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## Perinatal stress and early life programming of lung structure and function

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### Abstract

Exposure to environmental toxins during critical periods of prenatal and/or postnatal development may alter the normal course of lung morphogenesis and maturation, potentially resulting in changes that affect both structure and function of the respiratory system. Moreover, these early effects may persist into adult life magnifying the potential public health impact. Aberrant or excessive pro-inflammatory immune responses, occurring both locally and systemically, that result in inflammatory damage to the airway are a central determinant of lung structure-function changes throughout life. Disruption of neuroendocrine function in early development, specifically the hypothalamic-pituitary-adrenal (HPA) axis, may alter functional status of the immune system. Autonomic nervous system (ANS) function (sympathovagal imbalance) is another integral component of airway function and immunity in childhood. This overview discusses the evidence linking psychological factors to alterations in these interrelated physiological processes that may, in turn, influence childhood lung function and identifies gaps in our understanding.

### Keywords

maternal prenatal stress; postnatal stress; programming; hypothalamic-pituitary-adrenal axis; immunomodulation; autonomic nervous system; lung function; airway inflammation; airway hyperresponsiveness

### Introduction

Respiratory disorders are a leading cause of morbidity and mortality in children (Martin et al., 2008). Childhood lung function is important to the development of chronic obstructive pulmonary disease (COPD) in later life (von Mutius, 2002) with COPD projected to be the fourth leading cause of death worldwide by the year 2020 (Mannino and Braman, 2007). Thus, understanding early childhood factors that contribute to early airway inflammation and lung function that may be amenable to prevention and intervention is an important area of research.

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Due to significant developmental plasticity, exposure to environmental toxins during prenatal and/or early postnatal development may alter the normal course of lung morphogenesis and maturation, resulting in changes that affect both structure and function of the respiratory system (Pinkerton and Joad, 2006). Moreover, when normal development is altered, the early effects may persist into adult life, magnifying the public health impact (von Mutius, 2002). Longitudinal studies of the natural history of lung function have shown that a number of early life risk factors including asthma, active and passive smoking prenatally and during childhood, atopy, lower respiratory infections, low socioeconomic status (SES), and perinatal factors including birth weight, gestational age, and nutrition are associated with reduced lung function over the life course (Gern, 2000; Gern, Rosenthal et al., 2005; Hanrahan et al., 1992; Jackson et al., 2004a; Jackson et al., 2004b; O'Connor et al. 1987; Shaheen, 1997). However, these factors do not fully account for observed effects suggesting that as yet unidentified risk factors need exploration.

While the mechanisms of early life environmental influences on lung function are not completely understood, evidence suggests that aberrant or excessive pro-inflammatory immune responses, both locally and systemically, are a central determinant of lung structure-function changes (Holt et al., 2005; Prescott, 2006). Regulatory pathways that involve the collaboration of innate and acquired immune responses are involved. Influences of factors outside the immune system, such as neurohormonal and autonomic nervous system functioning (phenotypes), may also be important (Buijs et al., 2008; Elenkov et al., 2008). Both glucocorticoid action and sympathovagal balance play a role in regulating lung function and airway response, including having a major role in fetal and postnatal lung development.

Overlapping research suggests that negative emotion and psychological stress influence respiratory processes and lung function, having both transient and more long-term effects (Ritz, 2004; Ritz and Kullowatz, 2005). It has been estimated that between 20-40% of asthmatic subjects have significant changes in lung function induced by emotional stimuli (Isenberg et al., 1992). In one study, Ritz and Steptoe (2000) demonstrated clinically significant changes in forced expiratory volume in the first second ( $\Delta FEV_1$ ) (i.e.,  $\Delta FEV_1 > 15\%$ ) in association with negative mood states in 25% of the adult asthmatic subjects with mild to moderate disease. In another series of studies, Ritz and colleagues examined the effects of affective states on respiratory resistance (in both healthy and asthmatic subjects) using the forced oscillation technique during viewing of pictures, imagery, and films (for an overview of studies see Ritz, 2004). Collectively the findings from these studies showed the greatest effect was induced by negative states with within-individual effect sizes for an increase in respiratory resistance (relative to prestimulus levels) ranging between  $d = 0.37$  and 1.02 depending on the stimulus material. Cross-sectional (Jackson et al., 2007) and longitudinal epidemiological studies in adults (Kubzansky et al., 2006; Kubzansky et al., 2002) have linked negative emotion to reductions in maximally attained lung function as well as a more accelerated rate of lung function decline. While studies examining stress in the prenatal period are not available, one study in urban children found a relationship between exposure to chronic stress (violence) in early childhood and reduced lung function at age 6 years (Franco Sughia et al., 2007). Moreover, overlapping evidence demonstrating that psychological factors influence neurohormonal, and consequently, immune inflammatory processes, starting *in utero* suggest their possible role in early lung growth and development (Wright, 2007). Animal studies, and some early work in humans, have advanced our understanding of how perinatal stress may be linked to evolving immunocompetence and airway responses in early life, albeit research in humans remains sparse. In the discussion that follows, we review the evidence suggesting that stress elicited disruption of interrelated systems - neuroendocrine, autonomic, and immune - during the perinatal period, may lead to increased vulnerability to airway inflammation and reactivity

as well as reduced lung function in later life. Evidence is also reviewed suggesting that programming of early lung development may occur through epigenetic influences on gene regulation in critical periods of development starting prenatally.

