Effect of aging on cellular mechanotransduction

Miaozong Wu[1,2,4], **Jacqueline Fannin**[2,3], **Kevin M. Rice**[1,2,4], **Bin Wang**[1,5], and **Eric R. Blough**[1,2,3,4,*]

[1] Cell Differentiation and Development Center, Marshall University, Huntington WV

[2] Department of Biological Sciences, Marshall University, Huntington WV

[3] Department of Pharmacology, Physiology and Toxicology, Marshall University, Joan C. Edwards School of Medicine, Huntington WV

[4] Department of Exercise Science, Sport and Recreation, College of Education and Human Services, Marshall University, Huntington WV

[5] Department of Chemistry, Marshall University, Huntington WV

Abstract

Aging is becoming a critical heath care issue and a burgeoning economic burden on society. Mechanotransduction is the ability of the cell to sense, process, and respond to mechanical stimuli and is an important regulator of physiologic function that has been found to play a role in regulating gene expression, protein synthesis, cell differentiation, tissue growth, and most recently, the pathophysiology of disease. Here we will review some of the recent findings of this field and attempt, where possible, to present changes in mechanotransduction that are associated with the aging process in several selected physiological systems, including musculoskeletal, cardiovascular, neuronal, respiratory systems and skin.

1. Introduction

The aged population now comprises the fastest growing segment of people living in the United States. Living longer is associated with higher morbidity. Indeed, health care spending per capita for the elderly population is more than three and five times higher than that of a working-age person and child, respectively (Hartman et al., 2008). It is thought that mechanical forces are a primary regulator of several biological functions. Indeed, mechanical signals have been shown to mediate the development of a variety of tissues (e.g. skeletal muscle, bone, cartilage, blood vessels and heart), and can affect diverse cellular processes including cell growth, differentiation, cellular migration, gene expression, protein

[*]Author for correspondence: Eric Blough, Ph. D., Laboratory of Molecular Physiology, Room 241N, Robert C. Byrd Biotechnology Science Center, 1700 3rd Ave. Marshall University, Huntington, WV 25755-1090, Phone: 304-696-2708, Fax: 304-696-3766, blough@marshall.edu.

synthesis, and apoptosis (Alenghat and Ingber, 2002; Ingber, 2003a). Given the potential importance that mechanical signaling functions in maintaining cellular homeostasis it is not surprising that changes in mechanotransduction may also play a role in the pathophysiology of disease (Ingber, 2003a). Recent data strongly supports this notion as it is becoming recognized that many aspects of sarcopenia, cardiovascular and respiratory disease may be related to alteration in cellular mechanotransduction (Blough and Linderman, 2000; Rice et al., 2007b; Pardo et al., 2008; Hwee and Bodine, 2009a). The study of age-associated alterations in cellular mechanotransduction has only recently begun to be appreciated for its potential role in mediating cellular function and dysfunction. Here we will investigate the possibility that the ability of the cell to sense, process, and respond to mechanical stimuli is altered with aging and that these changes may be involved in the etiology of aging-associated disease. In an order to focus our discussion, we have chosen to examine how aging may affect mechanotransduction processes in: 1. musculoskeletal system (including skeletal muscle, bone and joint), 2. cardiovascular system, 3. neuronal system, 4. respiratory system, and 5. skin.

2. Mechanotransduction processes

The process of converting an external mechanical force into an internal cellular response is termed mechanotransduction (Wang et al., 1993; Davies, 1995; Alenghat and Ingber, 2002; Jaalouk and Lammerding, 2009). This process consists of three distinct phases 1) signal transduction, 2) signal propagation, and 3) cellular response (Figure 1). Mechanical forces exerted on cell/tissue can result in alterations of cell membrane tension. The integration of external forces into a biochemical signaling event involves four distinct elements: force transmission, mechanosensing, transduction and signal transmission (Figure 1). Force transmission is the result of various molecular components working together to transmit external loading onto or across the cell membrane. Cells in a differentiated, quiescent tissue have several types of external complexes (e.g. integrins, costameres, desmasomes, etc.) that function to position the cell in a specific orientation within the tissue architecture and act to set a "baseline" level of cell tension. This concept is called tensegrity and was first popularized by Ingber and his colleagues at Harvard Medical School during the early 1990s (Ingber, 1991, 1993; Wang et al., 1993). Although beyond the scope of the present review, several excellent reviews on this topic exist (Ingber, 1991, 1993, 1997, 2003b, 2003c, 2008). Changes in this baseline tension result in force transmission to the cellular membrane. Mechanosensing occurs as the cell realizes these external forces. This event sets the stage for the third and fourth elements of the signal transduction phase to occur (Figure 1). These latter processes of transduction and signal transmission are poorly understood, but are likely to include the participation of several proteins and protein complexes including the cytoskeletal elements (e.g. F-actin, intermediate filament, and microtubules), mechanically-activated ion channels, focal adhesions, integrins, adhesion receptors, ATP release, autocrine factor release, and G-protein coupled mechanoreceptors (Ingber, 2006; Geiger et al., 2009; Jaalouk and Lammerding, 2009). Because of the complexity of cell loading stimuli, the diversity of potential cellular transducers involved, and the potential interplay between different elements the initial phase of the mechanotransductive response is extremely complicated and quite difficult to mimic in the laboratory. Similarly, the contextual importance of physical contact between different structural hierarchies makes it nearly impossible to fully represent all of the critical features of the mechanoregulatory response seen in living tissues. As a result of these difficulties, very little research has been done in the area of age-related mechanotransduction. Indeed, a review of the literature demonstrates that a great proportion of the work done in this field has been focused on the signal propagation phase.

3. Effect of aging on skeletal muscle mechanotransduction

3.1. Aging is associated with declines in skeletal muscle mass and function

The age-related loss of skeletal muscle mass and muscle strength, known as sarcopenia, is a health problem that is expected to only increase in coming years as a greater portion of the population lives longer (Castillo et al., 2003; Dirks et al., 2006; Melov et al., 2007). The maintenance of muscle mass and function with increasing age is directly related to quality of life, disease prevention, and has profound socioeconomic significance. Sarcopenia is characterized by a loss of skeletal muscle mass that is greater in proportion than losses in fat-free mass. These alterations typically result in an increase in body fat content (Kyle et al., 2001). It is thought that the magnitude of these changes accelerate with aging (Kyle et al., 2001; Castillo et al., 2003; Lushaj et al., 2008). In humans in their eighth decade of life or older, lean body mass is significantly less than those age 70 to 79 (Kyle et al., 2001), and clinical studies have indicated that the prevalence of sarcopenia dramatically increases by more than 4-fold from ages 70–75 to 85 and older (Castillo et al., 2003). The loss of muscle mass is highly correlated to an age-related decrease in muscle strength. Population-based studies have demonstrated that sarcopenia is associated with decreases in balance, increased muscle weakness and fatigue, an increased incidence of falling and fracture, and a higher prevalence of disability (Castillo et al., 2003). Similar muscle-related changes have been observed in aged animals. For example, recent studies have shown that the Fischer 344/NNIaHSD × Brown Norway/BiNia (F344BN) rat accelerates age-associated decreases in muscle mass, muscle cross-sectional area, myosin and actin expression, and diminished muscle function after 30 months of age (Rice et al., 2005c; Lushaj et al., 2008; Wu et al., 2009b).

