

Evolution of ultraviolet vision in shorebirds (Charadriiformes)

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Diurnal birds belong to one of two classes of colour vision. These are distinguished by the maximum absorbance wavelengths of the SWS1 visual pigment sensitive to violet (VS) and ultraviolet (UVS). Shifts between the classes have been rare events during avian evolution. Gulls (Laridae) are the only shorebirds (Charadriiformes) previously reported to have the UVS type of opsin, but too few species have been sampled to infer that gulls are unique among shorebirds or that Laridae is monomorphic for this trait. We have sequenced the SWS1 opsin gene in a broader sample of species. We confirm that cysteine in the key amino acid position 90, characteristic of the UVS class, has been conserved throughout gull evolution but also that the terns *Anous minutus*, *A. tenuirostris* and *Gygis alba*, and the skimmer *Rynchops niger* carry this trait. Terns, excluding *Anous* and *Gygis*, share the VS conferring serine in position 90 with other shorebirds but it is translated from a codon more similar to that found in UVS shorebirds. The most parsimonious interpretation of these findings, based on a molecular gene tree, is a single VS to UVS shift and a subsequent reversal in one lineage.

Keywords: gulls; UV visual pigment; opsin; phylogeny

1. INTRODUCTION

There is a categorical, physiological difference in colour vision between groups of diurnal birds (Cuthill *et al.* 2000). These have short-wavelength sensitive type 1 (SWS1) cone-opsin-based visual pigments of either an ultraviolet sensitive (UVS) type with maximum absorbance wavelengths (λ_{\max}) between 355 and 380 nm or a violet sensitive (VS) type with λ_{\max} between 402 and 426 nm (reviewed by Ödeen *et al.* 2009). The VS type appears to be ancestral (Yokoyama 2002) and phylogenetically most widespread (Ödeen & Håstad 2003) in birds. Independent shifts to UVS may have occurred only three or four times (Håstad *et al.* 2009): in shorebirds (Charadriiformes), in a common ancestor of passerines and psittaciforms, in *Rhea americana* and possibly in trogons (Carvalho *et al.* 2007). Gulls are

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the only shorebirds reported to be UVS—demonstrated through the presence of cysteine in amino acid (aa) position 90 (bovine rod opsin numbering) in their SWS1 opsin gene and ocular media with spectral transmission similar to that of other UVS birds (Ödeen & Håstad 2003; Håstad *et al.* 2005, 2009).

There is an increased cost of UV transmittance owing to photo-oxidation of the retina (e.g. Boulton *et al.* 2001), making the presence of UV tuning in gulls rather puzzling. Unlike other UVS birds, no selective advantage of UV vision has been demonstrated in gulls, whether for detecting sexual signals (e.g. Bennett *et al.* 1997) or foraging (e.g. Church *et al.* 1998). It has been suggested to allow eavesdropping on UV communication in schools of fish swimming close to the surface (Ödeen & Håstad 2003) but evidence seems weak. All auks, waders and terns investigated so far have SWS1 opsins of the VS type (Håstad *et al.* 2005), and some, especially terns, are ecologically similar to gulls. However, too few species have been sampled to safely infer that no other charadriiforms have UV-tuned visual pigments, or that this character state does not vary within Laridae.

The SWS1 λ_{\max} pigments in avian retinae closely follows aa sequence variation in the SWS1 opsin gene. *In vitro* studies have demonstrated that λ_{\max} can be predicted on the basis of the opsin's aa sequence (Wilkie *et al.* 2000; Yokoyama *et al.* 2000; Carvalho *et al.* 2007). The cone sensitivity can then be estimated from the opsin sequence, owing to covariance between the visual pigment's λ_{\max} and the spectral absorptance properties of the cone oil droplets (Hart & Vorobyev 2005). As spectral tuning of the SWS1 single cone is under direct genetic control, it becomes possible to identify the type of short-wavelength sensitivity in a bird from a sample of genomic DNA. This method was conceived by Ödeen & Håstad (2003) and validated by Ödeen *et al.* (2009). We have used it here to search for gross differences in colour vision in a significantly widened sample of gulls and other shorebirds. The aim has been to assess how stable the SWS1 pigment sensitivity has been during the evolution of shorebirds in general and gulls in particular. As the basis for this analysis, we have inferred a phylogeny, mainly using GenBank sequence data.

