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## Targeting the cancer kinome through polypharmacology

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### Abstract

Kinase inhibitors are the largest class of new cancer drugs. However, it is already apparent that most tumours can escape from the inhibition of any single kinase. If it is necessary to inhibit multiple kinases, how do we choose which ones? In this Opinion article, we discuss some of the strategies that are currently being used to identify new therapeutic combinations of kinase targets.

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More than 10,000 patent applications for kinase inhibitors have been filed since 2001 in the United States alone<sup>1</sup>. This massive investment has been fuelled by the realization that kinases are intimately involved in cancer cell growth, proliferation and survival. Indeed, kinases and their direct regulators are among the most frequently mutated oncogenes and tumour suppressors<sup>2–4</sup>. Well known examples include the oncogenic kinases PIK3CA (the p110 $\alpha$  subunit of PI3K), epidermal growth factor receptor (EGFR) and BRAF; the Ras family of oncogenes, which activate both PI3Ks and Raf; and the PTEN tumour suppressor, which inhibits PI3K signalling.

Despite the excitement surrounding these targets, clinical progress has been uneven. Kinase inhibitors have revolutionized the treatment of a select group of diseases, such as chronic myeloid leukaemia (CML) and gastrointestinal stromal tumours (GIST), which are driven by a single oncogenic kinase; for these conditions, kinase inhibitors have achieved multi-year

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#### Competing interests statement

The authors declare no competing financial interests.

#### DATABASES

**Entrez Gene:** <http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=gene>

ERBB2 | KRAS

**National cancer institute Drug Dictionary:** <http://www.cancer.gov/drugdictionary/>

bevacizumab | cetuximab | dasatinib | erlotinib | flavopiridol | imatinib | irinotecan | lapatinib | nilotinib | rapamycin | sorafenib | trastuzumab

**UniProtKB:** <http://www.uniprot.org>

BRAF | EGFR | PIK3CA | PLK1 | PTEN | STK33 | TBK1 | VEGFR2

#### FURTHER INFORMATION

**PubMed database:** <http://www.ncbi.nlm.nih.gov/pubmed>

#### SUPPLEMENTARY INFORMATION

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increases in survival<sup>5–7</sup>. Smaller but significant responses have been observed for some cancers that are highly dependent on angiogenesis, and therefore sensitive to inhibitors of vascular endothelial growth factor (VEGF) signalling, such as renal cell carcinoma<sup>8–11</sup>.

Kinase inhibitors have been least effective in treating the types of cancer that have the highest mortality rates, such as lung, breast, colorectal, pancreatic and prostate cancer. Clinical trials show that the most effective kinase inhibitors prolong survival by only a few months for these cancers<sup>12–17</sup>. Results have been improved by identifying markers for patients that are more likely to respond to kinase inhibitor therapy — such as *EGFR* mutations in lung cancer<sup>18</sup>, *ERBB2* overexpression in breast cancer<sup>19</sup>, and wild-type *KRAS* in lung and colorectal cancer<sup>20,21</sup> — but even among these subgroups, relapse is inevitable for patients with disseminated disease.

Why has clinical progress been so challenging? One reason is that most tumours can escape from the inhibition of any single kinase (FIG. 1). This first became clear when resistance mutations in *BCR–ABL* were discovered in patients with CML who were resistant to imatinib<sup>22</sup>; similar mutations have now been detected in other kinases following treatment with kinase inhibitors<sup>23–26</sup>. Alternatively, tumours can acquire drug resistance through mechanisms that do not involve mutation of the target (FIG. 1a). These mechanisms include the activation of surrogate kinases that substitute for the drug target<sup>27</sup> and the inactivation of phosphatases to amplify the residual kinase activity that persists during drug treatment<sup>28</sup>. It is also clear that many tumours possess intrinsic resistance to kinase inhibitors at the time of initial therapy (FIG. 1b). This can result from the activation of multiple, redundant kinase signalling pathways<sup>29</sup> or the presence of activating mutations in downstream pathway components, such as *KRAS* or *PTEN*, which enable the tumour to bypass the drug target<sup>20,21,30</sup>.

Overcoming these resistance mechanisms will require targeting tumour cells at multiple levels, through either single drugs that bind to multiple proteins<sup>31</sup> or cocktails of highly selective inhibitors<sup>32</sup>. The challenge for the cancer research community is to learn how to predict the best combinations of targets and then prioritize those combinations for clinical testing. This is a daunting task, because the number of possible target combinations is almost limitless, but clinical trials are slow and expensive.

## Targeting one kinase with multiple drugs

If a tumour depends on the activity of a single kinase, then using multiple drugs to target that kinase can be effective. This was first demonstrated in CML, in which early clinical trials showed that more than 90% of patients with chronic phase disease responded to the *BCR–ABL* inhibitor imatinib<sup>5</sup> (TABLE 1), but that a subset of those patients relapsed while on the drug. Disease progression was associated with the emergence of leukaemic cells bearing mutations in *BCR–ABL* that block imatinib binding<sup>22</sup>, suggesting that drugs targeting these *BCR–ABL* mutants would be effective. Two second-generation *BCR–ABL* inhibitors were developed (dasatinib and nilotinib) that retain activity against most of the more than 50 clinically observed *BCR–ABL* resistance mutations, and these drugs are highly effective against imatinib-resistant disease<sup>33,34</sup>. However, a common *BCR–ABL* mutation (T315I) prevents the binding of all three drugs, and this has emerged as the default allele for many patients on long-term inhibitor therapy<sup>22,35</sup>. To address this problem, third-generation drugs have been developed that potentially inhibit *BCR–ABL* T315I. These agents are effective in preclinical models of drug-resistant CML<sup>36–39</sup>, and four such compounds are currently in clinical trials. Some patients have now survived more than 10 years since starting treatment by undergoing sequential therapy with three generations of *BCR–ABL* inhibitors<sup>40</sup>, proving that it is possible to extend the therapeutic response in CML by repeatedly targeting the same kinase.

A similar approach has been used to target the ERBB2 receptor tyrosine kinase in breast cancer. Trastuzumab is a monoclonal antibody that binds to the extracellular domain of ERBB2, thereby both inhibiting ERBB2 signalling and recruiting immune cells to the tumour<sup>19</sup>; however, patients with metastatic cancer who are treated with trastuzumab invariably relapse. The mechanism of trastuzumab resistance is not understood, but it is clear that resistant tumours remain dependent on ERBB2 signalling. This is because patients with breast cancer who have progressed on trastuzumab therapy nonetheless respond to lapatinib<sup>14</sup>, a small molecule inhibitor of the tyrosine kinase domain of ERBB2. Therefore, it is possible to induce a second response in these patients by targeting ERBB2 with a drug that binds to a different site on the protein. Unlike CML, however, the clinical response to lapatinib in metastatic breast cancer is brief, and disease progression typically occurs within a few months<sup>14</sup>.

