## Molecular cloning and sequencing of chicken liver fatty acid synthase cDNA

(multienzyme complex/enzyme structure/protein sequence)

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ABSTRACT The complete amino acid sequence of chicken liver fatty acid synthase [acyl-CoA:malonyl-CoA C-acyltransferase (decarboxylating, oxoacyl- and enoyl-reducing, and thioester-hydrolyzing), EC 2.3.1.851 has been determined from the corresponding cDNA sequence. A 5.3-kilobase-pair (kbp) region of cDNA coding for chicken fatty acid synthase has been cloned and sequenced that is contiguous to the 2.3-kbp region previously sequenced [Yuan, Z., Liu, W. & Hammes, G. G. (1988) Proc. Nall. Acad. Sci. USA 85, 6328-6331]. The cDNA codes for the remaining 1677 amino acids of the previously unsequenced region of the protein. The amino acid sequence contains peptides known to be associated with the NADPH binding site of the enoylreductase active center, the acetyl/ malonyltransacylase active site, the "waiting" site containing cysteine, and a pyridoxal 5'-phosphate binding site. Locations of the NADPH binding site of the  $\beta$ -ketoacylreductase active site and of the dehydratase active site are proposed on the basis of protein sequence homologies to catalytic sites in other enzymes. The molecular weight of the complete polypeptide chain is 267,288. A linear functional map of the chicken fatty acid synthase derived from its primary sequence is presented.

Chicken fatty acid synthase [acyl-CoA: malonyl-CoA C-acyltransferase (decarboxylating, oxoacyl- and enoyl-reducing, and thioester-hydrolyzing), EC 2.3.1.85] is a multienzyme complex responsible for the synthesis of palmitic acid from acetyl-CoA and malonyl-CoA with NADPH as the reducing agent. The kinetics and stereochemistry of many of the six separate enzymatic activities of the complex have been described (1). The complex consists of two identical polypeptides of  $M_r \approx 250,000$  (2). Limited tryptic digests of chicken liver fatty acid synthase lead to the identification of three major functional domains  $(2-4)$ . Domain I,  $M_r$  127,000, carries the acetyl- and malonyltransacylases as well as the  $\beta$ -ketoacylsynthase activities. Domain II,  $M_r$  107,000, contains the dehydratase,  $\beta$ -ketoacylreductase, and enoylreductase activities as well as the acyl carrier protein. Domain III,  $M_r$  33,000, carries the thioesterase active site. These molecular weights require <sup>a</sup> cDNA of about <sup>7</sup> kilobase pairs (kbp) to code for the entire polypeptide chain. The cDNA sequence of a 2.3-kbp 3'-terminal fragment of the fatty acid synthase coding region has been presented (5). The translated amino acid sequence contains peptides identified to be parts of the acyl carrier 4'-phosphopantetheine and thioesterase active sites. In this work the cDNA sequence of an additional 5.3 kbp, 5'-terminal to the previously described fragments, coding for domains <sup>I</sup> and II is presented.\*

## MATERIALS AND METHODS

Most of the materials and methods used in this work have been described (5-7). Escherichia coli strains used were JM101 and JM109 for M13 phage manipulation (8), Y1090 for Agtll phage library screening (9), and C600 and C600 hfl for work with  $\lambda$ gt10 phage (9).

cDNA libraries were prepared by Clontech from mRNA isolated from adult Leghorn rooster livers. The first-strand synthesis reaction for each library was primed with oligodeoxyribonucleotides synthesized by using sequence information from the <sup>5</sup>' terminus of the clone obtained from the preceding library.

