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## $n \rightarrow \pi^*$ Interaction and $n(\pi)$ Pauli Repulsion Are Antagonistic for Protein Stability

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The interplay between electronic effects and steric effects underlies molecular conformation. For example, the common C=O $\cdots$ H–N hydrogen bonds within protein main chains may be viewed as favored by the delocalization of an oxygen lone pair ( $n$ ) into the antibonding orbital ( $\sigma^*$ ) of the N–H bond, but disfavored by Pauli repulsion<sup>1</sup> between  $n$  and the N–H bonding orbital ( $\sigma$ ).<sup>2</sup> Here, we report on a second example of this type of dichotomy within protein main chains.

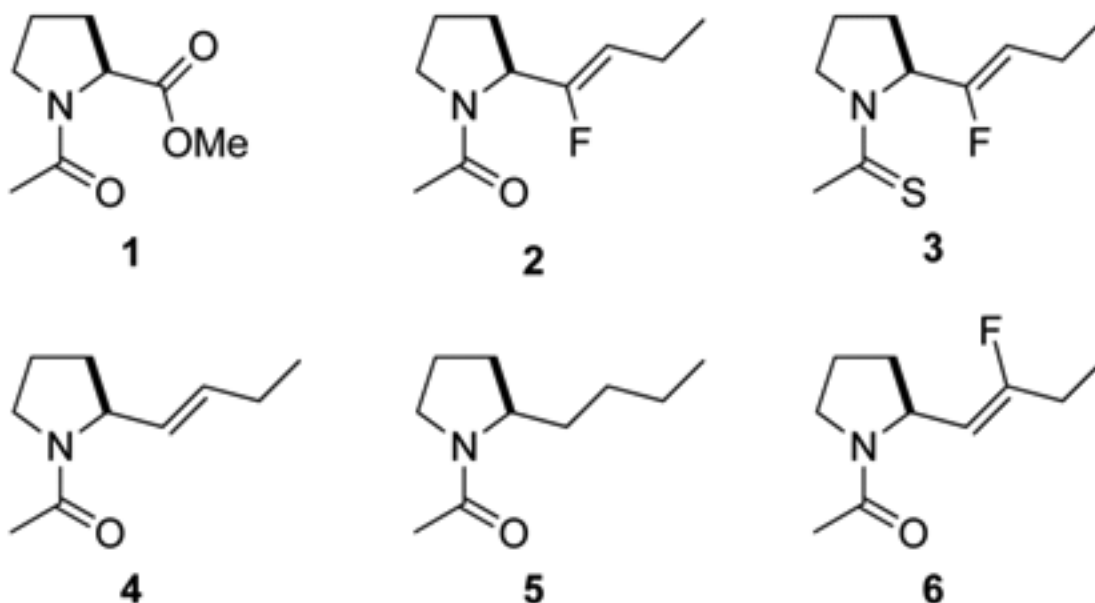
In common elements of protein secondary structure, the oxygen ( $O_{i-1}$ ) of a main-chain amide is proximal to the carbon ( $C_i'$ ) of the subsequent amide.<sup>3</sup> This short contact is promoted by  $n \rightarrow \pi^*$  electronic delocalization, wherein an oxygen lone pair ( $n$ ) overlaps with the  $C_i'=O_i$  antibonding orbital ( $\pi^*$ ) of the subsequent peptide bond.<sup>3–5</sup> We suspected that, as in a hydrogen bond, this electronic effect is antagonized by a steric effect, here arising from Pauli repulsion between  $n$  and the  $C_i'=O_i$  bonding orbital ( $\pi$ ).