These examples show that in certain cases sequential targeting of a single kinase with multiple drugs can prolong the therapeutic response. It is unclear how broadly this model applies, because it is unclear how many tumours are truly dependent on a single oncogene (a state referred to as 'oncogene addiction' (REF. 41)). The strongest evidence in favour of this hypothesis is the discovery of resistance mutations after kinase inhibitor treatment in CML, GIST, lung cancer and a myeloproliferative disorder known as hypereosinophilic syndrome<sup>22–26</sup>. Such mutations are definitive proof that the mutated kinase was required for the survival of that tumour. The oncogene addiction model is also supported by many preclinical studies showing that tumour cell lines containing an activating mutation or amplification of a kinase can be more sensitive to inhibitors of that kinase *in vitro*<sup>42–44</sup>.

Conversely, the detection of an oncogenic kinase mutation does not guarantee sensitivity to the corresponding kinase inhibitor. For example, mutations in *PIK3CA* or *PTEN* are poor predictors of the sensitivity of tumour cell lines to PI3K inhibitors<sup>31,45</sup>. Mutations in *KRAS* do not, in general, sensitize tumour cells to inhibitors of Raf or Mek<sup>43,46</sup>. Indeed, the response of most tumours to inhibition of an oncogene is much less dramatic than the response in CML, in which even transient inhibition of BCR–ABL irreversibly commits cells to apoptosis<sup>44</sup> (FIG. 2). In this respect, it is worth noting that the term oncogene addiction gained widespread use because it describes a paradox: inhibiting an oncogene would be predicted to reverse the gain of function caused by the oncogene, not kill all the tumour cells. It is only recently that this term has been conflated with the idea that oncogenes should be expected to be required for tumour survival. For this reason, there is clearly a need to identify additional vulnerabilities in tumours beyond the genes that are directly mutated.

## Targeting nodes in a signalling network

Three sets of targets collectively account for a large proportion of current efforts in kinase inhibitor drug discovery. These are the receptor tyrosine kinases (for example, EGFR, ERBB2, platelet-derived growth factor receptor (PDGFR) and VEGF receptor 2 (VEGFR2)), the kinases in the MAPK pathway (for example, BRAF, MEK1 and MEK2) and the kinases in the PI3K pathway (for example, PIK3CA, Akt and mTOR). These three groups of targets are mechanistically linked because most receptor tyrosine kinases activate the MAPK and PI3K pathways as their primary signalling function (FIG. 3).

There is a compelling biological rationale for targeting each of these groups in combination. For example, clinical resistance to tyrosine kinase inhibitors is often associated with reactivation of PI3K signalling<sup>28</sup>. Therefore, the effectiveness of tyrosine kinase inhibitors might be increased by combination with an inhibitor of the PI3K pathway. This combination has been shown to be effective in animal models and is undergoing extensive clinical testing: at least 21 clinical trials are currently evaluating the combination of a tyrosine kinase inhibitor and an mTOR inhibitor in several types of cancer. There has been a particular emphasis on the

use of PI3K inhibitors to sensitize tumours to inhibitors of EGFR or ERBB2 (such as erlotinib<sup>47</sup>, lapatinib<sup>48</sup> and trastuzumab<sup>49</sup>). This is because the anti-tumour activity of EGFR and/or ERBB2 inhibitors has been correlated with their ability to inhibit the phosphorylation of ERBB3, a kinase-inactive receptor that primarily functions to activate the PI3K pathway<sup>28</sup>.

Other target combinations are suggested by the connectivity of the signalling network. For example, mTOR activates a well-characterized negative feedback loop that inhibits the activity of PI3K (FIG. 3a). mTOR inhibitors such as rapamycin block this negative feedback loop, resulting in hyper-activation of PI3K that may counteract the anti-proliferative effect of mTOR inhibition. For this reason, it has been proposed that the dual inhibition of PI3K and mTOR may be more effective than inhibiting either target alone. Preclinical experiments support this idea<sup>45</sup>, and drugs such as PP121 that target multiple steps in this pathway have been designed<sup>31</sup> (FIG. 3a). Several dual PI3K and mTOR inhibitors are currently being evaluated in clinical trials (for example, NVP-BEZ235, BGT226 and XL765) alongside agents that selectively target either PI3Ks (for example, XL147 and GDC-0941) or mTOR (for example, OSI-027, AZD8055 and rapamycin analogues). As there are practical challenges associated with developing both multi-targeted single agents and multi-drug cocktails<sup>50</sup>, it will be interesting to see which approach emerges as the most successful from these clinical trials.

Combination therapy can be used in other cases to target an otherwise undruggable protein. For example, *KRAS* is one of the most commonly mutated oncogenes, but efforts to find Ras inhibitors have been unsuccessful. It was long believed that the MAPK pathway was the primary Ras effector in most tumours<sup>51</sup>, but Raf and Mek inhibitors have inconsistent activity against tumour cells with Ras mutations<sup>43,46</sup>. Ras also directly binds to and activates PI3K<sup>52</sup>, and the disruption of this interaction prevents *KRAS*-driven tumorigenesis in the mouse<sup>53</sup>. For this reason, it may be necessary to inhibit both the MAPK and PI3K pathways to block the growth of tumours with Ras mutations. This conclusion is supported by data showing that resistance to Mek inhibitors in some *KRAS*-mutant cells is caused by mutations in *PIK3CA* or *P TEN*, and that this resistance is reversed by PI3K inhibition<sup>54</sup>. Moreover, the combination of PI3K and Mek inhibitors is active in a mouse model of *KRAS*-driven lung cancer<sup>55</sup>. The rationale for this combination is so compelling that Merck and AstraZeneca recently announced a joint Phase I clinical trial that will test the combination of an Akt inhibitor (MK-2206) and a Mek inhibitor (AZD6244) against solid tumours (FIG. 3b).

