

Online Submissions: http://www.wjgnet.com/1007-9327office wjg@wjgnet.com doi:10.3748/wjg.v16.i20.2504 World J Gastroenterol 2010 May 28; 16(20): 2504-2519 ISSN 1007-9327 (print) © 2010 Baishideng. All rights reserved.

ORIGINAL ARTICLE

Clinicopathological features and prognosis assessment of extranodal follicular dendritic cell sarcoma

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Supported by Grants from National Natural Science Foundation of China, No. 30171052, 30572125 and 30772508

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Telephone: +86-10-87787508 Fax: +86-10-67713359 Received: February 23, 2010 Revised: March 24, 2010 Accepted: March 31, 2010

Published online: May 28, 2010

Abstract

AIM: To establish a model for prognosis assessment of extranodal follicular dendritic cell (FDC) sarcoma.

METHODS: Nine lesions were examined by routine and molecular approaches. Clinicopathological factors from the new cases and 97 reported cases were analyzed for their prognostic values.

RESULTS: The current lesions were found in five male and four female patients, located mainly in the head

and neck area and averaging 7.2 cm in size. Six patients had recurrence or metastasis and three remained free of disease. The 106 patients (male/female ratio, 1.1:1) were aged from 9 to 82 years (median, 44 years). The tumor sizes ranged from 1.5 to 21 cm (mean, 7.4 cm). Abdominal/pelvic region was affected most frequently (43%). Surgical resection was performed in 100 patients, followed by radiation and/or chemotherapy in 35 of them. Follow-up data were available in 91 cases, covering a period of 3-324 mo (mean, 27 mo; median, 19 mo). Of the informative cases, 38 (42%) had recurrence or metastasis, and 12 (13%) died of the disease. These tumors were classified histologically into low- and high-grade lesions. A size \geq 5 cm (P = 0.003), highgrade histology (P = 0.046) and a mitotic count $\ge 5/10$ HPF (P = 0.013) were associated with tumor recurrence. The lesions were defined as low-, intermediateand high-risk tumors, and their recurrence rates were 16%, 46% and 73%, and their mortality rates 0%, 4% and 45%, respectively.

CONCLUSION: Extranodal FDC tumors behave like soft tissue sarcomas. Their clinical outcomes are variable and can be evaluated according to their sizes and grades.

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Key words: Extranodal follicular dendritic cell sarcoma; Prognosis assessment; Histologic grade; Immunohistochemistry; *In situ* hybridization; Mutation detection

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Li L, Shi YH, Guo ZJ, Qiu T, Guo L, Yang HY, Zhang X, Zhao XM, Su Q. Clinicopathological features and prognosis assessment of extranodal follicular dendritic cell sarcoma. *World J Gastroenterol* 2010; 16(20): 2504-2519 Available from: URL: http://www.wjgnet.com/1007-9327/full/v16/i20/2504.htm DOI: http://dx.doi.org/10.3748/wjg.v16.i20.2504



INTRODUCTION

Follicular dendritic cell (FDC) sarcoma was described by Monda *et al*^[1] in 1986. It is considered to be derived from FDC, whose cell type normally forms a tight meshwork in the primary and secondary lymphoid follicles and participates in the immune system by interacting with B or T lymphocytes^[2]. Epstein-Barr virus (EBV) infection was demonstrated in majority of the hepatic and splenic lesions and its causative effect has been proposed for the pathogenesis of the lesion^[3-9], but the association is not evident in most of the tumors from other sites^[10-13]. Majority of the tumors were found in lymph nodes. However, about one-third of the lesions were identified in extranodal sites. It was considered that extranodal FDC sarcoma occurred preferentially in the head and neck area^[14].

FDC sarcoma is usually regarded as an indolent tumor with a tendency of local recurrence but a low risk of metastasis, behaving like a low-grade soft tissue sar $coma^{[2,15]}$. However, Perez-Ordonez *et al*^{16]} considered this tumor more aggressive through their observation of 13 cases. Because of the rarity of the tumor, assessment of its prognosis remains difficult, though intra-abdominal location, a high mitotic count ($\geq 5/10$ HPF), coagulative necrosis and marked cellular atypia were proposed to be predictors of an unfavorable outcome^[3]. With more cases encountered in our pathological practice and reported from literatures, various morphologic and clinical representations were noticed. Clearly, histological grade of this malignancy remains to be defined and a model is needed to evaluate the recurrence risk of individual tumors within this category.

In this study, we presented our experience in nine cases of FDC sarcoma from northern China. In addition, clinicopathological features and corresponding data extracted from 97 reported cases were analyzed, with special respect to prognosis assessment of this disease.

MATERIALS AND METHODS

Samples and histological examination

As listed in Table 1, the tissue specimens were resected from nine patients in three medical centers, including the Fourth Military Medical University Tangdu Hospital in Xi'an (Cases 1 and 2), Chinese Academy of Medical Sciences Cancer Hospital in Beijing (Cases 3-6, 8 and 9) and the First Affiliated Hospital of Inner Mongolia Medical College in Huhehot (Case 7). These patients were admitted for a solid occupation in head and neck areas (n = 6), and thoracic and abdominal cavities (n = 3) from 2002 to 2009. Cases 1 and 2 had been described in a brief report^[17], and follow-up data were complemented in the current study.

The formalin-fixed, paraffin-embedded tissue blocks were retrieved from the files. Sections of 4 μ m in thickness were prepared and stained with hematoxylin and eosin (HE). Three pathologists (Su Q, Yang HY and Li L) reviewed the slides independently, and the diagnosis was confirmed by their histological and immunohistochemical

phenotypes. As described previously^[2], whorl, storiform and fascicular arrangements of spindle tumor cells were regarded as architectural features for typical lesions, while epithelioid and pleomorphic patterns were considered anaplastic phenotypes in this study. Other morphologic parameters, including mitotic activity, coagulative necrosis, nuclear atypia, distribution of infiltrating small lymphocytes and tumor sites, were also assessed. In addition, we also evaluated proportions of the typical tumor components, as described above, in each lesion.

Immunohistochemistry

Immunohistochemical reactions were performed on paraffin sections following deparaffinization, rehydration and antigen retrieval. The antigen retrieval was performed by heating in citrate (pH 6.0) or ethylene-diamine-tetraacetic acid (EDTA) buffer (pH 9.0; Table 2). In addition to routine markers for tumor diagnosis, such as pan-cytokeratins (CKs; AE1/AE3), epithelial membranous antigen (EMA), vimentin, desmin and smooth muscle-type actin (SMA), a panel of antibodies were applied to demonstrate histiocytic and dendritic cell linage differentiation, including those against CD68, S100 protein, CD21, CD23, CD35, CD1a and podoplanin (D2-40). Ki-67 antigen was detected to show proliferative activity of each lesion, and p53 protein accumulation was demonstrated by reaction with the antibody DO-7. After incubation with primary antibodies at room temperature for 1 h, the antigen-antibody reaction was demonstrated by a horseradish peroxidase (HRP)-labeled secondary antibody (EnVisionTM Detection System, Dako A/S, Glostrup, Denmark) at room temperature for 15 min and visualized in a solution containing 0.5 mg/mL 3,3'-diaminobenzidine (DAB) and 0.01% hydrogen peroxide. Finally, the sections were counterstained slightly with hematoxylin and mounted in resin.

The expression levels were evaluated semiquantitatively according to the reaction intensities and percentages of immunoreactive cells, and expressed as strong (3+), moderate (2+), weak (+) and negative (-). Ki-67 antigen expression levels were assessed based on the number of positive cells/1000 nucleated tumor cells from randomly selected 5 high-power fields, and expressed as Ki-67 antigen-labeling indices (Ki-67-LI) in percentages. The p53 protein expression levels were evaluated by percentages of positive cells as described previously^[18] and expressed as high (3+, > 30%), moderate (2+, 5%-30%), low (1+, < 5%) and absent (-, 0%).

In situ hybridization

In situ hybridization was performed in seven cases to demonstrate EBV-encoded RNA (EBER) molecules using a detection kit (Triplex International Biosciences Co. Ltd., Fuzhou, China) according to instructions of the manufacturer. Briefly, sections were treated with proteinase K following deparaffinization and rehydration. After denaturation for 90 min at 55°C, the sections were incubated with a digoxin-labeled DNA probe overnight at 37°C, and washed with phosphate-buffered saline. The hybridization signals were demonstrated by consecutive reactions with



Table I Chinicopathological and therapeutic data of this new cases of extrahoual FDC salconia (Cases F	Table 1 Clin	copathological and	l therapeutic data o	f nine new cases o	f extranodal FDC sarcoma (Cases 1-	9)
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Case No.	Age (yr)	Gender	Sites	Size (cm)	HG	MC (/10 HPF)	Year of onset	Initial diagnoses	Initial treatment	Recurrence, DFT (mo)	Disease status	Follow- up (mo)
1	60	М	Tonsil	5.0	Low	<1	2002	Granuloma	Surg + RT		NED	86
2	35	F	Parapharyn-geal space	5.0	High	10	2003	NPC	Surg	LR, 2	DOD	12
3	63	М	Infratem-poral fossa	4.0	Low	1	2003	PNET	Surg + RT + ChT		NED	72
4	30	F	Pyform sinus	5.0	High	9	2004	FDC sarcoma	Surg + RT	DM (lung), 25	DOD	25
5	23	М	Mediastinum	8.0	Low	3	2006	MPNST	Surg + RT + ChT	DM (bone), 45	AWD	45
6	45	М	Liver	14.5	Low	<1	2007	FDC sarcoma	Surg		NED	27
7	36	F	Mesentery	15.0	High	7	2007	Malignant GIST	Surg	DM (liver, ovary), 4	AWD	27
8	28	F	Parapharyn-geal space	6.0	High	3	2008	FDC sarcoma	Surg + RT + ChT	DM (lung), 14	AWD	22
9	55	М	Tonsil	2.0	High	9	2008	FDC sarcoma	Surg + RT	LR, 18	AWD	21

AWD: Alive with disease; ChT: Chemotherapy; DFT: Disease-free time following operation; DM: Distant metastasis; DOD: Died of disease; GIST: Gastrointestinal stromal tumor; HG: Histological grades; LR: Local recurrence; MC: Mitotic counts; MPNST: Malignant nerve sheath tumor; NED: No evidence of disease; NPC: Nasopharyngeal carcinoma; PNET: Primitive neuroepidermal tumor; RT: Radiotherapy; Surg: Surgery; FDC: Follicular dendritic cell.