## RESULTS AND DISCUSSION

Repeated screening of the first library, which was constructed in phage  $\lambda$ gtll by priming the reverse transcription reaction with two oligodeoxyribonucleotides found 130 and 310 bp, respectively, from the <sup>5</sup>' terminus of the previously determined fatty acid synthase cDNA sequence (5), yielded <sup>75</sup> clones, which were analyzed by EcoRI restriction mapping. Selected clones were analyzed by Southern blotting (10). All clones hybridizing the synthetic oligodeoxyribonucleotide used for library screening contained an  $EcoRI$  insert  $\approx 800$  bp long. This fragment was subcloned and sequenced by the strategy shown in Fig. 1. The sequence of the fragment (Fig. 2) was found to overlap the <sup>5</sup>' end of the previously published chicken fatty acid synthase partial cDNA sequence (5).

Two oligodeoxyribonucleotides with sequences from the <sup>5</sup>' end of the 800-bp sequence described above were used to prime first-strand cDNA synthesis in the construction of <sup>a</sup> second library. The mRNA was denatured with <sup>2</sup> mM methylmercuric hydroxide prior to cDNA synthesis to remove possible secondary structures that may have been responsible for consistent termination of the first-strand synthesis reaction 800 bp away from the priming site in the construction of the first library. The cDNA was subcloned into Agt1O and amplified once. Screening with a synthetic oligodeoxyribonucleotide, which hybridizes to the noncoding strand of the fatty acid synthase cDNA in the <sup>5</sup>' region of the fragment cloned from the first library, yielded >20 independent clones. The sizes of the inserts in  $\lambda$ gt10 estimated by  $EcoRI$  restriction mapping and electrophoresis in 0.7% agarose ranged from 0.9 to 3.1 kbp. Inserts with sizes over 2.6 kbp yielded two non- $\lambda$ gt10 EcoRI fragments, of which one was 2.6 kbp. Southern analysis of the clones showed that the 2.6-kbp fragment hybridized to the oligodeoxyribonucleotide used to screen the library. This is consistent with the presence of an EcoRI restriction site 2.6 kbp to the <sup>5</sup>' side of the library priming site in the fatty acid synthase coding region. Restriction mapping of the longest insert with HindIII and Bgl II restriction endonucleases yielded the map shown in Fig. 1. The fragments indicated on the map were subcloned into appropriate sites of M13mp19 in the orientations indicated. The Bgl II fragments were subcloned into the BamHI site. Sequencing was performed according to the scheme in Fig. 1, and the resulting sequence is shown in Fig. 2.

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<sup>\*</sup>The sequence reported in this paper has been deposited in the GenBank data base (accession no. M22987).



quencing are shown above the restriction map.<br>Solid lines indicate clones in phage  $\lambda$ gt vectors.

Screening of the second library with an oligodeoxyribonu-<br>cleotide found near the 5' terminus of the sequence obtained yielded 2.6-kbp and 2.3-kbp non-Agt10 fragments when dicleotide found near the 5' terminus of the sequence obtained yielded 2.6-kbp and 2.3-kbp non-Agt10 fragments when di-<br>from the experiment described above, yielded 12 indepen-<br>gested with EcoRI. The 5'-terminal 2.3-kbp frag gested with  $EcoRI$ . The 5'-terminal 2.3-kbp fragment was

