

## Proposal for a Uniform Nomenclature for Defective Interfering Viruses of Vesicular Stomatitis Virus

M. E. REICHMANN, D. H. L. BISHOP, F. BROWN, J. CRICK, J. J. HOLLAND, C.-Y. KANG, R. LAZZARINI, S. MOYER, J. PERRAULT, L. PREVEC, C. R. PRINGLE, R. R. WAGNER, J. S. YOUNGNER, AND A. S. HUANG†\*

Fourth International Rhabdovirus Colloquium, University of Surrey, Guildford, Surrey, United Kingdom

Defective interfering particles of vesicular stomatitis virus have been named according to their parental derivation and to their genomic length and physical properties. This suggested uniform nomenclature can be adapted for other virus systems.

Defective interfering (DI) particles are found in every major animal virus family (10). Studies of them as deletion mutants are contributing to our understanding of the complex genetic rearrangements and essential sequences required for viral replication and packaging. Biologically, the heterogeneity in viral populations introduced by DI particles appears to affect virus-host interactions and thus, presumably, the outcome of viral diseases in intact animals. As more DI particles are isolated and characterized, it becomes necessary that their nomenclature be such that they can be readily identified and related one to the other.

A proposal for naming individual DI particles is made here. Examples are taken from the rhabdovirus vesicular stomatitis (VS) virus, from which many DI particles have been isolated and studied (23). The proposed principles for nomenclature are as follows.

(i) Name the parent virus and serotype according to the conventions established by the International Committee on Taxonomy of Viruses (2, 6); e.g., VS (Indiana) = VSI, and VS (New Jersey) = VSNJ.

(ii) Give the wild-type, temperature-sensitive (*ts*) mutant, or revertant designation, followed by the strain or isolate designation if known (4), e.g., VSI *ts*<sup>+</sup> ATCC and VSI *ts* G31.

(iii) Give the vernacular name of the DI particle which will be used in the text, and underline it. An example of a vernacular name is DI-T (11). The ideal name would be designated by a fractional number between 0 and 1.0 representing the known total length of the genome of the DI particle, e.g., VSI *ts*<sup>+</sup> ATCC DI 0.38 and VSI *ts* G31 DI 0.10.

(iv) Place other pertinent information within parentheses following this, such as: derivation from which end or from what part of the genome,

percent self-annealing, and any other important phenotype or genotype, e.g., VSI *ts*<sup>+</sup> ATCC DI 0.38 (5', 8%) and VSI *ts* G31 DI 0.10 (5', 65%).

(v) Follow with a reference giving the original isolation and any other references giving the characteristics of the DI particle, e.g., VSI *ts*<sup>+</sup> ATCC DI 0.38 (5', 8%) (13) and VSI *ts* G31 DI 0.10 (5', 65%) (15, 17, 18).

For the sake of uniformity, we recommend that fractional DI particle genomes should be calculated, whenever possible, from the data shown in Table 1 and using the value of  $4 \times 10^6$  as the molecular weight of virion RNA. Most of the data in Table 1 have been summarized previously (23).

This work was supported by a travel grant from the National Institute of Allergy and Infectious Diseases and a grant from the Little Fund, Animal Virus Research Institute, Pirbright, Surrey, United Kingdom.

### LITERATURE CITED

1. Bradish, C. J., J. B. Brooksby, and J. F. Dillion, Jr. 1956. Biophysical studies of the virus system of vesicular stomatitis. *J. Gen. Microbiol.* 14:290-314.
2. Brown, F., D. H. L. Bishop, J. Crick, R. I. B. Francki, J. J. Holland, R. Hull, K. Johnson, G. Martelli, F. A. Murphy, J. F. Obijeski, D. Peters, C. R. Pringle, M. E. Reichmann, L. G. Schneider, R. E. Shope, D. I. H. Simpson, D. F. Summers, and R. R. Wagner. 1979. Rhabdoviridae. *Intervirology* 12:1-7.
3. Brown, F., S. J. Martin, B. Cartwright, and J. Crick. 1967. The ribonucleic acids of the infective and interfering components of vesicular stomatitis virus. *J. Gen. Virol.* 1:479-486.
4. Clewley, J. P., D. H. L. Bishop, C.-Y. Kang, J. Coffin, W. M. Schnitzlein, M. E. Reichmann, and R. E. Shope. 1977. Oligonucleotide fingerprints of RNA species obtained from rhabdoviruses belonging to the vesicular stomatitis virus group. *J. Virol.* 23:152-166.
5. Crick, J., B. Cartwright, and F. Brown. 1966. Interfering components of vesicular stomatitis virus. *Nature (London)* 211:1204-1205.
6. Fenner, F. 1976. Classification and nomenclature of viruses. Second report of the International Committee on Taxonomy of Viruses. *Intervirology* 7:1-115.
7. Hackett, A. J. 1964. A possible morphologic basis for the autointerference phenomenon in vesicular stomatitis virus. *Virology* 24:51-59.