Table 2 Primary antibodies and antigen retrieving procedures									
Antibodies	Origins and clones	Dilutions	Antigen retrieval	Sources					
Pan-cytokeratin	MAb, AE1/AE3	1:80	MW-citrate	Invitrogen					
EMA	MAb, Mc5	1:100	MW-citrate	Invitrogen					
Vimentin	MAb, V9	1:200	MW-citrate	NeoMarker					
Desmin	MAb, ZC18	1:80	MW-citrate	Invitrogen					
SMA	MAb, IA4	1:100	Not treated	Invitrogen					
CD45	MAb, RP2/18	1:120	MW-citrate	Novacastro					
CD68	MAb, KP1	1:200	MW-citrate	Zeta					
S-100 protein	PAb, rabbit	1:400	MW-citrate	Dako					
CD21	MAb, 2G9	1:20	AC-EDTA	NeoMarker					
CD23	MAb, SP23	1:25	MW-citrate	NeoMarker					
CD35	MAb, KuN241	1:20	AC-EDTA	NeoMarker					
CD1a	MAb, MTB1	1:20	MW-citrate	Zeta					
Podoplanin	MAb, D2-40	1:50	AC-EDTA	Zeta					
p53 protein	MAb, DO-7	1:100	AC-EDTA	NeoMarker					
Ki-67 antigen	MAb, MIB-1	1:200	AC-EDTA	Zymed					

Antibody suppliers: Dako, Glostrup, Denmark; Invitrogen Corporation, Carlsbad, CA, USA; NeoMarkers Inc., Fremont, CA, USA; Novacastro Laboratories Ltd., Newcastle upon Tyne, UK; Zeta Corporation, Sierra Madre, CA; Zymed Laboratories, San Diego, CA, USA. EMA: Epithelial membrane antigen; MAb: Mouse monoclonal antibody; PAb: Polyclonal antibody; SMA: Smooth muscle-type actin; MW-citrate: Microwaving for 8 min in citrate buffer (pH 6.0); AC-EDTA: Autoclaving for 100 s in EDTA buffer (pH 9.0).

a mouse monoclonal antibody against digoxin and the polymerized HRP-labeled anti-mouse immunoglobulin G, and visualized by incubation in a solution containing DAB and hydrogen peroxidase. Finally, the slides were counterstained slightly with hematoxylin and mounted with resin. A case of Hodgkin lymphoma, known to be positive for EBER, was used as a positive control. Yellow or brown staining of tumor cell nuclei was considered positive.

Amplification and sequencing of P53 gene

As most of the documented mutations of P53 gene occur in its exons $5 \cdot 8^{[19,20]}$, these regions were examined for mutations by nested (for exons 6 and 7) or semi-nested polymerase-chain reaction (PCR; for exons 5 and 8). The primers used were adopted from literatures^[21,22]. Their sequences were as follows: exon5s, 5'-TCTGTCTCCTTCCTTCCTA-3'; exo-

n5as, 5'-AACCAGCCCTGTCGTCTCT-3'; exon6s, 5'-TTGCTCTTAGGTCTGGCCCC-3'; exon6as, 5'-CAGACCTCAGGCGGCTCATA-3'; exon7s, 5'-TT-GCTCTTAGGTCTGGCCCC-3'; exon7as, 5'-GGGT-CAGCGGCAAGCAGAGG-3'; exon8s, 5'-GA-CAAGGGTGGTTGGGAGTAGATG-3'; exon8as, 5'-GCAAGGAAAGGTGATAAAAGTGAA-3'. Selected tumor tissues were collected from serial paraffin sections of 10 µm in thickness. Following deparaffinization and rehydration, the tissues were digested by incubation with proteinase K (20 mg/mL) at 56°C for 72 h. Genomic DNA was extracted with a kit (QIAamp DNA Mini Kit, Qiagen GmbH, Hiden, Germany) following instructions of the manufacturer. Primer pairs used for the first-round amplification of exons 5, 6, 7 and 8 were exon5s/exon6as, exon5s/exon7as, exon6s/exon8as and exon7s/exon8as, respectively. The reactions were in a mixture of 10 µL containing 100 ng of DNA templates, 0.8 µL of deoxynucleotide triphosphate (2.5 mmol/L each), 1 µL of 1 μ mol/L sense and antisense primers each, 1 μ L 10 × buffer (100 mmol/L Tris-HCl, pH 8.3, containing 500 mmol/L KCl and 20 mmol/L MgCl₂) and 0.5 U of Taq DNA polymerase (Takara Biotechnology Co., Ltd., Dalian, China). Amplification was conducted for 35 cycles (94°C, 40 s; 55°C, 40 s; 72°C, 40 s for exons 5 and 7; 1 min for exons 6 and 8) following the initial denaturation at 94°C for 5 min. The final elongation was at 72°C for 10 min. The second-round PCR was performed in a mixture of 50 μ L containing 3 μ L of products of the first-round reaction and 1 µL of 5 µmol/L sense and antisense inner premiers each. Primer pairs used for amplifying exons 5, 6, 7 and 8 were exon5s/5as, exon6s/6sa, exon7s/7as and exon8s/8as, respectively. The cycling conditions were the same as the first-round reaction. Amplification efficiency and specificity were visualized by electrophoresis on an agarose gel (2%), and the products of exons 5, 6, 7 and 8 migrated at positions of 196, 81, 234 and 277 bp, respectively. The products were subjected to direct sequencing using an automatic system (Prism 3100 Genetic Analyzer, Applied Biosystems, Foster, CA, USA) as described previously^[22,23]. The sequences obtained were compared with those from Genbank (www.gdb.org).



Figure 1 Radiological features of extranodal follicular dendritic cell (FDC) sarcoma shown by computed tomographic scan. A: Case 8, a tumor (arrows) at the right parapharyngeal space showing soft tissue-like density and an expansive growth pattern, with the internal and external carotid arteries (arrowheads) engulfed; B: Case 6, a well-circumscribed mass (arrows) at the left lobe of liver, showing irregular enhancement at its periphery.

Literature review

Search of literatures was performed using MEDLINE on PubMed (www.ncbi.nlm.nih.gov/pubmed) with the terms "follicular dendritic cell tumor" or "follicular dendritic cell sarcoma" combined with "extranodal". Articles published in Chinese journals were found by searches on Wanfangdata (www.wanfangdata.com.cn). References of the collected articles were also reviewed to identify other relevant publications. Efforts were made to identify cases that were reported more than once in different settings and only one entry with the most updated information was included for such cases. Nodal tumors were not included in this study.

Statistical analysis

Statistical analyses were carried out using the Software Packages for Social Science 13.0 for Windows (SPSS, Inc, Chicago, IL, USA). Survival of patients was assessed using the data obtained from both the current study and the literatures. Overall and recurrence-free survival rates were calculated and analyzed using the Kaplan-Meier method. Associations of different pathological factors with clinical outcomes were described by the Chi-square test. Fisher's exact test was also used when necessary. *P* values below 0.05 were considered significant.

RESULTS

Current cases

Ages of the nine patients (five men and four women) ranged from 23 to 63 years (median, 36 years). A local mass was the primary complaint in eight cases (Figure 1), and a sore throat was the first symptom in one patient (Case 2). Sizes of the tumors ranged from 2.0 to 15.0 cm as measured by the longitudinal dimension, averaging 7.2 cm. A diagnosis of FDC sarcoma was made in four of the nine cases following pathologic examination of the resected specimens. For Cases 1, 2, 3, 5 and 7, the resected lesions were misdiagnosed as granulomatous inflammation suspicious of tuberculosis, nasopharyngeal carcinoma (NPC), primitive neuroectodermal tumor (PNET), malignant pe-

Table 3 Histological features of low- and high-grade FDC sarcomas

Parameters	Low-grade phenotypes	High-grade phenotypes
Architectures	Nodular patterns in whorl, storiform and fascicular arrangement	Diffuse and sheet-like patterns
Cellular features	Spindle or blunt cells with mild nuclear atypia and a small nucleolus in > 90% of area	Epithelioid or pleomorphic cells with marked nuclear atypia and large nucleoli
Mitotic counts	< 5/10 HPF	$\geq 5/10$ HPF
Ki-67-LI	< 10%	$\geq 10\%$
Necrosis	Absent	Frequently present
Lymphocyte	Sporadic and throughout	Focal or regional
infiltration	the lesion	

ripheral nerve sheath tumor (MPNST) and malignant gastrointestinal stromal tumor (GIST), respectively. The diagnosis of FDC sarcoma was established by re-examination and immunohistochemistry with the delayed periods ranging from one to 66 mo. Their clinicopathological features are listed in Table 1. No precursor lesions were recorded in any of them.

Typically, the lesions were composed of spindle and ovoid tumor cells arranged in whorl, fascicular and storiform patterns, showing mild nuclear atypia and sparkled with small lymphocytes (Figure 2). Of the nine tumors, three (Cases 1, 5 and 6) were composed uniformly of the typical components, and one (Case 3) showed typical components in > 90% of the area and epithelioid appearance in small foci (Figure 3A). In the remaining five cases, large areas of epithelioid (Cases 2 and 8) and pleomorphic tumor cell components (Cases 4, 7-9), arranged in a diffuse or sheet-like pattern and with moderate to marked nuclear atypia, were also identified (Figure 3B-D). Sporadic small lymphocytes were present throughout tumor tissues in all the four typical cases (Figure 2), and this feature was less prominent (Figure 3B) or even absent (Figure 3C and D) in the rest five lesions. Focal coagulative necrosis was identified in six cases (Cases 2, 3, 5-8). The hepatic lesion (Case 6)





Figure 2 Typical features of follicular dendritic cell sarcoma of the conventional (A-F) and inflammatory pseudotumor-like types (G and H). Spindle and ovoid tumor cells, frequently growing in nodules as in Case 1 (A), arrange in whorl (Case 3, B and C) and storiform (Case 3, D) and fascicular patterns (Case 6, H), with sprinkling small lymphocytes throughout the former type of lesions and numerous plasma cells and lymphocytes in the latter. Perivascular sclerosis was noted in Case 3 (E), with foci of osteoid matrix deposition resembling osteosarcoma (F). HE: A, × 40; B, E and G, × 100; C, D, F and H, × 200.



Figure 3 Atypical morphology of FDC sarcoma (A-D) and expression of p53 protein (E) and Epstein-Barr virus-encoded RNA (EBER) (F) in tumor cells. A-D: Epithelioid (A and B) and pleomorphic tumor cells (C and D) are arranged in a sheet-like or diffuse pattern. Lymphocyte infiltration is less prominent (A and B) or absent (C and D) in these areas. HE: A, C and D, × 400; B, × 200; E: Nuclear immunoreactivity for p53 protein in majority of tumor cells. S-P, × 400; F: *In situ* hybridization signal for EBER in tumor cells, × 400.