To unveil any  $n(\pi)$  Pauli repulsion, we sought a  $\pi$  system that is isosteric with a carbonyl group but provokes little  $n \rightarrow \pi^*$  interaction. We suspected that alkenyl groups, which lack the polarity of carbonyl groups, could have this attribute. To enable quantitative comparisons, we chose the AcProOMe (**1**) model system,<sup>6</sup> in which  $n$  is directed towards  $\pi^*$  in the trans conformation but not in the cis conformation (Figure 1). The value of  $K_{\text{trans/cis}}$  reports on the differential stability of the trans and cis conformations and can be measured by using NMR spectroscopy. We suspected that replacing the ester of **1** with an isosteric fluoroalkene<sup>7</sup> would attenuate the  $n \rightarrow \pi^*$  interaction. Hence, we synthesized and analyzed **1** and its fluoroalkenyl isostere, **2**.

We found evidence that unfavorable Pauli repulsion can indeed antagonize a favorable  $n \rightarrow \pi^*$  interaction. Replacing the carbonyl acceptor with a fluoroalkene switches the conformational preference of the amide bond from trans to cis (Table 1). We resorted to hybrid density functional theory and Natural Bond Orbital (NBO)<sup>8</sup> analyses to reveal the basis for this dramatic shift

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**Supporting Information Available:** Procedures for syntheses and analyses reported herein, and computational data. This material is available free of charge via the internet at <http://pubs.acs.org>.



in conformational preference. We performed geometry optimizations, frequency calculations, and NBO analyses at the B3LYP/6-311+G(2d,p) level of theory on eight conformations of **1** and **2** (see: Supporting Information, Tables S1 and S2). We estimated the stabilization afforded by  $n \rightarrow \pi^*$  electronic delocalization by using second-order perturbation theory, as implemented with NBO 5.0. In accord with our expectation, we found that fluoroalkene isostere **2** does not partake in an appreciable  $n \rightarrow \pi^*$  interaction (Table 1). The  $\pi^*$  orbital of the carbonyl group in **1** is oriented properly for extensive  $n \rightarrow \pi^*$  overlap, but that of the fluoroalkenyl group in **2** is not (Figure 2). Additionally, the energy difference between the  $n$  and the  $\pi^*$  orbitals of **2** (33.2 kcal/mol) is  $\sim 10$ -fold greater than that of **1** (3.5 kcal/mol). While the  $\pi^*$  orbital of the carbonyl is located primarily on the single carbonyl carbon, the  $\pi^*$  of the fluoroalkene isostere is distributed evenly between the two alkenyl carbons. Moreover, the distance between the donor oxygen ( $O_{i-1}$ ) and acceptor carbon ( $C_i'$ ) is short in all low energy conformations of **1** but long in **2** (Table S1). Finally,  $O_{i-1}$  in the low energy conformations of **1** is along the Bürgi–Dunitz trajectory<sup>9</sup> ( $\theta \sim 100^\circ$ ), but  $O_{i-1}$  of **2** is off of that trajectory ( $\theta \sim 125^\circ$ ) (Table S1).

The conformational differences between **1** and **2** are evident in their computational energy landscapes (Figure 3A and 3B). As the value of  $d$  decreases, the interpenetration of the van der Waals surfaces of the donor and acceptor groups increases. That endows **1** but not **2** with conformational stability. In **1**, the  $n(\pi)$  Pauli repulsion is offset by a strong  $n \rightarrow \pi^*$  interaction; in **2**, the  $n \rightarrow \pi^*$  interaction does not overcome that repulsion. Natural Steric Analyses (NSA) supports the existence of the antagonistic Pauli repulsion in low energy conformations (Table S1).

Fluoroalkene **2** lacks a favorable  $n \rightarrow \pi^*$  interaction despite restricted rotation of its  $C^\alpha-C_i'$  bond ( $\psi$  in Figure 1). The anti rotamer is stabilized by a hyperconjugative interaction between the bonding orbital ( $\sigma$ ) of  $C^\alpha-H$  and the antibonding orbital ( $\sigma^*$ ) of  $C_i'-F$  (Figure 4).<sup>10</sup> This rotamer gives rise to a larger value of  $^3J_{H,F}$  for the trans (16 Hz) than the cis (8 Hz) conformation.