10 20 20 30 40<br>AGAACCTGCT CAATGGGGTT GATATGGTCA CAGAGGACGA TCGGAGGTGG AAGCCAGGGA TTTATGGACT GCCCAAAAGA AATGGAAAGC<br>100 100 110 120 120 130 130 140 151 151 160 CA GTT CGC TTG 175<br>TCAAGGACAT AAAAAAAATTC GATGCCTCCT TCTTTGGGTC TTG GAA GTT TCT TAT GAG GCT ATT TTG GAT GCT ATT GAT GCC CTC CT GCT GAG GCT TAT ALL GV1 VALUED 250<br>TEP VALUED VALUED LATE CON AGC ATT ATT CON ACT GCC CTC CT GCC ACT GCC ACT ACT GCT GTG<br>TTG GTT GCT GCA AGT GCC TCA GAA GCT GC Leu Thr Lys Lys Ser Met Ala Lys Arg Val Tyr Ala Thr Ile Val Ash Ala Gly Ser Ash Thr Asp Gly Phe Lys<br>GAG CAA GGT GTG ACA TCC CCA TCT GGA GAG ATG CAG CAG CAG CTG GTT GGT TCT CTG TAC AGA GAA TGT GGT ATC<br>Glu Gln Gly Val Thr Ph ATT GTA AAT GTC TTC TGC CAG TGT GAG AGA GAG CCT CTG TTA ATT GGA TCA ACC AAG TCA AAC ATG GGT CAT CCA<br>11e Val Asn Val Phe Cys Gln Cys Glu Arg Glu Pro Leu Leu Ie Gly Ser Thr Lys Ser Asn Met Gly His Pro<br>1000 970 985 GAC CT GT TOT GGG CTT CT AG GAR CTT CT AT CTT CT AT CTT CT AT CHE AND 1005 CT GAR AT CAN CHE AND 1005 CT CAN ONE AND 1005 CT CAN ONE AND 1005 CHE CHE AND 1005 CHE CHE AND 1005 CHE CHE AND 1005 CHE AND 1005 CHE AND 1007 CHE 1615 1630 1645 1660 1675 ATA CAG ATT GCC CAA ATT GAT GTG CTA AAG GCT GCG GGT CTG CAA CCT GAT GGG ATT TTG GGC CAC TCA GTG GGA<br>
110 G1n Ile Ala Gli basp Val Leu 1705<br>
690 120 1720<br>
690 1275 1275 1275 128 128 128 129 1212 1225 1235 1245 1276 1276 128

FIG. 2. (Figure continues on the next page.)

