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Altering dietary lysine:arginine ratio has little effect on cardiovascular risk factors and vascular reactivity in moderately hypercholesterolemic adults

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Abstract

Background—Information is scarce regarding the effect of dietary protein type, with specific focus on the lysine to arginine (Lys:Arg) ratio, on cardiovascular risk factors and vascular reactivity in humans.

Objective—Determine effect of dietary Lys:Arg ratio on cardiovascular risk factors and vascular reactivity in moderately hypercholesterolemic adults.

Design—Randomized cross-over design of two 35-day diet phases; thirty adults (21 females and 9 males, \geq 50 y, LDL cholesterol \geq 120 mg/dL). Diets had 20% energy (E) protein, 30%E fat, 50%E carbohydrate and were designed to have low (0.7) or high (1.4) Lys:Arg ratio. Measures included fasting and postprandial lipid, lipoprotein, apolipoprotein concentrations; fasting high sensitivity C-reactive protein (hsCRP), small dense LDL (sdLDL)-cholesterol, remnant lipoprotein cholesterol (RemLC), glycated albumin, adiponectin and immunoreactive insulin concentrations, endogenous cholesteryl ester transfer protein (CETP) and lecithin:cholesterol acyl transferase (LCAT) activities; cholesterol fractional synthesis rate (FSR); and flow mediated dilation (FMD) and peripheral artery tonometry (PAT).

Results—No differences were observed in fasting and/or postprandial total, LDL, HDL and sdLDL cholesterol, RemLC, Lp(a) or apo B concentrations, LCAT and CETP activities, FSR, glycated albumin, immunoreactive insulin, FMD or PAT. The low, relative to the high, Lys:Arg ratio diet resulted in lower postprandial VLDL cholesterol (-24%, *P*=0.001) and triglycerides (-23%,

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P=0.001), and small but significant differences in fasting (-3%, P=0.003) and postprandial (-3%, P=0.018) apo AI, and fasting adiponectin concentrations (+7%, P=0.035). Fasting and postprandial hsCRP concentrations were 23% lower after the low Lys:Arg ratio diet (P=0.020 for both).

Conclusions—Diets differing in Lys:Arg ratios had no or small effects on cardiovascular risk factors and vascular reactivity.

Keywords

lysine:arginine ratio; lipoproteins; small dense LDL (sdLDL)-cholesterol; remnant lipoprotein cholesterol (RemLC); cholesterol fractional synthesis rate (FSR); flow mediated dilation (FMD); peripheral artery tonometry (PAT)

Introduction

Early studies in a variety of experimental animal models of atherosclerosis suggest that proteins from vegetable sources are less hypercholesterolemic and atherogenic than proteins from animal sources [1–4]. In humans, observational and interventional studies suggest a cardioprotective effect of plant-based diets relative to those containing animal based products [5]. Comparative studies have documented lower blood pressure and concentrations of total cholesterol, LDL-cholesterol, triglycerides, high sensitivity C-reactive protein (hsCRP), and glucose among individuals consuming a vegetarian diet relative to omnivores [6–9]. The majority of intervention studies in humans conducted to assess the cholesterolemic effect of dietary protein were carried out using soy protein and appeared to confirm the findings in animals [10]. For the most part, clinical trials have compared soy protein to casein; only a few studies have evaluated the effects relative to other protein sources [11–14]. However, recent well controlled intervention trials comparing soy protein to an animal protein, independent of the fatty acid profile of the diet, have not supported the original observations [15]. The effects have either been modest [16–22] or null [23–30].

Throughout the mid and latter part of the 20th century, there was sporadic interest in the effect of dietary amino acid profile, independent of protein source or associated fatty acids, on cardiovascular risk factors. The primary focus was on the lysine-to-arginine (Lys:Arg) ratio. Rats or rabbits fed diets with a higher Lys:Arg ratio were reported to have higher total, LDL, and HDL cholesterol concentrations than animals fed a lower Lys:Arg ratio [3,4,31–38]. Moreover, addition of lysine to diets containing soy or cottonseed protein, such that the resulting Lys:Arg ratio was comparable to that of casein, increased atherogenicity, whereas the addition of arginine to case in-containing diets reduced the atherogenicity [4,31,37,38]. These data suggested that dietary lysine was a hypercholesterolemic amino acid whereas arginine had the opposite effect. In addition, altering the L-Arg-nitric oxide (NO) pathway has b een reported to alter flow mediated dilatation (FMD), a surrogate measure of endothelial function [39]. An acute oral dose of L-Arg was found to increase FMD in patients with coronary artery disease [40,41], hypercholesterolemia [42,43] and hypertension [44]. Nonetheless, these findings have not been observed consistently and few other cardiovascular risk factors were reported in these studies [45–47]. Primary sources of dietary arginine include nuts and legumes, whereas most grains have limited quantities of this amino acid. The main dietary sources of lysine are foods of animal origin, including dairy products, fish, eggs, poultry, and beef.