## Limitations of rational drug combinations

The challenge associated with developing these types of rationally designed drug cocktails is that preclinical experiments do not predict their efficacy in humans. This is true even when the individual agents have already shown clinical anticancer activity. For example, preclinical experiments supported the combination of gefitinib and trastuzumab in breast cancer<sup>56,57</sup>, erlotinib and bevacizumab in renal cell carcinoma<sup>58</sup>, and cetuximab and bevacizumab in colorectal cancer<sup>59</sup>, but all of these failed in clinical trials<sup>58,60,61</sup>. In the case of cetuximab and bevacizumab, the drug combination reduced survival compared with the single agents<sup>60</sup>.

In some cases, these discrepancies may be due to misinterpretation of the preclinical data, rather than a failure of the preclinical model itself. For example, careful studies have shown that the addition of gefitinib to trastuzumab therapy in xenograft models of breast cancer results in only modest additional efficacy<sup>62</sup>, and that this additional benefit requires gefitinib concentrations that may be toxic in humans<sup>63</sup>. In other cases, subtle changes in the dosing regimen can have a large effect on the activity of the combination. For example, preclinical studies of the cyclin-dependent kinase inhibitor flavopiridol and the topoisomerase inhibitor irinotecan showed that this combination can effectively induce apoptosis in colon cancer cells when administered in

a specific sequence (irinotecan followed by flavopiridol, resulting in apoptosis in 43% of the cells)<sup>64</sup>. The reverse sequence of drugs (15% apoptosis) and concurrent therapy (30% apoptosis) were both less effective. This finding was rationalized by a model in which pretreatment with flavopiridol arrested cells in the G1 phase of the cell cycle and thereby reduced the number of cells progressing through S phase and therefore irinotecan sensitivity<sup>64</sup>. A subsequent clinical trial validated the safety and preliminary efficacy of this sequential dosing regimen<sup>65</sup>.

Preclinical studies of drug combinations are probably biased towards validating the targets that are already believed to be important, and this bias limits their ability to prioritize new drug combinations for clinical testing. For example, all kinase inhibitors have some toxicity to cells, and for this reason two kinase inhibitors can usually be shown to be more toxic than either compound alone. For these comparisons it is often unclear what should be used as the normal cell to measure therapeutic index<sup>66</sup>, and in many cases the survival of the mouse in a xenograft experiment is the only evidence of differential toxicity. This can be addressed to some degree by correlating lethality with genotype across many tumour cell lines<sup>42</sup> or by using pairs of isogenic cell lines that differ at a single locus<sup>66</sup>, but this becomes challenging when comparing drug combinations.

For a small group of kinase targets with an undisputed role in cancer — such as the oncogenic receptor tyrosine kinases and the core components of the PI3K and MAPK pathways — numerous clinical trials of drug combinations are planned or underway. It is uncertain, however, that these kinases are the best cancer drug targets<sup>67</sup>, and the route to clinical testing for combinations of drugs that target other kinases is less straight-forward. One major obstacle is that it is difficult to conduct clinical trials combining two investigational drugs, and even more difficult if the two drugs originate from different pharmaceutical companies<sup>68,69</sup>. Companies are reluctant to conduct joint clinical trials of early-stage compounds because of fears about loss of intellectual property and the possibility of an unforeseen side effect from the combination<sup>68,69</sup>. This creates a Catch-22 scenario: many kinase inhibitors are likely to be effective only as part of a combination therapy, but it will be difficult to test those combinations until after the drugs are approved as single agents. Indeed, the joint venture mentioned above between AstraZeneca and Merck to test Akt and Mek inhibitors in combination was reported in national media, such as *The Wall Street Journal*, partly because such early stage collaborations are so rare<sup>70</sup>. In the field of AIDS research, this problem was addressed in 1993 by the formation of the Inter-Company Collaboration for AIDS Drug Development that coordinated the testing of drug cocktails by 15 pharmaceutical companies<sup>71</sup>. However, there is not yet a comparable mechanism for companies to collaborate to test new combinations of investigational drugs in oncology, where there is arguably the greatest need and opportunity.

## Using RNAi to discover new targets

The development of RNA interference (RNAi) has made it possible to directly screen for the genes required for tumour proliferation in mammalian cells. These screens have two advantages. First, they can identify new drug targets, as any gene that selectively blocks tumour growth when knocked down by RNAi is a candidate. Second, these screens provide an unbiased test of models of tumour signalling, because they directly examine which genes are most important to the tumour. This perspective is valuable, because most combination therapies are based on simple models of tumour signalling; however, there is little evidence that such models capture the most crucial interactions in the tumour cell, which could be highly indirect and inaccessible to simple reasoning.

Three recent papers illustrate the power of large-scale RNAi screens to address this problem by looking for genes that are selectively required for the growth of tumour cells expressing an



activated KRAS mutant<sup>72–74</sup>. Luo *et al.*<sup>72</sup> screened ~75,000 short hairpin RNAs (shRNAs) and found 83 shRNAs targeting 77 genes that preferentially impaired the growth of *KRAS*<sup>G13D</sup> cells compared with control cells in which the *KRAS*<sup>G13D</sup> allele had been disrupted by homologous recombination<sup>72</sup>. Analysis of these hits revealed an unexpected enrichment of a network of genes involved in mitosis. A small molecule inhibitor of the mitotic kinase polo-like kinase 1 (PLK1) had increased cytotoxicity to *KRAS*-mutant cells *in vitro* and *in vivo*<sup>72</sup>.

Scholl *et al.*<sup>73</sup> screened a smaller set of shRNAs (5,024 targeting 1,011 genes) against a broader panel of cells that included 8 tumour cell lines (4 *KRAS*<sup>G13D</sup> mutant and 4 *KRAS* wild-type) and 2 control cell lines<sup>73</sup>. The top hit was STK33, a serine threonine kinase in the calmodulin kinase family with no previous connection to Ras signalling or cancer. shRNAs targeting STK33 induced *KRAS* mutation-dependent toxicity in a broad panel of tumour cell lines, through a mechanism that may involve modulation of S6K1 kinase activity<sup>73</sup>.

Barbie *et al.*<sup>74</sup> screened a panel of shRNAs targeting kinases, phosphatases and oncogenes against a panel of 19 tumour cell lines and then extracted from these data the genes selectively required for the survival of *KRAS*-mutant cells<sup>74</sup>. The top hit from this screen was TBK1, a protein kinase that activates nuclear factor- $\kappa$ B (NF- $\kappa$ B) signalling by phosphorylating the NF- $\kappa$ B inhibitory protein I $\kappa$ B $\alpha$ . A companion paper showed that genetic inhibition of NF- $\kappa$ B signalling was sufficient to block tumour development in a mouse model of *KRAS*-driven lung adenocarcinoma<sup>75</sup>.

