



Published in final edited form as:

Minerva Chir. 2009 October ; 64(5): 489–500.

Pancreatic cancer stem cells and EMT in drug resistance and metastasis

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Abstract

Pancreatic cancer (PC) is an aggressive malignancy with one of the worst outcomes among all cancers. It is the fourth leading cause of cancer death in the United States with a very low five-year survival rate. The high mortality of PC could, in part, be due to their drug resistance characteristics and high propensity for metastasis. Recently, cancer stem cells (CSCs) and epithelial-mesenchymal transition (EMT)-type cells, which shares molecular characteristics with CSCs, have been believed to play critical roles in drug resistance and cancer metastasis as demonstrated in several human malignancies including PC. Thus, the discovery of molecular knowledge of drug resistance and metastasis in relation to CSCs and EMT in PC is becoming an important area of research, and such knowledge is likely to be helpful in the discovery of newer drugs as well as designing novel therapeutic strategies for the treatment of PC with better outcome. In this brief review, we will summarize the current knowledge regarding the CSCs and EMT in the context of drug resistance and metastasis in PC, the molecular events occurring in CSCs and EMT, and the design of novel therapeutic strategies targeting CSCs and EMT-type cells to increase drug sensitivity and suppression of metastasis toward better treatment outcome of patients diagnosed with PC.

Keywords

Pancreatic cancer; cancer stem cells (CSCs); epithelial-mesenchymal transition (EMT)-type cells; drug resistance; metastasis

Introduction

Pancreatic cancer (PC) is an aggressive malignancy with one of the worst outcomes among all cancers. It is the fourth leading cause of cancer death in the United States¹. For all stages combined, the relative rate of 1-year survival is 24% and only 5% of all patients with PC will survive five years after diagnosis¹. Even for those people diagnosed with local disease, the 5-year survival is as low as 20%¹. Approximately, 42,470 people are expected to be diagnosed with PC and 35,240 people are expected to die from this disease in the US in 2009². The incidence rates of PC have been increasing in women by 0.6% per year since 1994 and the death rate for PC in women has been increasing by 0.1% per year since 1984¹. The lethal nature of PC stems from its propensity to rapidly disseminate to the lymphatic system and distant organs. The presence of occult or clinical metastases at the time of diagnosis together with the lack of effective chemotherapies contributes to the high mortality of patients diagnosed with PC, which indeed believed to be due to drug-resistant behavior of PC. The aggressiveness

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of PC could, in part, be due to their intrinsic and extrinsic drug resistance characteristics, and thus the cancer cell resistance to chemotherapeutic agents is a major cause of treatment failure in PC. Therefore, increasing drug sensitivity and inhibiting metastasis are the key steps toward successful treatment of patients diagnosed with this devastating disease.

Recently, cancer stem cells (CSCs) and epithelial-mesenchymal transition (EMT)-type cells have been believed to play critical roles in drug resistance and cancer metastasis. Thus, the molecular knowledge of drug resistance and metastasis with respect to CSCs and EMT in PC is considered very important, and the gain of such knowledge is likely to be helpful not only in the discovery of newer drugs but also in the design of novel therapeutic strategies for the treatment of PC with better outcome. The following sections will summarize what we know regarding CSCs and EMT in the context of drug resistance and metastasis in PC, the molecular events that occurs in CSCs and EMT, and the design of novel therapeutic strategies targeting CSCs and EMT for increasing drug sensitivity and the suppression of metastasis toward better treatment outcome of patients diagnosed with PC.

Pancreatic cancer stem cells (CSCs)

It has been well known that human cancer is composed of a heterogeneous population of cells which have different proliferative potential and they response differentially to chemotherapy and radiotherapy. Among cancer cells, a small population of cancer cells has been identified as cancer stem cells (CSCs) which possess the ability to self-renew and generate the diverse cell population that comprise the cancer mass³. A growing body of evidence now supports the concept that cancers are diseases driven by subpopulation of self renewing CSCs, which have been found in hematopoietic^{4,5} and solid tumors including brain tumor⁶, breast⁷, head and neck^{8,9}, colon¹⁰, lung¹¹, prostate¹², ovarian¹³, and PC^{14,15}. The CSCs not only possess the capacity for self-renewal but also have the potential and the ability to drive continued expansion of the population of malignant cells with invasive and metastatic propensity¹⁶. Moreover, CSCs also show resistance to a number of conventional therapies including chemotherapy and radiotherapy^{17,18}, which may explain why it is difficult to completely eradicate cancer and why recurrence is an ever-present threat.

To identify and isolate CSC from whole tumor cell population, several cell surface markers including CD24, CD34, CD44, CD133, CD139, CD166, and ESA have been used together with cell sorting^{13,19–22} by flow cytometry (FACS analysis). Among them, CD44, CD133, and ESA are more frequently used for the isolation of CSCs from different type of tumors. However, it is important to note that no unique marker for isolation of CSCs from different types of tumors has been found so far, suggesting that combination of several markers could increase the purity of isolated CSCs.

In 2007, two groups of investigators found the presence of CSCs in human PC using two different sets of cell surface markers^{14,15}. Li et al firstly described that the CD44⁺/CD24⁺/ESA⁺ PC cells, which counted for 0.2–0.8% of PC cells, showed the stem cell properties of self-renewal, the ability to produce differentiated progeny, and increased expression of the developmental signaling molecule sonic hedgehog¹⁴. They also found that PC cells with the CD44⁺/CD24⁺/ESA⁺ phenotype isolated from PC tissue had a 100-fold increased tumorigenic potential compared with marker-negative cancer cells. Furthermore, 50% of animals injected with as few as 100 CD44⁺/CD24⁺/ESA⁺ cells formed tumors that were histologically indistinguishable from the original human tumors¹⁴. Shortly thereafter, Hermann et al reported that human PC tissue contained CSCs defined by CD133 expression¹⁵. These CD133⁺ PC cells were exclusively tumorigenic. The results from these two groups clearly demonstrated that there is a small population of PC cells which possess the distinct cell surface markers commonly found in other organ-specific CSCs, and these cells showed properties of self-

renewal, and the increased tumorigenic potential, suggesting that these cells are authentic PC-specific CSCs. Moreover, their data suggest that a combination of the cell surface marker CD44⁺/CD24⁺/CD133⁺/ESA⁺ could be used to isolate the pancreas-specific CSCs from human PC tissues.

Pancreatic CSCs are drug resistance with propensity to metastasis

Over the past years, emerging evidences have shown that CSCs are largely involved in the drug resistance and metastasis^{23–29}. The CSCs also contribute to radioresistance through preferential activation of the DNA damage checkpoint response and an increase in DNA repair capacity¹⁷. It has been found that the fraction of glioma cells expressing CD133, a marker for brain cancer stem cells, was enriched after radiation in gliomas¹⁷. The CD133⁺ glioma cells survived after ionizing radiation showed increased proportions relative to most tumor cells which lack CD133 expression. These results suggest that the CD133⁺ tumor cells represent glioma CSCs that confers glioma radioresistance. It has been reported that CSCs in mouse mammary tumors may contribute to cisplatin resistance in a mouse model³⁰. Similarly, CSCs in colorectal cancers are also believed to be responsible for the resistance to chemotherapeutic drugs^{10,31}. Charafe-Jauffret et al have isolated breast cancer cell lines which contain functional CSCs with high metastatic capacity and a distinct molecular signature of CSCs²⁷, suggesting that breast cancer CSCs are involved in breast cancer metastasis.