The aim of the study was to determine the effect of the dietary Lys:Arg ratio, rather than focusing on single amino acid supplementation or type of protein, on a range of CVD risk factors, including plasma lipids, lipoproteins, apolipoproteins and lipoprotein particle concentrations, inflammatory factors, and endothelial function in moderately hypercholesterolemic adults. Our hypothesis was that, within the context of similar dietary

fatty acid profiles, a low Lys:Arg ratio diet would result in a risk factor profile consistent with a lower cardiovascular disease risk, relative to a high Lys:Arg ratio diet.

Methods

Subjects

Thirty-nine study participants were recruited from the greater Boston area. Inclusion criteria included >50 years, LDL cholesterol >120 mg/dL, free of apparent chronic disease, and for women, postmenopausal status. Exclusion criteria included use of medications or dietary supplements known to affect lipid metabolism; abnormal kidney, liver, thyroid, or cardiac function; abnormal fasting glucose concentration; irritable bowel syndrome; chronic use of anti-inflammatory medications; smoking; alcohol consumption >7 drinks per week; unwillingness to maintain body weight throughout the study; hypertriglyceridemia (TG >400 mg/dL); and BMI > 35 kg/m². All study participants gave written consent. The study protocol was approved by the Human Investigation Review Committee of Tufts University and Tufts Medical Center and was registered in the ClinicalTrials.gov registry (Identifier #NCT00175084). Thirty participants, 21 postmenopausal women and nine men, completed the study. Nine participants initially recruited did not complete the study, six dropped out during phase 1 and three during phase 2, citing the following reasons: scheduling conflict (phase 1, n=1; skin cancer diagnosis (phase 1, n=1); change in medical status (phase 2, n=1); dislike of study food (n=1 during phase 1, n=1 during phase 2); intolerance to protocol diet (phase 1, n=1) and no longer interested (n=2 during phase 1, n=1 during phase 2). Five of these participants terminated participation within the first week of their start date. Data from participants who did not complete the study were excluded from the statistical analysis. Staggered enrollment allowed for the replacement of participants who terminated participation prematurely so that the target of 30 participants was achieved. There were no significant differences in baseline characteristics between female and male participants (Table 1).

Experimental Design

This was a randomized cross-over design study consisting of two 35-day diet phases separated by a break period that lasted 14 to 21 days. Laboratory personnel were blinded with regard to the identification of the study samples. Participants visited our Metabolic Research Unit three times per week for a review of changes in exclusion criteria or medical condition, consumption of one meal, and food pickup. Blood pressure and body weight were monitored at each visit. All foods and beverages were provided to the participants in quantities sufficient to maintain initial body weight. Energy requirements were calculated using the Harris-Benedict equation [48], and amounts of food were provided to study participants in increments of 250 kcal to the closest amount to their energy needs for weight maintenance. Participants were instructed to consume the meals entirely, not to substitute any food or beverage item, and not to add any calorie- or monosodium glutamate-containing condiments. Caloric intake was adjusted, when necessary, to maintain a stable body weight throughout the study period (\pm 1.0 kg from initial weight). Body weight and blood pressure values from the three test days (see below) were used for this report. BMI was calculated as kg/m². Study participants were encouraged to maintain their usual level of physical activity and report any changes in approved medication use.

On three separate days after day 28, and within the last week of each diet phase, blood samples were collected after a 12-h fast into evacuated tubes for measurement of serum lipids or EDTA-containing tubes for the other biochemical or enzymatic assays. Twenty-four hours prior to one of these blood collections participants were asked to consume 0.48 g deuterated water $(D_2O)/kg$ estimated body water (60 percent body weight) to measure fractional cholesterol synthesis rate (FSR). Consistent with humans spending a greater amount of time in a postprandial than fasting state, biochemical measures were evaluated both fasting and four

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hours after consumption of the mid-day meal of a composition consistent with the appropriate diet phase. Fasted and postprandial lipids, lipoproteins, apoproteins and hsCRP concentrations were measured. During one of the three blood collection days, waist and hip circumference was measured. Prior data from our laboratory has indicated that under the specified study conditions plasma lipid concentrations were stable after day 28 of a feeding protocol [49]. Serum and plasma were separated from red blood cells by centrifugation at 1100 × g at 4°C, and each component was aliquoted and stored at -80° C for subsequent analysis.

