An Intuitive Safety Factor for Cardiac Propagation

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ABSTRACT Safety factor is a useful concept for analyzing the propagation of impulses through cardiac tissue, which may have compromised ion channel function or electrical connectivity. Several formulations for its calculation have been proposed and have proved useful in one dimension; however, as we demonstrate, recent attempts to use the same formulation in multiple dimensions have led to questionable conclusions. In this study, we mathematically analyze the latest formulation of safety factor and explain its puzzling behavior. We propose a new formulation that is suitable for any dimension and can be estimated from experimental measurements. Its applicability is verified in two-dimensional simulations.

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Cardiac safety factor (SF) is a putative quantifier of the robustness of propagation in heart tissue (1). It quantifies the surplus of current delivered to a cell relative to the amount required to depolarize the membrane to threshold. Leon and Roberge (2) put forth a version based on local cellular excitation. To address several shortcomings, Shaw and Rudy (3) developed a version that was later tweaked (4). This most recent formulation shall be referred to as SF_{R} .

A recent study by Aslanidi et al. (5) developed an excellent new ionic model for Purkinje cells, which was sorely needed as it replaces a pioneering-but-decades-old model (6). It also used $SF_{\rm R}$ to investigate propagation through Purkinje-ventricular (PVJs), which led to several counterintuitive observations:

- 1. SF_{R} was maximal surrounding the PVJ, even though propagation was most likely to fail in that region.
- 2. Tissue boundaries had a low $SF_{\rm R}$, even though cells in that region lost no current to downstream cells.
- 3. $SF_{\rm R}$ decreased with distance from the PVJ, even though the radius of curvature of the propagation wavefront increased, requiring cells to excite fewer downstream cells.
- 4. For Purkinje strand widths above a certain threshold, $SF_{\rm R}$ decreased.

Similarly, in other studies SF_R did not change at points where wavefronts collided or changed in size (4).

Because SF_R was only verified for one dimension (3,7), the question of whether SF_R properly captures the dynamics of successful propagation in higher dimensions remains unanswered. Although the concept of SF is simple enough, its definition may not be so straightforward.

DERIVATION OF SAFETY FACTOR

The equation describing cellular currents is derived from Kirchoff's current law, and is expressed in the monodomain formulation as

$$\nabla \cdot \overline{\sigma}_m \nabla V_m = \beta I_m - I_s, \tag{1}$$

where $\overline{\sigma}_m$ is the harmonic mean of the intracellular and extracellular conductivities, V_m is the transmembrane voltage, β is the cell-surface-area/volume ratio, I_m is the total membrane current (outward positive), and I_s is an intracellular stimulus. Integrating over the volume of a cell, this becomes

$$I_{\rm in} - I_{\rm out} = C_{\rm m} \frac{\partial V_{\rm m}}{\partial t} + I_{\rm ion} - I_{\rm s},$$
 (2)

where $C_{\rm m}$ is the membrane capacitance, $I_{\rm in}$ and $I_{\rm out}$ are the currents entering and leaving a cell through gap junctions, $C_{\rm m}$ is the membrane capacitance, and $I_{\rm ion}$ is the current passing through membrane channel proteins. Currents are expressed as whole cell quantities.

The expression for SF used by Romero et al. (4) and Aslanidi et al. (5) is given by

$$\delta F_{\rm R} = \frac{C_{\rm m} \Delta V_{\rm m} + Q_{\rm out}}{Q_{\rm in}},$$
(3)

where the charge is computed over the time of rise of the membrane voltage ($\Delta V_{\rm m}$). Equation 2 can be integrated over the rise time, and terms rearranged to obtain

$$\frac{C_{\rm m}\Delta V_{\rm m} + Q_{\rm out} + Q_{\rm ion}}{Q_{\rm in}} = 1, \qquad (4)$$

which, combined with Eq. 3, yields

$$SF_{\rm R} = 1 - \frac{Q_{\rm ion}}{Q_{\rm in}}.$$
 (5)

Thus, SF_R is >1 if total ionic charge, which is the active component, is inward. For this condition to occur, the cell must open enough Na⁺ or Ca² channels to counteract

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outward K^+ channels, thereby initiating a positive feedback loop, causing an action potential to fire. Thus, as has been demonstrated (4,7), SF_R must be less than unity when propagation fails, but the question remains whether it relates correctly to the surplus or deficit of charge relative to a threshold value. The relationship is not readily apparent.