1840<br>Cys Pro Pro AAC GTG GTA CCA GCA TGT CAC ACC TCT GAG GAT ACT GTG GAG CCT CTG GAT TCT GTG<br>Cys Pro Pro Asn Val Val Pro Ala Cys His Asn Ser Glu Asp Thr Val Thr Val Ser Gly Pro Leu Asp Ser Val<br>TCT GAG TTT GTA ACC AAA CTG A Set Glu Phe Val In Lys Leu Lys Lys Glu Val Ine Also Glu Val Arg Set Ala Gly Val Ala Phe His Set Tyr Tyr Me Ala Set IIe Ala Pro COS COST COST ATG CON COST CON CONTRESS Set Tyr Tyr Me Ala Set IIe Ala Pro Ala Leu Leu Set Ala Val Glu Ile Ala Pro His Ala Leu Leu 2005<br>CTG ATG AAG AAG GAC CAC AAA AAT AAC TTG GAG TTC TTC CTA ACG CAG ACT GGA AAG ATT CAT TTA ACT GGG ATA<br>Leu Met Lys Lys Asp His Lys Asn Asn Can Glu Phe Phe Leu Thr Gly Lys Ile His Leu T AAT GTT CTT GGA AAT AAC TTG TTC CCA CCT GTG GAA TAC CCT GTC CCT GTG GGA ACA CCT CTC ATT TCT CCA TAT<br>Asn Val Lou Gly Asn Asn Lou Phe Pro Pro Val Glu Tyr Pro Val Pro Val Gly Thr Pro Lou Ile Ser Pro Tyr 2400 2440<br>ATC AAA TGG GAC CAC AGC CAA GAC TGG GAT GTT CCA AAA GCT GAA GAC TTC CCC TCA GGC TCC AGA GGC TCT CCC<br>11e Lys Trp Asp Tre Hasp Trp Asp Trp A<br>2 TCT GCT TCA GTC TAC AAC ATC GAT GTG AGT CCT GAC TCT CCT GAC CAT TAC TTG GTT GGC CAT TGC ATT GAT GGC<br>Sor Ala Sor Val Tyr Asn Ile Asp Val Sor Pro Asp Sor Pro Asp His Tyr Leu Val Gly His Cys Ile Asp Gly<br>2650 2695 2695 2697 AGA GTC CTG TAC CCA GCA ACT GGG TAC TTA GTG CTG GCG TGG CGA ACT CTG GCA CGA TCT CTT GGC ATG GTC ATG<br>Arg Val Leu Tyr Pro Ala Thr Gly Tyr Leu Val Leu Ala Trp Arg Thr Leu Ala Arg Ser Leu Gly Met Val Met<br>2725 2725 2865 GAA CA ACA GCT GTT ATG TTT GAA GAA GTT ACA ATC CAT CAG GCA ACT ATC CTT CCC AAA AAG GGA TCA ACA CAG<br>
SIU GIn Thr Ala Wai Met Phe Glu Glu Glu Thr Ile His Gin Ala Thr Ile Leu Pro Lys Lys Gly Ser Thr Gin<br>
2740 2755 2750 2760 2 ATC TCC CTC CTA GAA AAC GAT GCT CTG AAG AAC TTT CAT AAC CAG CTG GCT GAC TTT CAG AGT CAA GCA AAC GTG<br>11e Ser Leu Leu G1u Asn Asp Ala Leu 2905<br>2990 2905 2920<br>ACT GCG AAG TCT GGC CTC TTG ATG GAA GAT GTT TAC CAA GAG CTG CAT CT Phe Gln Gly Val Leu Glu Cys Asn Ser Glu Gly Ser Ala Gly Lys Ile Leu Trp Asn Gly Asn Trp Val Thr Phe<br>3055 3040<br>CTT GAC ACC CTG CA CAC TTG ATA GTC TTA GCA GAG ACT GGG CACT CTA CGA TTG CCC AGC AGO ATT CGC TCA Leu Asp Thr Leu Leu His Leu Ile Val Leu Ala Glu Thr Gly Arg Ser Leu Arg Leu Pro Thr Arg Ile Arg Ser<br>3115 - 3130<br>GTG TAT ART GAC CCT GTG CTT CAT CAG GAG GTG TAC CAG TAC CAG GAC AAT GTA GAA GCT TTT GAT GTT GTT<br>Val Tyr Ile As 3250 3250<br>GTT GAC CGC TGT CTT GAT AGC CTC AAA GCA GGA GGT GTT CAG ATC AAT GGA CTT CAT GCC TCG GTG GCA CCA CCG<br>Val Asp Arg <u>Cys Leau Asp Ser Leu Lys</u> Ala Gly Gly Val Gln Ile Asn Gly Leu His Ala Ser Val Ala Pro Arg<br>3265 3265 CGA CAA CAG GAG COG ATC TCT CCC ACT CTG GAA AAA TTC TCC TTT OTT CCC TAT ATT GAO AOT GAC TOT TTO TCT Arg Gln Gln Glu Arg Ile Ser Pro Thr Leu Glu Lys Phe Ser Phe Val Pro Tyr Ile Glu Ser Asp Cys Leu Ser 3340 3355 3370 3385 3400 TOC AGT ACC CAS CAS CAT CAT CAT CAT CAT CAT CAT CAS ARE CAN ACC ARE CAN ACC AND CAN ACC AND STATES OF A SAN CAN CAN ACC AND STATES OF A SAN ACC AND ST Pro Ser Gly Asn Leu Thr Asn Ala Asp Leu Ala Val Cys Asn <u>Cys Ser Thr Ser Val Leu Gly Asn Thr Ala Glu</u><br>3940 3995 3985 4000 ATT ATC TCT AAC TTA GCA GCT GCA GTG AAA GAA GGA GGG TTT GTT TTG CTG CAC ACC CTT CTT AAA GAG GAA ACT ALA ALA ALA<br>1015 11e Ser Asn Leu Ala Ala Ala Ala Val Vys Glu Gly Oly Phe Val Leu Leu His Thr Leu Leu Lys Glu Glu 4075<br>4030 CTT GGA GAA ATT GTC AGC TTT CTT ACA AGT CCA GAC CTA CAG CAA GAG CAC AGC TTC CTG TCT CAG GCA CAG TGG<br>Lou Gly Glu Ile Val Ser Phe Lou Thr Ser Pro Asp Leu Gln Gln Glu His Ser Phe Leu Ser Gln Ala Gln Trp 4150<br>
GAG GAG TTA TTC AGC AAG GCC TCA TTG ATG THE GCC AGA ATC TTC TTT GCC TCA GCC TCA TTAT TTC CTG CTG ATG<br>
4150<br>
G1u G1u Leu Phe Ser Lys Ala Ser Leu Asn Leu Val Ala Met Lys Arg Ser Phe Phe Gly Ser Val Ile Phe Leu<br>
4195<br> Pro Ser Ser Thr Val Pro Ala Thr Ser Leu Ser Ser Leu Glu Met Gln Lys Ile Ile Glu Arg Asp Leu Val Met<br>4485 4495 4525 4465 4480 4489 4495 Asn Val Tyr Arg Asp Gly Lys Trp Gly Ser Phe Arg His Leu Pro Leu Gln Gln Ala Gln Pro Gln Glu Leu Thr<br>4500 4540<br>GAA TAT GCC TAG AAT GTG TTG ACT COSA GAT CTC TCT TCC CTT CGT TGG ATT TCC CCA CTT CGA CAC<br>Glu Tyr Ala Tyr Val Asn