Diets

Diets were designed to contain 20% of energy from protein and to have either a low Lys:Arg ratio (Lys:Arg = 0.7) or a high Lys:Arg ratio (Lys:Arg = 1.4) using commonly available foods. No amino acid supplements were used. Most of the protein sources for the low Lys:Arg ratio diet were from vegetable origin, including nuts and legumes. No attempt was made to avoid animal protein. Most of the protein sources for the high Lys:Arg ratio diet was from animal sources, including dairy products, fish, eggs, poultry, and beef. Diets were matched to the extent possible for polyunsaturated to saturated fat ratio, *trans* fatty acids, fiber, cholesterol, and micronutrients. The macronutrient and amino acid composition of the diets was confirmed by chemical analysis (Covance Laboratories, Madison, WI, USA) (Table 2). Micronutrients were calculated using the Nutrition Data System for Research software version 4.05 (2004), developed by the Nutrition Coordinating Center, University of Minnesota (Minneapolis, MN) [50]. Both diets provided adequate amounts of micronutrients (Table 6; Online appendix).

Biochemical measurements

Serum total cholesterol, LDL cholesterol, HDL cholesterol, and triglyceride concentrations were measured using an Olympus AU400 with enzymatic reagents and calibrators (Olympus America Inc., Melville, NY). VLDL cholesterol concentrations were calculated as the difference between total cholesterol and LDL cholesterol plus HDL cholesterol. Plasma apoprotein (apo) A-I and apo B (KAMIYA Biomedical Company, Seattle, WA) and Lp(a) concentrations (Wako Chemicals USA, Inc., Richmond, VA) were measured using an Olympus AU400 immunoturbidimetrically. Plasma hsCRP was measured using a Roche Cobas Fara centrifugal clinical chemistry analyzer (Roche Diagnostics, Indianapolis, IN) immunoturbidimetrically (DiaSorin, Inc., Stillwater, MN). Proficiency testing for these procedures was done through the College of American Pathologists (CAP) Interlaboratory Comparison and Survey Proficiency Program (Northfield, IL). Linearity studies for all procedures were done through the Verichem Laboratories Linear Testing Program (Providence, RI).

Measurement of triglyceride-rich remnant lipoprotein cholesterol (RemLC) was performed as described by Miyauchi et al. [51] using a homogenous assay that allows for measurement of cholesterol in chylomicron remnants, VLDL remnants, and intermediate density lipoproteins (IDL). Plasma small, dense LDL (sdLDL) cholesterol was measured using a simple heparinmagnesium precipitation method as previously described [52]. Plasma glycated albumin was measured enzymatically as previously reported [53]. A latex particle-enhanced turbidimetric immunoassay was used for the measurement of plasma adiponectin [54]. Plasma immunoreactive insulin was measured with a latex immunoassay [55]. Endogenous cholesteryl ester transfer protein (CETP) and lecithin:cholesterol acyl transferase (LCAT) activities were measured in plasma as previously reported [56].

Cholesterol Fractional Synthesis

Free cholesterol from approximately 0.5g of red blood cells was extracted and analyzed by gas chromatography-thermal conversion-isotope ratio mass spectrometry (GC-TC-IRMS; Delta V Plus, Thermo Electron Corporation) to determine the ${}^{2}H/{}^{1}H$ ratio versus VSMOW [57]. The

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cholesterol precursor pool was taken as the mean plasma water deuterium enrichment which was determined by TCEA-IRMS from plasma prepared by membrane filtration and centrifugation removing proteins > 5 kilodaltons [58]. Cholesterol FSR rates were calculated using the following equation:

 $FSR (\%/day) = \frac{[\delta \text{ cholesterol}/(\delta \text{ plasma} \times 0.478)]}{\text{deuterium incorporation period}} \times 24 \text{ hours} \times 100\%$

Where δ is deuterium enrichment of cholesterol or plasma water above baseline and time refers to the 2h deuterium incorporation period. The factor 0.478 is the fraction of hydrogen atoms per cholesterol molecule possibly labeled by a deuterium [58].

Endothelial function measurement

At the end of each dietary period, participants underwent a fasting brachial artery reactivity test (BART) to assess endothelium-dependent flow mediated dilatation (FMD). The diameter of the brachial artery was measured as described by Celermajer et al. [59] by high-resolution external vascular ultrasound during two conditions: baseline (after a 10-min supine rest) and in response to the inducement of hyperemia by inflation to 250 mm Hg and subsequent deflation of a sphygmomanometer cuff around the forearm to occlude arterial flow for 5 minutes. Peripheral vascular endothelial function was assessed simultaneously to the BART by peripheral arterial tonometry (PAT) as previously described [60].