2. MATERIAL AND METHODS

We studied a broad selection of species in the order Charadriiformes (table 1) with special emphasis on including a representative sample of various Laridae taxa. We sequenced a fragment of the SWS1 opsin gene containing the aa residues of positions 84–94 in up to six species each of seven shorebird families (table 1). We estimated λ_{\max} from three key positions, as outlined by Ödeen *et al.* (2009).

For the phylogenetic analyses, we used GenBank sequences of three mitochondrial and three nuclear loci, plus own β fibrinogen (FGB7) and myoglobin sequences of *Anous tenuirostris* and *Gygis alba*. Phylogenetic trees were constructed by maximum likelihood for one locus at a time and by Bayesian inference in the following combinations: (i) mitochondrial CO1, ND2 and cytochrome *b*, (ii) nuclear RAG-1, FGB7 and myoglobin intron 2 (myoglobin), and (iii) all loci together. For details, see the electronic supplementary material.

3. RESULTS

(a) Opsin gene analysis

All gull species investigated in this and previous studies (table 1) carry the UVS-pigment-conferring Ser to Cys

Table 1. SWS1 opsin gene fragments from shorebirds. The spectral tuning amino acid (aa) sites 86, 90 and 93 are marked in bold. Type of SWS1 opsin is either VS (violet sensitive) or UVS (UV-sensitive), as predicted from the gene fragment aa sequences. NRM, Swedish Museum of Natural History; LSUMZ, LSU Museum of Natural Science; UWBM, Burke Museum, University of Washington; ZMUC, Zoological Museum, University of Copenhagen.

family	species	common name	aa seq. 84–94	aa90 codon	type effect	unknown	origin/voucher no.	tissue no.	GenBank acc. no.	reference
Scolopacidae	<i>Tringa glareola</i>	Wood Sandpiper	FIA CIF S VFTV	agc	VS	Department of Animal Ecology, U. U.	031218-2	GU129664	this study	
	<i>Actitis hypoleucos</i>	Common Sandpiper	FIA CIF S VFTV	agc	VS			AY960716	Håstad <i>et al.</i> (2005)	
	<i>Phalaropus fulicarius</i>	Red Phalarope	FIA CIF S VFTV	agc	VS			AY960718	Håstad <i>et al.</i> (2005)	
	<i>Gallinago gallinago</i>	Common Snipe	FIA CIF S VFTV	agc	VS			AY960717	Håstad <i>et al.</i> (2005)	
	<i>Ruff</i>	Ruff	FIA CIF S VFTV	agc	VS	F. Widemo	11 individuals	GU129665	this study	
Thinocoridae	<i>Thinocorus orbignyianus</i>	Grey-breasted Seedsnipe	FIA CIF S VFTV	agc	VS	UWBM 54407	DAB 808	GU129666	this study	
Laridae	<i>Larus atlanticus</i>	Olrog's Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93	P. Yorio, J.-M. Pons	GU129667	this study	
	<i>Larus marinus</i>	Great Black-backed Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		AY227162	Ödeen & Håstad (2003)	
	<i>Larus argentatus</i>	European Herring Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		AY227160	Ödeen & Håstad (2003)	
	<i>Larus michahellis</i>	Yellow-legged Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		FJ790325	Håstad <i>et al.</i> (2009)	
	<i>Larus fuscus</i>	Lesser Black-backed Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		AY227161	Ödeen & Håstad (2003)	
	<i>Ichthyaetus hemprichii</i>	Sooty Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		AY960710	Håstad <i>et al.</i> (2005)	
	<i>Leucophaeus pipixcan</i>	Franklin's Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		GU129668	this study	
	<i>Chroicocephalus hartlaubii</i>	Hartlaub's Gull	FIC CVLC I STV	tgc	UVS	Ile86, 93		AY960709	Håstad <i>et al.</i> (2005)	
	<i>Chroicocephalus ridibundus</i>	Black-headed Gull	FIC CVLC I STV	tgc	UVS	Ile86, 93		AY960711	Håstad <i>et al.</i> (2005)	
	<i>Rhodostethia rosea</i>	Ross's Gull	FV ICVFC I SLV	tgc	UVS	Ile86,	ZMUC 139640	GU129669	this study	
	<i>Rissa tridactyla</i>	Black-legged Kittiwake	FIC CVFC I STV	tgc	UVS	Ile86, 93		AY960712	Håstad <i>et al.</i> (2005)	
	<i>Pugnophila eburnea</i>	Ivory Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		GU129670	this study	
	<i>Xema sabini</i>	Sabine's Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		GU129671	this study	
	<i>Creagrus furcatus</i>	Shallow-tailed Gull	FIC CVFC I STV	tgc	UVS	Ile86, 93		GU129672	this study	
Rynchopidae	<i>Rynchops niger</i>	Black Skimmer	FV ACVFC I IFTV	tgc	UVS			GU129673	this study	
Sternidae	<i>Anous minutus</i>	Black Noddy	FIA CIF C IFTV	tgc	UVS			GU129674	this study	
	<i>Anous tenuirostris</i>	Lesser Noddy	FIA CIF C IFTV	tgc	UVS			GU129675	this study	
	<i>Onychoprion anaethetus</i>	Bridled Tern	FV TCVFS I IFTV	tcc	VS	Thr86		GU129676	this study	