It has been known that pancreatic CSCs are implicated in drug resistance and that the drug resistant cells from PC are more tumorigenic and metastatic *in vitro* and *in vivo*. Hermann et al have found that human PC tissues contains pancreatic CD133⁺ CSCs that are exclusively tumorigenic and highly resistant to standard chemotherapy¹⁵. They also observed a distinct subpopulation of CD133⁺/CXCR4⁺ CSCs in the invasive front of pancreatic tumors, suggesting the metastatic phenotype of the individual tumor cells. Depletion of the CSC pool for these migrating CSCs virtually abrogated the metastatic phenotype of pancreatic tumors without affecting their tumorigenic potential. These findings demonstrate that pancreatic CSCs contribute to the drug resistance and metastasis. In another study investigating pancreatic CSCs in drug resistance, high-dose gemcitabine treatment was used to eliminate most of the cancer cells³². After the treatment, CD44⁺ cells proliferated and reconstituted the population of resistant cells, suggesting that pancreatic CSCs are gemcitabine resistant. In human PC samples, CD44 expression was correlated with histologic grade, and the patients with CD44-positive tumors showed poor prognosis³². These data suggest that PC stem-like cells were expanded during the acquisition of gemcitabine resistance.

EMT in pancreatic cancer

EMT is a process by which epithelial cells undergo remarkable morphological changes characterized by a transition from epithelial cobblestone phenotype to elongated fibroblastic phenotype. EMT was originally identified as a crucial differentiation and morphogenetic physiological process during embryogenesis³³. Currently, EMT is recognized as pathological mechanism during the progression of various diseases including inflammation, fibrosis and cancers. Alterations in the expression of critical molecules have been observed during the acquisition of EMT phenotype, consistent with their association in cellular signal transduction pathways. In the processes of EMT, cells lose epithelial cell-cell junction, actin cytoskeleton reorganization, and the expression of proteins that promote cell-cell contact such as E-cadherin, γ -catenin, and zonula occludens-1 (ZO-1), and gain mesenchymal molecular markers such as vimentin, fibronectin, α -smooth muscle actin (SMA), fibroblast-specific protein-1, and N-cadherin³⁴. Under most circumstances, the cells that are undergoing EMT show down-regulation of E-cadherin which is a cell-cell adhesion molecule and a calcium-dependent transmembrane glycoprotein present in the most epithelial cells in adult tissues. Emerging

evidence also suggest that the process of EMT is triggered by complex molecular interplay between extra-cellular signals and growth factors. The extra-cellular signals and growth factors involved in the EMT process include type I and III collagen, matrix metalloproteinases-2 (MMP-2), MMP-3, MMP-9, transforming growth factor- β (TGF- β), fibroblast growth factor (FGF), epidermal growth factor (EGF) and platelet-derived growth factor (PDGF) A and B^{35–39}. Moreover, a number of factors that transcriptionally repress E-cadherin and regulate other EMT markers have emerged as potent EMT drivers during normal development and cancer progression. The transcription factors involved in the regulation of EMT markers include the zinc finger binding transcription factors Snail homologues (Snail1, Snail2/Slug, and Snail3) and several other basic helix-loop-helix transcription factors such as Twist, ZEB1, ZEB2/SIP1, and TCF3/E47/E12^{40,41}. It is now more clear that the down-regulation and relocation of E-cadherin and ZO-1, the translocation of β -catenin from cell membrane to nucleus, the overexpression of vimentin and fibronectin, and the activation of Slug, Twist, and ZEB1 transcription factors, result in the induction of EMT phenotype^{40,42–47}.

We and other investigators have reported the EMT phenomenon in several PC cell lines and surgically resected pancreatic cancers^{48–55}. We found that several PC cell lines including L3.6pl, Colo357, BxPC-3, and HPAC cells showed strong expression of epithelial marker E-cadherin while another group of PC cells (MiaPaCa-2, Panc-1, and Aspc-1 cells) exhibited strong expression of mesenchymal makers including vimentin and ZEB-1 at mRNA and protein levels⁴⁸. Arumugam et al also reported similar observations showing that L3.6pl, BxPC-3, CFPAC-1, and SU86.86 cells had high expression of E-cadherin and low expression of ZEB-1 while low level of E-cadherin and high level of ZEB-1 were observed in PANC-1, Hs766T, AsPC-1, MIAPaCa-2, and MPanc96 cells⁴⁹. These results suggest that a population of distinct PC cells, which show EMT characteristics, exists in pancreatic tumors. These EMT cells in PC could be responsible for the progressive characteristics of PC. In a study investigating the expression of EMT markers such as low E-cadherin, high fibronectin, and vimentin in resected human PC, immunohistochemical staining was performed on formalin-fixed paraffin-embedded tissues using monoclonal antibodies against vimentin, fibronectin, E-cadherin, and p-Erk⁵³. It has been found that fibronectin overexpression correlated with the presence of vimentin ($p = 0.0048$) and activated Erk ($p = 0.0264$). There was a borderline association of fibronectin with worsening grade ($p = 0.06$). Importantly, increased fibronectin or vimentin and decreased E-cadherin correlated with poor survival, suggesting that EMT is associated with poor survival in surgically resected pancreatic adenocarcinoma⁵³. The expression level of E-cadherin, N-cadherin, vimentin, and TGF- β was also investigated in pancreatic primary and metastatic tumors⁵⁵. N-cadherin expression was observed in 43% (13/30) of primary tumors and in 53% (8/15) of metastatic tumors. N-cadherin expression correlated with neural invasion ($P = 0.008$) and histological type ($P = 0.043$). Vimentin was observed in a few cancer cells of primary tumor but was substantially expressed in liver metastasis. Moreover, it is known that TGF could stimulate N-cadherin and vimentin protein expression and decrease E-cadherin expression in Panc-1 cells with morphological changes⁵⁵. These studies provide the morphological and molecular evidence of EMT in PC cell lines and clinical pancreatic carcinoma tissues.

EMT is implicated in drug resistance and metastasis in PC

Recent studies have shown that EMT is associated with drug resistance and cancer cell metastasis^{34,56–60}. Emerging evidence have implicated EMT with the conversion of early stage tumors into invasive malignancies accompanied by increased cell motility and invasion⁶¹, and these processes are consistent with the acquisition of “cancer stem-like cell” phenotype that is also known as “stemness”⁶². For most epithelial tumors, progression toward malignancy is accompanied by a loss of epithelial differentiation and a shift towards mesenchymal phenotype, leading to enhanced cancer cell migration and invasion⁶¹. Studies have also shown

that cancer cells in the tumor center remain positive for the expression of E-cadherin and cytoplasmic β -catenin; however, the tumor cells in the periphery with propensity for invasion display loss of surface E-cadherin and up-regulation of vimentin, the typical characteristics of EMT phenotype⁶³. Highly invasive breast cancer cells have been studied and selected, and these selected invasive cells displayed EMT characteristics and dramatically enhanced invasive ability with decreased level of E-cadherin and increased vimentin, fibronectin, Twist, and AKT2⁶⁰. Recent findings also suggest that metastasis may be critically dependent on the ability of cancer cells to acquire EMT characteristics^{33,64}. EMT has been shown to be important on conferring drug resistance characteristics to cancer cells against conventional therapeutics including taxol, vincristine, oxaliplatin, or epidermal growth factor receptor (EGFR) targeted therapy^{59,65}, which is consistent with drug resistance characteristics of CSCs.

