

Mitochondrial protease AtFtsH4 protects ageing Arabidopsis rosettes against oxidative damage under short-day photoperiod

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Mitochondrial AtFtsH4 protease, whose catalytic site is exposed to the intermembrane space, is one of four inner membrane-bound FtsH proteases in Arabidopsis. We found that the loss of AtFtsH4 altered Arabidopsis leaf morphology at the late stage of rosette growth under short-day photoperiod, while such changes were not observed in *ftsH4* mutants grown under long days. These morphological changes were correlated with elevated levels of both reactive oxygen species (ROS) and carbonylated proteins, which strongly suggested that ageing *ftsH4* plants experienced oxidative stress. This view was supported by the accumulation of electron-dense material, presumably containing aggregated oxidized proteins, in mitochondria of *ftsH4* plants with the most strongly malformed leaf blades. Taken together, our data published in the May issue of *Plant Journal*¹ suggest a link between the lack of AtFtsH4 protease, oxidative stress and altered leaf morphology at the late rosette stage under short days. Here, we present evidence that the onset of altered leaf morphology in *ftsH4* correlates with an increase in the abundance of AtFtsH4 transcript observed in wild-type Arabidopsis growing under the same conditions. We also discuss how the lack of AtFtsH4 may cause oxidative stress towards the end of the vegetative growth in short days.

In Arabidopsis, four membrane-bound FtsH proteases, members of the ATP-dependent metalloprotease family, reside in mitochondria.² One of them is AtFtsH4 with catalytic sites exposed to

the intermembrane space.³ In yeast and mammals, FtsH proteases control mitochondrial protein quality, act as processing peptidases or serve as chaperones independently of their proteolytic function.⁴⁻⁷ Using two T-DNA null mutants we investigated the role of AtFtsH4 in plant growth and development.¹ We found that the loss of this protease significantly affects morphogenesis of Arabidopsis rosette leaves at the late stage of vegetative growth in short-day conditions (SD, 8 h light per day). In contrast, no morphological consequences of the lack of AtFtsH4 were detected at the early stage of vegetative growth in SD as well as during the entire rosette vegetative growth under long day conditions (LD, 16 h light per day). The most striking features of ageing *ftsH4* grown under SD conditions were a distinct asymmetry and irregular serration of leaf blades. The strength of these developmental disturbances increased towards the end of the vegetative growth and was well correlated with elevated levels of reactive oxygen species (ROS) and carbonylated proteins, which strongly indicated that ageing *ftsH4* plants suffered from oxidative stress. This view was supported by the presence of tiny accumulations of electron-dense material, presumably aggregates of oxidized proteins, in the mitochondrial matrix of *ftsH4* plants with the leaf-blade morphology severely affected. However, no such accumulations were visible in mitochondria of young *ftsH4* plants. In this addendum, we present additional data supporting the role of AtFtsH4 in the late phase of Arabidopsis vegetative growth under short-day conditions. The time-course of the AtFtsH4 transcript accumulation was followed in

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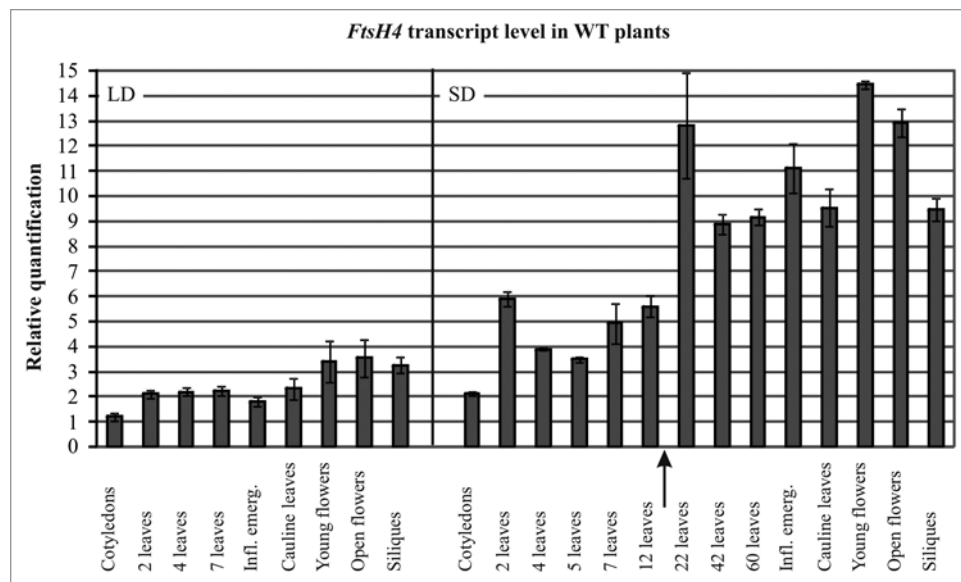