† Address correspondence to: A. S. Huang, Division of Infectious Diseases, Children's Hospital Medical Center, Boston, MA 02115.

TABLE 1. Physical properties of some DI particles of VS virus and of their RNA

Isolate	Particle length (nm)	Particle width (nm)	Sedimentation coefficient (S)			Diffusion coefficient ( $\times 10^{-8}$ cm $^2$ /s)	Mol wt ( $\times 10^{-6}$ )		
			Particle	RNA in gradients	RNA in gels		Particle	RNA in gradients	RNA in gels
<b>Indiana wild types</b>									
Ind C	62 <sup>a</sup>	62 <sup>a</sup>	330 <sup>a</sup>						
Hu	65 <sup>b</sup>	65 <sup>b</sup>		25 <sup>c</sup>	23 <sup>d,e</sup>			1.3 <sup>c</sup>	1.1 <sup>d,e</sup>
How	63 <sup>f</sup>	66 <sup>f</sup>		24 <sup>e</sup>	26 <sup>d,e</sup>			1.2 <sup>h</sup>	1.4 <sup>d,e</sup>
PS	65 <sup>i</sup>		344 <sup>j</sup>		22 <sup>e,k</sup>				1.1 <sup>e,h</sup>
ts <sup>+</sup> G	79 <sup>i</sup>	72 <sup>i</sup>	330 <sup>i</sup>	14 <sup>e,m</sup>	31 <sup>e,k,m</sup>	3.28 <sup>n</sup>	111 <sup>n</sup>	0.4 <sup>d,m</sup>	2.0 <sup>e,k,m</sup>
MS				19 <sup>o</sup>	21 <sup>d,e</sup>			0.8 <sup>h</sup>	0.9 <sup>d,e</sup>
MS (011)				10 <sup>m,o</sup>				0.2 <sup>k,m</sup>	0.8 <sup>o</sup>
PI	65 <sup>p</sup>	65 <sup>p</sup>		19 <sup>o</sup>				0.8 <sup>h</sup>	
<b>Indiana mutants</b>									
HR	100 <sup>i</sup>		422 <sup>i</sup>	30-32 <sup>i</sup>	31 <sup>e,k</sup>			1.7-2.0 <sup>i</sup>	2.1 <sup>e,k</sup>
ts G11	98 <sup>i</sup>	73 <sup>i</sup>	410 <sup>i</sup>	28 <sup>i,m</sup>	29 <sup>e,k,r</sup>	2.85 <sup>n</sup>	127 <sup>n</sup>	1.7 <sup>h,m</sup>	1.8 <sup>e,k,r</sup>
					39 <sup>e,k,m</sup>				3.4 <sup>e,k,m</sup>
ts G12		68 <sup>i</sup>	280 <sup>i</sup>						
ts G22	99 <sup>i</sup>	69 <sup>i</sup>	435 <sup>i</sup>		36 <sup>e,k,m</sup>				2.9 <sup>e,k,m</sup>
ts G31	50 <sup>i</sup>	50 <sup>i</sup>	150 <sup>i</sup>	15 <sup>i</sup>	14 <sup>d,e</sup>	3.95 <sup>n</sup>	42 <sup>n</sup>	9.5 <sup>h</sup>	0.4 <sup>d,e</sup>
Hol			470 <sup>i</sup>	34 <sup>e,k,m</sup>					2.4 <sup>e,k,m</sup>
<b>New Jersey wild types</b>									
Ogden short	82 <sup>i</sup>	65 <sup>i</sup>			20, 24 <sup>e,u,v</sup>				0.9, 1.2 <sup>e,u,v</sup>
Ogden long					29, 31 <sup>e,u,v</sup>				1.8, 2.0 <sup>e,u,v</sup>
Missouri short	98 <sup>w</sup>	74 <sup>w</sup>	393 <sup>w</sup>		20 <sup>e,u</sup>	3.46 <sup>w</sup>	163 <sup>w</sup>		0.9 <sup>e,u</sup>
Missouri long	135 <sup>w</sup>	71.5 <sup>w</sup>	523 <sup>w</sup>		27, 32 <sup>e,u,v</sup>	2.97 <sup>w</sup>	252 <sup>w</sup>		1.6, 2.3 <sup>e,u,v</sup>
Indiana wild-type virion (G)	172 <sup>i</sup>	68 <sup>i</sup>	610 <sup>i</sup>	43 <sup>e</sup>	42 <sup>e,k</sup>	2.32 <sup>n</sup>	290 <sup>n</sup>	4.2 <sup>h</sup>	4.1 <sup>e,k</sup>