3.2. Aging decreases the ability of load-induced growth in skeletal muscle

Mechanical loading and contractile activity play a critical role in the regulation of muscle mass and altered loading can lead to structural remodeling of muscle. Although human and animal studies have demonstrated that aged muscles maintain the ability to undergo hypertrophy, the capacity to sense and subsequently respond to the mechanical stimuli is diminished (Carson et al., 1995; Blough and Linderman, 2000; Pehme et al., 2004; Kosek and Bamman, 2008; Hwee and Bodine, 2009b). In addition it is thought that the aged muscle cell membrane is also more prone to mechanically-induced damage (Rice et al., 2006). Whether these changes are due to alteration in the muscle dystrophin-associated glycoprotein complex (DGC) or change to other structures are currently unknown. Nevertheless, understanding the mechanism(s) of impaired mechanotransduction in aging muscle is likely to shed light on strategies designed to prevent and treat aging-associated skeletal muscle dysfunction. Here we will review the primary intramuscular signaling cascades thought to regulate muscle mass and comment, where possible, on how aging may affect these pathways (Figure 2).

3.3. Aging impairs the ability of skeletal muscle to activate Akt, mammalian target of rapamycin (mTOR) and ribosomal S6 kinase (p70S6k) signaling

Protein kinase B (Akt/PKB) is a serine/threonine protein kinase that plays a central role in integrating anabolic and catabolic responses. The Akt signaling pathway transduces signals emanating from growth factors, nutrients, cytokines and mechanical stimuli via changes in phosphorylation (Bodine et al., 2001; Funai et al., 2006). Activation of Akt stimulates protein synthesis, muscle hypertrophy and cell survival while it antagonizes muscle protein loss (Bodine et al., 2001; Funai et al., 2006; Miyazaki et al., 2008; Hwee and Bodine, 2009b). Akt is highly mechanosensitive, and its activity is thought to be mediated, at least in part, by the phosphatidylinositol-3-kinase (PI3K) (Xiong et al., 2009). It has been demonstrated that muscle loading increases Akt phosphorylation (activation) while

unloading decreases Akt phosphorylation (Miyazaki et al., 2008; Hwee and Bodine, 2009b). Importantly, these alterations in Akt activation have been shown to be highly correlated with the addition (Hwee and Bodine, 2009b) or loss (Miyazaki et al., 2008) of muscle mass. Animal studies have demonstrated that the ability of mechanical loading to induce Akt signaling may become altered with age. For example, Hwee and Funai each demonstrated an age-associated uncoupling of Akt phosphorylation and the amount of Akt substrate phosphorylation following the increased muscle loading (Hwee and Bodine, 2009b) or a bout of electrically stimulated muscle contraction (Funai et al., 2006). These data are consistent with the work from our laboratory showing that aged skeletal muscle exhibits a mismatch between Akt kinase activity and the degree of Akt phosphorylation at Ser473 and Thr308 (Wu et al., 2009b). This impairment in Akt kinase activity is, in turn, associated with diminished insulin responsiveness, as higher levels of Akt phosphorylation (Ser474 and Thr308) were shown to be required to phosphorylate the Akt substrates glycogen synthase kinase (GSK)-3α and GSK3β (Wu et al., 2009b). Given the importance of Akt function in regulating muscle mass, it is likely that these changes in the ability of Akt to elicit downstream signaling could contribute to age-associated decrease in muscle mass. Similar to Akt, the mTOR signaling pathway has been shown to be involved in the regulation of load-induced skeletal muscle growth both *in vitro* and *in vivo* (Parkington et al., 2004; Hornberger and Chien, 2006). Up to a few years ago it was widely suggested that mTOR signaling was dependent, at least in part, on the activation of Akt (Bodine et al., 2001). However, recent studies using transgenic mice expressing a mutated Akt and pharmacological inhibitors specific to PI3K have demonstrated that the mechanical activation of mTOR can occur independently of the PI3K/Akt pathway (Hornberger et al., 2004). It is thought that the mechanical regulation of mTOR is mediated by activation of phospholipase D (PLD) (Hornberger et al., 2006). The regulation of PLD by mechanical stimuli is suggested by its localization at the Z-band (Hornberger et al., 2006), a site within the sarcomere that is particularly suited for the transmission of force (Goldstein et al., 1987). The PLD catalyzes the hydrolysis of phosphatidylcholine to form the lipid second messenger phosphatidic acid (PA) (Hornberger et al., 2006; Hornberger et al., 2007). Attesting to the role of mTOR in muscle growth, studies have demonstrated that mTOR inhibition by rapamycin can directly inhibit the load-induced activation of mTOR signaling and prevent mechanically-induced protein synthesis and muscle growth (Hornberger et al., 2004; Hornberger et al., 2006). In addition, it has also been demonstrated that PA can compete with rapamycin for binding to the FKBP-rapamycin-binding (FRB) domain of mTOR, and hence activate mTOR signaling (Hornberger et al., 2006; Hornberger et al., 2007). Mechanically-activated mTOR can further induce phosphorylation (activation) of ribosomal S6 kinase (p70S6k) (Morris et al., 2004; Parkington et al., 2004; Hornberger et al., 2007). Similar to that observed for Akt, aging has also been shown to negatively impact the ability of skeletal muscle to activate mTOR/p70S6k signaling. Indeed, Morris and others (2004), using a model of hindlimb suspension followed by muscle reloading, have clearly shown that the load-induced phosphorylation of p70S6k is lower in aged animals compared to adults (Morris et al., 2004). Interestingly, several reports have suggested that the basal phosphorylation of p70S6k is increased in older muscle (Parkington et al., 2004; Kinnard et al., 2005) and that this occurs at an age where muscle mass is actually decreasing. Although not clearly defined, this apparent contradiction may imply that p70S6k activity may become uncoupled from downstream 'p70S6k signaling'. The existence of such dysfunction, if present, may contribute to the diminished capacity for mechanical stimuli to induce muscle hypertrophy with aging.

3.4. Load induced mitogen-activated protein kinases signaling may be impaired in aging muscle

The mitogen activated protein kinases (MAPK) are serine/threonine protein kinases that respond to growth factors, environmental stressors and inflammatory cytokines such as TNF-α, IL-1 and IL- 6 (Goodyear et al., 1996; Wretman et al., 2001; Zhan et al., 2007). MAPK signaling is involved in regulating gene expression, cellular metabolism, cell growth, and cell differentiation. In addition, the inappropriate activation of MAPK signaling can trigger inflammation, protein degradation and cellular apoptosis (Wretman et al., 2001; Keren et al., 2006; Wu et al., 2009a). MAPK proteins such as the extracellular signal-regulated kinases (ERK), c-Jun N-terminal kinases (JNK) and p38 MAPK are mechanically sensitive (Goodyear et al., 1996; Aronson et al., 1997; Williamson et al., 2003; Mylabathula et al., 2006). In muscle, the activation of MAPK have been shown to be involved in the regulation of glucose transport, gene expression, and muscle hypertrophy (Sakamoto and Goodyear, 2002; Gibala et al., 2009; Wu et al., 2009a). It is thought that the Ras protein plays an important role in the mechanical regulation of MAPK activation. Mechanical stimuli have been shown to activate Ras, which in turn stimulates the phosphorylation of Raf-1. This event is thought to lead to the activation of MAPK kinase (MEK) cascades resulting in phosphorylation of MAPK proteins (Lange-Carter et al., 1993; Aronson et al., 1997; Barton, 2006). In addition to the direct activation of MAPK proteins by stretch, muscle contraction can also induce MAPK phosphorylation by stimulating the release of the inflammatory cytokine TNF-α (Zhan et al., 2007) or through metabolic alterations such as increased reactive oxygen species (ROS) and acidosis (Wretman et al., 2001).