If  $n(\pi)$  Pauli repulsion destabilizes the trans conformation of **2**, then its amplification should reduce further the population of that conformation. Some of us had shown previously that the sulfur of a thioamide is a better  $n \rightarrow \pi^*$  donor than is the oxygen of an amide.<sup>4e</sup> But because sulfur is larger than oxygen and C=S bonds are longer than C=O bonds, sulfur should engender

greater  $n(\pi)$  Pauli repulsion. To search for that manifestation, we replaced the donor oxygen ( $O_{i-1}$ ) in amide **2** with sulfur. We found the value of  $K_{\text{trans/cis}}$  for thioamide **3** to be less than that for amide **2** (Table 1). An origin in increased  $n(\pi)$  Pauli repulsion is supported by NSA (Table S1).

Likewise, we reasoned that attenuating any  $n(\pi)$  Pauli repulsion should stabilize the trans conformation. We suspected that a comparison of alkene **4** with alkane **5**, which lacks the acceptor  $\pi$  orbital, would allow us to test our reasoning. Again, we found evidence for  $n(\pi)$  Pauli repulsion, as the value of  $K_{\text{trans/cis}}$  for alkane **5** is greater than that for alkene **4** (Table 1).

Compound **4** offers another opportunity to probe for  $n(\pi)$  Pauli repulsion. The pendant fluoro group that is present in **2** but absent in **4** polarizes the  $\pi$  orbital, reducing the electron density on the acceptor carbon ( $C'_i$ ). The net effect is to diminish  $n(\pi)$  Pauli repulsion as evidenced by a larger value of  $K_{\text{trans/cis}}$  for **2** than **4** (Table 1; Figure 3C). Accordingly, we reasoned that polarizing the  $\pi$  bond in the opposite direction could *increase* the electron density on the acceptor carbon, thereby increasing any  $n(\pi)$  Pauli repulsion. Indeed, the value of  $K_{\text{trans/cis}}$  for **6** is less than that for both **2** and **4**. The correlation between the value of  $K_{\text{trans/cis}}$  for compounds **2**, **4**, and **6** and the  $^{13}\text{C}$  NMR chemical shift of each acceptor carbon (Table 1), which reports on its electron density, provides additional validation for our conclusions.

Some of us have argued<sup>4e</sup> that intimate carbonyl–carbonyl interactions, which are ubiquitous in many protein secondary structures,<sup>3</sup> involve  $n\rightarrow\pi^*$  interactions and cannot be interpreted in terms of classical electrostatic models, such as dipole–dipole<sup>11</sup> or charge–charge interactions.<sup>12</sup> The results herein support this argument. First, if the interaction between adjacent carbonyl groups were manifested as a classical dipole-dipole interaction, replacing the C=O group with an  $C(sp^2)$ –F group would not elicit a reversal in the conformational preference from trans to cis. Second, the value of  $K_{\text{trans/cis}}$  for **3** is less than that for **2**, despite the dipole moment of C=S being greater than that of C=O.<sup>13</sup> Third, the  $\phi$  and dihedral angles of **2** and **4** (which lacks a dipole) are almost identical and are distinct from those of **1** (Table 1; Figure 3C).

The  $O_{i-1}\cdots C'_i=O_i$  distance is especially small in  $\alpha$ -helices.<sup>3</sup> These short contacts position distal C=O and H–N groups in the main chain to form the canonical  $i\rightarrow i+4$  hydrogen bond (Figure 5). Our data indicate that  $n(\pi)$  Pauli repulsion deters such short contacts and would, unless counteracted by an  $n\rightarrow\pi^*$  interaction, impair  $\alpha$ -helix formation. Indeed, others have shown that replacing a single amide bond with an alkene or a fluoroalkene isostere severely disrupts  $\alpha$ -helical structure.<sup>14</sup> Moreover, we put forth  $n(\pi)$  Pauli repulsion as the basis for the anomalous polarization of the  $C'_i=O_i$   $\pi$  bond towards  $O_i$  that has been observed in  $\alpha$ -helices.<sup>15</sup> Analogous repulsion has been observed directly by atomic force microscopy at much larger donor–acceptor distances.<sup>16</sup>