We have reported that EMT in pancreatic cancer influences the sensitivity of cancer cells to drug treatment. We found that gemcitabine-sensitive PC cells including L3.6pl, Colo357, BxPC-3, and HPAC cells showed strong expression of epithelial marker E-cadherin while gemcitabine-resistant PC cells, MiaPaCa-2, Panc-1, and Aspc-1 exhibited strong expression of mesenchymal makers including vimentin and ZEB-1 at mRNA and protein levels, suggesting that the gemcitabine-resistant cells possess EMT characteristics⁴⁸. Arumugam et al also reported that L3.6pl, BxPC-3, CFPAC-1, and SU86.86 cells were sensitive while PANC-1, Hs766T, AsPC-1, MIAPaCa-2, and MPanc96 cells which showed EMT characteristics were resistant to three conventional chemotherapeutic agents, gemcitabine, 5-fluorouracil (5-FU), and cisplatin⁴⁹, consistent with our findings. EMT is also related to the high metastatic ability of PC cells. To study the role of EMT in PC metastasis, highly metastatic PC cell populations were selected by serial *in vivo* passaging of parental cells with low metastatic potential and characterized by global gene expression profiling, chromatin immunoprecipitation, and *in vivo* metastatic assay⁶⁶. The results showed that *in vivo* selection of highly metastatic PC cells induced EMT with the loss of E-cadherin expression and the up-regulation of mesenchymal marker Snail. Inactivation of E-cadherin in parental cells also induced EMT and increased metastasis *in vivo*. Silencing of E-cadherin in highly metastatic cells was mediated by a transcriptional repressor complex containing Snail, HDAC1, and HDAC2. These results suggest that EMT is responsible for the high propensity of metastasis in PC.

Molecular alterations in the formation of CSCs and EMT

Because CSCs and EMT have been believed to be responsible for drug resistance and cancer metastasis, it is important to uncover the molecular mechanism(s) involved in the formation of CSCs and EMT so that novel therapeutic strategies could be designed for the inhibition of cancer progression and metastasis. The formation of CSCs and EMT is a dynamic process and it is triggered by the interplay of multiple cellular signaling pathways such as Hedgehog, Notch, PDGF, Wnt, TGF- β , Akt, and NF- κ B signaling pathways^{50,51,67-71}. Emerging evidence also implicated the critical role of several microRNAs (miRNAs) in the processes of EMT^{72,73}. In addition, experimental evidence has shown that the presence of hypoxia can also induce EMT^{43,74}.

Deregulation of Hedgehog signaling has been found in the formation of CSCs and EMT. Hedgehog (Hh) signaling is an essential pathway for embryonic pancreatic development. It has been known that Hh signaling plays important roles in proper tissue morphogenesis and organ formation during the developing gastrointestinal tract. Hedgehog ligands including sonic hedgehog (Shh) are expressed throughout the endodermal epithelium at early embryonic stages but excluded from the region that forms the pancreas. Deregulation of the Hh pathway has been implicated in a variety of cancers including PC. It has been reported that overexpression of Shh may contribute to pancreatic tumorigenesis. Thayer et al found that no Shh was detected

in the islets, acini, or ductal epithelium of normal pancreas while Shh was aberrantly expressed in 70% of specimens from the patients with pancreatic adenocarcinoma, suggesting that Shh is a mediator of pancreatic cancer tumorigenesis⁷⁵. The down-regulation of Shh by cyclopamine, an inhibitor of Shh, can reduce the growth and viability of PC cells, suggesting that targeting Shh signaling may be an effective novel approach for the treatment of PC⁷⁶. Importantly, pancreatic CSCs demonstrate up-regulation of sonic hedgehog¹⁴, and the down-regulation of hedgehog signaling by the inhibitors of hedgehog resulted in the inhibition of CSCs and EMT with down-regulation of Snail and up-regulation of E-cadherin, leading to the inhibition of invasion and metastasis in PC^{77,78}.

In addition to the hedgehog signaling, another important developmental signaling pathways such as Notch signaling is involved in cell proliferation, survival, apoptosis, and differentiation which affects the development and function of many organs⁷⁹. To date, in mammals, the Notch family of trans-membrane receptors consists of four members: Notch-1-4. Mammals also express Notch ligands including Dll-1 (Delta-like 1), Dll-3 (Delta-like 3), Dll-4 (Delta-like 4), Jagged-1 and Jagged-2⁷⁹. Notch signaling is initiated when Notch ligand binds to an adjacent Notch receptor between two neighboring cells. Upon activation, Notch is cleaved, releasing the intracellular domain of the Notch (ICN) through a cascade of proteolytic cleavages. Recently, Notch signal pathway has been found to be a key regulator in the induction of EMT^{70,71,80}. Notch activation in endothelial cells results in morphological, phenotypic, and functional changes consistent with mesenchymal transformation. These changes include down-regulation of endothelial markers (VE-cadherin, Tie1, Tie2, platelet-endothelial cell adhesion molecule-1, and endothelial NO synthase) and up-regulation of mesenchymal markers (α -SMA, fibronectin, and platelet-derived growth factor receptors)⁸¹. Therefore, it is believed that Notch-induced EMT is restricted to cells expressing activated Notch. Moreover, Jagged-1 stimulation of endothelial cells is known to induce a similar mesenchymal transformation, suggesting that Jagged-1 mediated activation of Notch signaling is important during the induction of EMT⁸¹. Notch also interacts with several transcription and growth factors including Snail, Slug, and TGF- β in the processes of EMT. Notch has been shown to promote EMT through the regulation of Snail. For example, over-expression of Notch-1 in immortalized endothelial cells *in vitro* could induce Snail⁷⁰, which could bind to the E-boxes of human E-cadherin promoter and function as a repressor of E-cadherin gene expression⁸². Thus, Notch induced Snail over-expression is likely to down-regulate E-cadherin expression, resulting in the acquisition of EMT. In addition, Notch could induce EMT through stabilizing Snail-1 protein under hypoxic condition⁸⁰. It has been reported that Slug is a direct target of Notch and that the Notch signaling directly stimulates Slug promoter, resulting in the up-regulation of Slug and the formation of EMT⁸³. In addition, Slug was found to be essential for Notch-mediated repression of E-cadherin, resulting in β -catenin activation and EMT⁸⁴. It has been found that TGF- β can induce the expression of Notch ligands⁸⁵ and that TGF- β -induced EMT could be blocked by Hey-1 or Jagged-1 RNA silencing or by chemical inactivation of Notch⁷¹. We have also found that Notch-2 and its ligand, Jagged-1, are highly up-regulated in gemcitabine resistant PC cells, which show the acquisition of EMT phenotype⁵¹. These findings clearly suggest the important roles of Notch signaling in the formation of EMT.