The effects of aging on the mechanical regulation of MAPK phosphorylation in skeletal muscle (Williamson et al., 2003; Parkington et al., 2004; Hornberger et al., 2005; Mylabathula et al., 2006; Kosek and Bamman, 2008; Ljubicic and Hood, 2009) appear to be MAPK isoform- and fiber type-specific (Table 1). Whether the differences observed between studies is due to differences in the age of the animals studied, experimental protocol or other factors remains to be determined. Nonetheless, compared to that observed in the young/adult animal, the basal phosphorylation of MAPK is higher in aged muscle from both human and animals (Williamson et al., 2003; Mylabathula et al., 2006; Wu et al., 2009a). The increase in MAPK basal phosphorylation appears to be associated with age-associated increases in oxidative stress (Hollander et al., 2000; Ji, 2002; Wu et al., 2009a), as interventions aimed to diminish ROS can effectively normalize age-associated MAPK hyper-phosphorylation (Wu et al., 2009a). It is possible that age-related muscle "stress" may impair the ability of aging skeletal muscle to sense and adapt to mechanical stimuli, resulting in dysregulation of the MAPK signaling pathways. Such changes, if present, could blunt anabolism and limited hypertrophic adaptation (Jozsi et al., 2000; Williamson et al., 2003; Kosek and Bamman, 2008). Additional studies to investigate this possibility will no doubt be useful in increasing our understanding of how aging may affect the ability of mechanical loading to activate MAPK signaling.

3.5. Alteration of mechano growth factor (MGF) and its signaling pathway in aged skeletal muscle

Insulin-like growth factor-I (IGF-I) is an important regulator with anabolic properties for somatic growth and cellular metabolism. In skeletal muscle, at least two different IGF-I isoforms are expressed due to alternative splicing of the primary IGF-I transcript (Owino et al., 2001; Hameed et al., 2003; Cheema et al., 2005; Goldspink, 2007). Of these isoforms, one is quite similar to liver-derived IGF-I (IGF-IEa) and is important for the fusion of myoblasts to form myotubes (Owino et al., 2001; Hameed et al., 2003; Goldspink, 2004; Cheema et al., 2005; Goldspink, 2007). Expression of the other isoform, also known as mechano growth factor (MGF or IGF-IEc), is induced in response to physical activity. It is

thought that MGF is required for secondary myotube formation and the activation of muscle satellite cells for local muscle repair. MGF is also likely to be involved in load-induced muscle hypertrophy (Owino et al., 2001; Hameed et al., 2003; Goldspink, 2004; Cheema et al., 2005; Goldspink, 2007). With aging, the basal level of MGF is decreased (Owino et al., 2001; Hameed et al., 2003). Although aged muscle retains the capacity to induce MGF expression in response to mechanical stimuli, the magnitude of load-induced MFG expression has shown to be diminished by about 50% of that seen in adult animals exposed to a similar degree of overload (Owino et al., 2001). Moreover, aged muscle has also been shown to exhibit a diminished ability to up-regulate expression of the IGF-I receptor following increased loading (Owino et al., 2001). Taken together, these data suggest that alterations in load-induced MGF expression or impairments in the MGF receptor signaling pathway may contribute to the attenuation of muscle growth and repair in aged subjects.

3.6. Myostatin expression is increased in aging muscle

Myostatin is a key negative regulator of the skeletal muscle growth (Grobet et al., 1997; McPherron and Lee, 1997). Myostatin gene inactivation and knockout studies have shown that mutated myostatin protein or reduced myostatin expression are coupled to marked increases in skeletal muscle mass (Grobet et al., 1997; McPherron and Lee, 1997). It is believed that myostatin signaling participates in the regulation of age-associated muscle atrophy as both serum myostatin and muscle myostatin expression are increased with age (Kawada et al., 2001; Yarasheski et al., 2002). These age-associated increases in myostatin are in turn associated with decreases in fat-free mass and diminished muscle mass (Kawada et al., 2001; Yarasheski et al., 2002). The expression of myostatin can be mechanically regulated with increased loading resulting in decreased myostatin expression (Kawada et al., 2001). Interestingly, it has been reported that inhibition of myostatin by PF-354 enhances aerobic performance in aged mice (Lebrasseur et al., 2009). This increase in muscle endurance was associated with decreased phosphorylation/activation of myostatin downstream molecule Smad3 (Lebrasseur et al., 2009) and impaired Smad4 translocation to the nucleus (Joulia-Ekaza and Cabello, 2007). Therefore, manipulation of myostatin signaling may be a useful strategy to ameliorate the aging-related defects in muscle mechanotransduction and muscle growth.

4. Effect of aging on bone mechanotransduction

4.1. Effects of aging on bone structure

Bone health and structure, like muscle, is regulated by mechanical loading. During growth, body weight and muscular forces increase the loading on the growing bone which in turn acts to augment bone strength and mass. In the third decade of life muscle strength generally begins to decrease which is associated with gradual decline in bone loading and bone mass (Sumner and Andriacchi, 1996). In addition to changes in bone loading, non-mechanical factors are also thought to contribute to age-related bone loss. Left unchecked, bone loss can approach decreases of up to 70% by the age of 70 (Frost, 1997).

The factors controlling bone mass are not fully understood. It is thought osteoblasts function to construct bone tissue while osteoclasts maintain mineral homeostasis by reabsorbing bone (Zaidi, 2007). Working together, these two cell types act to maintain the balance between bone deposition and resorption. Nonetheless, as we age the number of osteoblast cells tend to decrease, impairing the natural processes of bone tissue renovation and a reduction in bone mass (D'Ippolito et al., 1999).

4.2. Mechanisms of load-induced signal transduction in bone

Calcium influx is considered to be one of the primary responses to changes in osteoblast and osteocyte loading (Mikuni-Takagaki, 1999; Chen et al., 2000; Ryder and Duncan, 2001; You et al., 2001). Although not well understood, it is thought that Ca^{2+} influx is regulated, at least in part, by fluid flow through the lacuno-canicular system. Attesting to this possibility, fluid flow has been shown to increase intracellular calcium in cultured osteoblastic cells and this response can be suppressed by gadolinium, a stretch-activated calcium channel blocker (Hung et al., 1996). In addition, voltage gated L-type calcium channels have also been show to play a role in bone mechanotransduction as changes in gene expression following calcium influx can be suppressed by incubating the cells with the L-type calcium channel blockers verapamil and nefedipine (Rawlinson et al., 1996; Li et al., 2002b; Li et al., 2003). Mechanical stimulation can also lead to the release of intracellular stores of calcium through a process that appears to be modulated by inositol 1, 4, 5-triphosphate (IP$_3$) (Chen et al., 2000; You et al., 2001). Increased intracellular calcium levels in turn have been linked to the activation of the MAPK signaling pathway and an increased expression of osteopontin and other matrix proteins (You et al., 2001). In addition to the activation of calcium channels, fluid flow and mechanical loading has also been shown to result in the recruitment of integrins to focal adhesions and modulate fluid-flow induced gene expression (Pavalko et al., 1998). How focal adhesions regulate load-induced bone growth is not fully understood.

It is likely that paracrine and autocrine signaling may also contribute to the ability of bone to respond to mechanical signals. For example, recent reports have shown that certain non-steroidal anti-inflammatory drugs can suppress mechanically-induced bone formation by blocking the syntheses of prostaglandins (Forwood, 1996; Chow et al., 1998; Li et al., 2002a). Other data has suggested that skeletal remodeling can also be mediated through parathyroid hormone (PTH), the 1–34 PTH fragment, estrogen, and insulin-like growth factors (Sato et al., 2000; Zaman et al., 2000; Gross et al., 2002; Bakker et al., 2003; Li et al., 2003). To date, the molecular mechanisms that regulate bone mechanotransduction have not been fully elucidated. More research perhaps using cultured cells and molecular approaches will be invaluable to further our understanding how bone "health" is regulated by loading.