A common finding from all three papers was that, although many genes were required for the survival of *KRAS*-mutant cells, few of those genes could have been predicted in advance on the basis of known biochemical interactions or models of Ras signalling. Among the three kinases (PLK1, STK33 and TBK1) that were the focus of follow-up experiments, only TBK1 had been previously linked to Ras (through a pathway involving the exocyst complex, RALB and RALGDS), and this protein could hardly be described as a well-known Ras effector. This is even more remarkable when we consider that Ras and its downstream targets are among the most intensely studied proteins in biology.

Similar results were described in a series of papers that attempted to define the ‘essential kinome’ that is required for cell proliferation and survival<sup>67,76,77</sup>. This was done by carrying out kinome-wide shRNA screens on a large panel of tumour cell lines, primary cells and pairs of isogenic cells that differed in the expression of a single gene. The primary conclusion from these papers was that there was little overlap in the kinases that are required for cell proliferation across many different cell lines. Indeed, there was no correlation between the number of PubMed citations for a kinase and the likelihood that the kinase was important for tumour cell proliferation. In the words of the authors<sup>77</sup>: “Although the regulation of cell proliferation and survival are heavily studied areas, we did not see a bias in these screens toward the identification of previously known and well studied kinases, suggesting that our knowledge of the molecular events in these areas is still meager.” (D. A. Grueneberg *et al.*, 2008).

Given the unpredictable sensitivities of tumour cells to shRNAs targeting a single kinase, it may be possible to identify new pairs of targets by screening shRNAs in the presence of a drug. An early experiment in this area looked for shRNAs that synergistically killed cancer cells in the presence of A-443654, a small molecule inhibitor of Akt<sup>78</sup>. This was motivated by the surprisingly weak anti-tumour activity of A-443654 as a single agent in preclinical models<sup>79</sup>. Two kinases were identified in this screen: casein kinase 1,  $\gamma$ 3 (CSNK1G3) and inositol polyphosphate multikinase (IPMK). Neither of these kinases had previously been linked to Akt signalling or cancer. However, knock down of both genes potentiated the inhibition of phosphorylation of Akt and ribosomal protein S6, suggesting that these kinases may have a cryptic role in regulating signalling through the PI3K pathway.

## Barriers to translating RNAi into drugs

RNAi screens can help challenge our assumptions about the genes that are most important in cancer. However, there are considerable obstacles to translating any hit from one of these screens into a new drug. Most RNAi screens measure only cell proliferation *in vitro*, which ignores most of the capabilities of a tumour. Therefore, it will be necessary to validate the large number of genes that emerge from these screens in more complex and time-consuming models. Once these hits are validated, they become subject to the same caveats that accompany potential drug targets identified in any other way.

In this respect, it is important to emphasize that there is not a direct correlation between RNAi knockdown of a gene and the identification of a potential drug target. Most drugs cannot be replicated by an shRNA because, for example, the drug interferes with multiple targets or inhibits a single domain of a multidomain protein only. Likewise, most shRNAs cannot be replicated by a drug, because most proteins are undruggable. Indeed, there are many examples in which an shRNA (or gene knockout) and a drug targeting the same protein give different phenotypes, and the reasons for these differences have been extensively documented (for a review of this topic see REF. 80). As a result, RNAi screens may be more likely to expose the gaps in our knowledge of cancer biology than to directly point the way to new therapeutic approaches.

## Using drugs to discover kinase targets

Historically, most drugs were discovered because they possessed activity in cells or animals, and their targets and mechanism of action were elucidated only later. This is sometimes called 'phenotype-based' drug discovery because the phenotype was discovered before the target. By contrast, almost all modern drug discovery is 'target-based', meaning that the target is selected first, on the basis of a hypothesis about its role in disease.

Nonetheless, there are instances in which phenotype-based drug discovery has contributed to the development of kinase inhibitors for cancer, albeit unintentionally. One example is sorafenib, which was originally designed as an inhibitor of Raf based on the logic that Raf inhibition might be effective for Ras-mutant tumours<sup>81</sup>. Sorafenib has yet to show clinical benefit for tumours that contain frequent Ras mutations, such as lung and pancreatic cancer, and has also failed in clinical trials for the treatment of melanoma<sup>82</sup>, a disease that has a high rate of BRAF mutations<sup>43,83</sup>. However, in early clinical trials of sorafenib (which were designed to establish safety and therefore contained a diverse patient population) responses were observed in two unexpected tumour types<sup>84</sup>: renal cell (kidney) and hepatocellular (liver) cancer. One patient with kidney cancer in an early Phase I trial achieved stable disease for 2 years<sup>84</sup>, leading to the broader testing and approval of sorafenib for kidney cancer (and more recently liver cancer). The efficacy of sorafenib in kidney cancer is now attributed to the inhibition of VEGFR2 in endothelial cells, which blocks angiogenesis, rather than the inhibition of Raf in the tumour. Preclinical studies have shown that the inhibition of an additional target, PDGFR in pericytes, may be important<sup>85</sup>. Therefore, sorafenib probably blocks tumour growth through the inhibition of two kinases, expressed in different tissues, neither of which was the intended target of the drug.

Imatinib provides a second example of serendipitous target discovery. After its initial approval for the treatment of CML, imatinib was tested in five patients with hypereosinophilic syndrome, a disease of unknown molecular origin, based on the reasoning that treatments that are effective in CML are sometimes also effective in patients with hypereosinophilia<sup>86</sup> (even though the mechanism of action of those other treatments, such as hydroxyurea and interferon- $\alpha$ , is unrelated to the mechanism of imatinib). Remarkably, four of the five patients treated with imatinib showed a complete haematological response (normalized eosinophil counts), such

that they were able to discontinue other therapies. Analysis of DNA from the leukocytes of these patients led to the discovery of a chromosomal rearrangement that generated a fusion between *PDGFRA* and *FIP1L1*, producing a constitutively active PDGFR kinase<sup>24</sup>. As PDGFR is one of a small number of kinases inhibited by imatinib, this suggested that PDGFR activation was probably the cause of the disease. This was confirmed by the discovery of a T674I resistance mutation in PDGFR in a patient who had relapsed from imatinib therapy<sup>24</sup>.