## Developmental Plasticity of the Respiratory System

Plasticity is a consequence of environmental exposures during critical life periods affecting key physiological systems that operate in orchestrating underlying developmental processes (Feinberg, 2007). Lung development commences *in utero* and progresses through to adolescence and young adulthood when maximally attained lung function is achieved. Due to rapid growth and plasticity of physiological characteristics during gestation and early childhood, the fetus and young children may be particularly vulnerable to environmental toxins (Kajekar, 2007). Adolescence is another period of rapid growth in lung function and also warrants attention in this regard.

Epidemiological studies demonstrate that prenatal factors that contribute to intrauterine growth restriction (IUGR) and low birth weight increase the risk of respiratory problems and reduced lung function in infants, children and adults as noted above. Factors that play a role include maternal prenatal stress and maternal-fetal cortisol disruption (e.g., Kivlighan et al., 2008). In animal models, restrictive growth *in utero* has been linked to reduction of internal surface area relative to lung volumes (Harding et al., 2000) and also airway development – specifically, impaired growth of bronchial walls which may have implications for airway compliance in later life (Wignarajah et al., 2002). The mechanisms underlying such structural changes are not completely elucidated.

While the origins of chronic lung diseases are multi-factorial, the central underlying mechanisms leading to reduced lung function and exaggerated responsiveness to bronchoconstrictor stimuli involve chronic airway inflammation associated with a cycle of injury, repair and remodeling (Holgate, 1997; Holt et al., 2005). The fundamental cause of the underlying airway inflammation is aberrant and/or excessive immune responses to various environmental agents (Holt et al., 2005). While such changes have been well documented in older children with clinical manifestations of asthma and reactive airways disease, it has become evident that airway inflammation and early remodeling occur and progress even in the presymptomatic and/or subclinical state (Pohunek et al., 2005; van den Toorn et al., 2001). This further underscores the importance of the early childhood period as lung function may be programmed or ‘set’ in the first years of life (Bush, 2008; Morgan et al., 2005; Rasmussen et al., 2002). Other research has focused on the role of the nervous system and neuropeptides in airway inflammation and response (Undem and Weinreich, 2003). Studies have also documented autonomic nervous system (ANS) dysregulation (increased heart rate variability, increased vagal tone) in asthmatic children (Anthracopoulos et al., 2005; Kazuma et al., 2000; Miller and Wood, 1997; 2003). Recent advances in methodology allow for the assessment of autonomic measures in even young infants suggesting that the disruption of cardiorespiratory regulation may be evident in the first year of life (Alkon et al., 2006; Bosquet-Enlow et al., 2009) and vulnerable to *in utero* and early postnatal programming related to environmental factors (Cohen et al., 2008; Proper et al., 2008).

Taken together, these data suggest that the developmental origins of the structural and functional organization of the lungs involve, in part, the coordinated maturation of the immune, neural, and endocrine systems. Environmental toxins that disrupt these interrelated systems during prenatal and/or early postnatal development may thus alter the normal course of lung morphogenesis and maturation, potentially resulting in long-term changes in the respiratory system (Kajekar et al., 2007; Bavis and Mitchell, 2008; Mitchell and Johnson,

2003). Psychological stress is an important candidate for such environmental programming although this remains virtually unexplored in human studies.

## Key Physiological Systems Linked to Lung Development and Airway Responses

### Immune function

Research continues to delineate the relationships among early environmental influences, immunodeviations, and developmental outcomes in the lungs (Heaton et al., 2005; Prescott, 2006). Immunophenotypes linked to airway inflammation and reactivity show significant interindividual variability and are dependent on the particular environmental challenge as well as timing of exposure(s). The most common cause of chronic airway inflammation in early childhood is asthma. Although diverse pathogenic mechanisms are likely involved in different forms of asthma and the resulting airway changes [i.e., airway hyperreactivity (AHR), which is thought to be present in all patients with asthma (Wills-Karp, 1999)], over the past 10 years there has been much focus on the influence of the systemic propensity for type 2 T-helper (Th2) allergic responses and eosinophils (Umetsu et al., 2002). Since the description of Th1 and Th2 clones, characterized by different patterns of cytokine secretion (Mossman and Coffman, 1989; Mossman and Sad, 1996), it has become evident that chronic airway inflammation and associated clinical syndromes (e.g., allergic asthma) are associated with a systemic Th2-biased response (Robinson et al., 1992). The Th2 paradigm involves a complex interaction of T and B lymphocytes, and antigen presenting cells, resulting in the production of higher levels of cytokines, such as interleukin-4 (IL-4) or IL-13, lower levels of interferon gamma (IFN- $\gamma$ ) and increased immunoglobulin E (IgE) production. These mechanisms have their roots in early life with an immunological bias towards a Th2 phenotype *in utero* (Devereux et al., 2002; G Devereux et al., 2001; Liao et al., 1996; Miles et al., 1996; Prescott et al., 1998; Tang et al., 1994). Epidemiological studies suggest that perinatal environmental conditions determine whether this propensity persists or is redirected toward Th1 and/or T cell regulatory (Treg) pathways, even in those with a genetic predisposition to allergy (Garn and Renz, 2007; Ownby et al., 2002; Taussig et al., 2003; Wang and McCusker, 2006; Willwerth et al., 2006). Experimental findings suggest that perinatal stress may influence the evolving systemic propensity for type Th2 responses (i.e., enhanced adaptive immunity) as discussed below.

