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Human bone marrow stromal cells display variable anatomic site-dependent response and recovery from irradiation

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Abstract

Objectives—Orofacial bone is commonly affected by osteoradionecrosis (ORN) during head and neck cancer radiotherapy possibly due to interactions of several factors including radiation damage to resident bone marrow stromal cells (BMSCs). Irradiation causes DNA damage, triggers p53-dependent signaling resulting in either cell-cycle arrest or apoptosis. In some individuals, disproportionately higher rapid growth of orofacial BMSCs relative to those of axial/appendicular bones suggests their response to radiation is skeletally site-specific. We hypothesized that survival and osteogenic recovery capacity of irradiated human BMSCs is site-dependent based on anatomic skeletal site of origin.

Methods—Early passage BMSCs from maxilla, mandible and iliac crest of four normal volunteers were exposed to 2.5 to 10 Gy gamma radiation to evaluate clonogenic survival, effects on cell cycle, DNA damage, p53-related response and *in vivo* osteogenic regenerative capacity.

Results—Orofacial bone marrow stromal cells (OF-MSCs) survived higher radiation doses and recovered quicker than iliac crest (IC-MSCs) based on clonogenic survival, proliferation and accumulation in G₀G₁ phase. Post-irradiation p53 level was relatively unchanged but expression of p21, a downstream effector was moderately increased in OF-MSCs. Re-establishment of *in vivo* bone regeneration was delayed more in irradiated IC-MSCs relative to OF-MSCs.

Conclusions—Effect of irradiation on human BMSCs was skeletal site-specific with OF-MSCs displaying higher radio-resistance and quicker recovery than IC-MSCs.

Keywords

Bone marrow stromal cells; bone regeneration; osteoradionecrosis; ionizing radiation; jaws

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Introduction

Radiotherapy of head and neck cancers often results in jaw osteoradionecrosis (ORN). About 10% of oro-pharyngeal cancer survivors treated with low dose radiation develop ORN, but increases when doses exceeds 60 Gy and irradiated field is subsequently traumatized (1). Pathogenesis of ORN is attributed to radiation-induced hypoxic-hypocellular-hypovascular tissue because osteoblast proliferation, osteocyte number and vascularity are suppressed in irradiated bone (2). Ionizing radiation induces G₂/M cell cycle arrest and triggers a p53-dependent signaling pathway that may induce apoptotic cell death (3). However, this response is variable and cell type-dependent because p53 response causes cell cycle arrest in untransformed cells (4) and apoptosis in transformed cells (5). The dynamic equilibrium between different p53 downstream effectors including p21^{Waf1/Cip1} determines cell cycle progression and cell fate (6).

Specific cellular mechanisms initiating site-specific osteonecrosis are unclear making it challenging to heal osteonecrosis without surgical intervention. While radiation alters cellular survival mechanisms in bone cells (7–9), it is still unclear if bone marrow stromal cells (BMSCs) respond disparately to radiation in a skeletal site-dependent fashion to hinder bone healing. We hypothesized that BMSCs respond to irradiation disparately based on anatomic skeletal site of origin. Using maxilla, mandible and iliac crest BMSCs and therapeutically-relevant radiation doses (10), we evaluated disparity of cell survival, p53-related response and *in vivo* osteogenic regenerative capacity of irradiated BMSCs. We report that in same individuals, orofacial BMSCs were apparently more radio-resistant than those of iliac crest.

Materials and Methods

Isolation and culture of adult bone marrow stromal cells

Bone marrow samples from iliac crest and trabecular bone from maxilla and mandible were harvested from four healthy teenage volunteers according to an Institutional Review Board-approved protocol at the University of Pennsylvania, Philadelphia, PA as previously described (7). Nucleated cells from each sample were cultured to establish primary BMSC using α -modified minimum essential medium (α -MEM; Gibco-Invitrogen, Carlsbad, CA) supplemented with 20% fetal bovine serum (FBS) (Equitech Bio, Kerville, TX), 100 U/ml penicillin, 100 mg/ml streptomycin sulfate and 2 mM glutamine incubated at 37 °C in a humidified atmosphere of 5% CO₂ and air as previously described (7). Primary BMSCs from iliac crest (IC-MSCs) and maxilla and mandible orofacial bones (OF-MSCs) were further expanded or stored in liquid nitrogen until tested. All experiments were performed with early passage 2 to 4 BMSCs from four normal volunteers.