Finally, we note the effect of  $n(\pi)$  Pauli repulsion on the conformation of other molecules. The collagen triple helix has an  $n\rightarrow\pi^*$  interaction between adjacent residues.<sup>17</sup> Each peptide bond in the triplet repeat of collagen strands has been replaced with an alkene isostere, and each substitution greatly diminishes triple-helix stability.<sup>18</sup> Likewise, an altered conformational energy landscape could be responsible for the diminished biological activity of some small-molecule ligands containing an alkene or fluoroalkene isostere.<sup>19</sup> These isosteres appear to be excellent mimics only for amides and esters that are not engaged in  $n\rightarrow\pi^*$  interactions. Implications for structural perturbations within more global elements of protein secondary structure remain an important avenue for further study.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

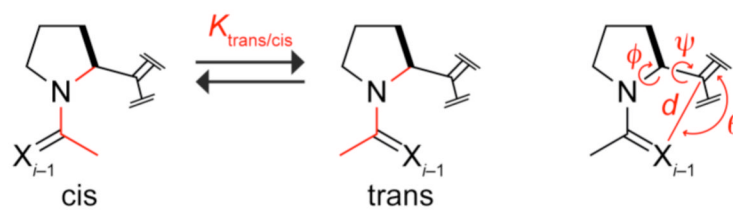
## Acknowledgments

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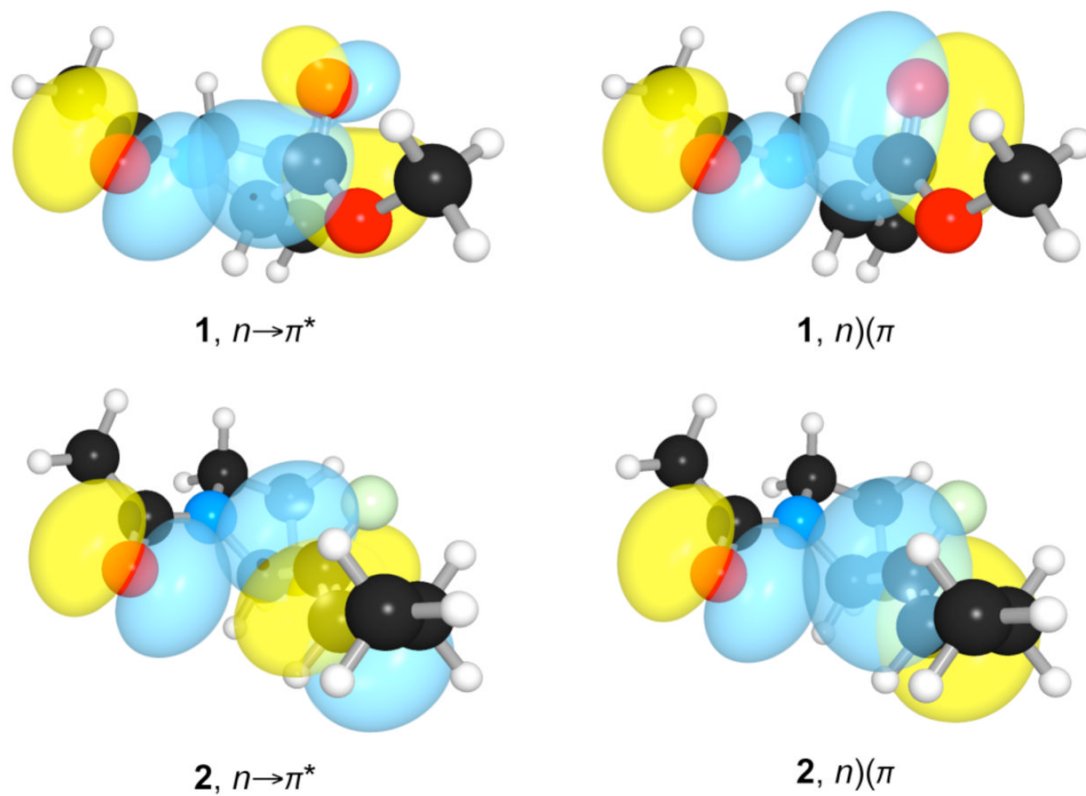
## References