While the Th2 paradigm has been useful in understanding the subjects with allergic asthma and airway inflammation, this is clearly not able to explain all forms. Early wheeze and asthma is increasingly understood to be a heterogeneous syndrome associated with diverse factors that involve not only allergen sensitization, but also infection, air pollution, tobacco smoke, and perinatal stress (Gold and Wright, 2005; Pinkerton and Joad, 2006). These pathways may be relevant to atopic wheeze and asthma, however a significant proportion of recurrent airway obstruction in the preschool years is nonatopic wheeze (Bush, 2008). These children often do not have a family or personal history of atopy. Airway changes are marked by neutrophilic inflammation (Wenzel and Busse, 2007) albeit increased eosinophilia and IgE production are common to both atopic and nonatopic wheeze (Turato et al., 2008).

It is also evident that non-Th2 factors, e.g., IFN- $\gamma$  (a Th1 cytokine) and neutrophils, are frequently found in the lungs of many patients with asthma, particularly in those with more severe disease (Wenzel and Busse, 2007). Indeed, Th1 cells also play a role in allergic inflammation (Hansen et al., 1999). It is now accepted that the Th2 biased polarization of adaptive immunity may be only one of several axes that result in enhanced susceptibility to airway inflammation and altered reactivity. Antigen-independent responses including innate immune cells [e.g., bronchial epithelial cells, alveolar macrophages, and dendritic cells

(DCs)] may also be important in modifying airway inflammation (Suarez et al., 2008). Environmental influences may impact maturation process of dendritic cells, thus determining their phenotype and function.

Still other evidence has identified a novel T cell subset associated with pulmonary inflammation as a result of particular environmental exposures (e.g., cigarette smoke exposure, ozone) (Harrison et al., 2008; Miossec et al., 2009). A number of investigators have examined several distinct mouse models of asthma and AHR, including allergic and non-allergic forms, searching for common disease mechanisms. What has been found is that a component of innate immunity, natural killer T (NKT) cells, are required for the development of multiple forms of AHR, induced with allergen (Akbari et al., 2003), exposure to ozone (a component of air pollution) (Pichavant et al., 2008), and exposure to respiratory viruses (Kim et al., 2008). Moreover, different subsets of NKT cells are involved in these different forms of AHR. It has also been demonstrated that NKT cells produce IL-17A in the context of oxidative stress, and it is known that oxidative stress can exacerbate allergen induced AHR (Rangasamy et al., 2005).

### Neuroendocrine function

Influences of factors outside the immune system, such as neurohormonal phenotypes, may also influence these processes (Buijs et al., 2008; Elenkov and Chrousos, 1999; Elenkov et al., 2008) albeit such relationships are virtually unexplored in human early life respiratory disorders, at least directly. Glucocorticoids play a major role in regulating fetal and postnatal lung development (Grier and Halliday, 2004). Many studies have demonstrated that fetal and early postnatal exposure to endogenous or exogenous corticosteroids affect lung development and that resulting structural alterations persist (Harding et al., 2000). Evidence also links hormonal imbalances (in particular, cortisol) with altered cytokine and Th1/Th2 balance. For example, glucocorticoids (GCs) are known to inhibit the production of IL-12, IFN- $\gamma$ , by antigen presenting cells and Th 1 cells, but up-regulate the production of IL-4, IL-10, and IL-13 by Th2 cells (Wonnacott and Bonneau, 2002), and to prevent the development of regulatory T cells (Stock et al., 2004). This may induce selective suppression of the Th1-mediated cellular immunity and trigger a shift towards Th2-mediated humoral immunity (Elenkov and Chrousos, 1999; Elenkov et al., 2005). It has been proposed that alterations in maternal cortisol levels *in utero* may influence fetal immune system development and Th2 cell predominance, perhaps through direct influence of stress hormones on cytokine production (von Hertzen, 2002; Wright, 2005). In addition, glucocorticoid resistance may be an important determinant of lung development and function (Barnes, 2004; McKinley et al., 2008).

### Autonomic nervous system

Our understanding of how the nervous system may be involved in the organization of the immune response continues to evolve (Sternberg, 1997, 2001; Sternberg, 2006; Tracey, 2002). CNS-mediated regulation of the peripheral immune response is mediated through vagal output (e.g., suppressing the innate immune defense to pathogens; altering pro-inflammatory cytokine balance). The efferent vagus nerve is proposed as an immune-to-brain pathway that may directly modulate the airway immune response to pathogenic invasion or to injury by irritants and toxins. The cholinergic vagus nerves participate in the regulation of the airway inflammatory response, in part, through efferent vagal endings present in the airway smooth muscles. Cholinergic mechanisms represent the predominant constrictor neural pathway in human airways (Barnes, 1986). Differences in expression of muscarinic acetylcholine receptors in asthma suggest that cholinergic system may participate in the molecular framework influencing airway function in this context (Lutz and Sulkowski, 2004). Conversely, inflammatory processes could exacerbate allergic cholinergic

airway narrowing. A current model of excessive airway narrowing in allergic asthma highlights inflammation-induced damage of m2-autoreceptors which downregulate cholinergic transmission at the level of the postganglionic nerve terminal in health and thereby limit the constriction of airway smooth muscles (Barnes, 1992; Fryer and Jacoby, 1998). Notably, a number of animal studies suggest that neural control of airway smooth muscle and the irritant receptor systems is established during the perinatal period (Card et al., 2005). It thus seems reasonable to suggest that disruption of the vagal anti-inflammatory pathway may predispose some individuals to excessive inflammatory responses in early life.