Notch ligand, Jagged-1 has been found to be an evolutionarily conserved target of Wnt/ β -catenin signaling pathway in progenitor cells, suggesting the participation of Wnt signaling in CSCs and EMT⁸⁶. Indeed, Wnt signaling has been shown to play important roles in both self renewal and carcinogenesis in a variety of cancer. Jagged-1 expression on progenitor cells induces self-renewal of stem cells due to canonical Wnt signaling activation and Notch signaling activation⁸⁶, demonstrating that Wnt and Notch signaling are important in maintaining the homeostasis of stem and progenitor cells. Moreover, inhibition of Wnt signaling resulted in the re-expression of breast epithelial differentiation markers and repression of EMT transcription factors Slug and Twist⁵⁷. In colorectal cancer cell lines,

overexpression of Snail leads to increased expression of Wnt target genes through the interaction with β -catenin whereas down-regulation of endogenous Snail by siRNA reduces target gene expression⁴². Collectively, these results provide a molecular link between self-renewal, EMT, and Wnt signaling.

Moreover, PDGF signaling has been reported to regulate the expression of the Notch-1 and play important roles in cancer cell growth, invasion and metastasis. Importantly, we have found that PDGF signaling contributes to EMT phenotype, resulting in the aggressiveness of tumor cells such as increased invasion and angiogenesis characteristics⁸⁷. We observed that overexpression of PDGF-D resulted in a significant induction of EMT as shown by changes in cellular morphology concomitant with the loss of E-cadherin and ZO-1, and gain of vimentin⁸⁷, suggesting the important roles of PDGF signaling in the induction of EMT phenotype.

In recent years, emerging evidence implicated the critical role of microRNAs (miRNAs) in the processes of EMT. The miRNAs elicit their regulatory effects by imperfectly binding to the 3' untranslated region (3'UTR) of target mRNA, causing either degradation of mRNA or inhibition of their translation to functional proteins^{88,89}. The expressions of miRNAs have been recognized as integral components of many normal biological processes, such as those involving cell proliferation, differentiation, apoptosis, and stress resistance⁹⁰. Moreover, it has been recently suggested that aberrant expression of miRNAs is associated with the development and progression cancer. More importantly, miRNAs have been found to be the regulators of EMT through the regulation of E-cadherin and other molecules⁷², and thus miRNA appears to play important roles in cancer development and progression^{91,92}. We have investigated the expression level of miR-200 and let-7 in pancreatic EMT-type cells. We found that miR-200b, miR-200c, and miR-200a were down-regulated in gemcitabine-resistant cells, which is consistent with the EMT phenotype of gemcitabine-resistant cells. We also found that many members of the tumor suppressor let-7 family were down-regulated in gemcitabine-resistant cells (EMT-type cells) and these findings are consistent with the aggressiveness of gemcitabine-resistant PC cells⁴⁸. In order to investigate the role of miR-200 family in the reversal of EMT phenotype of gemcitabine-resistant cells, we transfected miR-200a, miR-200b, and miR-200c pre-microRNAs into MiaPaCa-2 cells (gemcitabine resistant) for up to 14 days. We found that the re-expression of miR-200 family in the MiaPaCa-2 cells resulted in the up-regulation of epithelial marker E-cadherin and down-regulation of mesenchymal markers including ZEB1 and vimentin both at the mRNA and protein levels. After 14 days of transfection, the morphology of miR-200 transfected MiaPaCa-2 cells was partially changed from elongated fibroblastoid to epithelial cobblestone-like appearance, and the cells appeared to grow in close contact with each other. These results suggest that the loss of miR-200 family is critical for the acquisition of EMT characteristics and that the re-expression of miR-200 could reverse the EMT phenotype of gemcitabine-resistant cells. Moreover, we have also found that re-expression of miR-200b in PDGF-D overexpression cells led to the reversal of EMT phenotype, which was associated with the down-regulation of ZEB1, ZEB2 and Slug expression and these results were consistent with increased gene expressions of epithelial markers⁹³. Moreover, transfection of PDGF-D overexpression cells with miR-200b also inhibited cell migration and invasion with concomitant repression of cell adhesion to culture surface and cell detachment⁹³. These findings suggest that miRNAs play important roles in the formation of EMT in cancers including PC. The deregulation of other signaling pathways and molecules (i.e. TGF- β , Akt, PTEN, GSK-3 β , NF- κ B, etc) also plays roles in the development of CSCs and EMT. The *in vitro* and *in vivo* findings reviewed above demonstrate that a complex signaling network regulates the formation of CSCs and EMT.

Targeting CSCs and EMT to increase drug sensitivity and inhibition of metastasis

Because CSCs and EMT-type cells play important roles in drug resistance and metastasis, novel inhibitors of EMT or agents that could either reverse the EMT phenotype or kill CSCs or EMT-type cells would be a novel strategy for the treatment of cancers including PC, which are highly drug resistant. Agents that could either reduce drug resistance or reverse it would likely be useful for increasing therapeutic activity of conventional therapeutics. Therefore, discovery of precise mechanisms that govern the processes of EMT and the maintenance of cancer stem cell reservoir would likely lead to the exploitation of novel strategies for the inhibition of tumor progression and/or the killing of cancer stem cells in order to achieve complete eradication of tumors. Indeed, very recently, Gupta et al. identified a selective agent that kills cancer stem-like cells (EMT-type cells) very efficiently⁹⁴. They found that salinomycin destroyed cancer stem-like cells selectively. When compared to paclitaxel, which is a commonly used drug for the treatment of many malignancies including breast cancer, salinomycin reduced the proportion of cancer stem cells by more than 100-fold and inhibited mammary tumor growth in mice, which was associated with increased epithelial differentiation of tumor cells⁹⁴.