(Continued.)

Table 1. (Continued.)

family	species	common name	aa seq. 84–94	aa90 codon	type effect	unknown no.	origin/voucher tissue no.	GenBank acc. no.	reference
	<i>Onychoprion fuscatus</i>	Sooty Tern	FV T CVF S IIFTV	tcc	VS	Thr86	ZMUC 113320	GU129677	this study
	<i>Gygis alba</i>	White Tern	F IACIFC I IFTV	tgc	UVS		ZMUC 113340	GU129678	this study
	<i>Sternula albifrons</i>	Little Tern	FV T CIF S IIFTV	tcc	VS	Thr86	NRM 20076398	GU129679	this study
	<i>Chlidonias niger</i>	Black Tern	FV T CIF S IIFTV	tcc	VS	Thr86	NRM 20066936	GU129680	this study
	<i>Chlidonias hybrida</i>	Whiskered Tern	FV T CIF S IIFTV	tcc	VS	Thr86	ZMUC 131916	GU129681	this study
	<i>Sterna hirundo</i>	Common Tern	FV T CIF S IIFTV	tcc	VS	Thr86	Department of Animal Ecology, U.U.	GU129682	this study
Alcidae	<i>Thalasseus sandvicensis</i>	Sandwich Tern	FV T CI F S IIFTV	tcc	VS	Thr86		AY960720	Håstad <i>et al.</i> (2005)
	<i>Sterna paradisaea</i>	Arctic Tern	FV T CI F S IIFTV	tcc	VS	Thr86		AY960719	Håstad <i>et al.</i> (2005)
	<i>Uria aalge</i>	Common Murre	F IACI F S VFTV	agc	VS			AY227163	Ödeen & Håstad (2003)
	<i>Alca torda</i>	Razorbill	FV A CI F S VFTV	agc	VS			AY227159	Ödeen & Håstad (2003)
Sturnariidae	<i>Stercorarius longicaudus</i>	Long-tailed Jaeger	FV A CI F S VFTV	agc	VS		NRM 976537	GU129683	this study
	<i>Stercorarius parasiticus</i>	Parasitic Jaeger	FV A CI F S VFTV	agc	VS		NRM 20066101	GU129684	this study
	<i>Stercorarius macrominctus</i>	South Polar Skua	FV A CI F S VFTV	agc	VS		ZMUC 131807	GU129685	this study
	<i>Glaucidium brasilianum</i>	Collared Pratincole	F LA C VF S VFTV	agc	VS		M. Wilson, A.Ö. ZMUC 113386	GU129686	this study
Glareolidae	<i>Glaenda nuchalis</i>	Rock Pratincole	F LA C IF S VFTV	agc	VS		ZMUC 131583	GU129687	this study
	<i>Rhinoptilus chalcopterus</i>	Bronze-winged Courser	CLAC L IF S VFTV	agc	VS			GU129688	this study
	<i>Himantopus himantopus</i>	Black-winged Stilt	FV A CI F S VFTV	agc	VS			AY227156	Ödeen & Håstad (2003)
	<i>Haematopodidae</i>	<i>Haematopus ostralegus</i>	Eurasian Oystercatcher	F IACI F S VFTV	agc	VS		AY227155	Ödeen & Håstad (2003)
Charadriidae	<i>Pluvialis apricaria</i>	Eurasian Golden Plover	F IACI F S VFTV	agc	VS				
	<i>Charadrius dubius</i>	Little Ringed Plover	F IACI F S VFTV	agc	VS		Department of Animal Ecology, U.U.	AY227154	Ödeen & Håstad (2003)