Statistical analyses

Prior to the analysis, descriptive statistics and graphs (PROC UNIVARIATE and PROC MEANS) (SAS v 9.1 for Windows, Cary, N.C.) were used to summarize the overall effects of diets and distributions of the outcome measures. Data were tested for normality; when basic testing assumptions were violated, log10- transformations of the data were conducted to achieve normality prior to analysis and are so indicated in the tables. Data were analyzed using a paired t-test. When no transformation was appropriate, a nonparametric signed-rank test was used to compare means. Untransformed data are presented in text and tables as mean \pm SD. Differences were considered significant at the 0.05 alpha level.

Results

The mean energy intake (mean \pm SD) of the participants was 2617 \pm 468 kcal when consuming the low Lys:Arg ratio diet and 2662 \pm 490 kcal when consuming the high Lys:Arg ratio diet (n.s.; transformed to rank data before statistical analysis); energy intake for males was ~700 kcal greater than for women during both experimental diet phases (P_{Sex} =0.001). There were no significant differences in body weight, BMI, waist:hip ratio, or systolic and diastolic blood pressure at the end of the two diet phases (Table 3).

Consumption of the two experimental diets resulted in no significant differences in fasting total, LDL, VLDL, HDL and sdLDL cholesterol, RemLC or triglyceride concentrations and the total cholesterol:HDL cholesterol ratio (Table 4). At the end of the two diet phases there were no significant differences in Lp(a) or apo B concentrations. Compared to the high Lys:Arg ratio diet, the low Lys:Arg ratio diet resulted in a modestly lower apo AI (3%, P=0.003) and higher adiponectin concentration (7%; P=0.035). There were no significant differences in fasting concentrations of glycated albumin or immunoreactive insulin at the end of the two experimental diet phases. Relative to the high Lys:Arg ratio diet, the low Lys:Arg ratio diet resulted in 23% lower fasting hsCRP concentrations (P=0.020). The mean values for both were within the normal range. Endogenous LCAT and CETP activities were not significantly

different at the end of the two diet phases. Similarly, cholesterol FSR was not significantly altered by the Lys:Arg ratio of the diet (Figure 1)

In the postprandial state, consumption of the two experimental diets resulted in no significant differences in total, LDL, and HDL cholesterol concentrations (Table 5) although the total cholesterol:HDL cholesterol ratio was marginally lower after participants consumed the low Lys:Arg ratio diet compared to the high Lys:Arg ratio diet (-1.5%; P=0.05). Interestingly, compared to the high Lys:Arg ratio diet phase, postprandial serum VLDL cholesterol and triglyceride concentrations were 24% (P=0.001) and 23% (P=0.001) lower, respectively, following the low Lys:Arg ratio diet phase. This difference in triglyceride concentrations between the two diets was not reflected in HDL cholesterol concentrations. There were no significant differences in Lp(a) or apo B concentrations at the end of the two diet phases. As observed in the fasting state, compared to the high Lys:Arg ratio diet, the low Lys:Arg ratio diet resulted in modestly lower postprandial apo AI (3%, P=0.018) and lower hsCRP concentrations (29%, P=0.020).

Vascular reactivity was not significantly different at the end of the two experimental diet phases as measured by flow mediated dilation or peripheral artery tonometry (Figure 2).

Discussion

Previous work had suggested that a shift in the diet to lower Lys:Arg ratios had a favorable effect on cardiovascular risk factors in animal models and humans. By design, the intent of this study was to evaluate diets with different amino acid profiles, rather than focusing on a single amino acid or type of protein. This end was achieved by using commercially available food products and without amino acid supplements. Taking this approach the Lys:Arg ratio was shifted from 0.7 to 1.4. Nuts and legumes displaced foods containing animal protein from the diet to decrease the Lys:Arg ratio. By substituting nuts and legumes for animal protein the Arg content of the low Lys:Arg diet was 1.5-times higher than in the high Lys:Arg ratio diet. For moderately hypercholesterolemic participants, the difference in Lys:Arg ratio resulted in a null or small effect on cardiovascular disease risk factors. When differences were observed they were, for most part, in the non-fasting state.

From observational data, daily Arg intake has been estimated to range between 4.3 and 13.8 g/d, dictated by different regional dietary habits [61,62]. The Arg intake of our participants ranged from 8-12 g/day and 5-8 g/day, when consuming the low Lys:Arg and high Lys:Arg ratio diets, respectively, depending on energy requirements needed to maintain a stable body weight. These values are well within the range from observational studies.