1. (a) Pauli W. Z. Phys 1925;31:373–385. (b) Pauli W. Z. Phys 1925;31:765–783. (c) Massimi, M. Pauli's Exclusion Principle: The Origin and Validation of a Scientific Principle. Cambridge University Press; Cambridge, UK: 2005.
2. (a) Reed AE, Curtiss LA, Weinhold F. Chem. Rev 1988;88:899–926. (b) Weinhold, F.; Landis, CR. Valency and Bonding: A Natural Bond Orbital Donor–Acceptor Perspective. Cambridge University Press; Cambridge, UK: 2005. For another perspective, see: (c) Khaliullin RZ, Bell AT, Head-Gordon M. Chem. Eur. J 2009;15:851–855.
3. Hinderaker MP, Raines RT. Protein Sci 2003;12:1188–1194. [PubMed: 12761389]
4. (a) Bretscher LE, Jenkins CL, Taylor KM, DeRider ML, Raines RT. J. Am. Chem. Soc 2001;123:777–778. [PubMed: 11456609] (b) DeRider ML, Wilkens SJ, Waddell MJ, Bretscher LE, Weinhold F, Raines RT, Markley JL. J. Am. Chem. Soc 2002;124:2497–2505. [PubMed: 11890798] (c) Horng J-C, Raines RT. Protein Sci 2006;15:74–83. [PubMed: 16373476] (d) Hodges JA, Raines RT. Org. Lett 2006;8:4695–4697. [PubMed: 17020280] (e) Choudhary A, Gandla D, Krow GR, Raines RT. J. Am. Chem. Soc 2009;131:7244–7246. [PubMed: 19469574]
5. (a) Sonntag L-S, Schweizer S, Ochsenfeld C, Wennemers H. J. Am. Chem. Soc 2006;128:14697–14703. [PubMed: 17090057] (b) Gao J, Kelly JW. Protein Sci 2008;17:1096–1101. [PubMed: 18434500]
6. As a comparator, we chose a methyl ester rather than an amide to avoid the complications of  $\gamma$ -turn formation, as has been observed in AcProNHMe. See: Liang G-B, Rito CJ, Gellman SH. Biopolymers 1992;32:293–301. [PubMed: 1581548]
7. (a) Abraham RJ, Ellison SLR, Schonholzer P, Thomas WA. Tetrahedron 1986;42:2101–2110. (b) Boros LG, Corte BD, Gimi RH, Welch JT, Wu Y, Handschumacher RE. Tetrahedron Lett 1994;35:6033–6036. (c) Bartlett PA, Otake A. J. Org. Chem 1995;60:3107–3111. (d) Wipf P, Henninger TC, Geib SJ. J. Org. Chem 1998;63:6088–6089. [PubMed: 11672228] (e) Jakobsche CE, Peris G, Miller SJ. Angew. Chem., Int. Ed 2008;47:6707–6711.
8. (a) Weinhold, F. Encyclopedia of Computational Chemistry. Schleyer, P. v. R.; Allinger, NL.; Clark, T.; Gasteiger, J.; Kollman, PA.; Shaefer, HF., III; Schreiner, PR., editors. Vol. 3. John Wiley & Sons; Chichester, UK: 1998. p. 1792-1811. (b) Glendening ED, Badenhoop JK, Reed AE, Carpenter JE, Bohmann JA, Morales CM, Weinhold F. NBO 5.0. 2001
9. (a) Bürgi HB, Dunitz JD, Shefter E. J. Am. Chem. Soc 1973;95:5065–5067. (b) Bürgi HB, Dunitz JD, Lehn JM, Wipff G. Tetrahedron 1974;30:1563–1572. (c) Bürgi HB, Lehn JM, Wipff G. J. Am. Chem. Soc 1974;96:1965–1966. (d) Bürgi HB, Dunitz JD. Acc. Chem. Res 1983;16:153–161. (e) Eliel, EL.; Wilen, SH. Stereochemistry of Organic Compounds. Wiley Interscience Publication; New York: 1996. (f) Kirby, AJ. Stereoelectronic Effects. Oxford University Press; New York: 1996. (g) Clayden, J.; Greeves, N.; Warren, S.; Wothers, P. Organic Chemistry. Oxford University Press; New York: 2000. (h) Anslyn, EV.; Dougherty, DA. Modern Physical Organic Chemistry. University Science Books; Sausalito, California, USA: 2006.
10. Pophristic V, Goodman L. Nature 2001;411:565–568. [PubMed: 11385566]
11. (a) Paulini R, Müller K, Diederich F. Angew. Chem., Int. Ed 2005;44:1788–1805. (b) Fischer FR, Wood PA, Allen FH, Diederich F. Proc. Natl. Acad. Sci. U.S.A 2008;105:17290–17294. [PubMed: 18981424]
12. (a) Maccallum PH, Poet R, Milner-White EJ. J. Mol. Biol 1995;248:361–373. [PubMed: 7739046] (b) Maccallum PH, Poet R, Milner-White EJ. J. Mol. Biol 1995;248:374–384. [PubMed: 7739047]