### Integration of systems

A bidirectional network of interactions between the central nervous system, the endocrine system and the immune system is well documented (Butts and Sternberg, 2008; Wrona, 2006). The immune and nervous system are closely related in both physiological and pathological reactions in the lung. For example, extensive communications between neurons and immune cells are responsible for the magnitude of airway inflammation and the development of airway hyperreactivity, a consequence of neuronal dysregulation (Nockher and Renz, 2006; Veres et al., 2007). These systems likely act cooperatively to maintain homeostasis. To date, the majority of studies examining the impact of stress on physiological systems and subsequent health have examined one system in isolation from others. However, research increasingly suggests that it is the disturbed balance of these systems that predicts susceptibility to disease. For example, recent findings related to hypothalamic-pituitary-adrenal (HPA) axis and ANS functioning highlight the need to consider these systems simultaneously due to their interactive influences in predicting outcomes. Specifically, two recent studies have shown a modifying effect of high vs. low salivary alpha-amylase (as a surrogate marker of sympathetic nervous system functioning) on the influence of high vs. low cortisol on behavioral outcomes in young children (El-Sheikh et al., 2008) and adolescents (Gordis et al., 2006).

### Pre- and Postnatal Stress and Physiologic Programming

This section provides an overview of how stress may be involved in the early programming of these systems.

#### General Stress Paradigm

In response to stress, physiological systems may operate at higher or lower levels than in normal homeostasis. It is the disturbed balance of these systems that is relevant to disease. Immune and neuroendocrine defensive biological responses important for the short-term response to stress, may produce long term damage if not checked and eventually terminated (McEwen, 2002). Disturbed regulation of stress systems [e.g., HPA axis, sympathetic-adrenal-medullary (SAM) system] due to maternal stress may, in turn, modulate immune function in the offspring beginning *in utero* (Arck et al., 2006; de Weerth and Buitelaar, 2005). That is, offspring of mothers with biobehavioral sequelae may inherit a biological vulnerability to disrupted stress regulatory systems altering their reactivity to subsequent challenges (Yehuda and Bierer, 2008). Likewise, non-optimal early childhood environments and caregiving experiences may also impact these processes (e.g., maternal psychopathology, maternal insensitivity) (Anisman et al., 1998; Liu et al., 1997; Vallee et al., 1997). Developmental findings show that the acquisition of the ability to regulate one's response to stress ("self-regulation") progresses through several stages in the early years of development. Neonates and young infants are highly dependent on their caregivers as regulators. The HPA axis and ANS seem particularly susceptible to early-life programming.

## Perinatal Programming of the HPA Axis

Prenatal stress has been associated with early and long-term developmental effects resulting in part from altered maternal and/or fetal glucocorticoid exposure. Maternal and fetal stress also stimulates placental secretion of corticotrophin-releasing hormone (CRH), which in turn is elevated in the neonatal circulation (Goland et al., 1993; Reinisch et al., 1978; J. Seckl, 1997; Seckl, 2001). This may stimulate the fetal HPA axis to amplify fetal GC excess as well as activate additional elements of the fetal stress response (i.e., catecholamines and neurotrophins) influencing the developing immune and autonomic nervous systems (Arck et al., 2006). While these *in utero* responses may be adaptive in the short term, geared toward coping with anticipated environmental challenges, ultimately they may exact a toll in contributing to increased risk of inflammatory diseases in later life.

The HPA system remains highly reactive and labile in early infancy and becomes organized between 2 and 6 months of age through transactions between the child and caregiver (Wright and Bosquet, 2008). Studies have consistently demonstrated that the quality of caregiving that the child receives during early development predicts the emergence of later self-regulation abilities, with sensitive caregiving associated with more adaptive self-regulatory abilities and more optimal functioning of the child's HPA system (Lyons-Ruth and Block, 1996). Increased maternal stress has been associated with lower levels of parenting sensitivity and higher levels of negative parenting behaviors (e.g. abuse, hostility) (Belsky, 1984). Not surprisingly then, perinatal maternal stress has been associated with poor stress regulation and other negative outcomes in both animal and human offspring. (Caldji et al., 2000; Cicchetti and Rogosch, 2001; Coplan et al., 1996; DeBellis et al., 1999; Essex et al., 2002; Field, 1994; Francis et al., 1999; M.R. Gunnar et al., 2001; Gunnar and Donzella, 2002; Heim et al., 2000; Hessel et al., 1998; Kaufman et al., 1997).