Analyzing Eq. 3, the paradoxical behavior of SF_R at the PVJ in Eq. 5 can be explained. Because the input wavefront width at the PVJ is much smaller than the output wavefront width, Q_{in} will be relatively small, increasing the SF_R . Similar reasoning explains the counterintuitive SF_R reductions with increasing wavefront curvature and PVJ width. At the boundaries, a lack of downstream cells to excite means that Q_{out} is zero in the numerator, reducing the SF_R .

NEW FORMULATION FOR SAFETY FACTOR

An intuitive expression for SF must explicitly consider the concept of the surplus of charge delivered relative to the minimum required to trigger an AP. Thus, we propose

$$SF_{\rm VB} = \frac{\frac{1}{\beta} \int \nabla \cdot \overline{\sigma}_m \nabla V_{\rm m} \, dt}{Q_{\rm thr}(t_{\rm A})}; \qquad A \in [t_{1\%}, t_{I_{\rm m}0}], \tag{6}$$

$$=\frac{C_{\rm m}\Delta V_{\rm m}+Q_{\rm ion}-Q_{\rm s}}{Q_{\rm thr}(t_{\rm A})}; \qquad t_{\rm A}=t_{I_{\rm m}0}-t_{1\%}, \quad (7)$$

where A is the interval from 1% take off $(t_{1\%})$ to zero $I_{\rm m}$ $(t_{I_{\rm m}0})$, during which $I_{\rm m}$ is positive, $Q_{\rm s}$ is the intracellular stimulus charge, and $Q_{\rm thr}$ is the minimum charge required to elicit an action potential in a single cell, which is dependent on the stimulus duration $(t_{\rm A})$. This is the period during which depolarizing ionic channels have just begun to open; thus, it approximates the duration of the depolarizing stimulus brought about through spatial interaction.

Unique Q_{thr} functions must be determined for each preexcitation tissue state. These can be determined from computationally inexpensive single-cell simulations, as shown for several ionic models in Fig. 1. These relationships are nearly linear with respect to t_A because the passive membrane acts like a lossy RC circuit; thus, linear continuous functions were fit to several measured values for each scenario. In addition to ionic model dependence, Q_{thr} is sensitive to changes in refractoriness and excitability, as shown for impaired sodium conductance (g_{Na}) and increased pacing. Although this does increase the amount of computation required to calculate SF_{VB} , it is a necessary component because the history of a cell strongly affects its response.

 SF_{VB} was tested using a similar experimental design to the Aslanidi et al. (5) study: a 2-mm-long strand of Purkinje cells was attached to a 2 × 4-mm piece of myocardial tissue (see Fig. 2). A monodomain finite element formulation was used, details of which can be found in Vigmond et al. (8). An edge length of 5 μ m was used for all elements. Ion dynamics were described by the modified Beeler-Reuter model in the

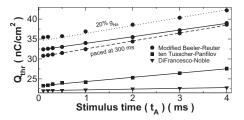


FIGURE 1 Threshold charge as a function of stimulus time for several preexcitation scenarios. Models were paced at 1000 ms with normal excitability except where otherwise indicated. Linear functions (*lines*) were fit to measured values (*points*).

myocardium (9) and the DiFrancesco-Noble model (6) in the Purkinje strand. As in Aslanidi et al. (5), the complex architecture known to exist at PVJs (10) was ignored. In Aslanidi et al. (5), strand width was varied from 500 μ m to 3.5 mm based on fiber bundle diameters (11); here, the range was reduced to 5–1000 μ m to encompass the finer-scale PVJ diameters shown in microscopic analysis (12).