Recently, dietary “natural” compounds have been found to regulate EMT in cancers. We have previously found that isoflavone genistein inhibited cell growth and induced apoptosis in various cancers including PC. We have also found that isoflavone genistein treatment significantly inhibited Akt activity, NF- κ B DNA-binding activity, and Notch activation in PC *in vitro* and *in vivo*^{95,96}. We have also found that 3,3'-diindolylmethane (DIM) inhibited cell proliferation and induced apoptotic cell death in a variety of cancers including PC through the inhibition of Akt and NF- κ B signaling⁹⁷. Therefore, we have investigated whether isoflavone and DIM could reverse EMT⁴⁸. Indeed, we found that DIM and isoflavone treatments increased the level of miR-200 family in MiaPaCa-2 cells (EMT-type cells). After the treatment of PC cells with low concentration of DIM (5 μ M) and isoflavone (10 μ M) for 21 days, we found up-regulation of epithelial marker E-cadherin and down-regulation of mesenchymal markers, ZEB1, vimentin and slug, at different levels. Immunofluorescence staining showed that E-cadherin was re-distributed in the cytoplasm closer to cell membrane after DIM and isoflavone treatments. Importantly, we also observed that the morphology of MiaPaCa-2 cells changed from elongated fibroblastoid to epithelial cobblestone-like appearance, and it seems that the cell-cell contact was increased after the treatments. These results are consistent with the changes in the expression of miRNAs and EMT markers. We also noted that some PC cells treated with isoflavone showed more elongated shape which was commonly seen in genistein treated cells; however, isoflavone showed stronger effect on E-cadherin expression while DIM exerted stronger effect on ZEB1 expression. These results suggest that both DIM and isoflavone could regulate the expression of EMT markers and indeed could be useful for the reversal of EMT phenotype. Interestingly, the combined effects of isoflavone and DIM could be even better, which requires further in-depth investigation. We further assessed the sensitivity of gemcitabine-resistant cells to gemcitabine after being transfected with miR-200b or treated with DIM or isoflavone. We found that the sensitivity to gemcitabine was significantly increased after miR-200 re-expression compared to control cells. The cells transfected with miR-200b showed 20.8% to 38.2% more inhibition compared to control. The pre-treatment of MiaPaCa-2 cells with DIM or isoflavone also increased the sensitivity of MiaPaCa-2 cells to gemcitabine. These results suggest that miR-200 re-expression, and DIM or isoflavone treatment could partially increase the sensitivity of gemcitabine-resistant cells to gemcitabine possibly through miR-200 mediated reversal of EMT status. These results suggest that isoflavone and DIM could be useful for the treatment of PC especially in combination with existing therapeutic agents.

We have also found that Notch-2 and its ligand, Jagged-1, are highly up-regulated in gemcitabine resistant PC cells, which show EMT phenotype as stated earlier⁵¹. Importantly, we found that down-regulation of Notch signaling by siRNA approach led to partial reversal of the EMT phenotype, resulting in the mesenchymal-epithelial transition (MET), which was associated with decreased expression of vimentin, ZEB1, Slug, Snail, and NF- κ B. These results suggest that the inactivation of Notch signaling by novel strategies could be a potential targeted therapeutic approach for overcoming chemoresistance toward the treatment of PC. In addition, novel strategies targeting Hedgehog signaling by using Hh inhibitors could also be used to suppress CSCs and EMT phenotype for the treatment of PC in the near future.

Conclusion and perspectives

The data from *in vivo* human and animal studies and *in vitro* experiments described above clearly suggest the existence of CSCs and EMT-type cells, which confer drug resistance and leads to metastasis in PC. The regulation of multiple signaling pathways including Hedgehog, Notch, Wnt, PDGF, TGF- β , Akt, NF- κ B, and miRNA contributes to the sustenance of CSCs and EMT phenotypic cells that shares many molecular characteristics with CSCs. With the discovery of more molecular knowledge of CSCs and EMT-type cells, novel therapeutic strategies could be designed to target CSCs and EMT-type cells to increase drug sensitivity, which will thereby suppress tumor progression and metastasis. However, more *in vivo* animal studies are needed to investigate the novel therapeutic strategies targeting CSCs and EMT. Recently, Jimeno et al reported a direct PC xenograft model which could be useful as a platform for cancer stem cell therapeutic development⁹⁸. In addition, there are still many unresolved issues regarding CSCs and EMT, such as highly impure population of CSCs, varied reported frequencies of CSCs in the same or different types of tumors, relatively poor overlap between the different makers reported for the CSCs in a given tumor type, the stability of CSC phenotype over time, the relationship between CSCs and EMT, and the complexity of the signaling networks that regulate EMT induction. Nevertheless, owing to the clinical importance of CSCs and EMT in PC, targeting CSCs and EMT-type cells would become an attractive and novel strategy for the treatment of patients diagnosed with PC for which better therapy is urgently needed.

Acknowledgments

The authors' work cited in this review was funded by grants from the National Cancer Institute, NIH (5R01CA083695, 5R01CA131151, and 1R01CA132794 awarded to FHS), a sub-contract award to FHS from the University of Texas MD Anderson Cancer Center through a SPORE grant (5P20-CA101936) on pancreatic cancer awarded to James Abbruzzese.

References

1. American Cancer Society. Cancer facts & figures 2009. Atlanta: American Cancer Society; 2009. p. 18-9.
2. Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ. Cancer statistics. *CA Cancer J Clin* 2009;59:225–49. [PubMed: 19474385]
3. Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001;414:105–11. [PubMed: 11689955]
4. Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat Med* 1997;3:730–7. [PubMed: 9212098]
5. Lapidot T, Sirard C, Vormoor J, Murdoch B, Hoang T, Caceres-Cortes J, et al. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature* 1994;367:645–8. [PubMed: 7509044]
6. Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, et al. Identification of a cancer stem cell in human brain tumors. *Cancer Res* 2003;63:5821–8. [PubMed: 14522905]