4615<br>TTC CAA ACA ACC AAT CCA AAT GTT CAG CTC TGC AAA GTC TAC TAT GCA TCT CTC AAT TTC CGG GAC ATT ATG CTG<br>Phe Gln Thr Thr Asn Pro Asn Val Gln Leu Cys Lys Val Tyr Tyr Ala Ser Leu Asn Phe Arg Asp Ile Met Leu<br>GCA ACA GGA AAG C Ala Thr Gly Lys Lou Ser Pro Asp Ala Ile Pro Gly Asn Trp Thr Lou Gln Gln Cys Met Lou Gly Met Glu Phe 4785 4810<br>TCA GGA CGG GAC CTG GCT GGA AGG AGA GTG ATG GGA TTG CTG CCA GCA AAA GGG CTG GCG ACA GTG GTG GAC TGT Ser Gly Arg Asp Lou Ala Gly Arg Arg Val Met GlY Lou Lou Pro Ala Lvs Glv Lou Ala Thr Val Val Asp Cys 4840 4855 4870 4885 4900 GAC AAG AGG TTT CTA TGG GAA GTG CCT GAA AAC TGG ACT CTG GAA GAA GCA GCT TCG GTG CCT GTG GTT TAT GCC ASD Lys Ara Phe Lou Trp Glu Val Pro Glu Asn Trp Thr Lou Glu Glu Ala Ala Ser Val Pro Val Val Tyr Ala 4915 4915<br>ACT GCT TAT TAT GCT TTG GTG GTT CGA GGT GCT ATG AAG AAG GGG GAG AGT GTC CTC ATT CAC TCT GGC TCA GGA<br>Thr Ala Tyr Tyr Ala Leu Val Val Arg Gly Gly Met Lys Lys Gly Glu Ser Val Leu Ile His Ser Gly Ser Gly<br>Thr Ala Tyr GGT GTG GGC CAA GCA GCC ATT GCC ATC GCC TTG AGC ATG GGC TGC CGT GTT TTT GCT ACT GTA GGC TCT GCT GAG Gly Val Gly Gln Ala Ala Ile Ala Ile Ala Lou Ser Met Gly Cys Arg Val Phe Ala Thr Val Glv Ser Ala Glu 5065 5080 5095 5110 5125 AAA CGT GAG TAT CTC CAA GCA AGG TTC CCA CAG CTG GAT GCT AAT AGC TTT GCC AGC TCC CGA AAT ACA ACC TTT Lys Ara Glu Tyr Lou Gln Ala Arg Phe Pro Gln Lou Asp Ala Asn Ser Phe Ala Ser Ser Arg Asn Thr Thr Phe 5140 5155 5170 5185 5200 GAG CAA CAC ATA CTO CGA GTT ACC AAT GGG AAA GOT GTC AAC CTT GTG TTA AAT TCC TTG GCA GAA GAG AAG CTC Glu Gln His Ile Lou Arg Val Thr Asn Gly Lys Gly Val Asn Lou Val Lou Asn Ser Lou Ala Glu Glu Lys Lou 5215 5230 5245 5254 CAA GCC AGT TTG CGT TGT CTT GCT CAA CAT GGO CGC TTC TTG GAA ATA GGC AAA Gln Ala Ser Lou Arg Cys Lou Ala Gln His Gly Arg Phe Lou Glu Ile Gly Lys