<sup>a</sup> Reference 1.<sup>b</sup> Reference 5.<sup>c</sup> Reference 12.<sup>d</sup> Reference 7.<sup>e</sup> Estimated by a comparison with BHK-21 rRNA markers.<sup>f</sup> Reference 16.<sup>g</sup> J. A. Lesnaw, R. N. Leamnon, J. T. Unger, and M. E. Reichmann, unpublished data.<sup>h</sup> Calculated by means of the Spirin equation,  $M = 1550S^{2.1}$ .<sup>i</sup> Reference 19.<sup>j</sup> Calculated from sucrose gradient profiles of reference 19, assuming 610S for the virion marker.<sup>k</sup> Reference 21.<sup>l</sup> Reference 20.<sup>m</sup> Aberrant values because of secondary structure in the molecule.<sup>n</sup> Reference 24.<sup>o</sup> Reference 14.<sup>p</sup> Reference 5.<sup>q</sup> Reference 3.<sup>r</sup> RNAs electrophoresed in 4% polyacrylamide gels and 99% formamide.<sup>s</sup> Reference 9.<sup>t</sup> Reference 7.<sup>u</sup> Reference 22.<sup>v</sup> Two RNA components separated on gels.<sup>w</sup> Reference 8.

8. Hartford, S. L., J. A. Lesnaw, W. H. Flygare, R. MacLeod, and M. E. Reichmann. 1975. Physical properties of New Jersey serotype of vesicular stomatitis virus and its defective particles. Proc. Natl. Acad. Sci. U.S.A. 72:1202-1205.
9. Holland, J. J., and L. P. Villarreal. 1974. Persistent non-cytopathic vesicular stomatitis virus infections mediated by defective T particles that suppress virion transcriptase. Proc. Natl. Acad. Sci. U.S.A. 71:2956-2960.
10. Huang, A. S., and D. Baltimore. 1977. Defective interfering animal viruses, p. 73-116. In H. Fraenkel-Conrat and R. R. Wagner (ed.), Comprehensive virology, vol. 10. Plenum Publishing Corp., New York.
11. Huang, A. S., J. W. Greenawalt, and R. R. Wagner. 1966. Defective T particles of vesicular stomatitis virus.

- I. Preparation, morphology, and some biologic properties. *Virology* 30:161-172.
- 12. Huang, A. S., and R. R. Wagner. 1966. Comparative sedimentation coefficients of RNA extracted from plaque-forming and defective particles of vesicular stomatitis virus. *J. Mol. Biol.* 22:381-384.
  - 13. Kang, C.-Y., T. Glimp, J. P. Clewley, and D. H. L. Bishop. 1978. Studies on the generation of vesicular stomatitis virus (Indiana serotype) defective interfering particles. *Virology* 84:142-152.
  - 14. Lazzarini, R. A., G. H. Weber, L. D. Johnson, and G. M. Stamminger. 1975. Covalently linked message and anti-message (genomic) RNA from a defective vesicular stomatitis virus particle. *J. Mol. Biol.* 97:289-308.
  - 15. Leamnson, R. N., and M. E. Reichmann. 1974. The RNA of defective vesicular stomatitis virus particles in relation to viral cistrons. *J. Mol. Biol.* 85:551-568.
  - 16. Nakai, T., and A. F. Howatson. 1968. The fine structure of vesicular stomatitis virus. *Virology* 35:268-281.
  - 17. Perrault, J., and R. W. Leavitt. 1978. Characterization of snap-back RNAs in vesicular stomatitis defective interfering virus particles. *J. Gen. Virol.* 38:21-34.
  - 18. Perrault, J., and R. W. Leavitt. 1978. Complementary terminal repeat sequences in single-stranded RNAs and snap-back RNAs from vesicular stomatitis defective interfering virus particles. *J. Gen. Virol.* 38:35-50.
  - 19. Petric, M., and L. Prevec. 1970. Vesicular stomatitis virus—a new interfering particle, intracellular structures, and virus-specific RNA. *Virology* 41:615-630.
  - 20. Reichmann, M. E., C. R. Pringle, and E. A. C. Follett. 1971. Defective particles in BHK cells infected with temperature-sensitive mutants of vesicular stomatitis virus. *J. Virol.* 8:154-160.
  - 21. Schnitzlein, W. M., and M. E. Reichmann. 1976. The size and the cistrionic origin of defective vesicular stomatitis virus particle RNAs in relation to homotypic and heterotypic interference. *J. Mol. Biol.* 101:307-325.
  - 22. Schnitzlein, W. M., and M. E. Reichmann. 1977. Interference and RNA homologies of New Jersey serotype isolates of vesicular stomatitis virus and their defective particles. *Virology* 77:490-500.
  - 23. Schnitzlein, W. M., and M. E. Reichmann. 1979. Defective interfering particles of rhabdoviruses. *Curr. Top. Microbiol. Immunol.* 86:123-168.
  - 24. Ware, B. R., T. Raj, W. H. Flygare, J. A. Lesnaw, and M. E. Reichmann. 1973. Molecular weights of vesicular stomatitis virus and its defective particles by laser light-scattering spectroscopy. *J. Virol.* 11:141-145.