Clonogenic cell survival assay

OF-MSCs and IC-MSCs seeded at 1×10^4 cells/cm² in T-75 flasks were grown to confluence before exposure to 0, 2.5, 5, 7.5 or 10 Gray (Gy) gamma radiation at room temperature using a self-shielded Mark I ¹³⁷Cs irradiator with moving source (J. L. Shepherd, San Fernando, CA) at a rate of 1.25 Gy /minute. Control cells were maintained in the same conditions without irradiation. After trypsinization, cells were re-plated at increasing densities in 6-well plates. At 2 weeks and weekly medium renewals, cells were stained with crystal violet, washed with water and air-dried. Colonies containing ≥ 50 cells were scored microscopically, surviving cells were analyzed using log-linear survival plot and linear quadratic modeling. D₀ was calculated from slope, while α and β were transformed using linear survival expression $-\ln(\text{Surviving fraction}/\text{Dose}) = \alpha + \beta(\text{Dose})$ (11–13).

Cell viability assay

OF-MSCs and IC-MSCs seeded in 96-well plates at 1×10^4 cells/cm² were subjected to 5 Gy radiation based on outcome of clonogenic survival assay above and clinical relevance. Time-dependent viability of irradiated versus non-irradiated cells was assessed at baseline, 6, 24 and 48 hours post-irradiation with WST-1 cell viability assay (Roche Applied Science, Indianapolis IN) at absorbance of 450 nm using a Multiskan MCC automated microplate reader (Thermo Fisher Scientific, Waltham, MA) as previously described (14,15).

Cell cycle analysis

Sub-confluent OF-MSCs and IC-MSCs in T-75 flasks were subjected to 5 Gy ionizing radiation. Cell populations in G₀/G₁, S and G₂/M phases of the cell cycle were analyzed 24 hours post-irradiation as previously described (16,17) using Becton Dickinson FacStar Plus flow cytometer (Becton Dickinson, Franklin Lakes, NJ).

DNA damage by Comet assay

OF-MSCs and IC-MSCs were harvested 30 minutes after 5 Gy irradiation as described above. Non-irradiated cells were used as control to determine baseline endogenous DNA damage and similarly-matched BMSCs treated with 100 μ M H₂O₂ were used as positive control for Comet tail formation. After alkaline unwinding, DNA was separated by alkaline electrophoresis for 30 min at 300 mA, stained with SYBR Green I and assessed with Nikon Eclipse 80i fluorescent microscope (Nikon Instruments, Melville, NJ). Comet tail moment of ≥ 50 randomly selected cells from each group was quantified with Comet Score software (TriTek Corp., Summerduck, VA) and DNA damage was analyzed by logarithmic transformation of comet scores.

Apoptosis

The *In-situ* Cell Death Detection Kit, Fluorescein (Roche Diagnostics, Indianapolis, IN) was used to assess apoptosis in three different experiments following the manufacturer's instructions. Briefly, OF-MSCs and IC-MSCs seeded at 8×10^3 cells/well in 8-well chamber slides (Nalge Nunc Int., Rochester, NY) were exposed to 5 Gy radiation as described above. Control cells received no radiation. Apoptosis was assessed at 24 hours or 3 days post-irradiation. Cells were washed with phosphate buffered saline (PBS pH 7.4) and fixed with 4% paraformaldehyde for 1 hour at room temperature. After permeabilization with 0.1% Triton X-100 in 0.1% sodium citrate buffer, cells were stained with TUNEL reaction mixture (18) and nuclei co-stained with 2 μ g/mL DAPI (4', 6-diamidino-2-pheylindole, Sigma-Aldrich, St. Louis, MO) for 10 minutes at room temperature in the dark. Number of TUNEL-positive cells per total cells was assessed with Nikon Eclipse 80i fluorescent microscope (Nikon Instruments, Melville, NJ).