FIG. 2. Sequence of the cDNA fragment coding for domains <sup>I</sup> and II of the chicken liver fatty acid synthase. The underlined DNA region overlaps the previously published sequence (5). Underlined amino acid sequences indicate protein sequences that have been obtained independently and are discussed in the text.

subcloned into M13mpl9, and the unknown part was sequenced according to the scheme presented in Fig. 1. The resulting sequence is shown in Fig. 2.

The amino acid sequence encoded by nucleotides 5183- 5254 of the sequence reported here matches exactly the <sup>5</sup>'-terminal sequence of the fatty acid synthase cDNA fragment reported previously (5). The open reading frame of both sequences is continuous (Fig. 2). The cDNA fragment sequenced by Yuan et al. (5) codes for domain III (thioesterase) and the acyl carrier protein region of domain II of fatty acid synthase. The sequence reported here codes for domain <sup>I</sup> and the remaining part of domain II.

The amino acid sequence coded by nucleotides 5024-5056 corresponds exactly to the sequence of the enoylreductase active site tryptic fragment isolated by Chang and Hammes (11). The amino acid sequence coded by nucleotides 1928- 1954 matches exactly the sequence of the other pyridoxal 5'-phosphate-labeled tryptic fragment of fatty acid synthase (11). The latter fragment is located in domain <sup>I</sup> of the enzyme in agreement with peptide mapping (4).

The amino acid sequence coded by nucleotides 1652-1678 corresponds exactly to the essential serine site of acetyl/ malonyltransacylase of chicken fatty acid synthase (12). This serine "loading" site is located in domain <sup>I</sup> of fatty acid synthase. The cysteine-containing "waiting" site peptide identified by iodoacetamide labeling (12) and located in domain <sup>I</sup> (13) is encoded by nucleotides 359-427.

Nucleotides 3185-3202 and 245-265 code for the amino acid sequences Cys-Leu-Asp-Ser-Leu-Lys and Val-Trp-Val-Gly-Ala-Ser-Gly, which match those of the fluorescent fragments isolated from the trypsin digest of chicken fatty acid synthase labeled with the nucleotide analog 2-[(4-bromo-2,3-dioxobutyl)thio]-1, $N<sup>6</sup>$ -ethenoadenosine 2,5-diphosphate (S.-I. Chang and G.G.H., unpublished results). The match of the known sequences with the protein sequence deduced from the cDNA

Table 1. Homology between the amino acid sequences of the regions containing the enoylreductase and the proposed  $\beta$ -ketoacylreductase sites\*

Active site	Amino acid sequence
Enovireductase	MG---CRVFATVGSAEKR
	xxxx X X X x x x X X XX.
$\beta$ -Ketoacylreductase	MGLLPAKGLATVVDCDKR

\*The sequences are aligned to give the best homology. No amino acids are omitted. The symbol - is used to denote a gap in a sequence introduced to improve alignment. Homologous residues are marked with X, and conservative substitutions are marked with x.

indicates that the reading frame of the sequence is correct and that no omissions in nucleotides are present.