## Perinatal Programming of Autonomic Reactivity

Several animal models as well as human studies support the connection between an adverse intrauterine environment as well as experiences in early postnatal life and alterations of autonomic nervous system balance (e.g., sympathovagal balance) (Card et al., 2005; Herlenius and Lagercrantz, 2004; Jansson and Lambert, 1999; Pryce, Ruedi-Bettschen, Dettling, and Feldon, 2002). Experimental rat models have shown that prenatal stress is associated with exaggerated cardiovascular reactivity to restraint stress (Igosheva et al., 2004). In humans, infants' autonomic responses show developmental changes with relative stability between 6 to 12 months of age (Alkon et al., 2006). The balance between functional parasympathetic and sympathetic activity in relation to stress, emotional stimuli, and immune function may also be important for the expression of allergic sensitization and atopic disorders as well as early airway inflammation and reactivity (Wright, 2005). Finally, a number of animal studies suggest that neural control of airway smooth muscle and the irritant receptor systems are established during this early life period and sensitive to environmental programming (Card et al., 2005).

## Linking stress, immunomodulation, respiratory function, and airway responses

The preceding discussion suggests that individuals may be susceptible to the development of immunologic deviations that contribute to both systemic and localized inflammation in the airways and/or airway hyperresponsiveness through a number of pathways linked to stress. Moreover, evidence suggests a role for imbalance in sympathovagal activity in airway responses and lung function which may also be influenced by stress and negative affect. The evidence linking these processes to respiratory function is summarized below.

## Perinatal Stress and Early Immunomodulation

Factors, including psychological stress, that alter the maturation of local immune networks (e.g., dendritic cells), epithelial cells [ECs], regulatory T cells) may predispose to a Th2 phenotype (Joachim et al., 2008). Psychological stress has been associated with increased proportions of both natural killer (NK) and NKT cells as well as the altering their functional mechanisms (Lutgendorf et al., 2005; Oya et al., 2000). These effects have been shown by others to be correlated with changes in cortisol and sympathetic nervous system input (Sagiyama et al., 2004). Evidence in rhesus monkeys suggests that prenatal stress impacts the infant monkey's response to antigens at birth (Coe et al., 1999). Others have documented dysregulated pathways of the cellular and humoral immune response upon antigen challenge in prenatally stressed adult mice, reflected by a Th2 greater than Th1 adaptive immune response and increased IgE levels in vivo (Pincus-Knackstedt et al., 2006). In a prospective urban pregnancy cohort in Boston [the Asthma Coalition on Community, Environment, and Social Stress (ACCESS) study], my laboratory has documented evidence that prenatal maternal stress is associated with increased cord blood IgE expression in the children (Sterntal et al., 2009) and enhanced reactivity to low-dose prenatal allergen exposure as indexed by elevated IgE in cord blood (Peters et al., 2008). Recent analyses in the Urban Environment and Childhood Asthma (URECA) study, demonstrated an association between increasing levels of cumulative maternal stress assessed prenatally and altered innate and adaptive immune responses in cord blood mononuclear cells (Wright et al., in press). My group has also prospectively linked early life caregiver stress to dysregulation of immune function in another Boston birth cohort predisposed to allergy (Wright et al., 2004). We found that increased maternal caregiving stress was associated with greater antigen-specific tumor necrosis factor (TNF)-alpha production in particular. A number of studies have found that stress induces the release of pro-inflammatory cytokines including TNF-alpha as well as others (e.g., IL-6) (LeMay et al., 1990; Nakano et al., 1992; Yamasu et al., 1992) and that TNF-alpha is important in asthmatic airway inflammation. Continued follow-up of these prospective studies will examine whether stress-induced perinatal immunomodulation impacts the expression of allergic disease in these children.

Chronic psychological stress is known to alter innate- and adaptive-immune responses to a variety of pathogenic challenges with Toll-like receptors (TLRs) playing a key role. Evidence in murine models suggests that psychological stress may operate in a TLR4-dependent manner. Powell and colleagues have demonstrated that stress modulates Toll-like receptor cytokine secretion in response to unmethylated CpG motif in bacterial deoxyribonucleic acid (DNA) and polyinosinic-polycytidylic acid (Poly I:C) in splenic DCs rendering them resistant to glucocorticoids (Powell et al., 2009). Zhang and others have demonstrated an association between stress and TLR4-mediated P13K/Akt signaling in mice (Zhang et al., 2008). Notably, recent data from a human pregnancy cohort showed that higher levels of prenatal maternal stress was associated with increased IL-8 and TNF-alpha production following microbial stimulation (e.g., polyinosinic-polycytidylic acid, Poly I:C) suggesting that prenatal stress may modify the neonatal immune response through Toll-like receptor (TLR)-dependent pathways (Wright et al., in press).

One notable challenge to linking stress-induced changes in immunity to lung function outcomes is assessing and understanding the complex relationships between local airway inflammation and systemic inflammatory processes in this context. While quantifying inflammation in the lungs is most directly accomplished through invasive methods including the analysis of bronchoalveolar lavage (BAL) fluid or bronchial biopsy obtained through bronchoscopy or semi-invasive methods including sputum induction (Montuschi, 2007), this is not always practical in human research studies. Less invasive techniques for assessing either the gaseous phase of exhaled breath (e.g., nitric oxide, carbon monoxide, hydrocarbons) or the liquid phase known as exhaled breath condensate (EBC) are being