Radiation-induced changes in p53 level by Western blot

Sub-confluent OF-MSCs and IC-MSCs seeded in duplicates 60 mm dishes at 1.5×10^4 cells/cm² were irradiated at 0, 2.5 or 5 Gy. Total protein extracted 2 hours post-radiation using M-PER Mammalian Protein Extraction Reagent (Pierce Biotechnology, Rockford, IL) was quantified with BCA protein assay (Pierce Biotechnology, Rockford, IL). 15 μ g protein per lane was loaded in 4–12% Bis-Tris gel (Invitrogen, Carlsbad, CA) under reducing conditions. Western blotting was performed as previously described (19). Membranes were probed with 1:200 dilution each of mouse monoclonal anti-p53 (sc-126) and rabbit anti-p21 (sc-397) (all from Santa Cruz Biotechnology, Santa Cruz, CA) followed by incubation with specie-specific secondary antibody: sheep anti-mouse or sheep anti-rabbit conjugated to horse radish peroxidase (GE Healthcare, UK). Monoclonal antibody to human β -actin (ab6276, Abcam, Cambridge, MA) at 1:1000 was used as protein loading control. Immunoreactive blots were

analyzed digitally with Kodak Image Station 4000MM (Molecular Imaging Systems, Carestream Health, Rochester, NY).

Real time-PCR and p53-dependent gene profiling by microarray analysis

Forty eight hours after irradiation of OF-MSCs and IC-MSCs with 0, 2.5 or 5 Gy as above, total mRNA was isolated using Trizol reagent (Invitrogen, Carlsbad, CA) following manufacturer's recommendations. cDNA was prepared from 2 µg of total mRNA using oligo (dT) and SuperScript First-Strand Synthesis System for RT-PCR (Invitrogen, Carlsbad, CA). Real-time PCR reaction was performed in ABI 7300 Real-Time PCR System using Power SYBR Green PCR Master Mix (Applied Biosystems, Foster City, CA) and cDNA-specific primers designed with Primer Express software (Applied Biosystems, Foster City, CA) based on GenBank sequence (human p53, GenBank # NM000546, forward, 5'-CCCAGCCAAAGAAGAAACCA-3', reverse, 5'-CAGCTCTCGGAACATCTCGAA-3'; human p21, GenBank # NM078467, forward, 5'-CCTGTCACGTCTTGTACCCTTGT-3', reverse, 5'-TTTGGAGTGGTAGAAATCTGTCATG-3' and human TATA binding protein (TBP), GenBank # M55654, forward, 5'-GGAGCTGTGATGTGAAGTTTCCTA-3', reverse, 5'-CCAGGAAATAACTCTGGCTCATAAC-3' used as normalizing control). Gene expression levels were normalized to TBP and presented as relative fold change using the $\Delta\Delta C_t$ method (Applied Biosystems, Foster City CA). Using cDNAs prepared as above, RT² Profiler PCR Array System (PAHS-0027, SuperArray Bioscience Corporation, Frederick, MD) was used to assess relative expression profiles of genes associated with p53-dependent signaling by RT-PCR reaction in ABI 7300 Real-Time PCR System (Applied Biosystems, Foster City, CA) (15).

In vivo regenerative capacity of irradiated BMSCs

Institutionally-approved animal protocol at University of Pennsylvania was used to assess *in vivo* bone regeneration of OF-MSCs and IC-MSCs exposed to 2.5 Gy, a sub-lethal dose to allow for transplantation of more abundant surviving cells. From 2 of 4 subjects, 2×10^6 cells attached to 40 mg of spheroidal hydroxyapatite/tricalcium phosphate (HA/TCP, particle size 0.5–1.0 mm, Zimmer, Warsaw, IN) were transplanted into the subcutis of 4-week-old immunocompromised nude female mice (NIH-III-nu, Charles River Laboratories, Wilmington, MA) as previously described (7). Transplants were harvested at 6 and 12 weeks, fixed in 4% paraformaldehyde in PBS (pH 7.4) and decalcified in 10% EDTA/PBS (pH 8.0) before paraffin embedding. Five micron sections were stained with hematoxylin/eosin for histological evaluation and semi-quantitative analysis of hard tissues regenerated as previously described (14,15).

Statistical analysis

Results were expressed as mean \pm standard deviation. Statistical analysis was performed by two-way analysis of variance (ANOVA) followed by post-hoc comparisons with Holm-Sidak test using SigmaStat 3.1 statistical package (Systat Software, Inc., Chicago, IL). Statistical significance was set at $P < 0.05$.