As these examples show, the advantage of using drugs to identify cancer targets is that they can reveal in an unbiased way the proteins most essential to the tumour. The major limitation of this approach is that it is difficult to identify the targets of a molecule that has an unknown mechanism of action<sup>87</sup>. If the target is unknown, then it is difficult to increase the potency of the compound by medicinal chemistry. It can also be challenging to determine whether the efficacy and toxicity of the drug are linked (because they reside in the same target) or separable (because they reside in different targets). For these reasons, it is often impossible to improve compounds that are identified in a screen but have an unknown mechanism of action.

## Targeted polypharmacology

In the case of sorafenib and imatinib, it was straightforward to identify the relevant targets of those drugs, because the targets were almost certain to be kinases. As these two drugs have a relatively small number of high-affinity targets in the human kinome (fewer than 20), the possibilities could be rapidly tested. Could this approach be generalized, so that kinase inhibitors could be used to search in an unbiased way for new combinations of therapeutic targets?

A unique feature of kinase inhibitors is that they have the potential for greater target promiscuity than almost any other type of drug. This is because the kinase superfamily (including the structurally related protein, lipid and small molecule kinases) is the largest family of druggable genes that binds to a common substrate (ATP). Kinases differ in this respect from other large gene families, such as G protein-coupled receptors, which interact in their druggable site with a wide range of structurally diverse ligands, including both peptides and small molecules. This fact has been emphasized<sup>50</sup> by noting that the kinase inhibitor sunitinib inhibits at least 79 kinases at low micromolar concentrations, whereas all the other approved drugs combined target only 320 proteins. Therefore, individual kinase inhibitors have an enormous potential for unpredicted target combinations and so new biological activities.

Despite this potential for promiscuity, it is increasingly feasible to enumerate the targets of kinase inhibitors in a systematic way. This is because most kinases can be heterologously expressed, either as a soluble kinase domain or on the surface of phage, and assayed for drug binding in a purified format. Although there are exceptions, the activity of most kinase inhibitors in cells correlates with biochemical parameters that can be measured *in vitro*, such as the dissociation constant ( $K_D$ ) of the drug and the Michaelis–Menten constant for ATP binding ( $K_{M,ATP}$ ) of the kinase<sup>88</sup>. As kinases have become increasingly important drug targets, the measurement of these biochemical parameters has been industrialized, and there are now many vendors that offer to screen compounds against panels of kinases that approach or exceed half of the kinome (FIG. 4a). As the cost of assaying compounds against these panels has decreased, it has transformed the types of experiments that are feasible (FIG. 4b). For example, a widely cited paper from 2000 reported the specificity of 24 commonly used kinase inhibitors against 28 kinases (approximately 700 kinase–drug pairs)<sup>89</sup>. In 2007, the same group published a follow-up paper that analysed the specificity of 65 common kinase inhibitors against 70 kinases (approximately 4,500 kinase–drug pairs)<sup>90</sup>. In 2008, scientists from GlaxoSmithKline reported the testing of a panel of 577 diverse kinase inhibitors against 203 kinases (more than



117,000 unique kinase–drug pairs)<sup>91</sup>; in this case, the aim was not to evaluate any specific compound but to characterize the selectivity properties of kinase inhibitors as a drug class.

Extrapolating from these trends, it is plausible that some drug discovery programmes in the near future will profile every kinase inhibitor that is synthesized against most of the kinome. This would occur before any biological testing, as a component of routine compound characterization. The availability of selectivity data on this scale would enable medicinal chemistry to focus on optimizing drug profiles against complex patterns of kinases that gave a desired phenotype, rather than attempting to maximize specificity for a single target. It is likely that drug discovery at some pharmaceutical companies already operates in this way to some degree, although it may not be explicitly acknowledged.

What would be the advantage of this approach? The primary advantage is that it allows for target serendipity — the discovery of target combinations that could not have been predicted, but that are optimal for killing tumour cells — while allowing medicinal chemists to optimize compounds based on biochemical measurements against purified proteins. This has the potential to address the limitations of both target-based drug discovery, which often fails because the target is wrong, and phenotype-based drug discovery, which often fails because the compounds cannot be optimized.

This type of ‘targeted polypharmacology’ would represent a considerable challenge to medicinal chemists, who would be asked to carry out chemical optimization against multi-dimensional target profiles. However, there is already evidence that this is possible for certain target combinations<sup>31,92</sup>, and kinase drug discovery seems to be the ideal setting to test this model. We analysed a large database of kinase inhibitor selectivity data<sup>93</sup> to discover whether certain combinations of kinase targets are enriched among known kinase inhibitors; whether the preference for these target combinations could be rationalized on the basis of sequence analysis; and whether this could be used to estimate the combinatorial druggability of most of the kinome that has not yet been targeted by a small molecule (FIG. 5; see Supplementary information S1 (figure)). We have found, consistent with previous analyses<sup>93</sup>, that there are clearly clusters of kinases that tend to be inhibited by similar drugs, but that there are also many target combinations that should be accessible but remain undiscovered. We interpret this to mean that there is an important opportunity to discover multi-targeted kinase inhibitors with new biological activities.

We have focused on approaches to identify combinations of kinase targets with increased anticancer activity, but understanding the basis for kinase inhibitor-mediated toxicity to normal cells is also valuable, as this information will improve efforts to increase therapeutic index. The broad kinome profiling of clinically approved and investigational kinase inhibitors is likely to help identify such problematic kinase targets. Removing these toxicity-associated kinases from new drug candidates may allow for more complete inhibition of cancer cell targets while avoiding systemic toxicity.