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Table 4	Expre	ssion of	lineage-spe	ecific and	l kinetic	associate	ed moleo	cules and EB	/-encode	d RNA (EBER)	in nine FDC :	sarcomas (Case	s 1-9)
Cases	СК	EMA	Vimentin	CD45	CD21	CD23	CD35	Podoplanin	CD68	S-100 protein	p53 protein ¹	Ki-67-LI (%)	EBER
1	-	1+	3+	-	3+	NT	1+	NT	NT	NT	3+	4	NT
2	-	-	NT	-	2+	NT	2+	NT	NT	-	2+	15	NT
3	-	1+	2+	-	2+	3+	-	3+	NT	-	2+	1	-
4	-	-	3+	-	3+	2+	2+	-	-	1+	2+	8	-
5	-	-	3+	-	2+	2+	3+	2+	1+	2+	2+	15	-
6	-	-	NT	-	1+	2+	-	-	1+	1+	-	10	+
7	-	-	2+	-	1+	3+	3+	-	-	-	2+	40	-
8	-	-	1+	-	3+	2+	-	2+	2+	1+	2+	20	-
9	-	-	2+	-	1+	2+	1+	-	-	-	2+	35	-

¹Evaluated by the percentages of positive tumor cells, although majority of the positive cells showing weak nuclear immunoreactivity. Immunoreactivities expressed as strong (3+), moderate (2+), weak (1+) and negative (-); NT: Not tested.

was composed of spindle cells with mild nuclear atypia and numerous inflammatory cells including plasma cells, resembling an inflammatory pseudotumor (Figure 2G and H). In addition, neutrophils infiltration was observed in two lesions (Cases 3 and 8). In some tumor areas (Case 3), vascular proliferation was prominent with perivascular sclerosis (Figure 2E) and focal deposition of osteoid matrix mimicking osteosarcoma (Figure 2F).

As described above, the histology varies greatly among the lesions. It is conceivable to predict clinical outcomes using pathological parameters. According to our own data and proposals by other authors^[3], we defined the criteria for low- and high-grade FDC sarcomas using six parameters. As shown in Table 3, the former four were found to be decisive and regarded as major factors for the grading. The lesions with typical architectural and cellular phenotypes, mitotic counts below 5/10 HPF and Ki-67-LI below 10%, as in Cases 1, 3, 5 and 6, were classified as low-grade tumors, and those with anaplastic morphology, mitotic counts up to 5/10 HPF and/or Ki-67-LI up to 10%, as in Cases 2, 4, 7-9, as high-grade tumors. The rest two parameters, including necrosis and loss or reduction of infiltrating lymphocytes, were found to be useful adjuvant factors, but not prerequisites for establishing a diagnosis of the high-grade lesion.

As listed in Table 4, all of the nine lesions were positive for vimentin and negative for CK and CD45. Weak immunoreactivity for EMA was observed in two cases. A diagnosis of FDC sarcoma was established by its immunohistochemical phenotypes including positivity for CD21 (9/9, 100%), CD23 (7/7, 100%) and CD35 (6/9, 67%). Podoplanin was detected in three (43%) of the seven cases. CD68 and S100 protein were detected in about half of the lesions, but their expression levels were low to moderate. Nuclear accumulation of p53 protein, albeit at relatively low levels, was observed in most of the tumor cells in eight (89%) of the nine lesions (Figure 3E). Amplification and sequencing were successful for P53 exons 5-8 in one lesion (Case 9), and we failed to show any mutation. Ki-67-LI in these lesions was quite variable, ranging from 1% to 40%. In situ hybridization was performed in seven of the lesions for EBER, with positive signals demonstrated only in the hepatic lesion (Case 6, Figure 3F).

As the initial treatment, surgery was performed for all of the lesions, with the tumor masses completely removed in seven cases. In Cases 1 and 8, complete resection was not fulfilled because of the involvement of the internal carotid artery. Adjuvant treatment was given to six of the patients (Cases 1, 3-5, 8 and 9) after the operation. Radiation was administered in all the cases and chemotherapy in three cases (Cases 3, 5 and 8). The remaining three patients (Cases 2, 6 and 7) rejected any adjuvant treatment. Follow-up was carried out in all the cases for a period of 12-86 mo, with a median of 27 mo. As shown in Table 1, seven (78%) patients remained alive, four of them had recurrence or metastasis and three were alive without event, and two (22%) died of the tumor recurrence. Of the six cases with events, two recurred locally and four had distant metastases, with a disease-free period below 3 years.

Clinicopathologic features of extranodal FDC sarcomas

Data of 106 extranodal FDC sarcomas, including nine new cases (Table 1) and 97 from literatures (Table 5), were extracted and reviewed carefully. Of the patients, 56 were men and 50 were women, with a ratio of 1.1:1. Ages of the patients ranged from 9 to 82 years at diagnosis, with their mean being 46 years and median being 44 years. The tumors were identified at different anatomic regions. The abdominal and pelvic region was affected in 46 cases (43%), including 14 in the liver, 13 in abdominal/pelvic soft tissues, five in the spleen, and 14 at other organs including pancreas (n = 3), stomach (n = 3), colon and rectum (n = 3), appendix (n = 2), ampulla of Vater (n = 3)1), small intestine (n = 1) and adrenal (n = 1). Of the five splenic cases, two also showed liver involvement. These hepatic lesions were regarded as metastasis in this study. Head and neck were another common region for FDC sarcoma (38/106, 36%), with tonsils and parapharyngeal space affected most frequently. Fourteen (13%) of the lesions were identified from thoracic cavity, of which nine were located at mediastinum, three at lung, one at pleura and one at the chest wall. The less frequent sites included breast (n = 3), soft tissues at thigh (n = 2), dura mater (n = 3)= 1), groin (n = 1) and skin (n = 1). Tumor sizes ranged from 1.5 to 21 cm as described in longitudinal dimensions, with a mean of 7.4 cm. A broad spectrum of histological

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Table 5 Clinicopathologic and therapeutic data of 97 cases of extranodal FDC sarcoma obtained from literatures (Cases 10-106)

Cases No.	Origins	Age (yr)	Gender	Size (cm)	Sites	Initial treatment (responses)	Recurrence, DFT (mo)	Status	Follow-up (mo)
10	Chan <i>et al</i> ^[24] , 1994	44	М	1.5	Tonsil	Surg		NED	36
11	Chan <i>et al</i> ^[24] , 1994	63	F	3	Palate	Surg + RT		NED	54
12	Hollowood et al ^[25] , 1995	23	F	6.5	Small intestine	Surg	LR (2 times), 6	AWD	24
13	Hollowood et al ^[25] , 1995	63	Μ	NA	Pancreas head	Surg	LR, 8	DOD	11
14	Nayler <i>et al</i> ^[26] , 1996	18	F	2	Tonsil	Surg + ChT		NA	NA
15	Perez-Ordonez et al ^[16] , 1996	40	F	NA	Abdomen	Surg + ChT	LR and DM (liver), 12	AWD	24
16	Perez-Ordonez <i>et al</i> ^[16] , 1996	62	F	NA	Tonsil	Surg		NED	12
17	Perez-Ordonez <i>et al</i> ^[16] , 1996	62	Μ	NA	Mediastinum	Surg + ChT	DM (lung), 24	AWD	24
18	Perez-Ordonez <i>et al</i> ^[16] , 1996	46	Μ	NA	Mediastinum	Surg + RT		NED	12
19	Perez-Ordonez <i>et al</i> ^[16] , 1996	31	Μ	NA	Mediastinum	Surg		NED	10
20	Perez-Ordonez <i>et al</i> ^[16] , 1996	75	F	NA	Spleen	Surg	LR, 11	DOD	11
21	Selves <i>et al</i> ^{$[27], 1996$}	68	F	NA	Liver	Surg + ChT		NED	30
22	Shek <i>et al</i> ^[28] , 1996	35	F	NA	Liver	Surg	LR (2 times), 30	AWD	60
23	Chan <i>et al</i> ⁽³⁾ , 1997	42	М	8	Mesocolon, involving LN and peritoneum	Surg + ChT	LR, 18	DOD	18
24	Chan <i>et al</i> ^[3] , 1997	32	Μ	NA	Tonsil	Surg + RT	LR and DM (LN), 54	AWD	54
25	Chan <i>et al</i> ^[3] , 1997	40	F	7	Parapharyngeal space	Surg	LR, 12	AWD	12
26	Chan et al ^[3] , 1997	17	М	11	Neck soft tissue	Surg + ChT		NA	NA
27	Chan <i>et al</i> ^[3] , 1997	25	F	7	Neck soft tissue	Surg + RT	DM (lung), 20	AWD	20
28	Beham-Schmid <i>et al</i> ^[29] , 1998	44	М	2	Nasopharynx	Surg + RT + ChT		NED	20
29	Shek <i>et al</i> ^[30] , 1998	37	М	1.5	Liver	Surg		NED	24
30	Shek <i>et al</i> ^[30] , 1998	61	F	4	Ampulla of Vater	Surg		NED	9
31	Araújo <i>et al</i> ^[31] , 1999	14	М	1.5	Hard palate	Surg		NED	5
32	Desai <i>et al</i> ^[32] , 1999	45	F	NA	Parapharyngeal space	Surg	LR, 31	NED	57
33	Fisher <i>et al</i> ^[33] , 1999	41	F	2.5	Breast	Surg	LR and DM (LN), 36	NED	36
34	Galati <i>et al</i> ^[34] , 1999	65	F	1.5	Thyroid	Surg + RT		NED	36
35	Schwarz <i>et al</i> ¹⁵⁵ , 1999	62	М	15	Abdominal wall	Surg + RT		NED	8
36	Choi <i>et al</i> ^[36] , 2000	28	F	4	Neck soft tissue	Surg	DM (lung), 324	AWD	324
37	Choi et $al^{[36]}$, 2000	66	F	NA	Neck soft tissue	Surg + RT	DM (lung), 24	AWD	24
38	Han <i>et al</i> ⁽³⁷⁾ , 2000	45	М	5	Stomach	Surg		NED	10
39	Chan <i>et al</i> ^[30] , 2001	34	М	NA	Nasopharynx	Surg		NED	36
40	Chang <i>et al</i> ^[59] , 2001	37	F	5	Ascending colon	Surg		NED	7
41	Chen <i>et al</i> ^[5] , 2001	57	F	9.5	Liver	Surveillance		AWD	36
42	Chen <i>et al</i> ⁽³⁾ , 2001	51	F	15	Liver	Surg		NED	12
43	Chiaramonte <i>et al</i> ⁽¹⁰⁾ , 2001	39	М	NA	Retroperitoneum	Surg	DM (LN and lung), 6	DOD	6
44	Shah <i>et al</i> ^[41] , 2001	33	M	9.5	Lung	Surg + ChT		NA	NA
45	Biddle <i>et al</i> ⁽¹¹⁾ , 2002	33	M	NA	Pharynx	Surg + RT	DM (lung), 10	AWD	10
46	Biddle <i>et al</i> ⁽¹¹⁾ , 2002	48	F	1.5	Ionsil	Surg		NED	6
47	Biddle <i>et al</i> ⁽²¹ , 2002)	48	M	3.5	Ionsil	Surg		NED	8
48	Pileri <i>et al</i> ⁽²⁾ , 2002	42	M	NA	Mediastinum	Surg		NED	NA
49	Vargas <i>et al</i> ⁽⁴²⁾ , 2002	54	F	3	Ionsil	Surg	ID (NED	8
50	Vargas <i>et al</i> 43 2002	54	F	6 NTA	Left parotid	Surg + KI	LK, 6	AWD	8
51	Cereson <i>et al.</i> $^{-7}$, 2003	35	M	NA 2	Terreil	Cn1 + R1 (Prog)		NED	24
52	Saton <i>et al.</i> $, 2003$	16	M	3 NTA	I onsil Chamach autith DM (limar)	Surg + KI + ChI		NED	24
55 E4	Geerts <i>et al.</i> 146 2004	40	Г М	NA	Stomacn, with Divi (liver)	Surg		AWD) 10
54	$Grogg et al.^{[46]} 2004$	62	IVI E	NA	Tensil involving IN	Surg		NED	18
55	$Grogg et ut^{-7}, 2004$	57	г	1A	Feloon with DM (liver)	Surveillance		NED	0
56	Grogg et al ^[47] 2004	64 76	Г М	14 N A	Spieen, with Divi (liver)	Surg + Chi		NED	о Э4
57	O'Mallov ^[48] 2004	28	M	INA 5	Cocum	Surg + KI		NED	Z4 NIA
50	Shi at $a^{[49]}$ 2004	30 27	M	15	Tongil	Surg		NED	1NA 26
60	Bradahavy at a ^[50] 2005	37	IVI M	1.5 E	Nock ooft tierue	Suig + Cili	DM(lum x) (0)	NED	06
61	$K_{azaleov}$ et al ^[13] 2005	29	M	19	Skip	Surg	Divi (tutig), 60	NLD	90 NIA
62	Kazakov et ut , 2005 Khalid et $al^{[51]}$ 2005	10	E	1.0	Livor	ChT (PP)		DOD	24
63	Torros <i>et al</i> ^[52] 2005	82	M	15	Liver	Surg		NED	18
64	Audin at $a^{[53]}$ 2005	76	E	25	Topgil	Surg + PT		NED	10
65	Choi <i>et al</i> ^[54] 2006	68	M	5.5 NIA	Dura mator	Surg + RT		NED	40
66	Clement et $aI^{[55]}$ 2006	27	E	4	Tonsil	Surg + RT		NED	6
67	Díaz de Liaño et $al^{[56]}$ 2006	NA	NA	NA	Abdominal cavity	Surg	DM (liver) 18	NED	18
68	Gan et $al^{[57]}$ 2006	32	E	4.5	Parotid	Surg	Divi (liver), 10	NED	20
60	Liang et al ^[58] 2006	46	F	4.J 8.4	Anterior modiactinum	ChT (stable)	DM (hopo) 12	NED	10
70	Jiang et al $^{[59]}$ 2006	40	F	0.4	Pelvic/abdominal cavity	Surg	Divi (bone), 12	NED	19
70	Junig et ut , 2000	40	r	21	multiple	Surg		NED	0
71	Kovács <i>et al</i> ^[00] , 2006	65	М	4	Lung	Surg		NED	18
72	Li et $al^{(61)}$, 2006	19	M	NA	Nasal cavity	Surg		NED	11
73	Lu <i>et al</i> ^{$1021, 2006$}	72	F	7	Groin	Surg		NA	NA
74	Shen <i>et al</i> ^{∞} , 2006	64	М	10.5	Pancreas	Surg	DM (liver), 18	NED	24