Results

Site-dependent radiation effects on BMSCs survival and cell cycle

Clonogenic survival analysis showed disparate site and dose-dependent responsiveness of BMSCs to irradiation. The iliac crest cells (IC-MSCs) were most sensitive to radiation relative to maxilla and mandible cells (OF-MSCs) ($P < 0.05$). Linear quadratic modeling, and linear transformation of α and β confirmed relative sensitivity of IC-MSCs over OF-MSCs [$D_0 = 1.052$ (iliac crest), -0.758 (maxilla) and -0.926 (mandible); $\bar{n} = 0.4345$ (iliac crest), 1.321

(maxilla) and 4.673 (mandible)]. (Figures 1A – C). OF-MSCs also recovered quicker from radiation effect and proliferated faster than IC-MSCs within 48 hours of 5 Gy irradiation (Figure 1D); a dose selected based on clonogenic survival data and clinical relevance. At baseline, there were more non-irradiated IC-MSCs in G₀G₁ at 81.31% compared to OF-MSCs at 67.57% and 65.44% for maxilla and mandible respectively (Figure 2A, top panel) but irradiation decreased S-phase cells in all cell types with corresponding increases in G₀G₁ phase (Figure 2A, lower panel) suggesting radiation-induced inhibition of S-phase entry. IC-MSCs had fewest S-phase cells and sustained apparently more DNA damage than OFMSCs as shown by Comet assay (Figures 2B) but there were no remarkable apoptotic responses in any of the cells based on either flow cytometry or *In-situ* cell death analysis (data not shown).

Radiation-induced changes in p53 responsiveness of BMSCs

p53 level by Western blot was relatively unchanged post-irradiation irrespective of skeletal site (Figures 2C and D) but the expression of p21^{WAF1/Cip1}, a downstream effector, increased notably in OF-MSCs (Figures 2E). Microarray gene panel included p53-dependent genes implicated in the regulation of apoptosis, growth, proliferation, differentiation and DNA repair. While several genes were strongly over-expressed in OF-MSCs only, there were a few minimally under-expressed genes in both OF-MSCs and IC-MSCs (Table 1). Moreover, some genes that inhibit p53 activity like MDM2, MDM4 and RELA (known as p65) were unchanged in irradiated IC-MSCs but disparately over-expressed in OF-MSCs (Table 1). These complex gene interactions may underlie disparate BMSC responsiveness to irradiation.

Bone formation by transplanted irradiated iliac crest, maxilla and mandible cells

Irradiated OF-MSCs formed more histologically observable bone than IC-MSCs *in vivo*. While *in vivo* bone regeneration by OF-MSCs was observable by 6 weeks, IC-MSCs bone regeneration was further delayed. Interestingly, the three cell types recovered by 12 weeks to form *in vivo* bone quantitatively similar to that regenerated by non-irradiated cells (Figures 3A and B).

Discussion

Unique skeletal site-specific responses of BMSCs to irradiation have not been previously reported. Using BMSC samples from four subjects, we provide new evidence of site-disparity in the response of BMSCs from different skeletal sites to gamma radiation. It is significant to further understand biological responses of bone cells to external insults like radiation and drugs based on clinical indications that osteonecrosis affects craniofacial bones disparately with variable incidence of 3–14% in head and neck cancer patients compared to 0.44% in axial/appendicular bones in gynecological cancers patients (14,20–22). Also, bisphosphonate-associated osteonecrosis occurs exclusively in the jaws apart from an isolated case report in the external auditory canal (22). Within the context of ORN, there are still no studies that addressed skeletal site-dependent cellular and molecular events that occur after irradiation.

Interestingly, despite the higher incidence of ORN in the jaws, we showed for the first time higher radio-resistance of OF-MSCs relative to IC-MSCs in same individuals. This suggests that OF-MSCs survived higher radiation doses and recovered quicker than IC-MSCs, a factor that may be moderating the damaging effects of irradiation in the jaws. p53 is a central element in cellular response to irradiation, and normal cells respond with a rapid but transitory accumulation of p53 protein; an essential process for cell cycle arrest at G₁ checkpoint (23). Irradiated OF-MSCs and IC-MSCs demonstrated similar response by accumulating more G₁-phase cells but OF-MSCs unlike IC-MSCs expressed higher p21^{WAF1/CIP1}, a downstream effector of p53 implicated in G₁ cell cycle arrest and DNA damage repair. This is consistent with the higher recovery capacity that we demonstrated in OF-MSCs and suggests the existence

of repair processes that make OF-MSCs less susceptible to irradiation. When activated, p21^{WAF1/CIP1}, acts as a cyclin-dependent kinase inhibitor to block cell cycle progression and support cell survival (24). Exposure of cells to ionizing radiation causes significant early accumulation of p21^{WAF1/CIP1} while loss of p21^{WAF1/CIP1} in ATM-deficient mice severely sensitized their fibroblasts to radiation (25,26). Previously described association of increased p21^{WAF1/CIP1} with ability to resist radiation-induced apoptosis indicate that radio-resistance of OF-MSCs may result from higher p21^{WAF1/CIP1} level (27).