The distribution of $SF_{\rm VB}$ during Purkinje-to-myocardium propagation with a 120- μ m strand is shown in Fig. 2. As expected, $SF_{\rm VB}$ was minimal near the PVJ, where the preparation was most vulnerable to propagation failure, and increased as the wavefront curvature became larger. At the tissue boundary, there was a sharp increase in $SF_{\rm VB}$ due to the lack of downstream cells to be excited.

Detailed numerical values for two points along the myocardial slab are shown in Table 1. SF_{VB} increases with distance from the junction and decreases with impaired g_{Na} .

Results for several other strand widths are presented in Fig. 3, which shows $SF_{\rm VB}$ from strand to myocardium, including the center of the PVJ. In cases where propagation was unsuccessful, $SF_{\rm VB}$ dropped below unity at the PVJ and the failed stimulus decayed exponentially into the surrounding tissue. For the critical strand width of 20 μ m, $SF_{\rm VB}$ dipped similarly at the PVJ but never crossed below unity; as expected, for wider strands, junctional $SF_{\rm VB}$ increased monotonically.

In summary, SF_{VB} has the following features:

1. It describes the surplus of charge delivered to single cells.

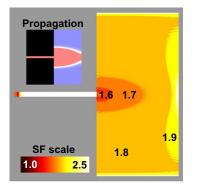


FIGURE 2 Distribution of SF_{VB} . Current was injected in the strand (width: 120 μ m) and excitation spread to the myocardium through a PVJ. (*Inset*) V_m pattern 6 ms after stimulation. Blue, at rest; red, depolarized; white, wavefront.

TABLE 1 SF_{VB} under normal and impaired conduction

	Control		$20\% g_{\rm Na}$		
<i>x</i> (mm)	2.5	3.5	2.5	3.5	
$C_{\rm m}\Delta V_{\rm m} + Q_{\rm ion}$	56.72	59.65	45.35	50.98	
$t_{\rm A} \ ({\rm ms})$	1.12	0.86	2.26	3.82	
$Q_{\rm thr}$	34.11	33.67	38.90	41.98	
SF_{VB}	1.66	1.77	1.17	1.21	

Numerical examples from two points along horizontal axis in the myocardial region, where $Q_s = 0$, for 120- μ m strand with normal and impaired g_{Na} . In both cases, SF_{VB} increases with distance along preparation (*x*). SF_{VB} decreases with impaired g_{Na} due to reduced Q_{ion} and increased Q_{thr} . Units for $C_m\Delta V_m + Q_{ion}$ and Q_{thr} are nC/cm².

- It avoids the decomposition of current into inward and outward components, which is particularly troublesome in threedimensional continuum approaches with unstructured grids.
- 3. With reduced excitability due to compromised inward membrane current, it will decrease due to increased Q_{thr} .
- 4. In tissue monolayers or in planar propagation where there is no depth component, it can be estimated experimentally with Laplacian electrode measurements (see Eq. 6).

 SF_{VB} behaved similarly when the more complex ten Tusscher and Panfilov (13) model was used for myocardial cells.

DISCUSSION

This study highlights how care must be taken when a metric developed in one dimension is applied to higher-order dimensions. Unexpected results may occur, leading to erroneous conclusions. Such is the case for SF_R , which had shown proper behavior in a strand where wavefront curvature effects are not present. An alternative expression, SF_{VB} , is derived here to address shortcomings of previous formulations. It is simple to compute with unstructured grids, and captures the essential features of SF under conditions known to affect propagation. Two-dimensional simulations verified its applicability, although it is suitable for any dimension.

 SF_{VB} is a ratio of the surplus or deficit between the charge available to a given node and the charge required for that node to achieve the depolarization threshold. Thus, if it has a value <1, propagation will fail because the cell will not undergo any active response.