4.3. The effects of aging on bone mechanotransduction are poorly understood

In the mature bone, a third basic cell type, the osteocyte begins to emerge. As we age, osteocytes soon become the most abundant cell type. Osteocytes are mechanically sensitive and are able to initiate signaling processes in response to changes in sheer stress and pressure (Knothe Tate et al., 2004). In addition, osteocytes may help to anchor osteoclasts to the extracellular matrix (ECM) by producing osteopontins (Zaidi, 2007). Whether or how aging affects the function of osteoclasts is not well understood. Research directly examining how aging affects the response of bone cells to various stimuli is almost entirely lacking. Donahue *et al* (2001) and Cao *et al* (2007) are amongst a small number of researchers that have examined mechanotransduction in aging bone. Donahue and colleagues demonstrated that aging is associated with diminished calcium influx in osteoblasts following increased loading induced by fluid flow (Donahue et al., 2001). Similarly, Cao and co-workers have demonstrated that aged osteoblasts exhibit a reduced ability for IGF-I to stimulate osteoprogenitor formation (Cao et al., 2007). Further understanding these age-associated changes may provide better insight into developing pharmacological agents for the treatment of age related-bone disease.

5. Effect of aging on joint mechanotransduction

5.1. Effects of aging on joint structure

Joints can be classified into three basic groups based upon how the bones within the joint are connected: fibrous, cartilaginous, and synovial (Benjamin et al., 1995; Khan et al., 2007). It is well accepted that mechanical loading can influence joint structure and function. These alterations can occur during development and aging. For example, the fibrous suture joints in the skull are eventually replaced by bone in early childhood (Mao, 2002). Similarly, increased loading over a lifetime can also result in deleterious changes such as the development of arthritis which is commonly seen during the 5th and 6th decades of life.

5.2. Mechanisms of load-induced signal transduction in joints

Fibrous joints represent a group of joints in which the adjacent bones are joined with fibrous connective tissue (Mao, 2002). These joints can be further divided into three types; sutures, found in the scull, the syndesmosis type which is found between long bones such as the radius and ulna, and the gomphosis type such as that found in the tooth sockets. To our knowledge, little is known regarding the effects of aging on fibrous joint mechanotransduction. Nonetheless, recent data has demonstrated that this joint exhibits mechanosensitivity. For example, the expression of fibroblast growth factor-2 (FGF-2) is increased following the application of tensile stress to the rat coronal suture (Yu et al., 2001). Similarly, osteocalcin, collagen I and alkaline phosphatase gene expression is increased following mechanically induced rat tooth movement (Pavlin et al., 2001). Examining the cranial suture, Ikegame and others showed that increased tensile stresses induced the up-regulation of BMP-4 gene expression and Cbfa1/Osf-2, an osteoblast-specific transcription factor (Ikegame et al., 2001). Like other tissues, the up-regulation of genes and transcription factors in sutures is often accompanied by increased protein synthesis. Indeed, increased suture stress has been found to increase the synthesis of type III collagen (Meikle et al., 1984; Tanaka et al., 2000) and alkaline phosphatase activity (Miyawaki and Forbes, 1987).

Cartilaginous joints represent a group of joints in which the bones are joined with cartilage. Cartilaginous joints are found in the intervertebral discs of the spinal column and in growth regions of the long bones. Although mechanical factors have been posited to be involved in the pathology of the intervertebral disc little information exists regarding the mechanism(s) of mechanotransduction or whether aging affects this process (Hadjipavlou et al., 2008). Likewise, limited research exists regarding how mechanotransduction may take place at the growth plate and if this process may play a role in long bone development and health.

Synovial joints represent a group of joints in which the adjacent bones are not directly joined. These joints consist of an articular surface that is covered with hyaline cartilage (Silver and Bradica, 2002; Pacifici et al., 2005). Because both tensile and compressive forces occur at the joint surface due to translational and rotational motion the cartilage residing within these joints is exposed to a complex pattern of loading (Silver and Glasgold, 1995). As we age, these joints experience a heightened level of injury and tend to be affected by arthritis. Although not well understood it is thought that this disorder may be due, at least in part, to an inability of the chrondrocytes to properly regulate the expression of collagens and proteoglycans (Goldring and Marcu, 2009). In the articular cartilage, anabolic tissue responses appear to be initiated by mechanical loading, however the precise mechanism underlying mechanical modulated matrix synthesis is unknown (Vincent and Saklatvala, 2006). The most favored mechanism of tissue mechanotransduction associated with articular cartilage is integrin ligation, however integrins do not have intrinsic signaling

capability (Millward-Sadler et al., 1999), leaving the exact mechanism of mechanocoupling unknown.

In general, static compression of articulate cartilage has been shown to decrease biosynthetic activity, while cyclic compression appears to increases protein synthesis (Wong et al., 1999). The amount of aggrecan and type II collagen mRNA has been shown to decrease with increasing compression over 24hr, however increases in aggregan and type II collagen mRNA have been reported with dynamic loading (Smith et al., 1996; Ragan et al., 1999; Millward-Sadler et al., 2000). Similarly, fibronectin and cartilage oliogomeric matrix protein have been shown to increase in response to cyclic compression (Steinmeyer et al., 1997; Wong et al., 1999). The signaling pathways regulating these changes are largely unknown however It has been demonstrated that the ERK-MAPK pathway is activated following cyclical loading and that this is most likely due to the release of basic fibroblast growth factor (bFGF) (Vincent et al., 2002; Vincent et al., 2004).

Although it is clear that joint loading is capable of eliciting a wide variety of different cellular responses the mechanisms responsible for these mechanically induced changes are, to date, largely unknown. Similarly, how aging may affect mechanotransductive processes within the different cells and tissues that comprise the joint have for the most part not been investigated. Additional research perhaps using cultured cells, tissue explants studies or molecular approaches will likely be invaluable to further our understanding how joint "health" is regulated by the interplay of loading and age.

6. Aging and cardiovascular mechanotransduction

6.1 Aging affects cardiovascular structure and function

Aging-associated heart diseases are major cause of death, accounting for about 29% of total death in the elderly (Heron et al., 2008). The cardiovascular system is exquisitely sensitive to mechanical stimuli. Indeed, mechanotransduction processes are important in the development of the heart and for the control of blood pressure and cardiac output. Like skeletal muscle and bone, increased cardiac loading can also result in cellular growth and hypertrophy. Diseases of mechanotransduction associated with the heart include angina, atherosclerosis, atrial fibrillation, heart failure, hypertension, intimal hyperplasia, and valvular disease (Ingber, 2003a).

Similar to other aging systems, aging in the heart is associated with a number of structural and functional alterations. The aged mammalian heart is characterized by diminished cardiomyocyte contractility (Harding et al., 1992), prolonged contraction and relaxation (Nair and Nair, 2001), and changes in calcium sensitivity (Lim et al., 2000; Nair and Nair, 2001). Although not well understood, these changes in cardiac function are thought to be mediated, at least in part, by age-associated changes in cardiac structure. Structural changes with aging include an increased deposition of ECM, increased sarcomere length and number, the loss of nuclear shape (Afilalo et al., 2007), lipofuscin accumulation, and cardiomyocyte hypertrophy (Terman and Brunk, 1998; Brunk and Terman, 2002; Nadal-Ginard et al., 2003; Terman and Brunk, 2005). It is also likely that cardiomyocytes number is decreased with aging (Bernhard and Laufer, 2008). The factor(s) responsible for these changes have not been fully elucidated but may include increased oxidative stress, reduced cellular repair, telomere shortening, altered cellular metabolism, the accumulation of post-translational modifications, increased apoptosis or necrosis, and inflammation (Bernhard and Laufer, 2008).