75	Shia <i>et al</i> ^[64] , 2006	30	М	NA	Rectum	Surg	LR, 10	AWD	15
76	Shia <i>et al</i> ^[64] , 2006	29	F	12	Lesser omentum	Surg		NED	17
77	Shia <i>et al</i> ^[64] , 2006	69	F	NA	Tonsil	Surg + RT	DM (hilar LN+lung), 96	AWD	108
78	Xu et al ^[65] , 2006	16	М	5	Adrenal	Surg		NA	NA
79	Chang <i>et al</i> ^[66] , 2007	64	М	16	Chest wall	Surg		NED	15
80	Leipsic <i>et al</i> ^[67] , 2007	43	М	13	Mediastinum	Surg		NA	NA
81	Padilla-Rodríguez et al ^[68] , 2007	35	М	21	Retroperitoneal space	Surg + RT		NED	24
82	Sander <i>et al</i> ^[69] , 2007	44	F	11	Spleen	Surg	DM (thorax, liver and	DOD	9
							kidney), 4		
83	Soriano <i>et al</i> ^[70] , 2007	25	F	6	Pelvis	Surg	LR, 2, 2nd Surg+RT	NED	14
84	Soriano <i>et al</i> ^[70] , 2007	56	М	2	Pancreas, involving LN	Surg + RT	LR, 2	AWD	7
85	Soriano <i>et al</i> ^[70] , 2007	33	М	NA	Nasopharynx	Surg + RT	LR, 10	DOD	14
86	Soriano <i>et al</i> ^[70] , 2007	64	F	4	Spleen, with liver involved	Surg + ChT		AWD	29
87	Soriano <i>et al</i> ^[70] , 2007	66	М	16	Pleura	ChT (Prog)		DOD	7
88	Tu <i>et al</i> ^[12] , 2007	63	М	15	Jejunum mesentery	Surg		NA	NA
89	Tu <i>et al</i> ^[12] , 2007	43	М	4	Appendix	Surg		NA	NA
90	Tu <i>et al</i> ^[12] , 2007	28	F	15	Stomach	Surg	DM (liver), 3	AWD	3
91	Yuan <i>et al</i> ^[71] , 2007	29	М	10	Liver	Surg	LR, 6	AWD	6
92	De Pas <i>et al</i> ^[72] , 2008	40	F	4	Breast	Surg		NED	62
93	De Pas <i>et al</i> ^[72] , 2008	53	F	NA	Liver	Surg	DM (LN), 11	DOD	22
94	De Pas <i>et al</i> ^[72] , 2008	64	F	2.1	Breast	Surg		NED	20
95	Granados <i>et al</i> ^[73] , 2008	57	F	13	Liver	Surg		NED	24
96	Liu <i>et al</i> ^[74] , 2008	42	М	12	Gastrocolic omentum	Surg		NA	NA
97	Peng <i>et al</i> ^[75] , 2008	60	М	7	Mesentery	Surg		NA	NA
98	Zhang <i>et al</i> ^[76] , 2008	36	М	NA	Nasopharynx	Surg		NED	5
99	Zhang <i>et al</i> ^[76] , 2008	32	F	NA	Spleen	Surg		NA	NA
100	An <i>et al</i> ^[77] , 2009	40	М	5	Liver	Surg		NED	3
101	Denning et al ^[78] , 2009	64	F	1.7	Lung	Surg		NED	24
102	Liu <i>et al</i> ^[79] , 2009	75	М	4	Liver	Surg		NA	NA
103	Romero-Guadarrama <i>et al</i> ^[80] , 2009	54	F	15	Thigh soft tissue	Surg	LR, 12	NED	48
104	Shen <i>et al</i> ^[81] , 2009	43	М	5	Appendix	Surg	LR, 8	NED	8
105	Vaideeswar et al ^[82] , 2009	50	М	2.5	Tonsil	Surg	,	NED	48
106	Xu et al ^[83] , 2009	57	F	11	Liver	Surg		NA	NA
						U			

LN: Lymph node; NA: Not available; Prog: Disease progression.

phenotypes were described, while the most helpful diagnostic features were the typical arrangement of spindle and ovoid tumor cells with eosinophilic cytoplasm, indistinct cytoplasmic borders and syncytial appearance, and presence of small lymphocytes in the tumor.

Morphologic information, indicative of the architectural and cellular anaplasia, was collected from the articles and re-evaluated carefully. Of the 63 informative cases, 37 (59%) were described as lesions with mild atypia, and 26 (41%) with moderate to severe cytological atypia throughout or in a certain area of the tumor. In the current study, these two groups were classified into low-grade and highgrade lesions, respectively. All of the tumors occurring in the liver resembled an inflammatory pseudotumor in morphology. Five of the eight informative cases were divided into low-grade group, and the other three into high-grade group due to identification of anaplastic components. Coagulative necrosis was recorded in 29 (45%) of the 64 informative cases. Mitotic counts were provided in 61 cases, with 31 (51%) up to 5/10 HPF. Ki-67-LI was calculated in 20 lesions, with 14 (70%) up to 10%. Preoperative tumor spreading was identified in 15 (23%) of the 66 informative cases through pathological examination of the resected samples, including local lymph node involvement in 11, distant metastasis to liver or peritoneal dissemination in three, and both nodal involvement and peritoneal spreading in one.

Management and clinical outcomes

One hundred and four patients received therapeutic procedures and two rejected any treatment. Of the patients, 100 (94%) were treated surgically to remove the tumor. Adjuvant treatment was administered in 35 cases, including radiation in 20, chemotherapy in 10 and both procedures in five. Chemotherapy was performed in most of the cases based on CHOP regimen including cyclophosphamide, adriamycin, vincristine and prednisone.

Follow-up data were available in 91 cases. The observation periods ranged from 3 to 324 mo, with a mean and a median of 27 and 19 mo, respectively. Overall, 38 patients (42%) had local recurrence and/or distant metastasis during the adjuvant treatment or surveillance period, with the events occurring within 36 mo in most (33/37, 89%) of the cases. Local recurrence occurred in 21 patients (23%) at a median of 12 mo after surgical removal (range, 3-31 mo). Distant metastases occurred in 19 patients (21%) at a median of 14 mo after operation (range, 4-324 mo). Metastatic sites included lung (n = 9), liver (n = 6), lymph node (n = 6)= 5), bone (n = 1), thorax (n = 1), ovary (n = 1) and kidney (n = 1). At the last follow-up, 12 patients (13%) died of the tumor, 25 (27%) were alive with disease, and 54 (59%) were alive with no evidence of disease. The 2-year and 5-year overall survival rates for the entire group were 82% and 79%, and their 2-year and 5-year disease-free survival rates were 57% and 32%, respectively (Figure 4).



Figure 4 Overall (A) and disease-free survival curves (B) of FDC sarcoma. The results are based on follow-up data of 91 informative cases.

Prognostic factors

Associations between nine clinicopathological factors and recurrence risk were studied, and the results are summarized in Table 6. Of the 60 informative lesions, 25 (42%) were smaller than 5 cm and 34 (58%) were up to 5 cm in their longitudinal dimensions. Four (16%) of the smaller lesions relapsed (two with local recurrence and two with distant metastasis) and 19 (56%) of the larger lesions relapsed (nine with local recurrence and 10 with metastasis). The recurrence rate was markedly elevated with the increase of tumor size (P = 0.003). In addition, a mortality rate of 17% was observed for cases with the larger lesions, and none of the patients with smaller lesions died of the disease (P = 0.036).