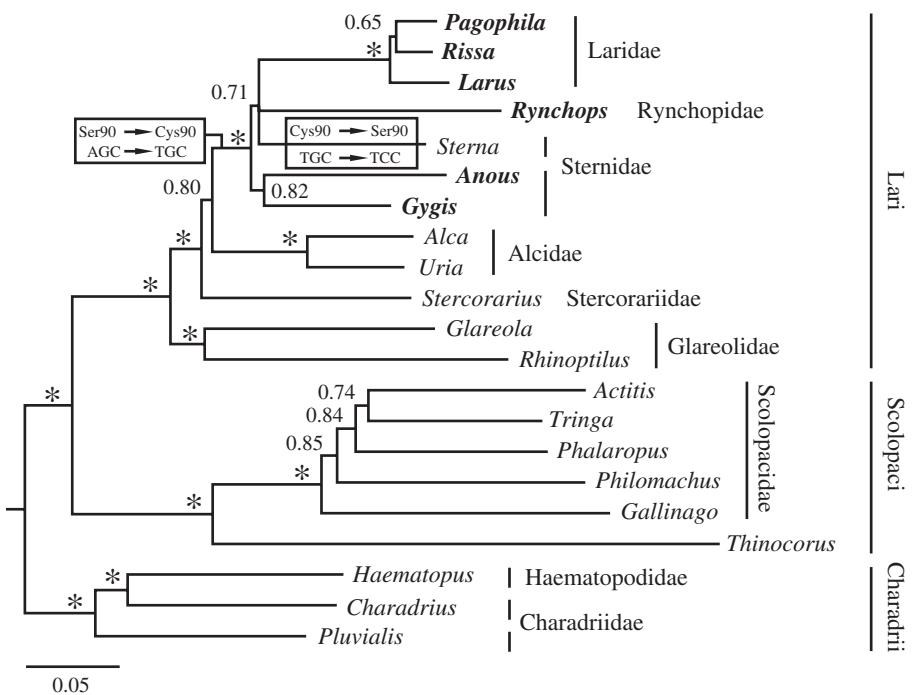


Figure 1. Bayesian inference tree of shorebirds based on mitochondrial ND2, cytochrome *b* and CO1, and nuclear RAG-1, FGB7 and myoglobin (approx. 7.4 kbp). The species having the UVS-pigment-conferring Ser90Cys substitution in their SWS1 opsin are in bold. The most parsimonious reconstruction of the sequence of mutations in the corresponding codon is shown in boxes. Numbers at nodes are posterior probabilities (values of 1.00 indicated by *). The name of the family Thinocoridae (comprising *Thinocorus*) is not marked owing to lack of space.

substitution at aa position 90 (Ser90Cys: Wilkie *et al.* 2000; Yokoyama *et al.* 2000), as specified by the codon TGC in the SWS1 opsin gene. We found this trait in four other shorebirds, namely *A. minutus*, *A. tenuirostris*, *G. alba* (Sternidae) and *Rynchops niger* (Rynchopidae). All other species examined carry Ser90, which is ancestral to Charadriiformes (Ödeen & Håstad 2003; Håstad *et al.* 2005), but translated from different synonymous codons. In general, VS shorebirds carry AGC; the exception is Sternidae (excluding *Anous* and *Gygis*), where all sequences read TCC instead. The distribution of these traits on the phylogeny is shown in figure 1 (and the electronic supplementary material, figures S1 and S2).