The difference in Lys:Arg ratio between the two diets was large in terms of the ratio and what could be reasonably achieved using habitually consumed foods alone (0.7 vs. 1.4), yet less than that used in animal studies. Differences in Lys:Arg ratios among studies involving animal models range from 0.3 to 2.2. In contrast to the current work, these animal studies reported a significant effect of Lys:Arg ratio on lipoprotein concentrations. For example, Sprague-Dawley rats fed a high Lys:Arg ratio (1.58 ratio) diet had the highest, whereas those fed a low Lys:Arg ratio (0.36 ratio) had the lowest total and HDL-cholesterol concentrations [31]. Similar results were observed in male albino rats (low Lys:Arg ratio, 0.67; high Lys:Arg ratio, 2.0) [32] and rabbits (high Lys:Arg ratio, 2.2 ratio; low Lys:Arg ratios, 0.9 and 0.3) [33]. In all these cases, the fatty acid profile of the diets did not differ as a result of manipulating the amino acid profile of the diets. Notably, these studies were conducted using animal species that lack analogies with human cholesterol and lipoprotein metabolism [63,64].

Several studies using rats and rabbits also focused on the effects of other amino acids and suggested that the sulfur-containing amino acids, methionine (Met) and cysteine (Cys), were

also hypercholesterolemic, whereas glycine (Gly) was hypocholesterolemic [35,65–67]. Similar to the low Lys:Arg ratio, proteins having a low Met:Gly ratio were purported to elicit a hypocholesterolemic effect [67]. Of note, in the present study the low Lys:Arg ratio diet had a Met:Gly ratio of 0.32, whereas the high Lys:Arg ratio diet had a Met:Gly ratio of 0.67. These values are comparable to those reported for soy protein and casein (Met:Gly ratio 0.32 and 0.64, respectively).

Postprandial triglyceride and apo AI concentrations were significantly lower at the end of the low Lys:Arg ratio diet phase. However, this difference in triglyceride concentrations was not reflected in higher HDL cholesterol concentrations, as is frequently observed [68]. The small absolute difference in apo AI concentration is of questionable clinical significance. Furthermore, no significant differences were observed in LCAT or CETP activities at the end of diet phases. There is no obvious explanation for this observation. We cannot rule out the possibility that different sources of carbohydrates and fiber in the two diets may have contributed to the observed postprandial triglyceride concentrations. Interestingly, the effect on triglyceride concentrations was not observed in the fasting state, which suggests an effect on chylomicron-triglyceride, rather than VLDL-triglyceride clearance. It is possible that consumption of the low Lys:Arg ratio diet may have induced greater postprandial insulin secretion relative to the high Lys:Arg ratio. Moreover, the concentration of adiponectin, known to affect triglyceride concentrations and increase insulin sensitivity [69], was significantly higher, albeit modestly, at the end of the low Lys:Arg ratio diet phase compared to the high Lys: Arg ratio diet phase. It is possible that the higher adiponectin concentrations resulting from the low Lys:Arg ratio diet may have contributed to the difference in insulin sensitivity and enhanced triglyceride clearance despite the lack of differences in fasting insulin concentrations.

The consumption of diets with different Lys:Arg ratios or absolute amounts of Arg had no significant effect on endothelial function assessed by measuring FMD or PAT. Reports suggesting that endothelial function was improved in humans by supplementary L-Arg were at intake levels for the amino acids that far exceeded the amount that could be achieved by manipulating dietary sources of protein as attained for this study. These observations have been made in control participants (21 g L-Arg/d for 4 weeks) [70], coronary heart disease patients (21 g L-Arg/d for 3 days) [41], and hypertensive patients (single dose of 6 g L-Arg) [44]. Notably, studies reporting an effect of supplementary L-Arg on FMD were of much shorter duration than the present intervention, and more likely represent acute effects of L-Arg supplementation. Conversely, more modest doses of supplemental L-Arg failed to have a significant effect on endothelial function in peripheral arterial disease patients (3 g L-Arg/d for 6 months) [71] and hypercholesterolemic patients (2 bar per day with 3.3 g L-Arg/bar for 1 week) [72] with the exception of one study involving hypercholesterolemic individuals (2 bar per day with 3.3 g L-Arg/bar for 1 week) in which a significant improvement was reported [73]. In contrast to the current work, studies in which L-Arg was observed to have an effect on FMD, supplementation resulted in an Arg intake greater than what is consumed through dietary protein alone and the potential effect of dietary lysine was not taken into consideration. Moreover, it has been suggested that the amount of dietary arginine that enters the NO synthesis pathway is relatively low and therefore is unlikely to affect endothelial function [74].