Irradiated BMSCs in this study apparently did not undergo apoptotic death because *In-situ* apoptotic assay was negative and we have found no significant changes in BAX expression (data not shown), the death promoting gene that serves as a p53-dependent apoptotic target (28). Hence, BMSC-susceptibility to radiation appears to be by cell cycle arrest similar to earlier report that p53 response in untransformed cells usually leads to cell cycle arrest rather than apoptosis (5). This cell-to-cell variability has been described in other studies that showed fibroblasts undergo cell cycle arrest in response to radiation whereas T cells undergo extensive apoptosis (29,30). Although microarray analysis showed more significant increase in expression of other pro-apoptotic genes (31) in OF-MSCs (Table 1), the relative OF-MSC radio-resistance to radiation-related apoptosis may be enhanced by over expression of counter players like HK2 (32) and WT1 (Table 1) that inhibit p53-dependent apoptotic pathway (33). Additionally, radio-resistance of OF-MSCs may be further enhanced by over-expression of MDM2, MDM4 and RELA genes that mutually repress p53 (34). However, the role of each of these p53-related genes needs further clarification.

In vitro radio-resistance of OF-MSCs relative to IC-MSCs was further confirmed by *in vivo* bone regenerative capacity post-irradiation. OF-MSCs formed *in vivo* bone early at 6 weeks while bone formation by irradiated IC-MSCs was further delayed (Figure 3) confirming better OF-MSC post-radiation recovery and superior osteogenic differentiation. Interestingly, IC-MSCs needed more recovery time (12 weeks) to regenerate equally appreciable bone *in vivo*. This pattern of higher *in vivo* osteogenic property of OF-MSCs that has been previously described in non-irradiated BMSCs (7) was maintained despite exposure to irradiation. Also, the three cell types post-irradiation, displayed appreciable *in vivo* osteogenic regenerative capacity similar to non-irradiated cells when given more recovery time of 12 weeks.

While age affects phenotypic and functional characteristics of human hematopoietic and bone marrow stromal progenitor cells (35), age effects were minimized in this study because the tissue samples were obtained from teenagers within a narrow age range. Our study also support previous reports using BMSCs from 4 individuals (7,14,15) to provide additional evidence of skeletal-site specificity of BMSCs as well as site-disparity in survival and recovery of irradiated BMSCs obtained from oral and hip bones. Further elucidation of underlying mechanism is still needed, but this impressive radio-resistance of OF-MSCs may have a modulating effect on ORN severity. Also, recovery capacity of irradiated OF-MSCs may make it easier for osseointegration at donor-recipient interface of osteoradionecrotic lesions treated with autologous bone grafts.

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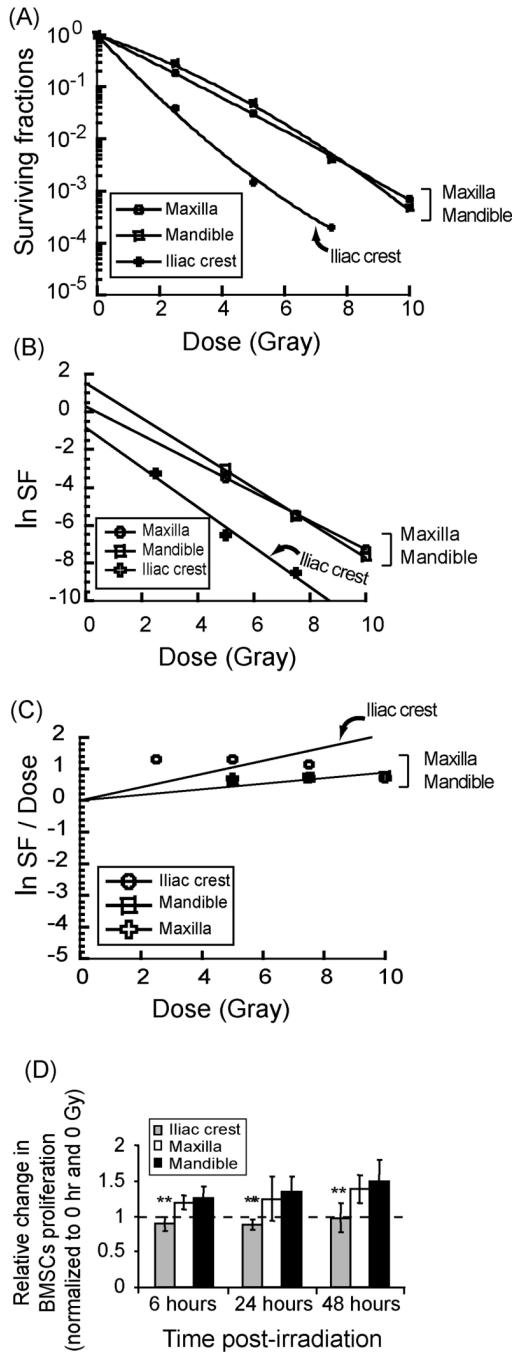