## Conclusions

Many different approaches will be necessary to identify the best combinations of targeted therapies for cancer. However, it is important to begin to consider the challenges that may be faced in the near future, when drugs targeting every kinase linked to cancer have been tested in clinical trials, but survival rates for most types of cancer have only marginally improved. It will not be sufficient in this case to simply pursue the next set of oncogenes, because tumour sequencing projects have already shown that such oncogenes do not exist, at least among the genes that are mutated with high frequency<sup>2–4</sup>. Therefore the burden will be on the cancer

research community to think of more creative ways to target important proteins such as kinases that have already been identified.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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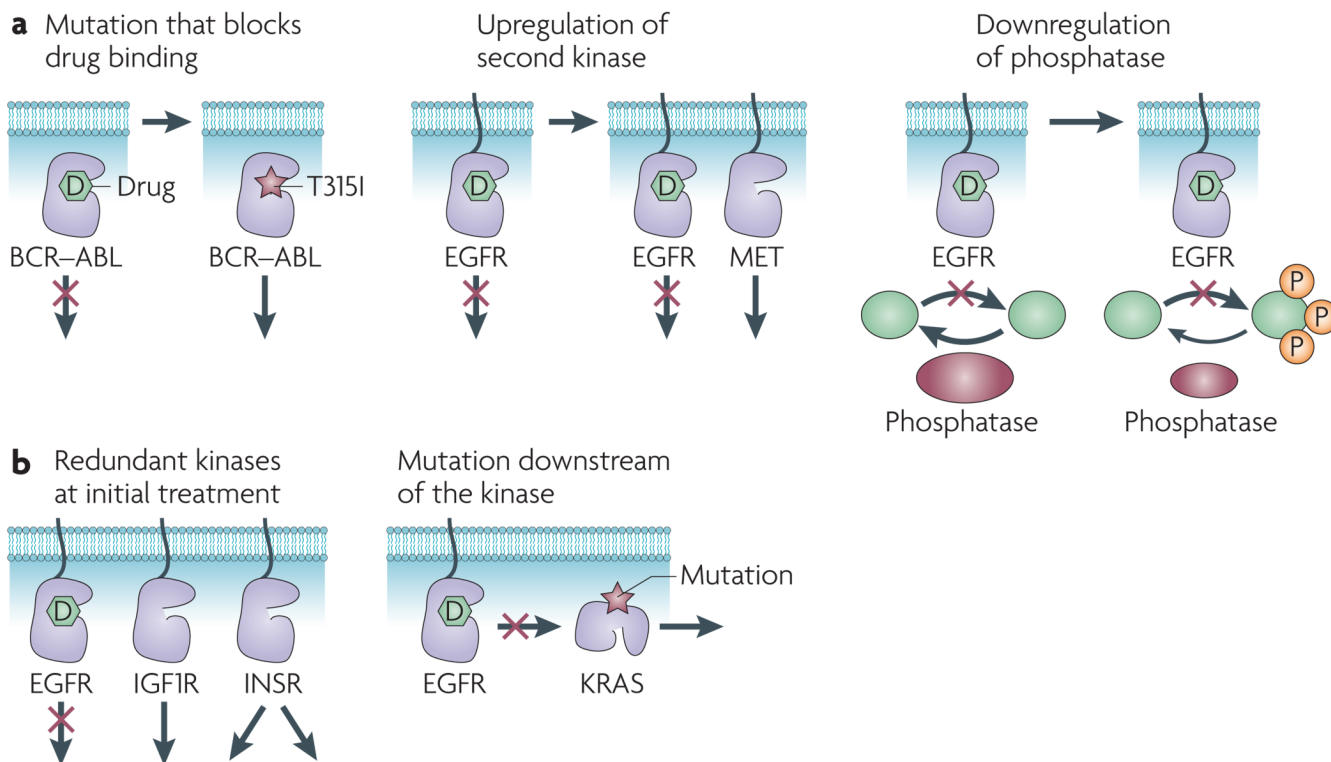
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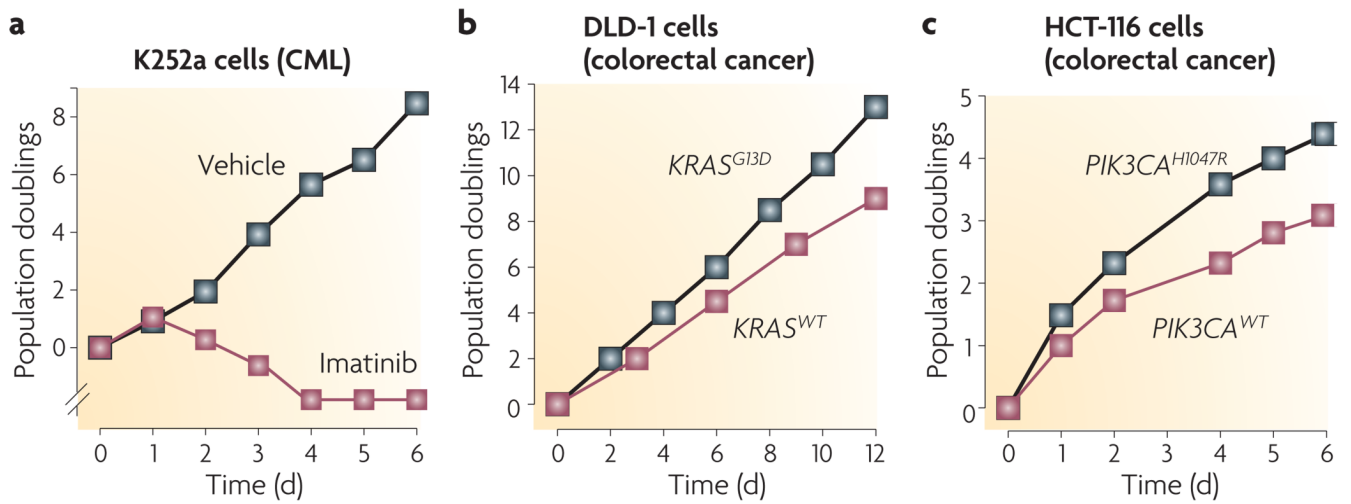


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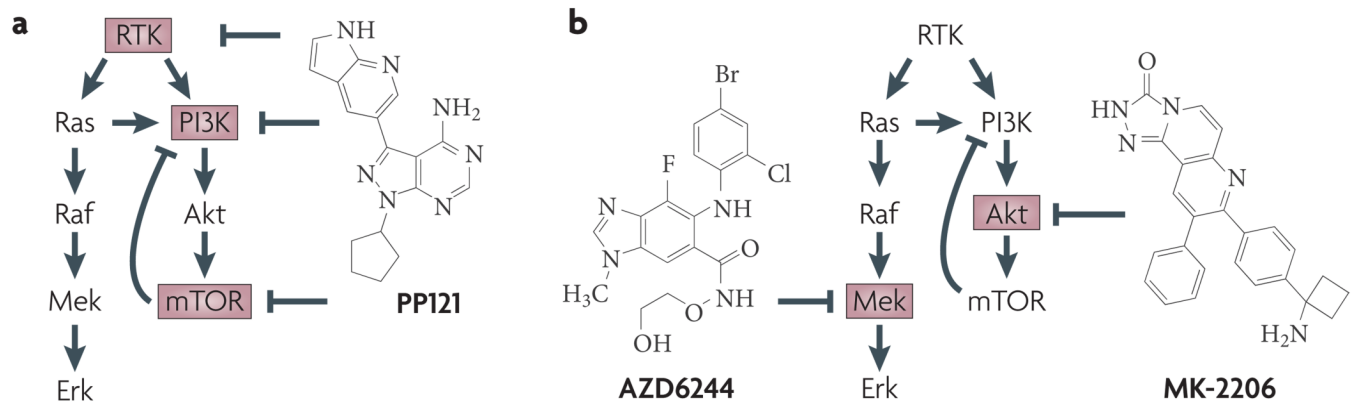
**Figure 1. Resistance to kinase inhibitors**