For most of the lesions, as listed in Table 5, histological grades were assessed mainly according to descriptions on architectural features and cellular atypia from respective literatures. For this reason, tumor necrosis, mitotic activity and Ki-67-LI were evaluated separately in this study. Of the 63 informative cases, 37 (59%) were evaluated as low-grade and 26 (41%) as high-grade lesions. Twelve (32%) of the low-grade lesions relapsed (seven with local recurrence, four with metastasis and one with both local recurrence and metastasis) and 15 (58%) of the high-grade lesions relapsed (seven with local recurrence, seven with metastasis and one with both

Table 6 Associations between clinicopathological parameters and recurrence risk in patients with extranodal FDC sarcoma

Pathologic parameters	Recurrence frequencies (%)	P values
Age (vr)		
< 50	26/51 (51)	0.072
≥ 50	11/35 (31)	
Gender	,	
Male	15/40 (38)	0.290
Female	22/46 (48)	
Size (cm)		
< 5	4/25 (16)	0.003
≥ 5	19/34 (56)	
Histological grades		
Low	12/37 (32)	0.046
High	15/26 (58)	
Mitotic counts		
< 5/10 HPF	8/30 (27)	0.013
$\geq 5/10$ HPF	18/31 (58)	
Ki-67-LI		
< 10%	2/6 (25)	0.628
$\geq 10\%$	8/14 (57)	
Necrosis		
Absent	17/35 (49)	0.307
Present	13/29 (45)	
Treatments		
Surgery alone	22/52 (42)	0.590
Surgery + adjuvant therapy	15/31 (48)	
Sites		
Abdominal/pelvic	18/37 (49)	0.372
Other sites	20/53 (38)	

local recurrence and metastasis). The recurrence rate was higher in the latter group than in the former (P = 0.046). In addition, the high-grade lesions resulted in death in nine (35%) of the cases, while only one (3%) of the patients with low-grade lesions died of the disease, the mortality rate being closely associated with tumor grade (P = 0.001). Eighteen (58%) of the 31 lesions with a mitotic count $\geq 5/10$ HPF relapsed, the frequency being higher than those with a count < 5/10 HPF (8/30, 27%, P = 0.013). The mortality rate (7/31, 23%) was also higher in the former group than in the latter (0/30, P =0.011). According to the limited number of informative cases, increased Ki-67-LI ($\geq 10\%$) appeared to indicate an unfavorable clinical outcome, but its impact failed to reach statistical significance (P = 0.628). The patients younger than 50 years tended to develop tumor recurrence more frequently (51%) than those over 50 years of age (31%), but without statistically significant difference (P = 0.072). Other factors, including gender, necrosis and therapeutic procedures following surgery, were not related to clinical outcomes (P > 0.05).

The associations of tumor size, histological grade and mitoactivity with overall and disease-free survival are described in Figure 5. Among them, tumor size and histological grade were the most important factors, respectively, for tumor recurrence (Figure 5B) and disease-associated death (Figure 5C). Thus, a model was established for the recurrence risk assessment by combining the former two parameters. Recurrence rates of lesions smaller than 5 cm, the larger lesions (\geq 5 cm) with low-grade histology and those with high-grade histology were 16% (4/25), 46%





Figure 5 Overall (A, C and E) and disease-free survival curves (B, D and F) of patients with FDC sarcoma, showing data of 120 mo.

(11/24) and 73% (8/11), respectively, their difference being significant (P = 0.000). Based on the recurrence potential, these three groups of lesions were designated as low-, intermediate- and high-risk FDC sarcomas, respectively (Figure 6). A markedly higher mortality rate was also observed in cases with a high-risk tumor (45%) than in those with the low-risk (0%, P = 0.001) and intermediate-risk lesions (4%, P = 0.006). The low-risk group had a longer survival and the high-risk group had the most unfavorable clinical outcomes considering both the tumor recurrence and mortality (Figure 7).

During the follow-up period, 18 (49%) of the 37 informative lesions from abdominal and pelvic cavities recurred (20/53, 38%, P = 0.302). Of the 37 abdominal/pelvic lesions, 12 occurred in the liver and 25 were

identified from other tissues. A lower recurrence risk (3/12, 25%) was observed in hepatic lesions than in the extrahepatic lesions (15/25, 60%, P = 0.046). A more favorable disease-free survival was also observed for the former group (P = 0.004). However, there was no significant difference between the hepatic (2/12, 17%) and extrahepatic groups (5/25, 20%) in mortality (P = 0.372, Figure 8). The size of the hepatic lesions (mean, 9.5 cm; range, 2-21 cm) was comparable to that of the extrahepatic lesions (mean, 10.4 cm; range, 1.5-16 cm, P = 0.644). Morphologically, all of the hepatic lesions belonged to the inflammatory pseudotumor-like variant, and most of the extrahepatic lesions were classified into conventional type. However, we failed to show a significant difference in frequency of occurrence of high-



Figure 6 Definitions of low- (low-risk), intermediate- (mediate-risk) and high-risk groups (high-risk) of extranodal FDC sarcomas. Mortality rates: overall, P = 0.000; low-risk vs mediate-risk, P = 1.000; low-risk vs high-risk, P = 0.001; mediate-risk vs high-risk, P = 0.006; Recurrence rates: overall, P = 0.001; low-risk vs mediate-risk, P = 0.032; low-risk vs high-risk, P = 0.002; mediate-risk vs high-risk, P = 0.167.



Figure 7 Overall (A) and disease-free survival curves (B) of patients with FDC sarcoma, estimated in groups with low-, intermediate- and high-risk tumors, showing data of 120 mo. A: Overall, P = 0.000; low-risk vs mediate-risk, P = 0.273; low-risk vs high-risk, P = 0.000; mediate-risk vs high-risk, P = 0.001; B: Overall, P = 0.000; low-risk vs mediate-risk, P = 0.003; low-risk vs high-risk, P = 0.003; low-risk vs high-risk, P = 0.273.

grade component between the hepatic (3/8, 38%) and extrahepatic lesions (7/18, 39%, P = 1.000).

The extrahepatic, abdominal/pelvic lesions were also compared with those occurring out of abdominal/pelvic cavity. Their recurrence rates were 60% (15/25) and 38%



Figure 8 Overall (A) and disease-free survival curves (B) of abdominal/pelvic FDC sarcomas. The lesions are divided into hepatic and extrahepatic groups.

(20/53), respectively, with the former lesions recurring more frequently (P = 0.000) and earlier (Figure 9). The former cases tended to have a higher mortality rate (5/25, 20%) than the latter group (5/53, 6%), but without statistically significant difference (P = 0.051, Figure 9). These two groups also showed difference in their sizes, with the average of the former group (9.5 cm) being larger than that of the latter group (5.3 cm, P = 0.003). They were not significantly different in histological grade (P = 0.204), with high-grade phenotypes observed in 57% (26/46) of the former lesions and 39% (7/18) of the latter lesions.

DISCUSSION

FDC sarcoma may occur in extranodal tissues from a variety of anatomical sites, including head and neck, liver, spleen, gastrointestinal tract, soft tissue, skin, lung and breast^{114]}. As shown in this survey, abdominal/pelvic cavity and head/neck areas are affected more frequently, where 43% and 36% of extranodal FDC sarcomas were identified. It usually presents with a solitary tumor, typically composed of spindle and ovoid cells with scattered mature lymphocytes. However, there is a broad spectrum of morphologic variations in this tumor, some of which overlap with those of other neoplasms including ectopic meningioma, thymoma, myoepithelial tumor, inflammatory myofibroblastic tumor and GIST. In our series, osteoid component was found in a low-grade FDC sarcoma



Figure 9 Overall (A) and disease-free survival curves (B) of patients with extrahepatic FDC sarcomas, showing data of 120 mo. The lesions are divided into abdominal/pelvic and non-abdominal/pelvic groups.

(Case 3). This is the first case showing presence of an osteosarcoma-like phenotype (Figure 2F). Misdiagnosis may occur when a pathologist is not aware of the tumor, as in Cases 1 and 2 of the current series (Table 1). The diagnosis can be established by immunohistochemical demonstration of CD21, CD23, CD35^[70], podoplanin and clusterin expression^[64,84,85].

FDC sarcoma is usually regarded as an indolent tumor with a tendency of local recurrence but a low risk of metastasis, behaving like a low-grade soft tissue sarcoma^[15]. In a study of 13 cases, Perez-Ordonez et al¹⁶ observed a substantial risk of metastases, prompting consideration of this malignancy to be of intermediate grade. An analysis by Shia et al^[64] found that the 2- and 5-year recurrencefree survival rates were 62% and 27%, respectively. In our study, a total number of 106 extranodal cases were analyzed. The recurrence rate was 42%, including local recurrence in 23% and distant metastasis in 21% of the cases, and the mortality rates were 13% over the periods from 3 to 324 mo (mean, 27 mo; median, 19 mo). The 2-year and 5-year disease-free survival rates were 57% and 32%, and the 2-year and 5-year overall survival were 82% and 79%, respectively. It appears that extranodal FDC sarcomas, as a whole, are more aggressive than a low-grade soft tissue sarcoma.

As demonstrated in this study, pathologic representations of extranodal FDC sarcoma are variable among different cases. New approaches are needed to find most important prognostic factors relevant to their clinical outcomes. It has been proposed that intra-abdominal involvement, a high mitotic count (\geq 5/10 HPF), coagulative necrosis and marked cellular atypia are potentially helpful predictors of an unfavorable outcome^[3]. In this study, nine clinicopathological parameters were analyzed, and large tumor size (\geq 5 cm in diameter), high-grade histology and high mitotic counts (\geq 5/10 HPF) were found to be closely associated with the clinical outcomes. The recurrence risk seemed to be increased in lesions with high Ki-67-LI, but the association showed no statistical significance. It needs to be classes.

While some histological phenotypes, including cellular atypia, mitoactivity and coagulative tumor necrosis, were considered important parameters for prognosis assessment^[3], a grading model remains to be established. Soriano et $al^{[70]}$ identified five high-grade lesions from 14 FDC sarcomas based on cellular atypia, but they failed to show its relevance of statistical significance to clinical outcomes. In this study, we examined nine new cases from northern China, extracted and reviewed data of 97 cases from literatures, and established histological criteria for lowgrade and high-grade FDC sarcomas according to four major parameters including architectural alteration, cellular atypia, mitoactivity and/or Ki-67-LI (Table 3). Using this model, 63 informative cases were classified into low- (n =37) and high-grade lesions (n = 26). Their recurrence rates were 32% and 58%, and their mortality rates were 3% and 35%, respectively. The results show a significant difference in their recurrence risks and mortality rates. Our data demonstrate that these two groups of lesions behave like low- and high-grade soft tissue sarcomas.