In the current study, both fasting and postprandial hsCRP concentrations were significantly lower after the low Lys:Arg ratio diet phase. Consistent with this finding, observational data from the Third National Health Nutrition and Examination Survey (NHANES III) suggested an association between higher consumption of Arg-rich foods and lower serum hsCRP concentrations [75]. Despite the statistically significant difference observed in the current study, both mean hsCRP concentrations were within the normal range at the end of each diet phase. At this time there is a dearth of data on the effect of dietary amino acids or protein types

on inflammatory markers. Additional data are needed prior to adequately interpret these findings.

We cannot rule out the possibility that factors associated with different dietary protein sources or phytochemicals were responsible to the differences in cardiovascular risk factors previously reported. Likewise, we cannot eliminate the possibility that undetected differences in the fatty acid profile of the diets in prior work had a significant effect on the outcome measures. In the current study we attempted to minimize differences between the two diets beyond that of the Lys:Arg ratios. However, due to constraints in the sources of amino acids and the intent to manipulate the Lys:Arg ratio with commonly consumed foods rather than amino acid supplements, we may have impacted study outcomes in unrecognized ways. Despite efforts towards maintaining comparable macronutrient and fiber contents between both diets, the lower Lys: Arg ratio diet had more dietary fat (31.8% energy vs. 25.1% energy) and less dietary fiber (17.8 g/1000 kcal vs. 21.5 g/1000 kcal) than the higher Lys: Arg ratio diet. We cannot rule out the possibility that changes due to these differences may have masked the absolute effect of differences in the Lys: Arg ratio. Lastly, whereas Lys is an essential amino acid, one that humans cannot synthesize adequate amounts to meet requirements, Arg is not. We were unable to determine the potential contribution of endogenous Arg synthesis to the total pool and how this may have affected the Lys:Arg ratio.

In conclusion, the amino acid profile of the diet, characterized by its Lys:Arg ratio, had no effect on fasting cardiovascular risk factors, vascular reactivity, or fractional cholesterol synthesis rate in mildly hypercholesterolemic adults. Overall, no convincing evidence was found to support the positive or negative modulation of cardiovascular disease risk factors based on characterizing dietary proteins by the Lys:Arg ratio.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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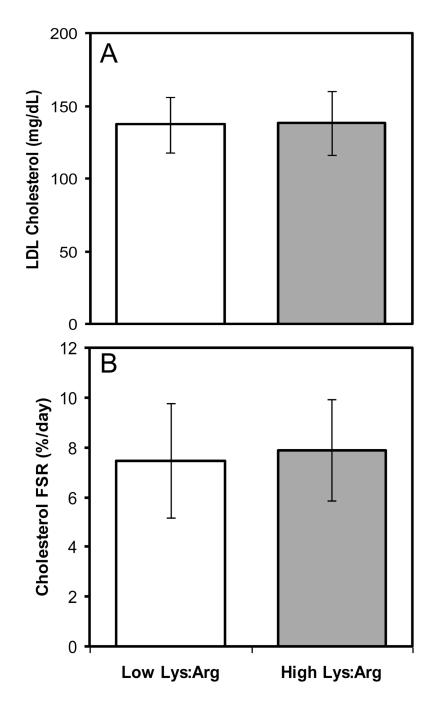


Figure 1.

Fasting LDL cholesterol concentrations (mg/dL, panel A) and cholesterol fractional synthesis rate (FSR; %/day; panel B) at the end of the two diet phases. Results at the end of the low and high Lys:Arg ratio experimental diet phases are shown in clear and dark bars, respectively. A paired t-test was carried out for each variable.

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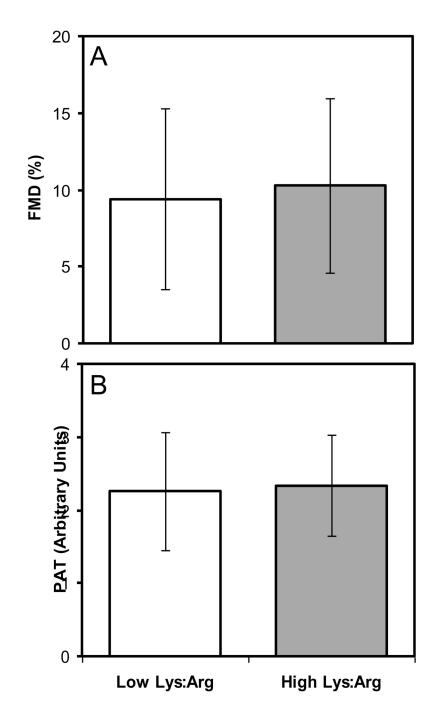


Figure 2.