developed. While measurement of exhaled nitric oxide (NO) has been standardized and validated for subjects as young as 4 years (Beydon et al., 2007), methods for EBC analysis in these younger age groups is still being worked out (Montuschi, 2007; Hoffmeyer et al., 2009). Other studies incorporate the measurement of inflammatory biomarkers in plasma [e.g., high sensitivity C-reactive protein (hs-CRP)]. While CRP from peripheral blood may be more reflective of systemic inflammation, there is correlational evidence that such markers may serve as a reasonable surrogate of airway inflammation. High sensitivity CRP, a systemic marker of low-grade inflammation has been negatively correlated with pulmonary function tests (FEV<sub>1</sub>) ( $r = -0.49$ ,  $p = 0.01$ ) and positively associated with percent eosinophils ( $r = 0.44$ ,  $p = 0.03$ ) in induced sputum from asthmatics (Allam et al., 2009) as well as an increased frequency of airway hyperresponsiveness (Ridker et al., 1997). While some studies in adult asthmatics demonstrate that hs-CRP in serum is correlated with hs-CRP in EBC ( $r = 0.74$  in stable asthma treated with inhaled corticosteroids,  $p = 0.0003$ ) as well as exhaled nitric oxide ( $r = 0.62$ ,  $p = 0.004$ ) (Zietkowski et al., 2009), other studies do not find this relationship (Sutherland et al., 2007). Overlapping lines of evidence in asthmatics (Navratil et al., 2009; Takemura et al., 2006) and those with chronic obstructive pulmonary disease (Sin and Man, 2006; Dahl et al., 2001) suggest that while systemic and airway inflammation may be correlates of one another, they may also operate independently in influencing lung structure and function. In a recent prospective study of young adults, increased levels of hs-CRP in blood predicted a greater decline in FEV<sub>1</sub> over a 9-year follow-up period [23 ml/yr vs. 1.6 ml/yr decline in men; 6.2 ml/yr decrease vs. 1.6 ml/yr increase in women (highest quartile hs-CRP vs. lowest quartile, respectively)] (Rasmussen et al., 2009). These differences remained significant when adjusted for a number of potential confounding factors including body mass index (BMI), cardiorespiratory fitness, smoking, asthma, AHR, and serum eosinophil cationic protein (ECP), an indicator of eosinophilic inflammation. Moreover, studies demonstrating an association between CRP genotype and lung function further support a primary role of systemic inflammation in airway disease. Thus, going forward, studies should examine interrelationships among stress-induced changes in both localized and systemic inflammation, lung growth and development, and lung function measures, extending what we have learned in these studies of older children with asthma and adult asthmatics to earlier developmental periods (at birth, in infancy, and early childhood).

### Perinatal Stress and Airway Pathophysiology

Perinatal stress and emotional arousal may influence airway narrowing through inflammatory pathways as well as neuronal activity (Quarcoo et al., 2009; Joachim et al., 2008; Pincus-Knackstedt et al., 2006). Nogueira and colleagues (Nogueira et al., 1999) were first to demonstrate that prenatal stress increased allergen-induced airway inflammation in adult mice offspring. Similarly, allergen aerosol challenge has been associated with increased airway hyperresponsiveness in mice exposed to prenatal stress (Pincus-Knackstedt et al., 2006). Moreover, total serum IgE was significantly increased in ovalbumin (OVA)-sensitized adult offspring in response to prenatal stress. Mice exposed to prenatal stress were also more likely to express a Th2 adaptive immune response. Additional studies have demonstrated exacerbations in airway inflammation in OVA-sensitized rats following chronic or repeated psychosocial challenge (Datti et al., 2002; Forsythe et al., 2004; Joachim et al., 2003; Joachim et al., 2004; Okuyama et al., 2007). One study in subjects with allergic asthma demonstrated that increased psychological stress was correlated with increased levels of brain-derived neurotrophic factor (BDNF) which, in turn, was negatively correlated with percent predicted forced expiratory volume in 1 second (FEV<sub>1</sub>) (Joachim et al., 2008). Notably, stress perception was also positively correlated with the percentage of TNF-alpha-producing T cells in these subjects. The authors speculated that this may point to a neuro-immunological interaction given evidence demonstrating the constitutive secretion of BDNF

in human peripheral blood monocytes which is enhanced when stimulated with TNF-alpha (Schulte-Herbruggen et al., 2005). This group has also demonstrated stress-induced increase in tachykinin-like substance P associated with allergic airway inflammation in a mouse model (Joachim et al., 2006). No studies to date have examined the influence of prenatal and early life stress on the expression of neuropeptides and their possible role on airway inflammation and response in early development.

### **Stress and acquired glucocorticoid resistance**

An alternative hypothesis linking stress, neuroendocrine and immune function considers a glucocorticoid resistance model (Miller et al., 2002). Glucocorticoid sensitivity plays a role in the developmental regulation of the lung (Ganalingham et al., 2006). Insight into the cellular and molecular mechanisms underlying stress-induced steroid resistance is provided in a number of recent studies. Oxidative stress pathways have been implicated in the link between psychosocial stress and asthma (Wright, 2005) as well as steroid resistant asthma (Adcock and Barnes, 2008; Marwick et al., 2008). This may particularly be relevant in airway inflammation where neutrophilic rather than eosinophilic inflammation predominates (Bush, 2008; Wenzel et al., 1999). Indeed, it was recently shown that oxidative stress contributes to steroid resistance in the context of neutrophilic inflammation in a mouse model of acute asthma exacerbations (Ito et al., 2008). Notably, psychological stress is also an oxidant and may thus operate through these same pathways (Wright, 2005). While human data in the context of lung disease are sparse in this regard, one recent cross-sectional analysis in adolescents demonstrated that peripheral blood mononuclear cells harvested from asthmatics who perceived low parental support (i.e., greater stress) were more resistant to hydrocortisone's effects on cytokine expression (IL-5, IFN- $\gamma$ ) and activation of eosinophils relative to asthmatics reporting higher parental support (Miller et al., 2009). Examination of mechanisms contributing to steroid resistance in relation to perinatal stress may provide insight into the link between stress and reduced lung function or airway hyperresponsiveness over subsequent development.