7. Al-Hajj M, Wicha MS, Ito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci U S A* 2003;100:3983–8. [PubMed: 12629218]
8. Ailles L, Prince M. Cancer stem cells in head and neck squamous cell carcinoma. *Methods Mol Biol* 2009;568:175–93. [PubMed: 19582427]
9. Prince ME, Sivanandan R, Kaczorowski A, Wolf GT, Kaplan MJ, Dalerba P, et al. Identification of a subpopulation of cells with cancer stem cell properties in head and neck squamous cell carcinoma. *Proc Natl Acad Sci U S A* 2007;104:973–8. [PubMed: 17210912]
10. Todaro M, Alea MP, Di Stefano AB, Cammareri P, Vermeulen L, Iovino F, et al. Colon cancer stem cells dictate tumor growth and resist cell death by production of interleukin-4. *Cell Stem Cell* 2007;1:389–402. [PubMed: 18371377]
11. Eramo A, Lotti F, Sette G, Pillozzi E, Biffoni M, Di VA, et al. Identification and expansion of the tumorigenic lung cancer stem cell population. *Cell Death Differ* 2008;15:504–14. [PubMed: 18049477]
12. Gu G, Yuan J, Wills M, Kasper S. Prostate cancer cells with stem cell characteristics reconstitute the original human tumor in vivo. *Cancer Res* 2007;67:4807–15. [PubMed: 17510410]
13. Zhang S, Balch C, Chan MW, Lai HC, Matei D, Schilder JM, et al. Identification and characterization of ovarian cancer-initiating cells from primary human tumors. *Cancer Res* 2008;68:4311–20. [PubMed: 18519691]
14. Li C, Lee CJ, Simeone DM. Identification of human pancreatic cancer stem cells. *Methods Mol Biol* 2009;568:161–73. [PubMed: 19582426]
15. Hermann PC, Huber SL, Herrler T, Aicher A, Ellwart JW, Guba M, et al. Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell* 2007;1:313–23. [PubMed: 18371365]
16. Jordan CT, Guzman ML, Noble M. Cancer stem cells. *N Engl J Med* 2006;355:1253–61. [PubMed: 16990388]
17. Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, et al. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature* 2006;444:756–60. [PubMed: 17051156]
18. Li X, Lewis MT, Huang J, Gutierrez C, Osborne CK, Wu MF, et al. Intrinsic resistance of tumorigenic breast cancer cells to chemotherapy. *J Natl Cancer Inst* 2008;100:672–9. [PubMed: 18445819]
19. Huang EH, Hynes MJ, Zhang T, Ginestier C, Dontu G, Appelman H, et al. Aldehyde dehydrogenase 1 is a marker for normal and malignant human colonic stem cells (SC) and tracks SC overpopulation during colon tumorigenesis. *Cancer Res* 2009;69:3382–9. [PubMed: 19336570]
20. Yi JM, Tsai HC, Glockner SC, Lin S, Ohm JE, Easwaran H, et al. Abnormal DNA methylation of CD133 in colorectal and glioblastoma tumors. *Cancer Res* 2008;68:8094–103. [PubMed: 18829568]
21. Yamashita T, Forgues M, Wang W, Kim JW, Ye Q, Jia H, et al. EpCAM and alpha-fetoprotein expression defines novel prognostic subtypes of hepatocellular carcinoma. *Cancer Res* 2008;68:1451–61. [PubMed: 18316609]
22. Patrawala L, Calhoun-Davis T, Schneider-Broussard R, Tang DG. Hierarchical organization of prostate cancer cells in xenograft tumors: the CD44+alpha2beta1+ cell population is enriched in tumor-initiating cells. *Cancer Res* 2007;67:6796–805. [PubMed: 17638891]
23. Bauerschmitz GJ, Ranki T, Kangasniemi L, Ribacka C, Eriksson M, Porten M, et al. Tissue-specific promoters active in CD44+CD24-/low breast cancer cells. *Cancer Res* 2008;68:5533–9. [PubMed: 18632604]
24. Matsui W, Wang Q, Barber JP, Brennan S, Smith BD, Borrello I, et al. Clonogenic multiple myeloma progenitors, stem cell properties, and drug resistance. *Cancer Res* 2008;68:190–7. [PubMed: 18172311]
25. Folkins C, Man S, Xu P, Shaked Y, Hicklin DJ, Kerbel RS. Anticancer therapies combining antiangiogenic and tumor cell cytotoxic effects reduce the tumor stem-like cell fraction in glioma xenograft tumors. *Cancer Res* 2007;67:3560–4. [PubMed: 17440065]
26. Gravdal K, Halvorsen OJ, Haukaas SA, Akslen LA. Proliferation of immature tumor vessels is a novel marker of clinical progression in prostate cancer. *Cancer Res* 2009;69:4708–15. [PubMed: 19487287]

27. Charafe-Jauffret E, Ginestier C, Iovino F, Wicinski J, Cervera N, Finetti P, et al. Breast cancer cell lines contain functional cancer stem cells with metastatic capacity and a distinct molecular signature. *Cancer Res* 2009;69:1302–13. [PubMed: 19190339]
28. Odoux C, Fohrer H, Hoppe T, Guzik L, Stolz DB, Lewis DW, et al. A stochastic model for cancer stem cell origin in metastatic colon cancer. *Cancer Res* 2008;68:6932–41. [PubMed: 18757407]
29. Kleeberger W, Bova GS, Nielsen ME, Herawi M, Chuang AY, Epstein JI, et al. Roles for the stem cell associated intermediate filament Nestin in prostate cancer migration and metastasis. *Cancer Res* 2007;67:9199–206. [PubMed: 17909025]
30. Shafee N, Smith CR, Wei S, Kim Y, Mills GB, Hortobagyi GN, et al. Cancer stem cells contribute to cisplatin resistance in Brca1/p53-mediated mouse mammary tumors. *Cancer Res* 2008;68:3243–50. [PubMed: 18451150]
31. Dylla SJ, Beviglia L, Park IK, Chartier C, Raval J, Ngan L, et al. Colorectal cancer stem cells are enriched in xenogeneic tumors following chemotherapy. *PLoS One* 2008;3:e2428. [PubMed: 18560594]
32. Hong SP, Wen J, Bang S, Park S, Song SY. CD44-positive cells are responsible for gemcitabine resistance in pancreatic cancer cells. *Int J Cancer*. 2009
33. Thierry JP, Sleeman JP. Complex networks orchestrate epithelial-mesenchymal transitions. *Nat Rev Mol Cell Biol* 2006;7:131–42. [PubMed: 16493418]
34. Christiansen JJ, Rajasekaran AK. Reassessing epithelial to mesenchymal transition as a prerequisite for carcinoma invasion and metastasis. *Cancer Res* 2006;66:8319–26. [PubMed: 16951136]
35. Ahmed N, Maines-Bandiera S, Quinn MA, Unger WG, Dedhar S, Auersperg N. Molecular pathways regulating EGF-induced epithelio-mesenchymal transition in human ovarian surface epithelium. *Am J Physiol Cell Physiol* 2006;290:C1532–C1542. [PubMed: 16394028]
36. Fischer AN, Fuchs E, Mikula M, Huber H, Beug H, Mikulits W. PDGF essentially links TGF-beta signaling to nuclear beta-catenin accumulation in hepatocellular carcinoma progression. *Oncogene* 2007;26:3395–405. [PubMed: 17130832]
37. Gotzmann J, Fischer AN, Zojer M, Mikula M, Proell V, Huber H, et al. A crucial function of PDGF in TGF-beta-mediated cancer progression of hepatocytes. *Oncogene* 2006;25:3170–85. [PubMed: 16607286]
38. Strutz F, Zeisberg M, Ziyadeh FN, Yang CQ, Kalluri R, Muller GA, et al. Role of basic fibroblast growth factor-2 in epithelial-mesenchymal transformation. *Kidney Int* 2002;61:1714–28. [PubMed: 11967021]
39. Yang L, Lin C, Liu ZR. P68 RNA helicase mediates PDGF-induced epithelial mesenchymal transition by displacing Axin from beta-catenin. *Cell* 2006;127:139–55. [PubMed: 17018282]
40. Moreno-Bueno G, Portillo F, Cano A. Transcriptional regulation of cell polarity in EMT and cancer. *Oncogene* 2008;27:6958–69. [PubMed: 19029937]
41. Klymkowsky MW, Savagner P. Epithelial-mesenchymal transition: a cancer researcher's conceptual friend and foe. *Am J Pathol* 2009;174:1588–93. [PubMed: 19342369]
42. Stemmer V, de CB, Bex G, Behrens J. Snail promotes Wnt target gene expression and interacts with beta-catenin. *Oncogene* 2008;27:5075–80. [PubMed: 18469861]
43. Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, et al. Direct regulation of TWIST by HIF-1alpha promotes metastasis. *Nat Cell Biol* 2008;10:295–305. [PubMed: 18297062]
44. Yang J, Mani SA, Donaher JL, Ramaswamy S, Itzykson RA, Come C, et al. Twist, a master regulator of morphogenesis, plays an essential role in tumor metastasis. *Cell* 2004;117:927–39. [PubMed: 15210113]
45. Muller T, Bain G, Wang X, Papkoff J. Regulation of epithelial cell migration and tumor formation by beta-catenin signaling. *Exp Cell Res* 2002;280:119–33. [PubMed: 12372345]
46. Graham TR, Zhou HE, Odero-Marrah VA, Osunkoya AO, Kimbro KS, Tighiouart M, et al. Insulin-like growth factor-I-dependent up-regulation of ZEB1 drives epithelial-to-mesenchymal transition in human prostate cancer cells. *Cancer Res* 2008;68:2479–88. [PubMed: 18381457]
47. Spaderna S, Schmalhofer O, Wahlbuhl M, Dimmler A, Bauer K, Sultan A, et al. The transcriptional repressor ZEB1 promotes metastasis and loss of cell polarity in cancer. *Cancer Res* 2008;68:537–44. [PubMed: 18199550]