It should be noted that, for reported cases, histological grades were evaluated mainly according to two parameters (the architectural features and cellular atypia) in this study. Mitotic counts were not available for about half of the cases from literatures, and Ki-67-LI was provided in only 21 of the reported cases. For this reason, influence of mitoactivity on clinical outcomes was evaluated separately, with the recurrence rates of the mitotically indolent (< 5/10 HPF) and active lesions ($\geq 5/10$ HPF) determined to be 27% and 58%, respectively. It is expected that, with the assessment based on all of the six parameters including four major and two adjuvant factors (Table 3), the predicting power of our grading model for tumor recurrence risk would be stronger. This has been reflected in our own cases: the recurrence rates of the low- and high-grade lesions were 25% (1/4) and 100% (5/5), respectively.

Among the three parameters associated with clinical outcomes of extranodal FDC sarcoma, tumor size was shown to be most important. It appears that tumor size is a main factor for predicting its recurrence potential, and histological grade is closely associated with mortality rate mainly in patients with larger tumors (Figures 5 and 6). Considering the independent impact of the size to tumor behaviors and close association between mitoactivity and histological grade, we combined the size and grade factors, and proposed a model for recurrence risk assessment (Figure 6). By this model, FDC sarcomas were classified into low-, intermediate- and high-risk groups. Recurrence rates in the three groups were 16%, 46% and 73%, and the mortality rates were 0%, 4% and 45%, respectively. This will provide a convenient approach for diagnostic pathologists to predict the clinical outcomes of the tumor more accurately following its surgical resection or even with a biopsy. It should be noted that it is impossible to know exactly whether an FDC sarcoma developed from lymph node or extranodal tissues at advanced stages, as in some of the cases in this study. We believe that this model may also be applied for prognosis assessment of the FDC sarcomas which develop from lymph nodes.

Chan et al³ pointed out that intra-abdominal location was associated with a higher recurrence rate. In this study, the recurrence rates of the intra-abdominal lesions and those from other sites were 49% and 38%, respectively, but their difference did not attain a statistical significance. Interestingly, a lower recurrence rate (25%) and a more favorable clinical course were observed in the hepatic lesions as compared with that of the extrahepatic, abdominal/pelvic tumors (60%). It seems to be true that, for extranodal FDC sarcomas of the conventional type, intra-abdominal lesions recur more frequently than those from other sites. The tumors of the former group were found to be larger at diagnosis (9.5 cm) than the latter lesions (5.3 cm). It is conceivable to ascribe their difference in clinical outcomes to their size difference, considering the fact that abdominal/pelvic FDC sarcomas are frequently concealed and diagnosed later compared with those from other sites including head, neck and some superficial areas.

While its pathogenesis has not been established, several factors were linked to development of FDC sarcoma. Some FDC sarcomas from lymph nodes appear to be associated with hyaline-vascular type Castleman disease^[86], and similar changes were also noted in tissues surrounding some rare extranodal tumors^[38,87,88]. Several reports have observed FDC proliferation and dysplastic changes in lymphoid tissues with Castleman disease where an FDC sarcoma develops^[38,89-91], apparently supporting the hypothesis that FDC sarcoma develops from Castleman disease in a hyperplasia - dysplasia - neoplasia sequence^[24]. However, the association was not evident in any lesion of our series.

There have been some data indicative of a link between persistent EBV infection and Castleman disease^[92,93]. EBER signals were also demonstrated in some FDC sarcomas, particularly the hepatic and splenic lesions^[4-9]. Most of these EBV-positive lesions contain numerous inflammatory cells, including plasma cells and lymphocytes, and dispersed tumor cells, thereby being described as inflammatory pseudotumor-like FDC sarcoma^[94]. In this study, EBER was demonstrated in the hepatic inflammatory pseudotumor-like lesion (Case 6), but not in any of the six conventional FDC sarcomas examined. It appears to be true that inflammatory pseudotumor-like FDC sarcomas are related to EBV infection.

Clonal cell composition was described in an extranodal

FDC sarcoma^[17], but greater efforts are required to elucidate its genetic background and establish its molecular pathogenesis. Nuclear immunoreactivity for p53 protein was noticed fortuitously in a few cases^[17,38]. In this series, eight extranodal FDC sarcomas without a background of Castleman disease were tested, and a low-level accumulation of p53 protein was observed in majority of tumor cells. Our data raises a possibility of involvement of the p53-mediated pathway during development or progression of FDC sarcoma. In addition, P53 exons 5-8, where most of the P53 gene mutations had been found^[19,95], were examined in one of the lesions, but without any mutation demonstrated. Based on these data, we consider that the nuclear immunoreactivity may reflect accumulation of wild-type p53 protein. This phenomenon may be an adaptive reaction responsive to elevated mitotic activity and/or increased stress for cell survival, as observed in other cell types under neoplastic^[96] and non-neoplastic conditions^[18,97-99].

ACKNOWLEDGMENTS

The authors thank Dr. Ling Li and Dr. Hong-Tu Zhang for the helpful discussions, Dr. Lin Yang for her help in statistical analysis, Guo-Lian Wei, Xiu-Yun Liu, Xin-Hua Xue and Yong-Qiang Xie for their technical assistance.

COMMENTS

Background

Follicular dendritic cell (FDC) sarcoma is a rare tumor, occurring in lymph nodes and extranodal tissues. It is usually regarded as an indolent tumor with a tendency of local recurrence but a low risk of metastasis. With more cases encountered during pathological practice and reported from literatures, various morphologic and clinical representations were noticed. Because of the rarity of the tumor, assessment of its prognosis remains difficult. Clearly, histological grade of this malignancy remains to be defined and a model is needed to evaluate the recurrence risk of individual tumors within this category.

Research frontiers

While FDC sarcoma is regarded as an indolent tumor, like a low-grade soft tissue sarcoma, a broad spectrum of pathologic phenotypes and more aggressive clinical representations were noticed for this tumor type. Intraabdominal involvement, elevated mitoactivity, coagulative necrosis and marked cellular atypia were considered potentially helpful predictors of an unfavorable outcome, but a grading model remains to be established. In this study, a total number of 106 extranodal FDC sarcomas, nine new cases and 97 from literatures, were analyzed, histological grades of this malignancy were defined and a model was established for recurrence risk assessment of the tumor.

Innovations and breakthroughs

In this study, data of 106 cases of extranodal FDC sarcoma were extracted and reviewed. Histological criteria for low-grade and high-grade FDC sarcomas were established according to four major parameters including architectural pattern, cellular atypia, mitoactivity and/or Ki-67-labeling index. Using this model, 63 informative cases were divided into low- (n = 37) and high-grade lesions (n = 26). Their recurrence rates were 32% and 58%, and the mortality rates were 3% and 35%, respectively. The results showed a significant difference in their recurrence risks and mortality rates. In addition, nine clinicopathological parameters were analyzed, and large tumor size (≥ 5 cm in diameter), highgrade histology and elevated mitotic counts ($\ge 5/10$ HPF) were found to be closely associated with the clinical outcomes. According to the tumor size and histological grade, a model was proposed for recurrence risk assessment. By this model, FDC sarcomas were classified into low-, intermediate- and high-risk groups, and their recurrence rates were 16%, 46% and 73%, and the mortality rates were 0%, 4% and 45%, respectively.

Applications

A grading approach was established for evaluating aggressiveness of extranodal FDC sarcomas, and the lesions were divided into low-grade and highgrade categories. According to the tumor size and histological grade, a model was proposed for recurrence risk assessment. This will provide a convenient procedure for diagnostic pathologists to predict the clinical outcomes of the tumor more accurately following its surgical resection or even with a biopsy. This model may also be applied to the prognosis assessment of the FDC sarcomas which develop from lymph nodes.

Peer review

It is a well written and important paper that likely represents the most thorough examination of a rare sarcoma. The authors are to be congratulated for an important work.

REFERENCES

- 1 **Monda L**, Warnke R, Rosai J. A primary lymph node malignancy with features suggestive of dendritic reticulum cell differentiation. A report of 4 cases. *Am J Pathol* 1986; **122**: 562-572
- 2 Pileri SA, Grogan TM, Harris NL, Banks P, Campo E, Chan JK, Favera RD, Delsol G, De Wolf-Peeters C, Falini B, Gascoyne RD, Gaulard P, Gatter KC, Isaacson PG, Jaffe ES, Kluin P, Knowles DM, Mason DY, Mori S, Müller-Hermelink HK, Piris MA, Ralfkiaer E, Stein H, Su IJ, Warnke RA, Weiss LM. Tumours of histiocytes and accessory dendritic cells: an immunohistochemical approach to classification from the International Lymphoma Study Group based on 61 cases. *Histopathology* 2002; **41**: 1-29
- 3 Chan JK, Fletcher CD, Nayler SJ, Cooper K. Follicular dendritic cell sarcoma. Clinicopathologic analysis of 17 cases suggesting a malignant potential higher than currently recognized. *Cancer* 1997; **79**: 294-313
- 4 Cheuk W, Chan JK, Shek TW, Chang JH, Tsou MH, Yuen NW, Ng WF, Chan AC, Prat J. Inflammatory pseudotumorlike follicular dendritic cell tumor: a distinctive low-grade malignant intra-abdominal neoplasm with consistent Epstein-Barr virus association. *Am J Surg Pathol* 2001; 25: 721-731
- 5 Chen TC, Kuo TT, Ng KF. Follicular dendritic cell tumor of the liver: a clinicopathologic and Epstein-Barr virus study of two cases. *Mod Pathol* 2001; 14: 354-360
- 6 Horiguchi H, Matsui-Horiguchi M, Sakata H, Ichinose M, Yamamoto T, Fujiwara M, Ohse H. Inflammatory pseudotumor-like follicular dendritic cell tumor of the spleen. *Pathol Int* 2004; 54: 124-131
- 7 Bai LY, Kwang WK, Chiang IP, Chen PM. Follicular dendritic cell tumor of the liver associated with Epstein-Barr virus. *Jpn J Clin Oncol* 2006; 36: 249-253
- 8 Laurent C, Meggetto F, de Paiva GR, Selves J, Palasse J, Laurent G, Brousset P. Follicular dendritic cell tumor of the spleen associated with diffuse large B-cell lymphoma. *Hum Pathol* 2008; **39**: 776-780
- 9 Gong QX, Fan QH, Zhou ZS, Zhang ZH, Yu MN, Wang Z, Wang C, Zhang WM. [Inflammatory pseudotumor-like follicular dendritic cell tumor of spleen] *Zhonghua Binglixue Zazhi* 2008; 37: 40-44
- 10 Agaimy A, Wünsch PH. Follicular dendritic cell tumor of the gastrointestinal tract: Report of a rare neoplasm and literature review. *Pathol Res Pract* 2006; **202**: 541-548
- 11 **Biddle DA**, Ro JY, Yoon GS, Yong YW, Ayala AG, Ordonez NG, Ro J. Extranodal follicular dendritic cell sarcoma of the head and neck region: three new cases, with a review of the literature. *Mod Pathol* 2002; **15**: 50-58
- 12 **Tu XY**, Sheng WQ, Lu HF, Wang J. [Clinicopathologic study of intraabdominal extranodal follicular dendritic cell sarcoma] *Zhonghua Binglixue Zazhi* 2007; **36**: 660-665
- 13 Kazakov DV, Morrisson C, Plaza JA, Michal M, Suster S. Sarcoma arising in hyaline-vascular castleman disease of skin and subcutis. *Am J Dermatopathol* 2005; 27: 327-332
- 14 Youens KE, Waugh MS. Extranodal follicular dendritic cell sarcoma. *Arch Pathol Lab Med* 2008; **132**: 1683-1687