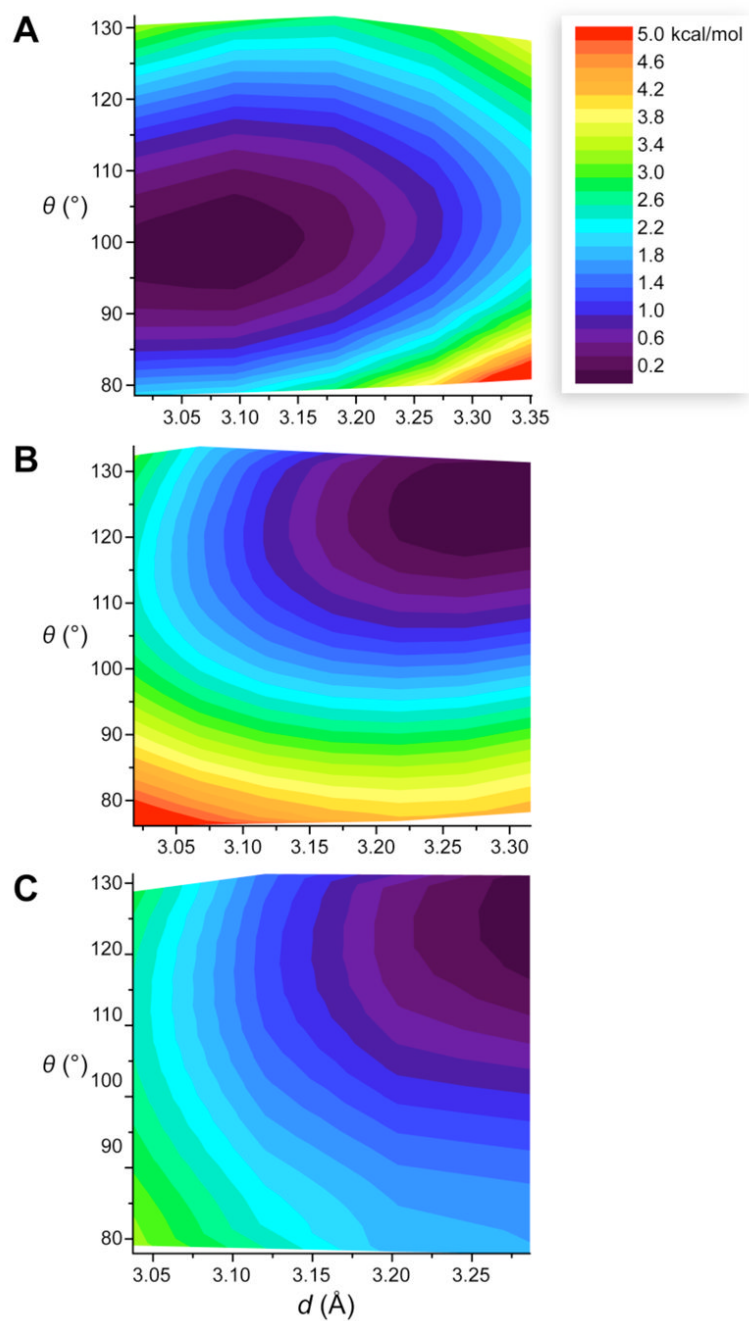
- (c) Allen FH, Baalham CA, Lommerse JPA, Raithby PR. *Acta Cryst. B* 1998;B54:320–329. (d) Deane CA, Allen FH, Taylor R, Blundell TL. *Protein Eng* 1999;12:1025–1028. [PubMed: 10611393]
13. Wiberg KB, Rush DJ. *J. Am. Chem. Soc* 2001;123:2038–2046. [PubMed: 11456827]
14. Oishi S, Kamitani H, Kodera Y, Watanabe K, Kobayashi K, Narumi T, Tomita K, Ohno H, Naito T, Kodama E, Matsuoka M, Fujii N. *Org. Biomol. Chem* 2009;7:2872–2877. [PubMed: 19582296]
15. Lario PI, Vrielink A. *J. Am. Chem. Soc* 2003;125:12787–12794. [PubMed: 14558826]
16. Gross L, Mohn F, Moll N, Liljeroth P, Meyer G. *Science* 2009;325:1110–1114. [PubMed: 19713523]
17. Shoulders MD, Raines RT. *Annu. Rev. Biochem* 2009;78:929–958. [PubMed: 19344236]