Aging in the vascular system is characterized by aging vessel elongation, torsion, enlargement of the lumen, and a thickening of the vascular walls (Ferrari et al., 2003). Under

a normal lifetime of operation, the pulsatile nature of blood flow causes the vasculature to undergo a tremendous number of cyclic expansions and contractions. Given the tempestuous demands placed on the vessel walls, the contractile-elastic unit, elastin, begins to fatigue by the sixth decade of life (Franklin, 2006). Eventually, this fatigue is counteracted by changes in the ECM which may include the proliferation of collagen and calcium deposition (Franklin, 2006). How changes in the vessel ECM may affect the ability of the vasculature to respond to mechanical stimuli is not well understood.

6.2. Age-related changes to the ECM are associated with increased cardiac stiffness

The ECM acts to support and align cardiomyocytes and is involved in transmitting mechanical forces (Burgess et al., 2001). The composition of the ECM is dynamic and can change in response to environmental and mechanical stimuli. There is also evidence that the ECM changes with age. For example, the amount of fibronectin mRNA has been shown to be regulated in a developmental and age-specific manner (Mamuya et al., 1992). Boluyt and colleagues using captopril, demonstrated that fibronectin expression is increased in the senescent spontaneous hypertensive rat heart (Boluyt and Bing, 2000). Similarly, as the heart ages, collagen expression has also been shown to increase (Li et al., 1992; Orlandi et al., 2004; Lieber et al., 2008). It is thought that these changes in fibronectin and collagen may be associated with increased diastolic stiffness (Burgess et al., 2001; Merx et al., 2005). Aging has also been shown to be associated with increases protein cross-linking (Spinale et al., 1996), increased protein glycation and alterations in the expression of matrix metalloproteinases (MMPs). The MMPs are zinc-dependent endopeptidases that degrade proteins in the extracellular matrix and are inhibited by tissue inhibitor of metalloproteinases (TIMPs) (Malemud, 2006). In aged mouse, recent data has suggested that aging is associated with an increase in the amount of MMP-3, MMP-8, MMP-9, MMP-12, and MMP-14 and decreased levels of TIMP-3 and TIMP-4 (Lindsey et al., 2005). Changes of this type would be expected to favor ECM accumulation which may be associated with decreased left ventricular diastolic function (Bonnema et al., 2007). Whether these changes alone or others working in concert with them are responsible to age-related changes in cardiac stiffness has yet to be fully elucidated.

6.3. Aging can alter the expression of cardiac mechanosensory molecules

The sarcomere is the basic contractile unit of the cardiomyocyte. Integrins are found in the sarcolemmal membrane next to the costameres (Sussman et al., 2002). Similar to skeletal muscle, integrins are thought to play a key role in regulating cardiac mechanotransduction (Ingber, 2003a). With advancing age, research has shown that the expression of integrin subunits in the heart is altered (Burgess et al., 2001). Whether these changes in integrin subunit expression lead to age-related changes in cardiac mechanotransduction has not, to our knowledge, been investigated. In addition to the integrins, the Z-discs of the sarcomere are considered to be mechanosensors (Goldstein et al., 1988, 1989). The Z-disc is linked to the sarcolemma by dystrophin (Pyle and Solaro, 2004). The organization and localization of dystrophin and other dystrophin associated proteins, such as utrophin, vinculin and talin, have been shown to be altered with aging (Mora et al., 1996). Although not well understood, it is possible that such changes could lead to change in cardiac stiffness or to load-induced signaling. Similarly, desmin is another protein that functions to connect different regions of the saromere. Whether or how aging affects desmin expression is unclear as different studies have shown to decrease (Diedrich et al., 2007; Lieber et al., 2008) or remain unaltered with aging (Diedrich et al., 2007). Like desmin, titin has also been implicated in sensing mechanical loading (Pyle and Solaro, 2004). Titin functions to connect the thick filament of the sarcomere to the Z-disc and is responsible for the passive tension of the cardiomyocytes (Pyle and Solaro, 2004). In addition to its structural role, titin also contains binding sites for other proteins such as telethonin, alpha actinin, calpain-3, obscurin, myosin-binding protein

C, calmodulin 1, CAPN3, and MURF1. Titin isoforms have been shown to change during heart development (Warren et al., 2004). How changes in the Z-disc, dystrophin, desmin or titin might affect cardiac mechanotransduction during aging is not yet clear.

Stretch-activated channels are activated by forces acting upon the ECM, integral membrane proteins, cytoskeleton, or the membrane itself. The impact of aging on stretch-activated channels and their signaling pathways in the heart is not well understood. Recent data has suggested that senescent mouse myocytes exhibit impaired calcium handling with increasing pacing frequency (Lim et al., 2000). There is also evidence to suggest the molecules involved in calcium signaling may change with age. For example, the calmodulin-dependent protein kinase II, which is involved in Ca^{2+} uptake into the sarcoplasmic reticulum, was significantly increased in ventricular extracts from old mice (Kim et al., 1998). Conversely, ATP-dependent Ca^{2+}-uptake activity of the sarcoplasmic reticulum and the relative amount of CaM kinase II was reduced significantly with aging (Xu and Narayanan, 1998). Likewise, the sarcoplasmic/endoplasmic reticulum Ca^{2+} ATPase (SERCA) gene which is down-regulated in several models of cardiac hypertrophy also exhibits lower levels of mRNA in the senescent heart. How these changes may affect the ability of the aging heart to sense and respond to mechanical stimulation is not clear.

6.4. The effect of aging on vascular endothelial cells

The blood vessels of the vascular system are lined with endothelial cells (EC). The prominent mediator of EC function is fluid shear stress. Increases in blood flow stimulate EC in the arteries and this can induce relaxation of the surrounding vascular smooth muscle cells (VSMC). This process occurs as a result of hyperpolarization of VSMC, and is mediated through the production and release of nitric oxide (NO), arachidonic acid metabolites, and/or prostacyclin (Vanhoutte et al., 1995; Bittar et al., 2006; Campbell and Falck, 2007). Although we do not yet understand how the response to shear stress is orchestrated in EC, integrins, vascular endothelial (VE)-cadherin, platelet/endothelial cell-adhesion molecule 1 (PECAM1), G-proteins, receptor tyrosine kinases, integrins, and ion channels have all been identified as potential mechanical sensors (Li et al., 2005; Orr et al., 2006). Similar to other cell types, the cytoskeleton is also thought to play a role as the microtubules, actin, and intermediate filaments of the EC can act to transmit forces throughout the cell cytoskeleton (Davies, 1995). Likewise, there is also evidence to suggest that EC contain mechanosensitive ion channels that can be activated by shear force (Hoger et al., 2002) resulting in membrane hyperpolarization through the inward flux of potassium ion (Zaritsky et al., 2000). In general, aging in the vasculature is associated with loss of endothelial function as well as disruption of the endothelia lining. How aging may alter EC mechanotransduction has not fully investigated.