**a** | Mechanisms of acquired resistance. treatment with kinase inhibitors can select for mutations that block drug binding (left panel). this was first demonstrated for the T315I mutation in BCR-ABL in chronic myeloid leukaemia<sup>22</sup>. treatment with a kinase inhibitor can induce upregulation of a second kinase that substitutes for the drug target (centre panel). The receptor tyrosine kinase MET (also known as hepatocyte growth factor receptor) has been shown to be overexpressed in lung cancer cells that acquire resistance to epidermal growth factor receptor (EGFR) inhibitors<sup>27</sup>. Tumour cells can respond to treatment with a kinase inhibitor by downregulating the phosphatase that normally dephosphorylates the substrates of that kinase (right panel). This has the effect of decreasing the cellular potency of the kinase inhibitor. this mechanism has been observed in acquired resistance to EGFR inhibitors in breast cancer cells<sup>28</sup>. **b** | Mechanisms of intrinsic resistance. Many tumours express multiple oncogenic kinases that signal redundantly to promote cell survival (left panel). For example, some gliomas show constitutive activation of multiple receptor tyrosine kinases<sup>29</sup>. Mutational activation of a downstream pathway component can reduce the effectiveness of a kinase inhibitor (right panel). *KRAS* mutations are associated with resistance to EGFR inhibitors in lung and colorectal cancer<sup>20-21</sup>. IGF1R, insulin-like growth factor 1 receptor; INSR, insulin receptor; P, phosphorylation.



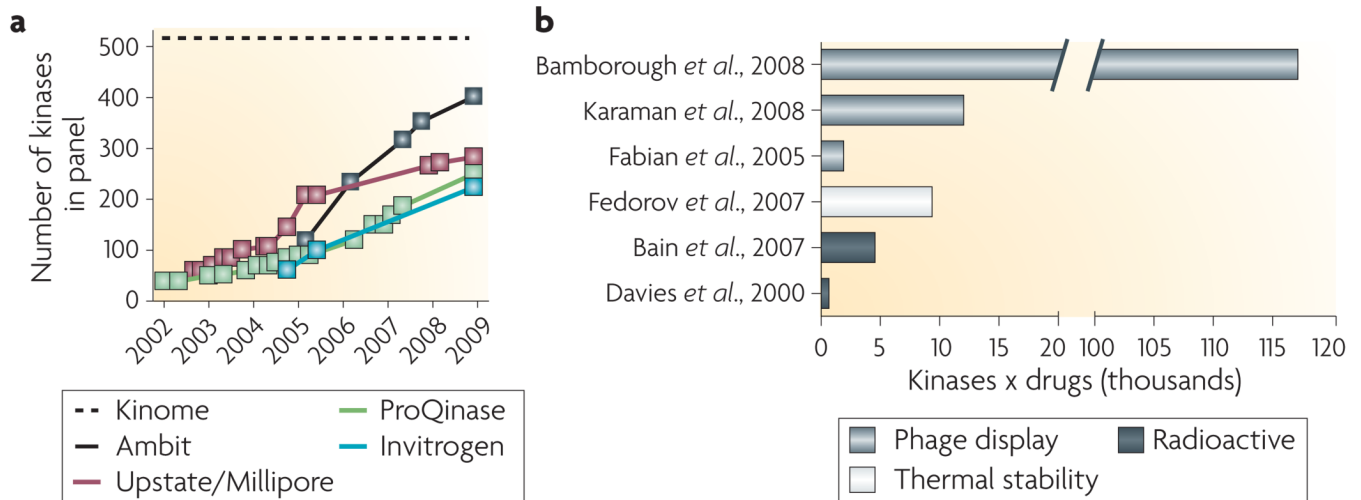
**Figure 2. Degrees of oncogene addiction**

Three examples of oncogene addiction drawn from the recent literature. **a** | Treatment of K252a chronic myeloid leukaemia (CML) cells with the BCR–ABL inhibitor imatinib results in complete cell death by day 4 (REF. 31). **b** | Disruption of the  $KRAS^{G13D}$  oncogene in DLD-1 colorectal cancer cells slows the rate of cell proliferation<sup>72</sup>. **c** | Disruption of the  $PIK3CA^{H1047R}$  oncogene in HCT-116 colorectal cancer cells slows the rate of cell proliferation<sup>94</sup>. WT, wild type.