18. (a) Jenkins CL, Vasbinder MM, Miller SJ, Raines RT. *Org. Lett* 2005;7:2619–2622. [PubMed: 15957905] (b) Dai N, Wang XJ, Etzkorn FA. *J. Am. Chem. Soc* 2008;130:5396–5397. [PubMed: 18366169] (c) Dai N, Etzkorn FA. *J. Am. Chem. Soc* 2009;131:13728–13732. [PubMed: 19725497]
19. (a) Kaltenbronn JS, Hudspeth JP, Lunney EA, Michniewicz BM, Nicolaidis ED, Repine JT, Roark WH, Stier MA, Tinney FJ, Woo PKW, Essenburg AD. *J. Med. Chem* 1990;33:838–845. [PubMed: 2405159] (b) Fincham CI, Higginbottom M, Hill DR, Horwell DC, O'Toole JC, Ratcliffe JC, Rees DC, Roberts E. *J. Med. Chem* 1992;35:1472–1484. [PubMed: 1573640] (c) Wai JS, Bamberger DL, Fisher TE, Graham SL, Smith RL, Gibbs JB, Mosser SD, Oliff AI, Pompliano DL, Rands E, Kohl NE. *Bioorg. Med. Chem* 1994;2:939–947. [PubMed: 7712129] (d) Venkatesan N, Kim BH. *Curr. Med. Chem* 2002;9:2243–2270. [PubMed: 12470245] (e) Welch, JT. *Fluorine and Health*. Elsevier B. V.; Amsterdam: 2008.