The results presented here differ markedly from those of Aslanidi et al. (5), but agree with intuitive expectations; namely, increasing PVJ width provides more current, leading to faster, more robust propagation, which is reflected by higher SF_{VB} . The PVJ is the most critical point in the propagation path, so it makes sense that SF in that region is nearest to unity.

Other discrepancies between the two studies may be due to differences in numerical methods. The finite element method used herein preserves no-flux boundary conditions regardless of shape, whereas the finite difference approach used in Aslanidi et al. (5) cannot satisfy no-flux conditions properly at corners, which is where the PVJ is located. Also, this study used a finer discretization, which may be important for computing propagation accurately with narrow Purkinje strands.

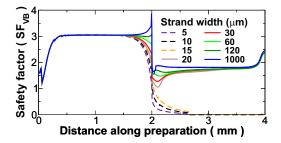


FIGURE 3 SF_{VB} along the horizontal axis, from stimulation site (0 mm) through PVJ (2 mm) to myocardial boundary (4 mm), for several strand widths. In failed propagation (*dashed lines*), SF_{VB} dropped below unity near the PVJ. Junctional SF_{VB} increased monotonically with strand width.

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REFERENCES and FOOTNOTES

- Delgado, C., B. Steinhaus, ..., J. Jalife. 1990. Directional differences in excitability and margin of safety for propagation in sheep ventricular epicardial muscle. *Circ. Res.* 67:97–110.
- Leon, L. J., and F. A. Roberge. 1991. Directional characteristics of action potential propagation in cardiac muscle. A model study. *Circ. Res.* 69:378–395.
- Shaw, R. M., and Y. Rudy. 1997. Ionic mechanisms of propagation in cardiac tissue. Roles of the sodium and L-type calcium currents during reduced excitability and decreased gap junction coupling. *Circ. Res.* 81:727–741.
- Romero, L., B. Trénor, ..., J. M. Ferrero, Jr. 2009. The relative role of refractoriness and source-sink relationship in reentry generation during simulated acute ischemia. *Ann. Biomed. Eng.* 37:1560–1571.
- Aslanidi, O. V., P. Stewart, ..., H. Zhang. 2009. Optimal velocity and safety of discontinuous conduction through the heterogeneous Purkinje-ventricular junction. *Biophys. J.* 97:20–39.
- DiFrancesco, D., and D. Noble. 1985. A model of cardiac electrical activity incorporating ionic pumps and concentration changes. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 307:353–398.
- Wang, Y., and Y. Rudy. 2000. Action potential propagation in inhomogeneous cardiac tissue: safety factor considerations and ionic mechanism. *Am. J. Physiol. Heart Circ. Physiol.* 278:H1019–H1029.
- Vigmond, E. J., R. Weber dos Santos, ..., G. Plank. 2008. Solvers for the cardiac bidomain equations. *Prog. Biophys. Mol. Biol.* 96:3–18.
- Skouibine, K. B., N. A. Trayanova, and P. K. Moore. 1999. Anode/ cathode make and break phenomena in a model of defibrillation. *IEEE Trans. Biomed. Eng.* 46:769–777.
- Tranum-Jensen, J., A. A. Wilde, ..., M. J. Janse. 1991. Morphology of electrophysiologically identified junctions between Purkinje fibers and ventricular muscle in rabbit and pig hearts. *Circ. Res.* 69:429–437.
- Vassalle, M., and C. O. Lee. 1984. The relationship among intracellular sodium activity, calcium, and strophanthidin inotropy in canine cardiac Purkinje fibers. J. Gen. Physiol. 83:287–307.
- Oosthoek, P. W., S. Virágh, ..., A. F. Moorman. 1993. Immunohistochemical delineation of the conduction system. II: The atrioventricular node and Purkinje fibers. *Circ. Res.* 73:482–491.
- ten Tusscher, K. H. W. J., and A. V. Panfilov. 2006. Alternans and spiral breakup in a human ventricular tissue model. *Am. J. Physiol. Heart Circ. Physiol.* 291:H1088–H1100.