(b) Phylogeny

All analyses produced results largely in agreement with previous studies (Ericson *et al.* 2003; Paton *et al.* 2003; Paton & Baker 2006; Baker *et al.* 2007; Fain & Houde 2007), and identify three main clades corresponding to Lari, Scolopaci and Charadrii. Only the tree based on all loci (figure 1) is shown (combined nuclear and mitochondrial gene trees in the electronic supplementary material, figures S1 and S2). All trees recover a strongly supported clade comprising Laridae, Sternidae and *Rynchops*. Within this clade, in the tree based on all loci, Laridae, *Rynchops* and *Sterna* form a poorly supported clade, which is sister to an insufficiently supported clade comprising *Anous* and *Gygis*. The nuclear tree has the same topology, but even weaker support. In contrast, in the mitochondrial tree, Laridae, *Rynchops* and *Anous* form a moderately well-supported trichotomy that is sister to a strongly

supported clade containing *Gygis* as sister to the rest of Sternidae (excluding *Anous*).

4. DISCUSSION

Our results agree with previous studies in showing that gulls have Cys90 in their SWS1 opsins, characteristic of the UVS class of colour vision. This trait has been conserved during gull evolution, as revealed by the denser taxon sampling of this study compared with previous ones (Håstad *et al.* 2005). Another new finding is that gulls are not unique among shorebirds in being UVS, as the terns *A. minutus*, *A. tenuirostris* and *G. alba* and the skimmer *R. niger* share the UVS opsin forming Ser90Cys substitution.

The evolutionary interpretation of our results hinges on the correct inference of the phylogeny. As the relationships in the Laridae/Sternidae/Rynchopidae clade are somewhat uncertain, different interpretations are possible. However, the most parsimonious reconstruction suggests a Ser90Cys substitution in the common ancestor to Laridae/Sternidae/Rynchopidae, and a subsequent reversal to Ser in the Sternidae lineage (except *Anous* and *Gygis*), in all formed by two consecutive point mutations in the codon at aa position 90, from AGC to TGC to TCC. This is true even if the uncertainty in the gene tree is taken into account by considering the Sternidae branch (except *Anous* and *Gygis*) in alternative positions. For example, the posterior probability that *Sterna* is outside the clade comprising Laridae, *Rynchops*, *Anous* and *Gygis*—which would support a single Ser90Cys substitution but no subsequent reversal—is lesser than 0.06. Accordingly, our favoured hypothesis based on the

available data is a single shift from VS to UVS in the common ancestor to Laridae/Sternidae/Rynchopidae and a subsequent reversal in the Sternidae lineage (except *Anous* and *Gygis*).

All key aas we report in positions 86, 90 and 93 have already been described (Wilkie *et al.* 2000; Yokoyama *et al.* 2000; Ödeen & Håstad 2003), but the spectral tuning effects of some of them are still unknown. The effect of Cys86 in birds is probably negligible (Ödeen *et al.* 2009), but neither the effects of Thr86 or Ile86 nor Ile93 has been investigated *in vitro*. However, the λ_{max} of SWS1 pigment of *Anas platyrhynchos*, which carries Ile93, is not significantly different from that of species with aas of known effect in this position (Ödeen *et al.* 2009).

In one aspect, avian colour vision is under simple genetic control: the VS/UVS pigment opsin character depends on the Ser90Cys substitution, which can be caused by a single nucleotide mutation. Still, this character is surprisingly conserved among birds, indicating strong stabilizing selection. This is a rare example of a system where two very similar and well-understood variants of the same gene, which have a significantly different effect on the phenotype and ecology of the carrier, have been equally successful in a large taxon.

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