Vascular reactivity at the end of the two experimental phases was assessed by measuring flow mediated dilation (FMD; %; panel A) and peripheral artery tonometry (PAT; Arbitrary Units; panel B). Results at the end of the low and high Lys:Arg ratio experimental diet phases are shown in clear and dark bars, respectively. A paired t-test was carried out for each variable. Flow mediated dilation data were transformed to rank data before statistical analysis.

Table 1

Baseline characteristics of participants¹

Variable	All participants N=30	Females N=21	Males N=9	P value ²
Age (y)	61.8 ± 6.5 (51–76)	61.8 ± 6.0 (51–76)	$\begin{array}{c} 62.0 \pm 7.9 \\ (5175) \end{array}$	0.935
Body mass index (kg/m ²)	26.7 ± 3.2 (21.3–32.9)	$\begin{array}{c} 26.9 \pm 3.29 \\ (22.0 32.9) \end{array}$	$\begin{array}{c} 26.2 \pm 3.2 \\ (21.3 31.7) \end{array}$	0.581
Weight (kg)	$74.5 \pm 11.5 \\ (56111)$	$\begin{array}{c} 72.4\pm9.3\\(5688)\end{array}$	$79.2 \pm 15.0 \\ (61 111)$	0.138
Systolic BP (mm Hg)	$\begin{array}{c} 126\pm16\\ (95160)\end{array}$	$\begin{array}{c} 125\pm15\\ (95150)\end{array}$	127 ± 18 (103–160)	0.726
Diastolic BP (mm Hg)	74 ± 9 (55–89)	$\begin{array}{c} 72 \pm 10 \\ (55 89) \end{array}$	77 ± 9 (63–88)	0.271
Plasma lipids, lipoproteins an	nd apoproteins			
TC (mg/dL)	223 ± 25 (182–300)	228 ± 24 (196–300)	211 ± 23 (182–258)	0.095
LDL-C (mg/dL)	$\begin{array}{c} 145 \pm 17 \\ (118 203) \end{array}$	148 ± 19 (126–203)	138 ± 12 (118–149)	0.132
VLDL-C (mg/dL)	$\begin{array}{c} 23\pm10\\(845)\end{array}$	$\begin{array}{c} 23\pm11\\(845)\end{array}$	22 ± 10 (14–42)	0.828
HDL-C (mg/dL)	55 ± 13 (35–96)	56 ± 14 (35–96)	51 ± 12 (36–68)	0.333
TG (mg/dL)	117 ± 52 (41–228)	119 ± 54 (41–228)	114 ± 48 (70–210)	0.816
TC/HDL-C	$\begin{array}{c} 4.26 \pm 0.97 \\ (2.48 6.64) \end{array}$	$\begin{array}{c} 4.25 \pm 1.05 \\ (2.48 6.64) \end{array}$	$\begin{array}{c} 4.27 \pm 0.81 \\ (3.31 6.12) \end{array}$	0.956

^IValues are expressed as mean±SD (range). BP: blood pressure; HDL-C: HDL cholesterol; LDL-C: LDL cholesterol; TC: total cholesterol; TG: triglycerides; VLDL-C: VLDL cholesterol.

 $^2\mathrm{An}$ independent samples t-test was carried out for the male: female comparison.

Composition of experimental diets¹

Constituent	Low Lys:Arg	High Lys:Arg		
	Percent of dail	Percent of daily energy intake		
Protein	19.8	21.8		
Carbohydrate	47.9	52.7		
Fat	31.8	25.1		
Saturated fatty acids	6.6	5.5		
Monounsaturated fatty acids	11.3	8.0		
Polyunsaturated fatty acids	13.0	10.6		
Trans fatty acids	0.4	0.3		
ω -6 fatty acids ²	12.1	10.5		
ω -3 fatty acids ³	1.4	0.6		
	Amount per 1000 Kcal			
Cholesterol (mg)	69	75		
Fiber (g)	17.8	21.5		
	Amino acids (mg/100g)			
Asp	540	470		
Thr	180	210		
Ser	270	240		
Glu	1120	970		
Pro	300	330		
Gly	220	180		
Ala	220	240		
Cys	60	60		
Val	230	260		
Met	70	120		
Ile	220	230		
Leu	390	420		
Tyr	170	180		
Phe	260	230		
His	130	150		
Lys	280	380		
Arg	400	270		
Trp	60	60		
Lys:Arg ratio	0.70	1.41		

 I Macronutrients, fiber, cholesterol, fatty acids and amino acids were determined by chemical analysis of food.