**Figure 1.** Definition of equilibrium constant  $K_{\text{trans/cis}}$ , distance  $d$ , planar angle  $\theta$ , and dihedral angles  $\phi$  and  $\psi$ . X = O in **1**, **2**, and **4-6**; X = S in **3**.



**Figure 2.** Overlap between  $n$  and the  $\pi^*$  and  $\pi$  orbitals of **1** and **2** in their optimized geometries.

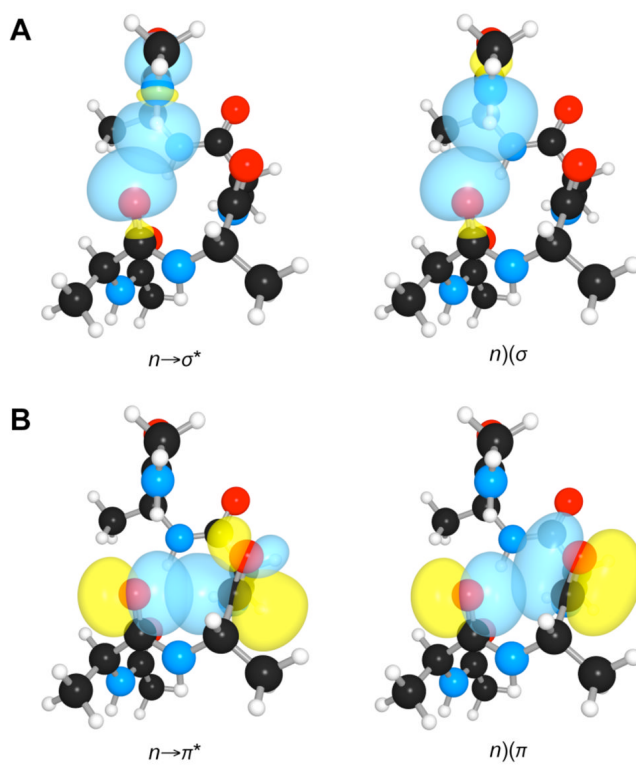


**Figure 3.** Conformational energy landscapes of (A) **1**, (B) **2**, and (C) **4**.





**Figure 4.** Overlap of the  $\sigma$  ( $C^{\alpha}-H$ ) and  $\sigma^*$  ( $C^{\beta}-F$ ) orbitals of **2** in its optimized geometry.



**Figure 5.** Orbital overlaps that stabilize (left) and destabilize (right) the  $\alpha$ -helical conformation of an AcAla<sub>4</sub>NHMe model system. (A)  $i \rightarrow i+4$  hydrogen bond. (B)  $n \rightarrow \pi^*$  interaction.

Table 1

Conformational properties of compounds **1–6**.

Compound	$K_{\text{trans/cis}}^a$	Chemical shift of $C_i$ (ppm)	$d$ (Å) <sup>b</sup>	$\theta$ (°) <sup>b</sup>	$\phi$ (°) <sup>b</sup>	$\psi$ (°) <sup>b</sup>	$n \rightarrow \pi^*$ (kcal/mol) <sup>b</sup>
<b>1</b>	3.7 : 1.0	ND	3.08	99.5	-71.12	152.67	0.40
<b>2</b>	1.0 : 1.7	156	3.28	124.9	-82.81	117.01	0.01
<b>3</b>	1.0 : 2.2	ND	3.59	126.3	-84.42	120.92	0.05
<b>4</b>	1.0 : 2.9	133	3.32	126.4	-84.02	116.56	0.02
<b>5</b>	1.4 : 1.0	ND	—	—	-78.89	167.16	—
<b>6</b>	1.0 : 4.0	105	3.25	104.1	-80.43	142.03	0.03

<sup>a</sup> Measured in CDCl<sub>3</sub> at 25 °C.<sup>b</sup> Computed in the optimized conformations (trans amide bond; C $\gamma$ -endo pyrrolidine ring pucker).