Figure 1.

Disparate site-dependent post-irradiation survival of BMSCs. **A.** Clonogenic radiation survival analysis showed site and dose-dependent radiobiological response of BMSCs from three skeletal sites. Maxilla and mandible BMSCs were more radio-resistant than iliac crest BMSCs ($P < 0.05$, triplicate experiments of $n = 4$ subjects). **B and C.** Data was fitted to linear quadratic equation, D_0 was calculated from slope, while α and β were transformed using linear survival expression $-\ln(\text{Surviving fraction}/\text{Dose}) = \alpha + \beta(\text{Dose})$. [$D_0 = 1.052$ (iliac crest), -0.758 (maxilla) and -0.926 (mandible); $\bar{n} = 0.4345$ (iliac crest), 1.321 (maxilla) and 4.673 (mandible)]. **D.** Maxilla and mandible also recovered quicker than iliac crest BMSCs as early as 6 hours post-irradiation (** = $P < 0.01$).

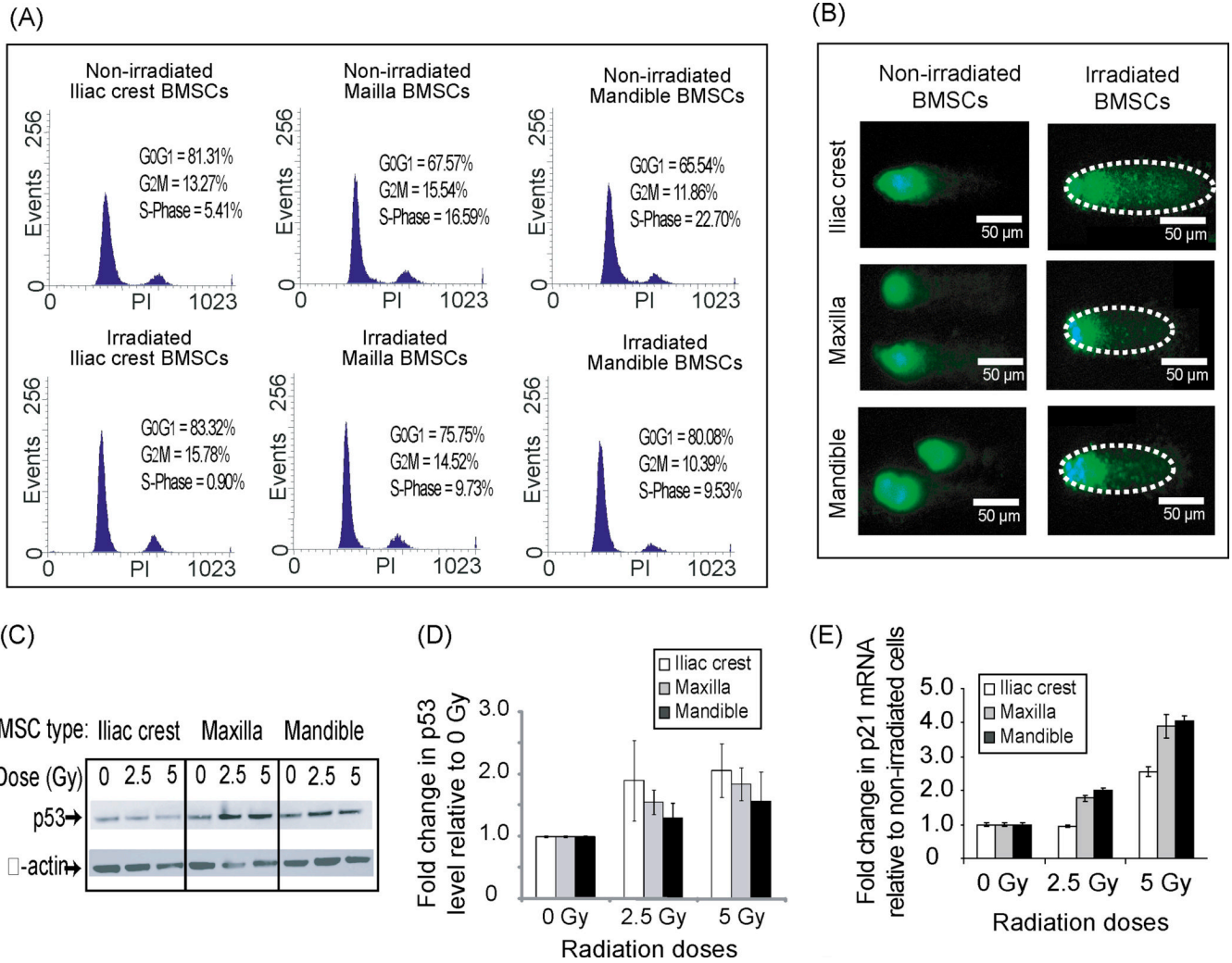


Figure 2. Cell cycle distribution, DNA damage and p53-related changes in irradiated BMSCs. **A.** Cell cycle distribution of non-irradiated and irradiated BMSCs from the three skeletal sites indicate radiation-induced G₀G₁ arrest based on more BMSCs accumulation in G₀G₁ post-irradiation. **B.** Representative images of the comet tails visualized by SYBR Green I staining 30 minutes post-irradiation indicate more DNA damage in iliac crest relative to maxilla and mandible BMSCs (Analysis of relative changes in tail moment of ≥ 50 cells indicate statistically significant differences between IC-MSCs and OF-MSCs, P < 0.01). **C and D.** Representative Western blots and p53 levels based on densitometric analysis from irradiated cells (n=4 subjects) showed disparate p53 levels based on immunoreactivity to mouse anti-human p53 antibody. **E.** The response to radiation based on p21^{Waf1/Cip1} mRNA was higher in maxilla and mandible BMSCs (n=4 subjects) post-irradiation suggesting underlying mechanisms may be downstream of p53 signaling.