48. Li Y, VandenBoom TG, Kong D, Wang Z, Ali S, Philip PA, et al. Up-regulation of miR-200 and let-7 by natural agents leads to the reversal of epithelial-to-mesenchymal transition in gemcitabine-resistant pancreatic cancer cells. *Cancer Res* 2009;69:6704–12. [PubMed: 19654291]
49. Arumugam T, Ramachandran V, Fournier KF, Wang H, Marquis L, Abbruzzese JL, et al. Epithelial to mesenchymal transition contributes to drug resistance in pancreatic cancer. *Cancer Res* 2009;69:5820–8. [PubMed: 19584296]
50. Singh A, Greninger P, Rhodes D, Koopman L, Violette S, Bardeesy N, et al. A gene expression signature associated with “K-Ras addiction” reveals regulators of EMT and tumor cell survival. *Cancer Cell* 2009;15:489–500. [PubMed: 19477428]
51. Wang Z, Li Y, Kong D, Banerjee S, Ahmad A, Azmi AS, et al. Acquisition of epithelial-mesenchymal transition phenotype of gemcitabine-resistant pancreatic cancer cells is linked with activation of the notch signaling pathway. *Cancer Res* 2009;69:2400–7. [PubMed: 19276344]
52. Cates JM, Byrd RH, Fohn LE, Tatsas AD, Washington MK, Black CC. Epithelial-mesenchymal transition markers in pancreatic ductal adenocarcinoma. *Pancreas* 2009;38:e1–e6. [PubMed: 18766116]
53. Javle MM, Gibbs JF, Iwata KK, Pak Y, Rutledge P, Yu J, et al. Epithelial-mesenchymal transition (EMT) and activated extracellular signal-regulated kinase (p-Erk) in surgically resected pancreatic cancer. *Ann Surg Oncol* 2007;14:3527–33. [PubMed: 17879119]
54. Yin T, Wang C, Liu T, Zhao G, Zhou F. Implication of EMT induced by TGF-beta1 in pancreatic cancer. *J Huazhong Univ Sci Technol Med Sci* 2006;26:700–2. [PubMed: 17357493]
55. Nakajima S, Doi R, Toyoda E, Tsuji S, Wada M, Koizumi M, et al. N-cadherin expression and epithelial-mesenchymal transition in pancreatic carcinoma. *Clin Cancer Res* 2004;10:4125–33. [PubMed: 15217949]
56. Tsuji T, Ibaragi S, Hu GF. Epithelial-Mesenchymal Transition and Cell Cooperativity in Metastasis. *Cancer Res*. 2009
57. DiMeo TA, Anderson K, Phadke P, Feng C, Perou CM, Naber S, et al. A novel lung metastasis signature links Wnt signaling with cancer cell self-renewal and epithelial-mesenchymal transition in basal-like breast cancer. *Cancer Res* 2009;69:5364–73. [PubMed: 19549913]
58. Kudo-Saito C, Shirako H, Takeuchi T, Kawakami Y. Cancer metastasis is accelerated through immunosuppression during Snail-induced EMT of cancer cells. *Cancer Cell* 2009;15:195–206. [PubMed: 19249678]
59. Fuchs BC, Fujii T, Dorfman JD, Goodwin JM, Zhu AX, Lanuti M, et al. Epithelial-to-mesenchymal transition and integrin-linked kinase mediate sensitivity to epidermal growth factor receptor inhibition in human hepatoma cells. *Cancer Res* 2008;68:2391–9. [PubMed: 18381447]
60. Cheng GZ, Chan J, Wang Q, Zhang W, Sun CD, Wang LH. Twist transcriptionally up-regulates AKT2 in breast cancer cells leading to increased migration, invasion, and resistance to paclitaxel. *Cancer Res* 2007;67:1979–87. [PubMed: 17332325]
61. Thiery JP. Epithelial-mesenchymal transitions in tumour progression. *Nat Rev Cancer* 2002;2:442–54. [PubMed: 12189386]
62. Hugo H, Ackland ML, Blick T, Lawrence MG, Clements JA, Williams ED, et al. Epithelial--mesenchymal and mesenchymal--epithelial transitions in carcinoma progression. *J Cell Physiol* 2007;213:374–83. [PubMed: 17680632]
63. Brabletz T, Jung A, Reu S, Porzner M, Hlubek F, Kunz-Schughart LA, et al. Variable beta-catenin expression in colorectal cancers indicates tumor progression driven by the tumor environment. *Proc Natl Acad Sci U S A* 2001;98:10356–61. [PubMed: 11526241]
64. Kang Y, Massague J. Epithelial-mesenchymal transitions: twist in development and metastasis. *Cell* 2004;118:277–9. [PubMed: 15294153]
65. Sabbah M, Emami S, Redeuilh G, Julien S, Prevost G, Zimmer A, et al. Molecular signature and therapeutic perspective of the epithelial-to-mesenchymal transitions in epithelial cancers. *Drug Resist Updat* 2008;11:123–51. [PubMed: 18718806]
66. von BJ, Eser S, Paul MC, Seidler B, Brandl M, Messer M, et al. E-cadherin regulates metastasis of pancreatic cancer in vivo and is suppressed by a SNAIL/HDAC1/HDAC2 repressor complex. *Gastroenterology* 2009;137:361–71. 371. [PubMed: 19362090]