- 15 Chan JKC, Pileri SA, Delsol G, Fletcher CDM, Weiss LM, Grogg KL. Follicular dendritic cell sarcoma. In: Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, Thiele J, Vardiman JW, editors. WHO classification of tumors of haematopoietic and lymphoid tissues. 4th ed. Lyon: International Agency for Research on Cancer, 2008: 364-365
- 16 Perez-Ordonez B, Erlandson RA, Rosai J. Follicular dendritic cell tumor: report of 13 additional cases of a distinctive entity. Am J Surg Pathol 1996; 20: 944-955
- 17 Su Q, Wang SF, Shang FX, Sun H, Gong L, Duan YY, Yuan LT, Wang YC. Extranodal follicular dendritic cell sarcoma, report of 2 cases and clonality analysis. *Zhonghua Binglixue Zazhi* 2004; 34: 183-185
- 18 Su Q, Schröder CH, Otto G, Bannasch P. Overexpression of p53 protein is not directly related to hepatitis B x protein expression and is associated with neoplastic progression in hepatocellular carcinomas rather than hepatic preneoplasia. *Mutat Res* 2000; 462: 365-380
- 19 Lamb P, Crawford L. Characterization of the human p53 gene. Mol Cell Biol 1986; 6: 1379-1385
- 20 Joerger AC, Fersht AR. Wild type p53 conformation, structural consequences of p53 mutations and mechanisms of mutant p53 rescue. In: Hainaut P, Wiman KG, editors. 25 Years of p53. Dordecht: Springer-Verlag, 2005: 377-398
- 21 Chau Y, Hongyo T, Aozasa K, Chan JK. Dedifferentiation of adenoid cystic carcinoma: report of a case implicating p53 gene mutation. *Hum Pathol* 2001; **32**: 1403-1407
- 22 Bharaj BS, Angelopoulou K, Diamandis EP. Rapid sequencing of the p53 gene with a new automated DNA sequencer. *Clin Chem* 1998; **44**: 1397-1403
- 23 **Detwiler MM**, Hamp TJ, Kazim AL. DNA sequencing using the liquid polymer POP-7 on an ABI PRISM 3100 Genetic Analyzer. *Biotechniques* 2004; **36**: 932-933
- 24 Chan JK, Tsang WY, Ng CS. Follicular dendritic cell tumor and vascular neoplasm complicating hyaline-vascular Castleman's disease. Am J Surg Pathol 1994; 18: 517-525
- 25 Hollowood K, Stamp G, Zouvani I, Fletcher CD. Extranodal follicular dendritic cell sarcoma of the gastrointestinal tract. Morphologic, immunohistochemical and ultrastructural analysis of two cases. *Am J Clin Pathol* 1995; **103**: 90-97
- 26 Nayler SJ, Verhaart MJ, Cooper K. Follicular dendritic cell tumour of the tonsil. *Histopathology* 1996; **28**: 89-92
- 27 Selves J, Meggetto F, Brousset P, Voigt JJ, Pradère B, Grasset D, Icart J, Mariamé B, Knecht H, Delsol G. Inflammatory pseudotumor of the liver. Evidence for follicular dendritic reticulum cell proliferation associated with clonal Epstein-Barr virus. *Am J Surg Pathol* 1996; 20: 747-753
- 28 Shek TW, Ho FC, Ng IO, Chan AC, Ma L, Srivastava G. Follicular dendritic cell tumor of the liver. Evidence for an Epstein-Barr virus-related clonal proliferation of follicular dendritic cells. *Am J Surg Pathol* 1996; 20: 313-324
- 29 Beham-Schmid C, Beham A, Jakse R, Auböck L, Höfler G. Extranodal follicular dendritic cell tumour of the nasopharynx. Virchows Arch 1998; 432: 293-298
- 30 Shek TW, Liu CL, Peh WC, Fan ST, Ng IO. Intra-abdominal follicular dendritic cell tumour: a rare tumour in need of recognition. *Histopathology* 1998; 33: 465-470
- 31 Araújo VC, Martins MT, Salmen FS, Araújo NS. Extranodal follicular dendritic cell sarcoma of the palate. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1999; 87: 209-214
- 32 **Desai S**, Deshpande RB, Jambhekar N. Follicular dendritic cell tumor of the parapharyngeal region. *Head Neck* 1999; **21**: 164-167
- 33 Fisher C, Magnusson B, Hardarson S, Smith ME. Myxoid variant of follicular dendritic cell sarcoma arising in the breast. Ann Diagn Pathol 1999; 3: 92-98
- 34 Galati LT, Barnes EL, Myers EN. Dendritic cell sarcoma of the thyroid. *Head Neck* 1999; **21**: 273-275
- 35 **Schwarz RE**, Chu P, Arber DA. Extranodal follicular dendritic cell tumor of the abdominal wall. *J Clin Oncol* 1999; **17**: 2290-2292



- 36 Choi PC, To KF, Lai FM, Lee TW, Yim AP, Chan JK. Follicular dendritic cell sarcoma of the neck: report of two cases complicated by pulmonary metastases. *Cancer* 2000; 89: 664-672
- 37 Han JH, Kim SH, Noh SH, Lee YC, Kim HG, Yang WI. Follicular dendritic cell sarcoma presenting as a submucosal tumor of the stomach. *Arch Pathol Lab Med* 2000; **124**: 1693-1696
- 38 Chan AC, Chan KW, Chan JK, Au WY, Ho WK, Ng WM. Development of follicular dendritic cell sarcoma in hyalinevascular Castleman's disease of the nasopharynx: tracing its evolution by sequential biopsies. *Histopathology* 2001; 38: 510-518
- 39 Chang KC, Jin YT, Chen FF, Su IJ. Follicular dendritic cell sarcoma of the colon mimicking stromal tumour. *Histopathology* 2001; 38: 25-29
- 40 **Chiaramonte MF**, Lee D, Abruzzo LV, Heyman M, Bass BL. Retroperitoneal follicular dendritic cell sarcoma presenting as secondary amyloidosis. *Surgery* 2001; **130**: 109-111
- 41 Shah RN, Ozden O, Yeldandi A, Peterson L, Rao S, Laskin WB. Follicular dendritic cell tumor presenting in the lung: a case report. *Hum Pathol* 2001; 32: 745-749
- 42 **Vargas H**, Mouzakes J, Purdy SS, Cohn AS, Parnes SM. Follicular dendritic cell tumor: an aggressive head and neck tumor. *Am J Otolaryngol* 2002; **23**: 93-98
- 43 **Ceresoli GL**, Zucchinelli P, Ponzoni M, Gregorc V, Bencardino K, Paties CT. Mediastinal follicular dendritic cell sarcoma. *Haematologica* 2003; **88**: ECR04
- 44 Satoh K, Hibi G, Yamamoto Y, Urano M, Kuroda M, Nakamura S. Follicular dendritic cell tumor in the oro-pharyngeal region: report of a case and a review of the literature. Oral Oncol 2003; 39: 415-419
- 45 Geerts A, Lagae E, Dhaene K, Peeters M, Waeytens A, Demetter P, Cuvelier C, Defreyne L, De Vos M, Pattyn P. Metastatic follicular dendritic cell sarcoma of the stomach: a case report and review of the literature. *Acta Gastroenterol Belg* 2004; 67: 223-227
- 46 Grogg KL, Lae ME, Kurtin PJ, Macon WR. Clusterin expression distinguishes follicular dendritic cell tumors from other dendritic cell neoplasms: report of a novel follicular dendritic cell marker and clinicopathologic data on 12 additional follicular dendritic cell tumors and 6 additional interdigitating dendritic cell tumors. *Am J Surg Pathol* 2004; 28: 988-998
- 47 Kröber SM, Marx A, Aebert H, Dohmen BM, Kaiserling E. Sarcoma of follicular dendritic cells in the dorsal mediastinum. *Hum Pathol* 2004; 35: 259-263
- 48 O'Malley DP. Diagnosis: follicular dendritic cell tumor mimicking GI stromal tumor. 2004. Available from: URL: http://www.socforheme.org/case-nov-04.htm
- 49 Shi QL, Zhou XJ, Ma J, Ma HH, Wu KM, Zhou M, Wang QP. Follicular dendritic cell sarcoma of tonsil: a case report and review of the literature. *Zhenduan Binglixue Zazhi* 2004; 11: 81-83
- 50 **Bradshaw EJ**, Wood KM, Hodgkinson P, Lucraft H, Windebank KP. Follicular dendritic cell tumour in a 9-year-old child. *Pediatr Blood Cancer* 2005; **45**: 725-727
- 51 Khalid T, Folman R. Symptoms in cancer patients and an unusual tumor: Case 3. Follicular dendritic cell sarcoma. *J Clin Oncol* 2005; **23**: 9425-9426
- 52 **Torres U**, Hawkins WG, Antonescu CR, DeMatteo RP. Hepatic follicular dendritic cell sarcoma without Epstein-Barr virus expression. *Arch Pathol Lab Med* 2005; **129**: 1480-1483
- 53 Aydin E, Ozluoglu LN, Demirhan B, Arikan U. Follicular dendritic cell sarcoma of the tonsil: case report. *Eur Arch Otorhinolaryngol* 2006; 263: 1155-1157
- 54 **Choi JW**, Lee JH, Kim A, Kim CH, Chae YS, Kim I. Follicular dendritic cell sarcoma arising in the dura mater of the spine. *Arch Pathol Lab Med* 2006; **130**: 1718-1721
- 55 Clement P, Saint-Blancard P, Minvielle F, Le Page P, Kossowski M. Follicular dendritic cell sarcoma of the tonsil: a case report. Am J Otolaryngol 2006; 27: 207-210
- 56 Díaz de Liaño A, Garde C, Artieda C, Yárnoz C, Flores L,

Ortiz H. Intra-abdominal follicular dendritic cell sarcoma. *Clin Transl Oncol* 2006; **8**: 837-838