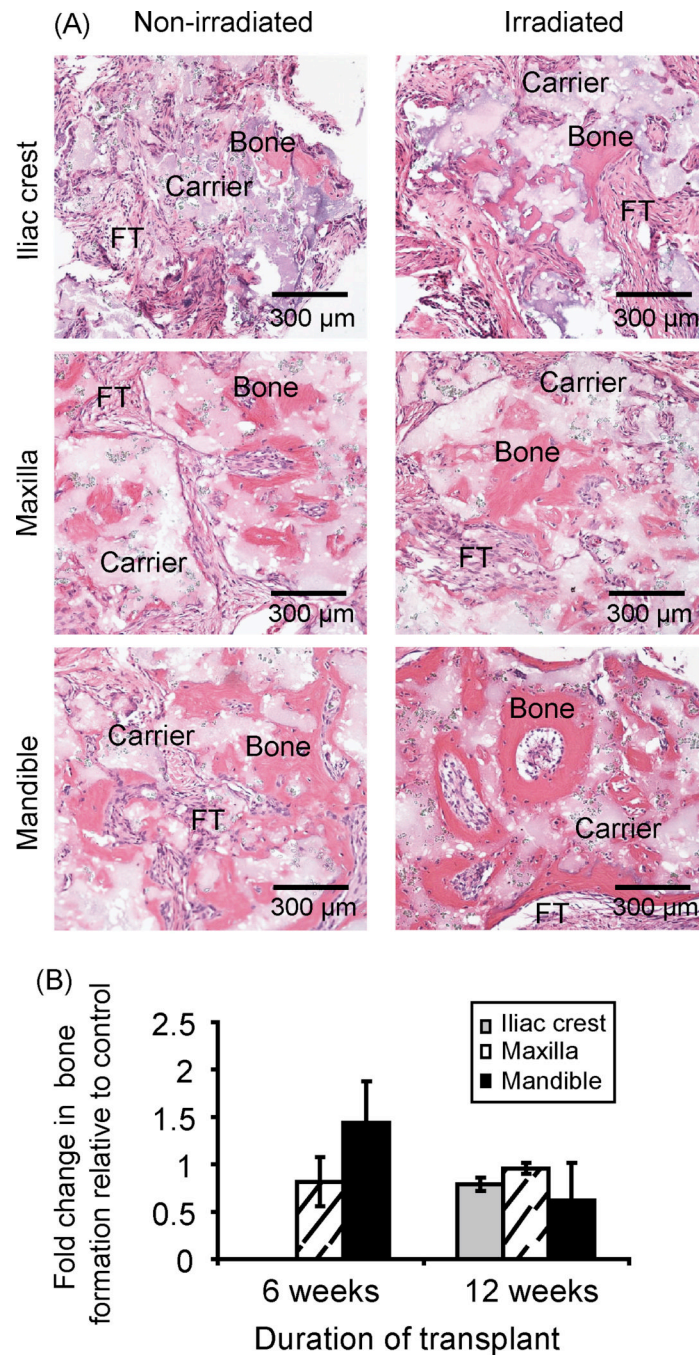


Figure 3. *In vivo* bone formation by irradiated BMSCs. **A.** Hematoxylin/eosin-stained sections *in vivo* bone showed irradiated BMSCs recovered and formed appreciable bone by 12 weeks. **B.** Despite irradiation, 12 weeks allowed the three cell types to recover and form bone quantitatively similar to non-irradiated cells. (Histology sections representatives of BMSCs transplants from n = 4 subjects; FT = fibrous tissue; carrier = hydroxyapatite/tricalcium phosphate).

Table 1
Microarray analysis showing disparate changes in p53-related genes post-BMSCs irradiation

Genes Over-Expressed in Irradiated BMSCs compared with Non-Irradiated Control

Cell Type	Gene Symbol	Gene Name	Fold Change	Group*
	FASLG	Fas ligand (TNF superfamily, member 6)	8.0	<i>a</i>
	GML	GPI anchored molecule like protein	8.4	<i>b</i>
	IFNB1	Interferon, beta 1, fibroblast	8.6	<i>a, b</i>
	LRDD	Leucine-rich repeats and death domain containing	4.9	<i>a</i>
	TNF	Tumor necrosis factor (TNF superfamily, member 2)	6.9	<i>a</i>
	TP73	Tumor protein p73	4.1	<i>b</i>
	WT1	Wilms tumor 1	4.4	<i>d</i>
	ATM	Ataxia telangiectasia mutated	4.0	<i>b</i>
Maxilla / Mandible	CASP2	Caspase 2	4.7	<i>a</i>