67. Huber MA, Azoitei N, Baumann B, Grunert S, Sommer A, Pehamberger H, et al. NF-kappaB is essential for epithelial-mesenchymal transition and metastasis in a model of breast cancer progression. *J Clin Invest* 2004;114:569–81. [PubMed: 15314694]
68. Moustakas A, Heldin CH. Signaling networks guiding epithelial-mesenchymal transitions during embryogenesis and cancer progression. *Cancer Sci* 2007;98:1512–20. [PubMed: 17645776]
69. Zavadil J, Bottinger EP. TGF-beta and epithelial-to-mesenchymal transitions. *Oncogene* 2005;24:5764–74. [PubMed: 16123809]
70. Timmerman LA, Grego-Bessa J, Raya A, Bertran E, Perez-Pomares JM, Diez J, et al. Notch promotes epithelial-mesenchymal transition during cardiac development and oncogenic transformation. *Genes Dev* 2004;18:99–115. [PubMed: 14701881]
71. Zavadil J, Cermak L, Soto-Nieves N, Bottinger EP. Integration of TGF-beta/Smad and Jagged1/Notch signalling in epithelial-to-mesenchymal transition. *EMBO J* 2004;23:1155–65. [PubMed: 14976548]
72. Gregory PA, Bracken CP, Bert AG, Goodall GJ. MicroRNAs as regulators of epithelial-mesenchymal transition. *Cell Cycle* 2008;7:3112–8. [PubMed: 18927505]
73. Peter ME. Let-7 and miR-200 microRNAs: guardians against pluripotency and cancer progression. *Cell Cycle* 2009;8:843–52. [PubMed: 19221491]
74. Cannito S, Novo E, Compagnone A, Valfre di BL, Busletta C, Zamara E, et al. Redox mechanisms switch on hypoxia-dependent epithelial-mesenchymal transition in cancer cells. *Carcinogenesis* 2008;29:2267–78. [PubMed: 18791199]
75. Thayer SP, di Magliano MP, Heiser PW, Nielsen CM, Roberts DJ, Lauwers GY, et al. Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis. *Nature* 2003;425:851–6. [PubMed: 14520413]
76. Berman DM, Karhadkar SS, Maitra A, Montes De OR, Gerstenblith MR, Briggs K, et al. Widespread requirement for Hedgehog ligand stimulation in growth of digestive tract tumours. *Nature* 2003;425:846–51. [PubMed: 14520411]
77. Feldmann G, Fendrich V, McGovern K, Bedja D, Bisht S, Alvarez H, et al. An orally bioavailable small-molecule inhibitor of Hedgehog signaling inhibits tumor initiation and metastasis in pancreatic cancer. *Mol Cancer Ther* 2008;7:2725–35. [PubMed: 18790753]
78. Feldmann G, Dhara S, Fendrich V, Bedja D, Beaty R, Mullendore M, et al. Blockade of hedgehog signaling inhibits pancreatic cancer invasion and metastases: a new paradigm for combination therapy in solid cancers. *Cancer Res* 2007;67:2187–96. [PubMed: 17332349]
79. Miele L. Notch signaling. *Clin Cancer Res* 2006;12:1074–9. [PubMed: 16489059]
80. Sahlgren C, Gustafsson MV, Jin S, Poellinger L, Lendahl U. Notch signaling mediates hypoxia-induced tumor cell migration and invasion. *Proc Natl Acad Sci U S A* 2008;105:6392–7. [PubMed: 18427106]
81. Nosedà M, McLean G, Niessen K, Chang L, Pollet I, Montpetit R, et al. Notch activation results in phenotypic and functional changes consistent with endothelial-to-mesenchymal transformation. *Circ Res* 2004;94:910–7. [PubMed: 14988227]
82. Becker KF, Rosivatz E, Blechschmidt K, Kremmer E, Sarbia M, Hofler H. Analysis of the E-cadherin repressor Snail in primary human cancers. *Cells Tissues Organs* 2007;185:204–12. [PubMed: 17587826]
83. Niessen K, Fu Y, Chang L, Hoodless PA, McFadden D, Karsan A. Slug is a direct Notch target required for initiation of cardiac cushion cellularization. *J Cell Biol* 2008;182:315–25. [PubMed: 18663143]
84. Leong KG, Niessen K, Kulic I, Raouf A, Eaves C, Pollet I, et al. Jagged1-mediated Notch activation induces epithelial-to-mesenchymal transition through Slug-induced repression of E-cadherin. *J Exp Med* 2007;204:2935–48. [PubMed: 17984306]
85. Niimi H, Pardali K, Vanlandewijck M, Heldin CH, Moustakas A. Notch signaling is necessary for epithelial growth arrest by TGF-beta. *J Cell Biol* 2007;176:695–707. [PubMed: 17325209]
86. Katoh M, Katoh M. Notch ligand, JAG1, is evolutionarily conserved target of canonical WNT signaling pathway in progenitor cells. *Int J Mol Med* 2006;17:681–5. [PubMed: 16525728]
87. Kong D, Wang Z, Sarkar SH, Li Y, Banerjee S, Saliganan A, et al. Platelet-derived growth factor-D overexpression contributes to epithelial-mesenchymal transition of PC3 prostate cancer cells. *Stem Cells* 2008;26:1425–35. [PubMed: 18403754]

88. Liu J, Valencia-Sanchez MA, Hannon GJ, Parker R. MicroRNA-dependent localization of targeted mRNAs to mammalian P-bodies. *Nat Cell Biol* 2005;7:719–23. [PubMed: 15937477]
89. Saxena S, Jonsson ZO, Dutta A. Small RNAs with imperfect match to endogenous mRNA repress translation. Implications for off-target activity of small inhibitory RNA in mammalian cells. *J Biol Chem* 2003;278:44312–9. [PubMed: 12952966]
90. Ambros V. MicroRNA pathways in flies and worms: growth, death, fat, stress, and timing. *Cell* 2003;113:673–6. [PubMed: 12809598]
91. Lee YS, Dutta A. The tumor suppressor microRNA let-7 represses the HMGA2 oncogene. *Genes Dev* 2007;21:1025–30. [PubMed: 17437991]
92. Takamizawa J, Konishi H, Yanagisawa K, Tomida S, Osada H, Endoh H, et al. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res* 2004;64:3753–6. [PubMed: 15172979]
93. Kong D, Li Y, Wang Z, Banerjee S, Ahmad A, Kim HR, et al. miR-200 regulates PDGF-D-mediated epithelial-mesenchymal transition, adhesion, and invasion of prostate cancer cells. *Stem Cells* 2009;27:1712–21. [PubMed: 19544444]
94. Gupta PB, Onder TT, Jiang G, Tao K, Kuperwasser C, Weinberg RA, et al. Identification of selective inhibitors of cancer stem cells by high-throughput screening. *Cell* 2009;138:645–59. [PubMed: 19682730]
95. Li Y, Ahmed F, Ali S, Philip PA, Kucuk O, Sarkar FH. Inactivation of nuclear factor kappaB by soy isoflavone genistein contributes to increased apoptosis induced by chemotherapeutic agents in human cancer cells. *Cancer Res* 2005;65:6934–42. [PubMed: 16061678]
96. Wang Z, Zhang Y, Banerjee S, Li Y, Sarkar FH. Inhibition of nuclear factor kappaB activity by genistein is mediated via Notch-1 signaling pathway in pancreatic cancer cells. *Int J Cancer* 2006;118:1930–6. [PubMed: 16284950]
97. Li Y, Chinni SR, Sarkar FH. Selective growth regulatory and pro-apoptotic effects of DIM is mediated by AKT and NF-kappaB pathways in prostate cancer cells. *Front Biosci* 2005;10:236–43. [PubMed: 15574364]
98. Jimeno A, Feldmann G, Suarez-Gauthier A, Rasheed Z, Solomon A, Zou GM, et al. A direct pancreatic cancer xenograft model as a platform for cancer stem cell therapeutic development. *Mol Cancer Ther* 2009;8:310–4. [PubMed: 19174553]