binding sites. The site at the encylreductase active center was found by comparison to a known peptide sequence, as described above. There is no available protein sequence information for the NADPH binding site of the β -ketoacylreductase active center. A homology search between the amino acid sequence surrounding the enoylreductase NADPH binding site and the rest of the translated amino acid sequence of the enzyme was performed. The comparison located a region of extensive homology (Table 1) 70 residues to the N terminus side of the enoylreductase site. The region is encoded by nucleotides 4781-4834 (Fig. 2). This sequence probably represents the β -ketoacylreductase NADPH binding site. Both reductase active centers are located within a polypeptide of M_r 10,000, which is consistent with trypsin-digest data (4). However, the studies of trypsin digestion suggested that enoylreductase is located to the N-terminal side of the β -ketoacylreductase, whereas our results suggest the reverse order (Fig. 3).



FIG. 3. Linear functional map of chicken fatty acid synthase constructed on the basis of primary sequence comparison with known peptide sequences. TE, thioesterase; ACP, acyl carrier protein; ER, enoylreductase; KR, β -ketoacylreductase; DH, dehy-dratase; AT/MT, acetyl/malonyltransacylase; and KS, ketoacylsynthase. The locations of the SH "waiting" site, the OH "loading" site, a pyridoxal 5'-phosphate (PLP) binding site, the 2-[(4-bromo-2,3-dioxobutyl)thio]-1, N^6 -ethenoadenosine 2,5-diphosphate (BDB-TeADP) binding sites, the dehydratase active site, the NADPH binding sites of enoyl reductase and β -ketoacylreductase, the 4'-phosphopantetheine group, and the thioesterase active site are indicated.

Table 2.	Comparison of the primary stru	cture of the propose	d dehydratase site	e of chicken liver	fatty acid synthase
with home	ologous regions from other dehy	dratases			

Dehydratase	Amino acid sequence	Ref.
Chicken liver fatty acid synthase	1247-C S T S V L G N T A E I I S N L A A A	This paper
Yeast fatty acid synthase β subunit	1612-F V D M V L P N T A L K T S I Q H V G	14, 15
Yeast threonine dehydratase ILV1	$132-C - S - \overline{A} G \overline{N H A} N G V \overline{A} F - A A K$	16
Rat liver L-serine dehvdratase SDH2	62-C S – S – A G N – A G M A T A Y A A R	17
	96- P S P L T A G N – A G M A T A Y A A R	18
E. coli biodegradative threonine dehydratase tdc	81-C — – S – A G N H Ā Q G V S L S C A M	19
E. coli threonine dehydratase ilvA	83-Ī T A Š — A G N H Ā Q G V Ā F S S Ā R	*
E. coli D-serine dehydratase	$171 - \mathbf{A} \mathbf{V} \mathbf{G} \mathbf{\overline{S}} \mathbf{T} - \mathbf{\overline{GN}} - \mathbf{\overline{L}} \mathbf{G} \mathbf{L} \mathbf{S} \mathbf{I} \mathbf{G} \mathbf{I} \mathbf{\overline{M}} - \mathbf{\overline{M}}$	20

The sequences are aligned to give the best homology. The symbol – denotes a gap in a sequence introduced to improve alignment. Residues homologous to those in the chicken liver sequence are underlined.

*EMBL/GenBank Genetic Sequence Database (1986) GenBank (Bolt, Beranek, and Newman Labs., Cambridge, MA), Release 58.0, accession no. KO3503.

The chicken liver fatty acid synthase dehydratase site was located by comparison with known dehydratase sequences (Table 2). Nucleotides 3893-3949 code for an amino acid sequence with significant homologies to five other dehydratases. The differences between the metabolic functions and origins of the different enzymes account for the sequence differences. The presence of the common motif Ser-Xaa₀₋₃-(Ala or Leu)-Gly-Asn-(Thr or His)₀₋₁-Ala-Xaa₆₋₇-Ala₁₋₂ points to the importance of these conserved residues in the dehydratase activity of these enzymes. The dehydratase activity is located in domain II of the chicken liver fatty acid synthase in agreement with trypsin digest data (21).

Analysis of the cDNA sequence revealed only one in-frame ATG start codon (nucleotides 149–151) between the sequence (nucleotides 245–265) encoding one of the 2-[(4-bromo-2,3dioxobutyl)thio]-1, N^6 -ethenoadenosine 2,5-diphosphate binding sites and an upstream inframe nonsense codon. Thus, the methionine encoded by this codon must represent the N terminus of the protein, unless it is removed by posttranslational modification. The calculated molecular weight of the polypeptide encoded by the cDNA is 267,288, in agreement with the approximate molecular weight of chicken liver fatty acid synthase determined from the electrophoretic mobilities of the enzyme and its fragments (2, 21).

The cDNA sequence of chicken liver fatty acid synthase, combined with the amino acid sequences of labeled tryptic fragments and computer analysis of the translated sequence, permits construction of the linear functional map of this multienzyme complex that is presented in Fig. 3.

The sequencing of the full cDNA coding for the rat fatty acid synthase has been reported recently in an abstract (22). Analysis of homologies between the chicken and rat fatty acid synthase sequences will yield information about conserved, functionally important domains and about the evolutionary relationship of the two enzymes.

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