Figure 1. Time-course of *AtFtsH4* transcript level detected by quantitative PCR in wild-type plants under long (LD) and short (SD) day conditions. At the early growth stage the transcript level was measured in cotyledons or two-leaved seedlings. Later, between the fourth leaf stage and inflorescence emergence (Inf.emerg.) the youngest leaf was always sampled. In the generative phase, cauline leaves, buds, open flowers and siliques were tested. The arrow indicates the time when subtle leaf malformations become visible in the *ftsH4* mutant. Error bars indicate SE.

wild-type plants grown under LD and SD conditions using quantitative PCR (Fig. 1). In the LD, *AtFtsH4* transcripts remained almost constant in rosette leaves during the entire vegetative growth. Under SD conditions, the abundance of *AtFtsH4* transcript was almost the same in leaves appearing at the early stage of rosette development but this level turned out to be substantially higher in leaves of ageing *Arabidopsis* plants. Moreover, the similar high level was also observed in all generative organs. Interestingly, the increase of the *AtFtsH4* transcript level in wild-type plants occurs at exactly the same developmental time, when first, however subtle, malformations in leaf-blade morphology become visible in *ftsH4* plants.

Taken together, our data suggest a link between the lack of *AtFtsH4* protease, oxidative stress and altered leaf morphology under certain developmental and environmental conditions. Our results indicate that the loss of *AtFtsH4* leads to the mild deficiency of the mitochondrial oxidative system caused by impaired assembly/stability of respiratory complexes regardless of the growth stage and the day-length.^{1,8} The impaired respiratory activity may result in an increased production of ROS and oxidized proteins with deleterious effects on mitochondrial function. On the

other hand, it is well known that mitochondria have a threshold below which they can accommodate the increased level of ROS and oxidized proteins as by-products of a dysfunctional oxidative system.⁹ We believe that the defense system devoid of the *AtFtsH4* protease is sufficiently efficient only in the short term and in optimal growth conditions (long days, early stage of growth in short days). However, in the SD photoperiod when the vegetative phase is much longer than in LD, plants have to deal with longer dark periods which enhance the role of mitochondria in energy production, which in turn results in increased oxidative parameters in ageing rosettes of the *ftsH4* mutant.

Another possibility that does not exclude the first one is that *AtFtsH4* may play a more specific role at the end of the vegetative growth in addition to the housekeeping function required during the entire *Arabidopsis* life. The *AtFtsH* protease, like its yeast or bacterial homologues, is expected to have numerous substrates and therefore may be involved in multiple molecular pathways preventing the oxidative stress. The increased abundance of *AtFtsH4* transcripts in ageing wild-type rosettes in the SD photoperiod supports the view that *AtFtsH4* may have an important function towards the end

of the vegetative growth. It was reported by Johansson et al.¹⁰ that a carbonylated protein level in *Arabidopsis* rosette leaves increases progressively with age and then declines rapidly at the end of the vegetative growth. Thus, it is possible that one of the putative functions of the *AtFtsH4* protease at this stage of plant development is to prevent accumulation of carbonylated proteins produced by ROS-generated oxidative stress. It has been suggested that protein oxidation leads to a partial loss of its secondary structure without disturbing the overall folding pattern, resulting in flexible regions that serve as targeting signals for degradation.¹¹ In agreement with this view, degradation of oxidized proteins requires enzymes sensing the local protein unfolding. Unlike other ATP-dependent proteases, *FtsH* proteases lack a robust unfoldase activity and use the folding state of their substrates as a criterion for degradation.¹² Thus, the *AtFtsH4* may recognize oxidized proteins and then degrade them or deliver them for degradation mediated by other proteases. In mammals, the Lon protease, an ATP-dependent serine protease, plays a crucial role in the degradation of oxidized proteins.¹³ Likely, the lack of *AtFtsH4* limits the capacity of mitochondrial system controlling the level of oxidized proteins. This defect can be

perceptible mostly under conditions favorable for accumulation of oxidized proteins, as in ageing rosettes of *ftsh4* mutant in SD, causing mitochondrial dysfunctions and, subsequently, impairing cell and leaf development. We believe that the abnormality in leaf formation could be associated with an accumulation of oxidatively damaged proteins in mitochondria of both meristematic and growing leaf cells, which is progressive with the age of *ftsh4* plants. This assumption could explain why the malformation of emerging leaves progressed with every initiated leaf of aging *ftsh4* rosettes. Furthermore, the developmental retardation and the inability of 85% of *ftsh4* plants to enter flowering phase under SD conditions could result from the impairment of the meristem function due to progressive accumulation of oxidatively damaged proteins. However, the exact relations between observed malformations at the organismal level and molecular and physiological reasons laying at their background require further investigations.

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