6.5. The effect of aging on vascular smooth muscle mechanotransductive signaling

In addition to EC, VSMC are also capable of sensing and responding to mechanical loading. Recent data using multi-axial stretch and uni-axial stretch (Figure 3A and 3B) has demonstrated that aging is associated with alterations in how the aorta respond to mechanical loading (Rice et al., 2005a; Rice et al., 2005b; Rice et al., 2007a; Rice et al., 2008). For example, pressure-induced Fak-related non-kinase (FRNK p41) translocation increased with aging while RhoA translocation appeared to be unaffected (Rice et al., 2007a). In adult F344BN aorta, both increased aortic intraluminal pressure and uni-axial stretch caused increased activation of the ERK1/2, p38 MAPK, and JNK proteins (Rice et al., 2005a; Rice et al., 2007b). In contrast, aging diminished the response of p38 MAPK and JNK to increased aortic pressure but did not affect the response of the aorta to uni-axial stretch (Rice et al., 2005a; Rice et al., 2007b). With aging the contents of Akt, protein tyrosine phosphatase (SHP-2), and phosphatase and tensin homolog (PTEN) were

significantly increased, while those of p70S6k and GSK3β were unchanged (Rice et al., 2005b). The pressure-induced phosphorylation of p70S6k in aged F344BN aorta was impaired compared to that observed in the adult aorta (Rice et al., 2005b). Together, these results suggest that VSMC can distinguish between different stretch types (multi-axial vs. uni-axial) and that the signaling response is altered with age. In addition, aging in the aorta of the F344BN has been shown to result in diminished contractile force generation and increased calcium-dependent stress relaxation (Blough et al., 2007). The reasons for differences with aging are not clear but may be related to increases in vascular stiffness (Blough et al., 2007). Indeed, an increase in vascular stiffness would be predicted to decrease the vessel deformation with loading. This decrease in deformation, in turn, may act to reduce mechanically-regulated signaling. In addition to changes in vascular stiffness, it is also possible that changes in the membrane composition may play a role in age-associated vasacular mechanotransduction. For example, high plasma membrane cholesterol levels have been shown to inhibit the function of stretch-activated ion channels (Fang et al., 2005). Similarly, age-associated reductions in the number of ion channel subunits or diminished nitric oxide synthase (NOS) activity and nitric oxide release could also play a role. Whether changes in vessel stiffness, alterations in membrane composition, or other factors are responsible for the attenuation of load-induced signaling with aging will require further investigation.

7. Effect of aging on neuronal mechanotransduction

7.1. Aging brain and Alzheimer's disease

The aging brain is characterized by a shrinkage of brain mass, the degeneration of synaptic transmission, a loss of neurons, and a decrease in the abundance of chemical messengers (Dickstein et al., 2007). These age-associated alterations, if localized to the hippocampus and cerebral cortex, can result in memory impairment and the decline of cognitive function. Alzheimer's disease (AD) is the most common progressive neurodegenerative disease, and is one of top five causes of death in the elderly (Heron et al., 2008). In Alzheimer's disease, plaques of aggregated amyloid β peptide (Aβ) are formed around nerve cells (Li et al., 2004; Sun et al., 2006; Tesco et al., 2007). It is thought that this accumulation of Aβ interrupts synaptic transmission and alters synaptic plasticity. In addition, aging-related increase in the hyperphosphorylation of Tau proteins may be associated with an increase the formation of neurofibrillary tangles (Olesen, 1994; Deutsch et al., 2006). Currently, the link between neuronal mechanotransduction and brain aging/neurodegenerative disease is not well understood.

7.2. Dysregulation of ion channel/calcium homeostasis in Alzheimer's disease development

Mechanosensitive cation channels, including stretch-activated cation channels (SACCs) and several transient receptor potential channels (TRPC), have been characterized both in sensory neurons and in non-specialized neurons (Erxleben, 1989; Jia et al., 2007). Mechanosensitive cation channels that conduct Ca^{2+} are present in neuronal dendrites, soma, and at the synaptic terminal. It is thought that these channels regulate neurite outgrowth, nerve growth cone motility, neurotransmitter release, synaptic plasticity, and in some cases, gene expression (Raza et al., 2007; Thibault et al., 2007; Bojarski et al., 2008; Wojda et al., 2008). Intracellular Ca^{2+} concentrations are modulated by Ca^{2+} influx from the extracellular environment via different types of channels, or by Ca^{2+} release from internal stores. Recent studies have demonstrated that aging increases the activity of voltage-gated Ca^{2+} channels in hippocampal neurons, and is associated with an elevation in intracellular Ca^{2+} concentration (Campbell et al., 1996; Landfield, 1996; Thibault and Landfield, 1996). Calcium influx through SACCs inhibits neurite outgrowth, whereas Ca^{2+} influx through

other channels in the plasma membrane (e.g., TRPC) or release from internal stores through inositol-1,4,5-triphosphate receptor (IP$_3$R) and ryanodine receptor (RyR) stimulation promotes neurite extension and steering (Jacques-Fricke et al., 2006). Although not well understood, it is thought that the dysregulation of intracellular Ca^{2+} levels is linked to the Aβ aggregation and changes in Tau phosphorylation (Pierrot et al., 2006; Bojarski et al., 2008).

7.3. Insulin resistance and Alzheimer's disease

Insulin signaling is essential for neuronal development, memory formation, synaptic plasticity, and neuroprotection (Cole and Frautschy, 2007; Li and Holscher, 2007; Zhao and Townsend, 2009). It has been reported that the insulin signaling pathway is disturbed with aging and AD. For example, the expression of insulin and insulin receptor are reduced in AD and aging, with a greater decrease occurring in AD (Frolich et al., 1998; Hoyer, 1998). There is a dramatic decrease in the phosphorylation of Src homology adaptor protein (Shc), the association of Shc/growth factor receptor binding protein-2 (GRB2), and the expression of SH protein tyrosine phosphatase-2 (SHP2) in the forebrain cortex and the cerebellum of aged rats (Fernandes et al., 2001). This impairment in the Shc/GRB2 signaling may cause reduced synthesis of insulin-degrading enzyme (IDE), a protein responsible for the clearance of Aβ fibril, and this may be one mechanism why aging is oftentimes associated with an accumulation of Aβ (Figure 4). In addition, there is evidence that aging may reduce activation of the PI3K/Akt pathway which may be associated with a decreased ability of the aged brain to inhibit apoptosis (Eldar-Finkelman et al., 1999; Schubert et al., 2003). This alteration may also lead to decreased inhibition of the downstream kinase GSK-3 and increases in the Tau hyperphosphorylation which have been associated with increases in Aβ (Figure 4) (Phiel et al., 2003; Rickle et al., 2004). Additional studies using gene knockout or other molecular approaches to investigate insulin signaling may lead to new knowledge regarding how aging affects brain structure and function.

8. Effect of aging on respiratory mechanotransduction

8.1. Age-associated changes in respiratory structure and function

Respiratory diseases cause approximately 6% of total deaths in people 65 years of age and older (Heron et al., 2008). Almost all components of the respiratory system, including lung parenchyma, pulmonary and bronchial vascular systems, and diaphragm, are affected by aging. As we age, the lung is thought to become less elastic, and muscles of the chest wall and diaphragm lose mass and strength (Mays et al., 1989). Therefore, the respiratory system experiences an increase in dead space and a decrease in arterial oxygen tension/ concentration. Aging airways have also been shown to close more readily and tend to collapse, while the aging diaphragm is more susceptible to injury and atrophy (Cantillon and Bradford, 2000; Suzuki et al., 2009). As a result, the aged exhibit an increased possibility of developing respiratory failure, pneumonia, asthma, fibrosis, chronic bronchitis, chronic obstructive pulmonary disease (COPD), emphysema, aspiration, influenza, tuberculosis, and pleurisy (Rossi et al., 1996).

8.2. The effects of aging on the respiratory system in response to mechanical loading

Cells in the respiratory system are subjected to a variety of mechanical forces, including tensile and compressive forces, which can influence airway and alveolar epithelial cell function during the respiratory cycle. Aging is thought to negatively affect the ability of the respiratory system to respond to mechanical loading as the aging lung is more susceptible to pulmonary injury following mechanical ventilation (Nin et al., 2008). Aging also impairs the sensitivity and reactivity of airway smooth muscle to mechanical stimuli (An et al., 2007; Fabry and Fredberg, 2007). Likewise, the aging diaphragm not only exhibits a lower

maximal isometric tension, but also an impaired ability to recover diaphragmatic force in response to unloading (Criswell et al., 2003). How these changes in the respiratory system respond to mechanical stimuli may affect load-induced signaling has to our knowledge not been investigated.