### **Epigenetics – A Fundamental Programming Mechanism**

Programming effects of stress on respiratory outcomes may operate at a more fundamental molecular level, i.e., through epigenetic programming. Determining the range of environmental exposures that impact the epigenome during development was a research priority identified at the recent National Heart, Lung, and Blood Institute (NHLBI) Pediatric Pulmonary Disease Strategic Planning Workshop (Castro et al., 2009). DNA methylation is an adaptable epigenetic mechanism that, in mammals, modifies genome function through the addition of methyl groups to cytosine to form 5-methyl-cytosine. DNA methylation marks are largely established early in life (Feinberg, 2007; Gluckman et al., 2008) and may ensure stable regulation that mediates persistent changes in biological and behavioral phenotypes over the lifespan (Miller and Ho, 2008; Weaver et al., 2002). DNA methylation of many genes changes with disease status and in response to environmental signals including chemical exposures such as diet, drugs and toxins (Baccarelli et al., 2009; Jaenisch and Bird, 2003; Li et al., 2003). Recent findings also implicate psychological stress given behavioral studies demonstrating epigenetic changes during fear conditioning (Levenson and Sweatt, 2006; Miller et al., 2007) and evidence for epigenetic programming related to maternal care (Meaney and Szyf, 2005; Szyf et al., 2008).

Genes involved in HPA axis functioning seem particularly susceptible to stress-related programming (Wright and Bosquet, 2008). These include glucocorticoid receptor expression, the activation of which alters HPA activity through negative feedback inhibition (De Kloet, 2004). The human glucocorticoid receptor (GR) promoter region is extensively methylated with diverse methylation profiles demonstrated in a nonclinical sample of

subjects (Turner et al., 2008). The intracellular access of glucocorticoids to their receptors is also modulated by the 11 beta-hydroxysteroid dehydrogenase (11 $\beta$ HSD) enzymes, which interconvert biologically active 11  $\beta$ -hydroxyglucocorticoids and inactive 11-ketosteroids (Krozowski et al., 1994). While compromised 11 $\beta$ HSD2 activity can be caused by loss of function mutations of the gene encoding 11 $\beta$ HSD2, the frequency of such mutations is extremely low (Zaehner et al., 2000). Thus, other mechanisms accounting for the inter-individual variability in 11 $\beta$ HSD2 enzyme activity should be considered (Alikhani-Koopaei et al., 2004). The 11 $\beta$ HSD2 promoter comprises a highly G + C-rich (or GC-rich) core, contains more than 80% GC, lacks a TATA-like element, and has two typical CpG islands raising the possibility that methylation may play a role in the epigenetically determined inter-individual variable expression of 11 $\beta$ HSD2. One candidate pathway implicated in both airway inflammation (Esposito and Cuzzocrea, 2007) and autonomic response (Danson et al., 2009) is the nitric oxide (NO) signaling pathways. Alterations of NO expression occur in the context of psychological stress and stress-related behaviors (McLeod et al., 2001). The inducible nitric oxide synthase (NOS) genes are also susceptible to epigenetic programming (Tarantini et al., 2009).

The notion that variability in methylation between subjects may reflect an important epigenetic mechanism is suggested by recent studies in both animals and humans. Epigenetic modulation of the 11 $\beta$ HSD2 gene has been recently demonstrated in a rodent model and cultured cell lines (Alikhani-Koopaei et al., 2004), albeit epigenetic regulation of this gene is not well characterized in humans. Weaver and colleagues have demonstrated differential methylation patterns of the Ngfi-A-binding site in GR promoter 1 $\gamma$  in the rat brain in offspring that had received poor maternal care versus those that had received better maternal care (Weaver et al., 2004). When pups were cross-fostered between mothers providing good or poor post-natal care (i.e., pups who had received poor caregiving from their biological mother were switched to the caregiving of the foster mothers who now provided positive caregiving), the pups developed the epigenome of the foster mother. This same group recently reported increased methylation in a neuron-specific GC receptor (NR3C1) promoter as well as decreased levels of GC receptor mRNA from hippocampus tissue obtained from suicide victims with a history of childhood abuse (McGowan et al., 2009). Recent human data demonstrates that methylation of exon 1F in fetal cord blood was sensitive to maternal mood in the perinatal period and the infants HPA stress reactivity (Oberlander et al., 2008). Whether alterations in DNA methylation underlie stress-induced altered phenotypic plasticity related to lung structure and function remains largely unexplored.

Finally, genetic and epigenetic studies tell us that exposure to altered glucocorticoid receptor response through early development, even beginning *in utero*, programs major changes in the endogenous neuroendocrine and immune mechanisms that may, in turn, lead to increased vulnerability to asthma (Wright, 2007). It will be important to begin to understand factors related to developmental programming of glucocorticoid sensitivity during critical periods of development which may play a role in disease etiology as well as subsequent morbidity.