- 57 Gan MF, Yu CK, Cai JF, Lu HS, Yu XR. Follicular dendritic cell sarcoma of parotid: a case report and review of literatures. *Linchuang Yu Shiyan Binglixue Zazhi* 2006; 22: 204-207
- 58 Jiang L, Admirand JH, Moran C, Ford RJ, Bueso-Ramos CE. Mediastinal follicular dendritic cell sarcoma involving bone marrow: a case report and review of the literature. *Ann Diagn Pathol* 2006; 10: 357-362
- 59 Jiang Y, Yao M, Liu Q. Follicular dendritic cell sarcoma occurring in the pelvic and abdominal cavity, a case report. *Shanghai Yixue Yingxiang* 2006; 15: 174-175
- 60 Kovács RB, Sattar HA, Krausz T, Kas J, Berta M, Sápi Z. Primary follicular dendritic cell sarcoma of the lung. *Histopathology* 2006; 49: 431-433
- 61 Li F, Yu YH, Yao LQ. Follicular dendritic cell sarcoma of nasal cavity: a case report and literature review. *Zhongguo Wuzhenxue Zazhi* 2006; 6: 4715-4717
- 62 Lu CH, Hong M, Luo YD, Zhang ZM, Xu L. Follicular dendritic cell sarcoma occurring in the left groin, a case report. *Fujian Yiyao Zazhi* 2006; **28**: 55-56
- 63 Shen SC, Wu CC, Ng KF, Wu RC, Chen HM, Chen TC. Follicular dendritic cell sarcoma mimicking giant cell carcinoma of the pancreas. *Pathol Int* 2006; **56**: 466-470
- 64 Shia J, Chen W, Tang LH, Carlson DL, Qin J, Guillem JG, Nobrega J, Wong WD, Klimstra DS. Extranodal follicular dendritic cell sarcoma: clinical, pathologic, and histogenetic characteristics of an underrecognized disease entity. *Virchows Arch* 2006; 449: 148-158
- 65 Xu KJ, Li HZ, Shi J, Jin HZ, Liu YH. A case of paraneoplastic pemphigus associated with follicular dendritic cell sarcoma. *Linchuang Pifuke Zazhi* 2006; 35: 794-796
- 66 Chang ZP, Liao SL, Jin Y, Song QP, Duan LJ. [Castleman's disease of chest wall complicated by follicular dendritic cell sarcoma/tumor: report of a case] *Zhenduan Binglixue Zazhi* 2007; 36: 430-431
- 67 Leipsic JA, McAdams HP, Sporn TA. Follicular dendritic cell sarcoma of the mediastinum. AJR Am J Roentgenol 2007; 188: W554-W556
- 68 Padilla-Rodríguez AL, Bembassat M, Lazaro M, Ortiz-Hidalgo C. Intra-abdominal follicular dendritic cell sarcoma with marked pleomorphic features and aberrant expression of neuroendocrine markers: report of a case with immunohistochemical analysis. *Appl Immunohistochem Mol Morphol* 2007; 15: 346-352
- 69 Sander B, Middel P, Gunawan B, Schulten HJ, Baum F, Golas MM, Schulze F, Grabbe E, Parwaresch R, Füzesi L. Follicular dendritic cell sarcoma of the spleen. *Hum Pathol* 2007; 38: 668-672
- 70 **Soriano AO**, Thompson MA, Admirand JH, Fayad LE, Rodriguez AM, Romaguera JE, Hagemeister FB, Pro B. Follicular dendritic cell sarcoma: a report of 14 cases and a review of the literature. *Am J Hematol* 2007; **82**: 725-728
- 71 Yuan J, Li XH, Lu YL, Song X. Pathologic diagnosis of hepatic inflammatory pseudotumor and inflammatory pseudotumor-like follicular dendritic cell tumor. *Linchuang Yu Shiyan Binglixue Zazhi* 2007; 23: 39-42
- 72 **De Pas T**, Spitaleri G, Pruneri G, Curigliano G, Noberasco C, Luini A, Andreoni B, Testori A, de Braud F. Dendritic cell sarcoma: an analytic overview of the literature and presentation of original five cases. *Crit Rev Oncol Hematol* 2008; **65**: 1-7
- 73 **Granados R**, Aramburu JA, Rodríguez JM, Nieto MA. Cytopathology of a primary follicular dendritic cell sarcoma of the liver of the inflammatory pseudotumor-like type. *Diagn Cytopathol* 2008; **36**: 42-46
- 74 Liu YH, Wang XH, Lu LM. Pathological features of follicular cell sarcoma of gastrocolic omentum: a case report and review of the literatures. *Wannan Yixueyuan Xuebao* 2008; 27: 118-120
- 75 **Peng WJ**, Yao LQ. Small mesenteric follicular dendritic cell sarcoma: a case report and literature review. *Zhongguo Wu*-



zhenxue Zazhi 2008; 8: 3040-3042

- 76 Zhang ZX, Cheng J, Shi QL, Ma J, Zhou XJ, Zhou HB, Ma HH. [Follicular dendritic cell sarcoma: a clinicopathologic study of 8 cases] *Zhonghua Binglixue Zazhi* 2008; 37: 395-399
- 77 An XJ, Zhang ZX, Shi QL, Wu B, Ma J, Zhou HB, Ma HH. Hepatic inflammatory pseudotumor-like follicular dendritic cell tmnor: repart of one case and review of literature. *Zhenduanxue Lilun Yu Shijian* 2009; 8: 63-66
- 78 Denning KL, Olson PR, Maley RH Jr, Flati VR, Myers JL, Silverman JF. Primary pulmonary follicular dendritic cell neoplasm: a case report and review of the literature. *Arch Pathol Lab Med* 2009; 133: 643-647
- 79 Liu J, Zhang ZT, Li JS, Wang Y. Hepatic follicular dendritic cell sarcoma: a case report and review of the literatures. *Zhongguo Shiyong Waike Zazhi* 2009; **29**: 421-424
- 80 Romero-Guadarrama MB, Reyes-Posada O, Hernández-González MM, Durán-Padilla MA. Follicular dendritic cell sarcoma/tumor: 2 cases of a rare tumor of difficult clinical and pathological diagnosis. *Ann Diagn Pathol* 2009; 13: 257-262
- 81 Shen DP, Ni XZ, Yin XL, Wu ZY. Clinical and pathological features of follicular dendritic cell sarcoma of appendix: a case report. *Chin Med J* (Engl) 2009; **122**: 1595-1597
- Vaideeswar P, George SM, Kane SV, Chaturvedi RA, Pandit SP. Extranodal follicular dendritic cell sarcoma of the tonsil case report of an epithelioid cell variant with osteoclastic giant cells. *Pathol Res Pract* 2009; 205: 149-153
- 83 Xu YQ, Dai CL, Jia CJ, Yi YF, Shu H, L. PS, Li XJ, Zhao L. Follicular dendritic cell sarcoma of the liver. *Zhonghua Putong Waikexue Wenxian* 2009; **3**: 48-49
- 84 Xie Q, Chen L, Fu K, Harter J, Young KH, Sunkara J, Novak D, Villanueva-Siles E, Ratech H. Podoplanin (d2-40): a new immunohistochemical marker for reactive follicular dendritic cells and follicular dendritic cell sarcomas. *Int J Clin Exp Pathol* 2008; 1: 276-284
- 85 Yu H, Gibson JA, Pinkus GS, Hornick JL. Podoplanin (D2-40) is a novel marker for follicular dendritic cell tumors. *Am J Clin Pathol* 2007; **128**: 776-782
- 86 **Cronin DM**, Warnke RA. Castleman disease: an update on classification and the spectrum of associated lesions. *Adv Anat Pathol* 2009; **16**: 236-246
- 87 Katano H, Kaneko K, Shimizu S, Saito T, Irié T, Mori S. Follicular dendritic cell sarcoma complicated by hyaline-vascular type Castleman's disease in a schizophrenic patient. *Pathol Int* 1997; 47: 703-706
- 88 Kazakov DV, Fanburg-Smith JC, Suster S, Neuhauser TS,

Palmedo G, Zamecnik M, Kempf W, Michal M. Castleman disease of the subcutis and underlying skeletal muscle: report of 6 cases. *Am J Surg Pathol* 2004; **28**: 569-577

- 89 Lin O, Frizzera G. Angiomyoid and follicular dendritic cell proliferative lesions in Castleman's disease of hyalinevascular type: a study of 10 cases. *Am J Surg Pathol* 1997; 21: 1295-1306
- 90 Cokelaere K, Debiec-Rychter M, De Wolf-Peeters C, Hagemeijer A, Sciot R. Hyaline vascular Castleman's disease with HMGIC rearrangement in follicular dendritic cells: molecular evidence of mesenchymal tumorigenesis. *Am J Surg Pathol* 2002; 26: 662-669
- 91 Pauwels P, Dal Cin P, Vlasveld LT, Aleva RM, van Erp WF, Jones D. A chromosomal abnormality in hyaline vascular Castleman's disease: evidence for clonal proliferation of dysplastic stromal cells. *Am J Surg Pathol* 2000; 24: 882-888
- 92 Hernández JL, Gómez-Román J, Ramos-Estébanez C, Nan D, Martín-Oviedo J, Riancho JA, González-Macías J. Human herpesvirus 8 and Epstein-Barr virus coinfection in localized Castleman disease during pregnancy. *Haematologica* 2005; 90 Suppl: ECR35
- 93 Chen CH, Liu HC, Hung TT, Liu TP. Possible roles of Epstein-Barr virus in Castleman disease. J Cardiothorac Surg 2009; 4: 31
- 94 Maeda E, Akahane M, Kiryu S, Kato N, Yoshikawa T, Hayashi N, Aoki S, Minami M, Uozaki H, Fukayama M, Ohtomo K. Spectrum of Epstein-Barr virus-related diseases: a pictorial review. Jpn J Radiol 2009; 27: 4-19
- 95 Hollstein M, Sidransky D, Vogelstein B, Harris CC. p53 mutations in human cancers. *Science* 1991; **253**: 49-53
- 96 Abdel-Fattah R, Challen C, Griffiths TR, Robinson MC, Neal DE, Lunec J. Alterations of TP53 in microdissected transitional cell carcinoma of the human urinary bladder: high frequency of TP53 accumulation in the absence of detected mutations is associated with poor prognosis. *Br J Cancer* 1998; 77: 2230-2238
- 97 Hainaut P, Milner J. Interaction of heat-shock protein 70 with p53 translated in vitro: evidence for interaction with dimeric p53 and for a role in the regulation of p53 conformation. *EMBO J* 1992; **11**: 3513-3520
- 98 Fritsche M, Haessler C, Brandner G. Induction of nuclear accumulation of the tumor-suppressor protein p53 by DNAdamaging agents. Oncogene 1993; 8: 307-318
- 99 Siegel J, Fritsche M, Mai S, Brandner G, Hess RD. Enhanced p53 activity and accumulation in response to DNA damage upon DNA transfection. *Oncogene* 1995; 11: 1363-1370

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