	FOXO3	Forkhead box O3	4.3	<i>a</i>
	HK2	Hexokinase 2	7.3	<i>d</i>
	LRDD	Leucine-rich repeats and death domain containing	16.0	<i>a</i>
	MDM2	Mdm2, transformed 3T3 cell double minute 2	5.5	<i>e</i>
	MDM4	Mdm4, transformed 3T3 cell double minute 4	4.5	<i>e</i>
	PCBP4	Poly(rC) binding protein 4	9.9	<i>a, b</i>
	RELA	V-rel re ticuloendotheliosis viral oncogene homolog A	12.0	<i>e</i>
	SESN2	Sestrin 2	13.5	<i>a</i>
	TNFRSF10D	TNF receptor superfamily, member 10d	4.3	<i>a</i>

Genes Under-Expressed in Irradiated BMSCs compared with Non-Irradiated Control

Cell Type	Gene Symbol	Gene Name	Fold Change	Group
Iliac crest	P53AIP1	p53- regulated apoptosis inducing protein 1	0.1	<i>a</i>
	PCAF	P300/CBP-associated factor	0.2	<i>a, b</i>
Maxilla / Mandible	BIRC5	Baculoviral IAP repeat containing 5 (survivin)	0.1	<i>c</i>
	CCNB2	Cyclin B2	0.2	<i>c</i>
	CDC2	Cell division cycle 2, G1 to S and G2 to M	0.2	<i>c</i>
	PRC1	Protein regulator of cytokinesis 1	0.2	<i>c</i>
	PTTG1	Pituitary tumor transforming 1	0.2	<i>e</i>
	CCNB2	Cyclin B2	0.2	<i>c</i>

* Groups:

a pro-apoptotic genes

b genes involved in cell cycle arrest

c genes implicated in progression through cell cycle

d anti-apoptotic genes

^e genes inhibiting p53 transcriptional activity