**Figure 3. Strategies for multi-targeted kinase inhibition**

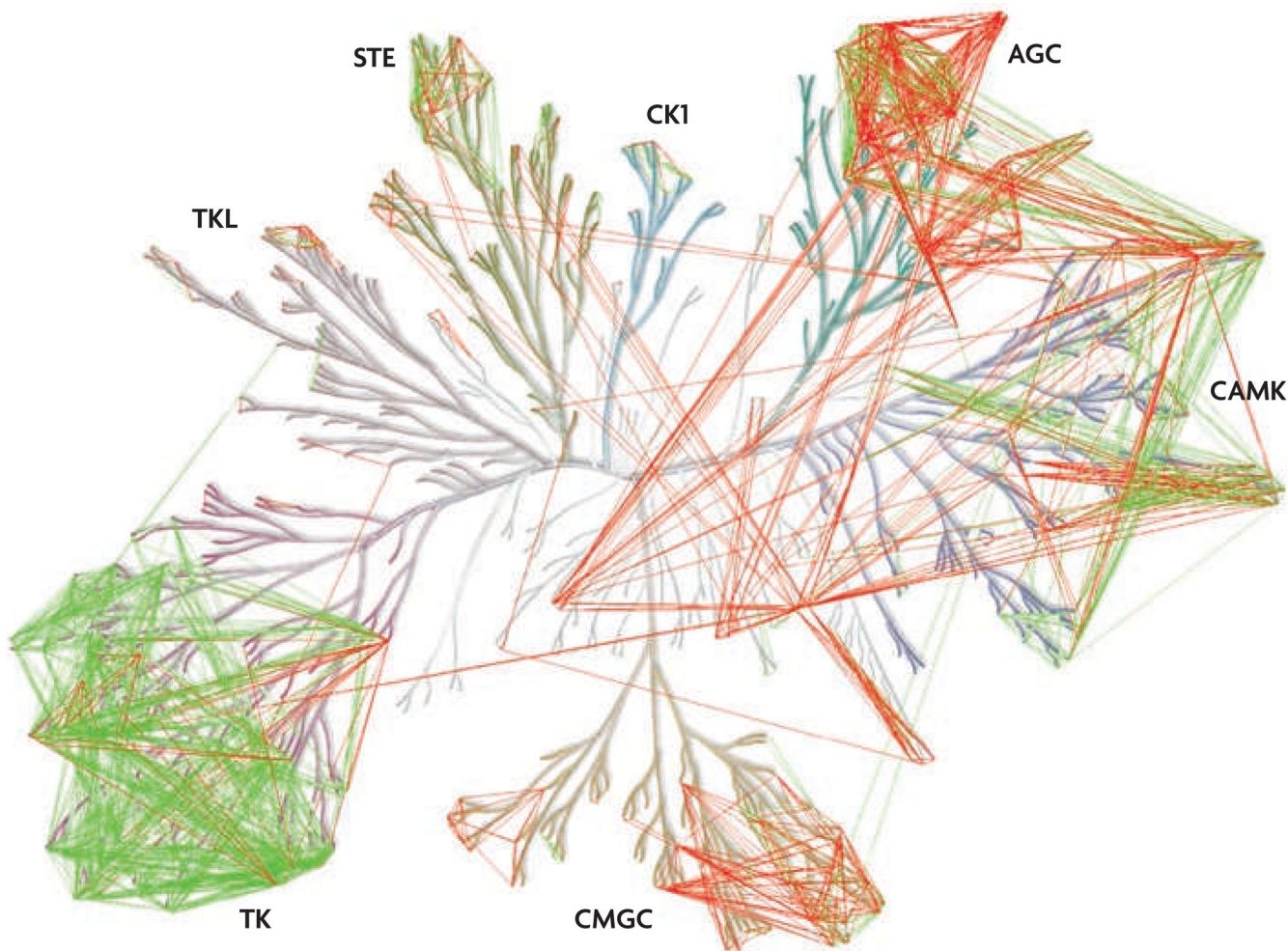
**a** | The single agent PP121 was shown to target both tyrosine kinases (such as vascular endothelial growth factor receptor 2, BCR-ABL and RET) and PI3K family members such as PIK3CA and mTOR (inhibitor targets are shown in red boxes). Note that the combined inhibition of mTOR and PI3K by PP121 disables a negative feedback loop in which mTOR inhibits PI3K. **b** | the combination of the MEK inhibitor AZD6244 and the Akt inhibitor MK-2206 results in the inhibition of both the MAPK and PI3K pathways. This combination is being evaluated in clinical trials. RTK, receptor tyrosine kinase.



**Figure 4. Selectivity profiling of kinase inhibitors**

**a** | The number of kinases available for screening from commercial vendors by year. the complete human kinome includes approximately 520 protein kinases<sup>95</sup> and a smaller number of lipid and small molecule kinases. **b** | Landmark papers in kinase inhibitor selectivity profiling<sup>89-91,93-96,97</sup>, plotted against the number of selectivity measurements (kinases × drugs) that were reported. Representatives from three different approaches that measure inhibitor binding are shown.





**Figure 5. Polypharmacology in the protein kinome**

Pairs of kinases were associated based on kinase selectivity data for a broad range of inhibitor scaffolds<sup>93</sup> and the sequence conservation between aligned sequences of an expanded ATP binding site. Pairs of kinases that were potently inhibited by a common inhibitor (common dissociation constant ( $K_D$ ) < 10 nM) (green lines) were used to determine a sequence similarity cut-off to predict pairs of kinases that can be inhibited by a common inhibitor (red lines). A much higher sequence similarity in the binding pockets was observed for those pairs of kinases with a common potent inhibitor compared with pairs of kinases with a common but less potent inhibitor or no common inhibitor at all. The level of sequence conservation necessary for the predictions was determined based on the sequence conservation distribution of the experimentally determined pairs. The predicted pairs of kinases (red lines) represent potential target combinations that may be more easily accessible with new kinase inhibitors. The image is shown in more detail in supplementary information s1 (figure). AGC, kinases from the protein kinase A, protein kinase G and protein kinase C families; CAMK, calcium/calmodulin-dependent protein kinases; CK1, casein kinase 1; CMGC, kinases from the cyclin-dependent kinase, MAP kinase, glycogen synthase kinase and casein kinase II families; STE, homologues of yeast sterile 7, sterile 11 and sterile 20 kinases; TK, tyrosine kinases; TKL, tyrosine kinase-like kinases.

**Table 1**

## US FDA-approved kinase inhibitors

<b>Drug</b>	<b>Key targets for therapeutic activity</b>	<b>US FDA-approved indication</b>
Imatinib	BCR–ABL, PDGFR and KIT	CML and GIST
Dasatinib	BCR–ABL	CML
Nilotinib	BCR–ABL	CML
Gefitinib	EGFR	Lung cancer
Erlotinib	EGFR	Lung and pancreatic cancers
Lapatinib	EGFR and ERBB2	Breast cancer
Sunitinib	VEGFR2, PDGFR and KIT	Kidney cancer and GIST
Sorafenib	VEGFR2 and PDGFR	Kidney and liver cancers
Pazopanib	VEGFR2, PDGFR and KIT	Kidney cancer
Everolimus	mTOR	Kidney cancer
<b><i>Antibody</i></b>		
Trastuzumab	ERBB2	Breast cancer
Cetuximab	EGFR	Colorectal, and head and neck cancers
Panitumumab	EGFR	Colorectal cancer
Bevacizumab	VEGF	Colorectal, lung and breast cancers

CML, chronic myeloid leukaemia; EGFR, epidermal growth factor receptor; FDA, Food and Drug Administration; GIST, gastrointestinal stromal tumour; PDGFR, platelet-derived growth factor receptor; VEGFR2, vascular endothelial growth factor receptor 2.