8.3 The effects of calcium channels on respiratory mechanotransduction

It is well accepted that the lung is sensitive to mechanical stimuli. For example, stretch can activate mechanosensitive cation channels to allow Ca^{2+} influx into lung cells (Bialecki et al., 1992; Winston et al., 1993; Boitano et al., 1994). This increase in cytoplasmic Ca^{2+} is associated with the activation of protein tyrosine kinase, which in turn activates phospholipase C-β and mediates the production of both IP_3 and diacylglycerol (DAG). The IP_3 induces Ca^{2+} release from intracellular stores, and along with DAG activates protein kinase C (PKC). PKC further activates transcriptional factors such as *c-fos*, which can bind to stretch response elements (SRE), shear stress-response elements (SSRE) or the shear 12-O-tetradecanoylphorbol 13-acetate-response element (TRE) to induce changes in gene expression (Resnick et al., 1993; Shyy et al., 1995; Garcia et al., 2006). How aging affects loading-induced Ca^{2+}-dependent signaling in the lung is not clear, although it has been postulated that this process is negatively affected by increasing age (Papazafiri and Kletsas, 2003). Physical forces exerted on the smooth muscle of airways can induce oscillation of the cytosolic Ca^{2+} concentration, and this plays a key role in excitation-relaxation coupling. The balance between the phosphorylation of the myosin light chain (MLC) by Ca^{2+}-activated calmodulin-dependent myosin light chain kinase (MLCK) and dephosphorylation of the MLC by myosin light chain phosphatase (MLCP) controls smooth muscle contraction and relaxation in the airway (Smith et al., 1995; Mehta et al., 2000; Totsukawa et al., 2000; Burridge and Doughman, 2006; Janssen and Killian, 2006; Fajmut and Brumen, 2008). Aging induces abnormal oscillation of the intracellular calcium concentration and dysregulation of the balance between MLCK and MLCP activities, which may be a cause of airway hypersensitivity and hyperreactivity (Smith et al., 1995; Ammit et al., 2000; Chitano et al., 2000; Mehta et al., 2000; Totsukawa et al., 2000; Parameswaran et al., 2002; Fajmut and Brumen, 2008).

Contraction-induced ROS generation in the diaphragm has been shown to be related to extracellular calcium concentration and is likely regulated in part by ROS production (Supinski et al., 1999). The oxidative challenge has also been linked to the impaired contractile function (Lawler et al., 1997b). These changes may be related to age-associated increases in diaphragmatic fatigability (Khawli and Reid, 1994; Lawler et al., 1997a). Further studies of the effects of aging on respiratory mechanotransduction may provide valuable insight regarding potential therapeutic treatments for age-associated respiratory dysfunction.

9. Effect of aging on skin mechanotransduction

9.1. Age-associated changes in skin structure and function

Although skin is a durable system, its structural character and physiological function undergo dramatic changes with age (Waller and Maibach, 2005; Farage et al., 2007; Puizina-Ivic, 2008; Robert et al., 2009). The decrease in epidermal thickness is accelerated with age, especially in exposed areas such as the face, neck, forearms, and hands, due to a slower renewal rate of epidermal cells (Boss and Seegmiller, 1981). Aged epidermis contains fewer melanocytes and immunocompetent Langerhans cells, resulting in uneven pigmentation and impaired immunity in aged skin (Waller and Maibach, 2005; Farage et al., 2007). The thickness of the dermis also decreases with age and this is accompanied by a decrease in number of mast cells and fibroblasts, and a decrease in the generation of

collagen, elastin, glycosaminoglycans, and hyaluronic acid all of which are thought to contribute to the development of wrinkles and a loss of elasticity (Duncan and Leffell, 1997; Carrino et al., 2003; Sudel et al., 2005; Waller and Maibach, 2005). Aged skin is drier than younger skin due to lack of sebaceous glands in the dermal layer (McCullough and Kelly, 2006; Farage et al., 2007). The dermal-epidermal junction flattens with age resulting in a decrease in the transfer of nutrients and oxygen between the two layers, making the skin more fragile and susceptible to damage (Sudel et al., 2005; Farage et al., 2007). The subcutaneous fat in the hypodermis (the skin layer below the dermis) diminishes with age and causes the skin to wrinkle and sag (Hashizume, 2004; Farage et al., 2007). Taken together, aged skin has a reduced capability to repair and proliferate, is more susceptible to sunburn, trauma, and bruising, and has diminished perception of external stimuli such as pressure and temperature changes (Puizina-Ivic, 2008; Robert et al., 2009).

9.2. Age-related changes in mechanical properties of skin tissue

Type I collagen is the most abundant protein in skin, and constitutes the extracellular matrix of the dermis (Varani et al., 2006; Fisher et al., 2009). In normal skin, fibroblasts bind to intact collagen fibrils through integrin receptors resulting in the formation of an integrated structure that exhibits a basal level of load or tension within the dermal layer (Wang and Ingber, 1994; Grinnell, 2003). As we age, it is thought that the skin contains an increased amount of fragmented and disorganized collagen fibrils which in turn acts to dramatically reduce the amount of fibroblasts-collagen fibril linkages (Oba and Edwards, 2006; Varani et al., 2006; Fisher et al., 2009). This decrease in linkages in turn results in a weakened extracellular matrix, a reduction in the amount of mechanical loading experienced by the dermal fibroblasts and although not yet tested, the possibility that aging diminishes the ability of the skin to respond to changes in mechanical load (Fisher et al., 2002; Varani et al., 2006; Fisher et al., 2009). Though not well understood, it is thought that these alterations in the extracellular matrix of aged skin are due, at least in part, to increased ROS and ROS-induced matrix metalloproteinase 1 (MMP-1) expression. MMP-1 is an enzyme that degrades collagen, resulting in fragmentation of collagen and disorganization of fibroblast-collagen (Varani et al., 2000; Fisher et al., 2009). Indeed, research from Voorhee and colleagues has demonstrated that aging induces an elevated level of ROS, which is accompanied by increased expression of the transcription factors cJun, AP-1, and the α2β1 integrin, all key regulators of MMP-1 gene expression (Varani et al., 2006; Fisher et al., 2009). Studies on human subjects have also shown that MMP-1 expression and collagen fibril fragmentation is significantly increased in aged human dermal fibroblasts (Fisher et al., 2009). Age-associated increase of fragmented collagen in the dermis not only causes a loss of strength and a reduction of mechanical tension in aged human skin, but also further induces increases in ROS (Fisher et al., 2009).

Beside increased fragmentation of collagen in aged skin, collagen expression is also decreased with aging (Varani et al., 2006). To our knowledge whether changes in the amount of collagen, alterations in tissue ROS or decreases in the amount of fibroblast-collagen linkage result in a diminished ability of the skin to detect or propagate mechanical stimuli has not yet been investigated.

10. Conclusions and future directions

The ability of the cell to sense, process, and respond to mechanical stimuli appears to be altered with aging and these changes have been shown to be associated with increased susceptibility to mechanical damage, increased apoptosis, alterations in intracellular signaling, and a dysregulation of gene expression. These age-associated impairments appear to be ubiquitous, but are not well understood given the complexity of cellular mechanotransduction and the difficulty of studying mechanotransduction process in aged

tissues. Understanding the cellular and molecular basis of physiological adaptation to mechanical loads may play a role in the frailty of aging. Additional research in this area will not only shed light on our understanding of mechanotransduction process in aging, but also may provide valuable clinical insight for the increasing aged population.