 $^2\omega$ -6 fatty acids: linoleic acid + arachidonic acid

 $^3\omega\text{-}3$ fatty acids: linolenic acid + eicosapentaenoic acid + docosahexaenoic acid

Anthropometric characteristics and blood pressure at end of experimental diet phases

	Low Lys:Arg	High Lys:Arg	P value
Body weight (kg)	73.4 ± 11.5	73.6 ± 11.6	0.358
Body mass index (kg/m ²)	26.3 ± 3.3	26.3 ± 3.3	0.375
Waist:Hip ratio	0.861 ± 0.078	0.865 ± 0.073	0.675
Systolic blood pressure (mm Hg)	113 ± 12	111 ± 10	0.101
Diastolic blood pressure (mm Hg)	70 ± 7	69 ± 7	0.291

A paired t-test was carried out for each variable.

*Transformed to log10 values before statistical analysis.

Fasting lipoprotein-related parameters and hsCRP concentration at the end of experimental diet phases.

	Low Lys:Arg	High Lys:Arg	P value
Total cholesterol (mg/dL)	207 ± 24	210 ± 27	0.236
LDL cholesterol (mg/dL)	137 ± 19	138 ± 22	0.593
VLDL cholesterol $(mg/dL)^{\#}$	15 ± 9	16 ± 10	0.189
HDL cholesterol (mg/dL)	55 ± 13	57 ± 13	0.133
Triglyceride (mg/dL)*	104 ± 35	109 ± 42	0.317
Total cholesterol:HDL cholesterol $^{\#}$	3.89 ± 0.85	3.86 ± 0.86	0.741
Lipoprotein (a) (mg/dL) #;	35.6 ± 25.3	35.6 ± 26.3	0.63
Apoprotein AI (mg/dL)	148.7 ± 22.8	152.7 ± 22.3	0.003
Apoprotein B (mg/dL)	101.5 ± 14.5	103.0 ± 15.8	0.267
sdLDL cholesterol (mg/dL)	36.1 ± 10.2	34.1 ± 9.3	0.152
RemLC (mg/dL)	5.99 ± 3.36	5.77 ± 2.60	0.601
hsCRP (mg/dL)	1.87 ± 2.45	2.44 ± 3.14	0.020
Glycated Albumin $(\%)^{\#}$	13.6 ± 0.95	13.6 ± 0.9	0.863
Adiponectin (µg/mL)*	12.4 ± 6.58	11.6 ± 5.4	0.035
Immunoreactive Insulin $(\mu U/mL)^*$	9.90 ± 3.96	10.8 ± 5.9	0.076
LCAT (μ mol chol • L ⁻¹ • h ⁻¹) [*]	48.0 ± 15.5	49.2 ± 13.9	0.647
CETP (µmol chol • L^{-1} • h^{-1})	33.2 ± 19.4	32.6 ± 17.6	0.931
Cholesterol FSR (%/day)	7.48 ± 2.28	7.89 ± 2.04	0.406

A paired t-test was carried out for each variable.

*Transformed to log10 values before statistical analysis.

 $^{\#}$ Transformed to rank data before statistical analysis.

Postprandial lipids, lipoproteins, apoproteins and hsCRP at the end of test phases

	Low Lys:Arg	High Lys:Arg	P value
Total cholesterol (mg/dL)	202 ± 24	207 ± 28	0.073
LDL cholesterol (mg/dL)#	128 ± 18	126 ± 20	0.904
VLDL cholesterol $(mg/dL)^*$	22 ± 11	29 ± 16	0.001
HDL cholesterol (mg/dL)	52 ± 12	53 ± 12	0.594
Triglyceride (mg/dL)*	137 ± 52	178 ± 80	0.001
Total cholesterol:HDL cholesterol [#]	4.03 ± 0.89	4.09 ± 0.93	0.050
Lipoprotein (a) (mg/dL)#	33.8 ± 25.0	34.3 ± 26.3	0.057
Apoprotein AI (mg/dL)	147.0 ± 21.9	151.4 ± 21.9	0.018
Apoprotein B (mg/dL)#	98.5 ± 13.6	100.3 ± 15.7	0.136
hsCRP (mg/dL)	1.81 ± 2.30	2.34 ± 2.89	0.020

A paired t-test was carried out for each variable.

*Transformed to log10 values before statistical analysis.

[#]Transformed to rank data before statistical analysis.