The chicken liver fatty acid synthase polypeptide contains two NADPH binding sites. The site at the enoylreductase active center was found by comparison to a known peptide sequence, as described above. There is no available protein sequence information for the NADPH binding site of the  $\beta$ -ketoacylreductase active center. A homology search between the amino acid sequence surrounding the enoylreductase NADPH binding site and the rest of the translated amino acid sequence of the enzyme was performed. The comparison located a region of extensive homology (Table 1) <sup>70</sup> residues to the N terminus side ofthe enoylreductase site. The region is encoded by nucleotides 4781-4834 (Fig. 2). This sequence probably represents the  $\beta$ -ketoacylreductase NADPH binding site. Both reductase active centers are located within a polypeptide of  $M_r$  10,000, which is consistent with trypsin-digest data (4). However, the studies of trypsin digestion suggested that enoylreductase is located to the N-terminal side of the  $\beta$ -ketoacylreductase, whereas our results suggest the reverse order (Fig. 3).



FIG. 3. Linear functional map of chicken fatty acid synthase constructed on the basis of primary sequence comparison with known peptide sequences. TE, thioesterase; ACP, acyl carrier protein; ER, enoylreductase; KR,  $\beta$ -ketoacylreductase; DH, dehydratase; AT/MT, acetyl/malonyltransacylase; and KS, ketoacylsynthase. The locations of the SH "waiting" site, the OH "loading" site, a pyridoxal 5'-phosphate (PLP) binding site, the 2-[(4-bromo-2,3-dioxobutyl)thio]-1, $N<sup>6</sup>$ -ethenoadenosine 2,5-diphosphate (BDB-TeADP) binding sites, the dehydratase active site, the NADPH binding sites of enoyl reductase and  $\beta$ -ketoacylreductase, the 4'phosphopantetheine group, and the thioesterase active site are indicated.





The sequences are aligned to give the best homology. The symbol - denotes a gap in a sequence introduced to improve alignment. Residues homologous to those in the chicken liver sequence are underlined.

\*EMBL/GenBank Genetic Sequence Database (1986) GenBank (Bolt, Beranek, and Newman Labs., Cambridge, MA), Release 58.0, accession no. K03503.

The chicken liver fatty acid synthase dehydratase site was located by comparison with known dehydratase sequences (Table 2). Nucleotides 3893-3949 code for an amino acid sequence with significant homologies to five other dehydratases. The differences between the metabolic functions and origins of the different enzymes account for the sequence differences. The presence of the common motif  $\text{Ser-Xaa}_{0-3}$ -(Ala or Leu)-Gly-Asn-(Thr or  $His$ )<sub>0-1</sub>-Ala-Xaa<sub>6-7</sub>-Ala<sub>1-2</sub> points to the importance of these conserved residues in the dehydratase activity of these enzymes. The dehydratase activity is located in domain II of the chicken liver fatty acid synthase in agreement with trypsin digest data (21).

Analysis of the cDNA sequence revealed only one in-frame ATG start codon (nucleotides 149-151) between the sequence (nucleotides 245-265) encoding one of the 2-[(4-bromo-2,3 dioxobutyl)thio]-l,N6-ethenoadenosine 2,5-diphosphate binding sites and an upstream inframe nonsense codon. Thus, the methionine encoded by this codon must represent the N terminus of the protein, unless it is removed by posttranslational modification. The calculated molecular weight of the polypeptide encoded by the cDNA is 267,288, in agreement with the approximate molecular weight of chicken liver fatty acid synthase determined from the electrophoretic mobilities of the enzyme and its fragments (2, 21).

The cDNA sequence of chicken liver fatty acid synthase, combined with the amino acid sequences of labeled tryptic fragments and computer analysis of the translated sequence, permits construction of the linear functional map of this multienzyme complex that is presented in Fig. 3.

The sequencing of the full cDNA coding for the rat fatty acid synthase has been reported recently in an abstract (22). Analysis of homologies between the chicken and rat fatty acid synthase sequences will yield information about conserved, functionally important domains and about the evolutionary relationship of the two enzymes.

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