Acknowledgments

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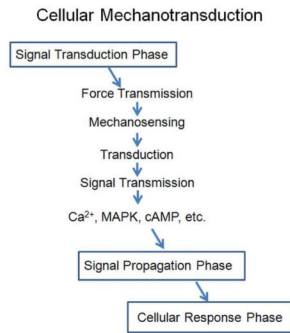
Figure 1.
A simplified schematic of the mechanotransduction processes. The mechanotransductive responses consist of three distinct phases: signal transduction, signal propagation, and cellular response.
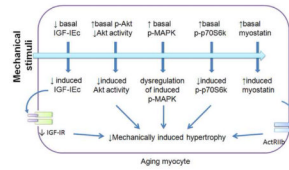
Figure 2.
Dysregulation of mechanotransduction in aging skeletal muscle. At rest the basal expression of mechano growth factor (MGF or IGF-IEc) and IGF-I receptor (IGF-IR) are decreased, while myostatin expression is increased. With aging, the basal phosphorylation of MAPK, Akt and p70S6k are higher, while Akt activity may be decreased. These alterations are thought to impair the ability of aged skeletal muscle to sense and respond to the mechanical stimuli, which may be associated with decreases in mechanically-induced hypertrophy. ActRIIb: activin receptor IIb.
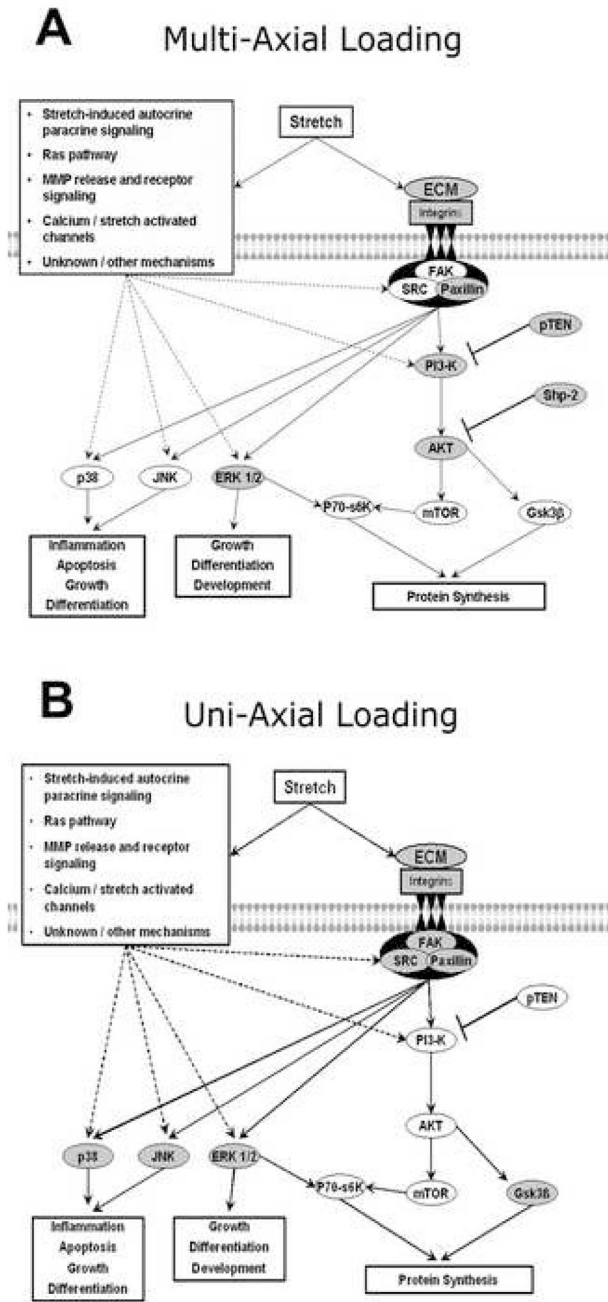
Figure 3.
Stress-induced signaling in aorta. **A.** Effect of aging on load-induced signaling following multi-axial stress in the aging F344BN aorta. **B.** Effect of aging on load-induced signaling following uni-axial stress in the aging F344BN aorta. Unfilled circles represent age-related differences (see text for details).
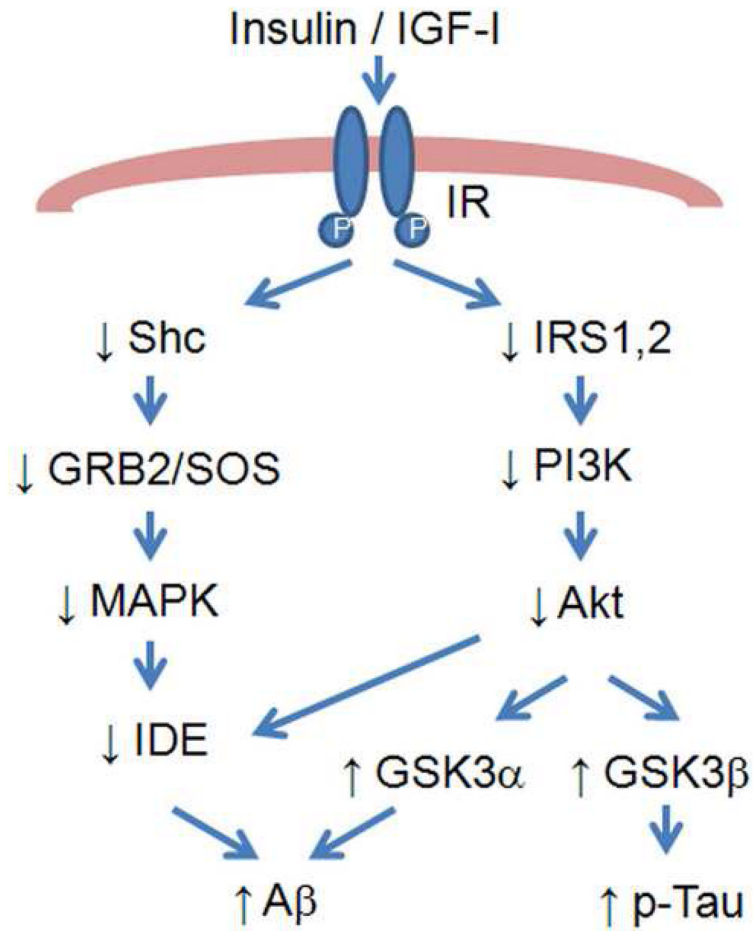
Figure 4.
An overview of the relationship between insulin/IGF-I desensitization and Alzheimer s disease. Insulin resistance is associated with diminished activation of Shc/GRB2/MAPK and IRS/PI3K/Akt signaling, which could lead to increased Tau phosphorylation and Aβ accumulation (see text for details).

Table 1

Response of MAPK phosphorylation to mechanical stimuli in aging skeletal muscle.

isoform	Tissue(s)	Stimulation	Aged (vs. young)	Reference
p38	Red TA	Electrical	Similar	(Ljubicic and Hood, 2009)
	EDL	Stretch	Similar	(Hornberger et al., 2005)
	EDL	Stretch	Similar	(Mylabathula et al., 2006)
	Soleus	Stretch	Lower	(Mylabathula et al., 2006)
	VL	Exercise	Lower	(Williamson et al., 2003)
	VL	Resistance training	Higher	(Kosek and Bamman, 2008)
JNK	EDL	Stretch	Similar	(Hornberger et al., 2005)
	EDL	Stretch	Lower	(Mylabathula et al., 2006)
	Soleus	Stretch	Similar	(Mylabathula et al., 2006)
	VL	Exercise	Lower	(Williamson et al., 2003)
ERK	EDL	Stretch	Higher	(Mylabathula et al., 2006)
	Soleus	Stretch	Higher	(Mylabathula et al., 2006)
	VL	Exercise	Higher	(Williamson et al., 2003)
	VL	Resistance training	Similar	(Kosek and Bamman, 2008)
	TA	Electrical	Lower	(Parkington et al., 2004)
	Plantaris	Electrical	Lower	(Parkington et al., 2004)

TA: tibialis anterior; EDL: Extensor digitorum longus; VL: vastus lateralis.