## Influence of Modifying Factors

A number of factors have been shown to modify the effects of perinatal stress on the early child developmental processes discussed here that may, in turn, influence lung function. While an in-depth discussion is beyond the scope of this review, it is worth acknowledging those factors that may be considered in research going forward. These include genetic susceptibility of the index child, family context (Morris et al., 2007), race/ethnicity (Cunningham et al., 2009), developmental stage (Boyce and Ellis, 2005), child's gender

(Hatzinger et al., 2007) and temperament (Talge et al., 2008). For example, data from my laboratory has shown gender differences in the relationships between family and community violence exposure and early childhood lung function (Franco Suglia et al., 2007).

## Influence of Stress on Prospective Development of Wheeze Phenotypes

In order to disentangle whether stress has a greater effect on lung architecture and growth versus processes involved in airway response and inflammation, it may be useful to examine the influence of stress on early wheeze phenotypes in childhood in a prospective design. While our understanding of the natural history of childhood wheezing is still evolving, epidemiological studies have characterized distinct patterns of wheezing based on age of onset, remission, and persistence with further classification based on allergic, immunologic and physiological parameters (Taussig et al., 2003). Wheeze phenotypes (transient, atopic, nonatopic) evolve over early childhood and are more clearly delineated by age 6 to 7 years. These distinctive patterns of wheezing may reflect differences in etiology and prognosis for asthma development as well as having differential relationship to lung function (Martinez et al., 1995). Typically, studies discriminate between transient wheezing (nonatopic children who wheeze during the first 3 years of life but not thereafter), nonatopic persistent wheezing [nonatopic children who wheeze beyond the third year of life after having a lower respiratory illness (LRI)], and atopic wheezing (atopic children with wheezing during the first 6 years of life). This latter group can be further divided into early atopic or persistent wheezers (those whose symptoms started during the first 3 years and wheezed at age 6) and late atopic wheezers (those who did not wheeze before age 3 but did have wheezing by age 6) albeit the latter group is relatively rare. These wheezing phenotypes, in turn, have been differentially associated with lung function changes (Lowe et al., 2009; Stern et al., 2007) including airway resistance (Brussee et al., 2004). Wheezing phenotypes, particularly persistent wheezing, are predictors of lung function by age 6 years. Stress may operate through different pathways to influence wheeze risk and lung growth and development including altered somatic growth (prematurity, birth weight), altered intrinsic airway responses, and immunomodulation beginning *in utero* that impacts consequent airway inflammation. Specifically, stress-elicited disruption of interrelated systems – neuroendocrine, autonomic, and immune – during the prenatal and early childhood periods may contribute to an increased risk for wheezing syndromes as well as potentiate an increased vulnerability to airway inflammation and reactivity resulting in altered childhood lung function. How stress influences distinct wheeze phenotypes may vary based on timing of exposure (prenatal, early postnatal, cumulative) underscoring the need to examine associations prospectively. For example, in prospective studies one could examine the influence of prenatal maternal stress on maternal cortisol disruption *in utero* and subsequent effects on subsequent wheeze phenotypes and childhood lung function. One could also examine whether this relationship is mediated through IUGR and low birth weight.

## Summary and Conclusion

Evidence that psychological factors influence neurohormonal and consequently immune inflammatory processes starting *in utero* suggest their possible role in lung growth and development albeit research on the influence of stress on childhood lung function outcomes remains sparse. Stress-elicited disruption of interrelated systems - neuroendocrine, autonomic, and immune - during perinatal development may lead to increased vulnerability to airway inflammation and reactivity as well as reduced lung function in later childhood and even into adulthood. Future studies need to examine how fetal exposure to maternal stress may influence programming of these interrelated systems and how such effects are independent of and moderated by postnatal stress as well as other environmental toxins. That is, stress-induced disruption of these interrelated systems may have important

independent effects on lung growth and development, but also may enhance vulnerability to other known lung toxins (i.e., air pollution, tobacco smoke) (Wright et al., 2005). An important step toward identifying children at risk for costly pediatric disorders is the identification of mechanisms that may lead to and maintain early predisposition. Identifying programming agents that help to shape functional competence of the immune system and interrelated systems may give new insights into the mechanisms by which they exert their effects on lung function growth and development.

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## References

- Adcock IM, Barnes PJ. Molecular mechanisms of corticosteroid resistance. *Chest* 2008;134:394–401. [PubMed: 18682458]
- Akbari O, Stock P, Meyer E, Kronenberg M, Sidobre S, Nakayama T, Taniguchi M, Grusby MJ, DeKruyff RH, Umetsu DT. Essential role of NKT cells producing IL-4 and IL-13 in the development of allergen-induced airway hyperreactivity. *Nature Medicine* 2003;9(5):582–588.
- Alikhani-Koopaei R, Fouladkou F, Frey FJ, Frey BM. Epigenetic regulation of 11 beta-hydroxysteroid dehydrogenase type 2 expression. *Journal of Clinical Investigation* 2004;114:1146–1157. [PubMed: 15489962]
- Alkon A, Lippert S, Vujan N, Rodriguez ME, Boyce WT, Eskenazi B. The ontogeny of autonomic measures in 6- and 12-month-old infants. *Developmental Psychobiology* 2006;48(3):197–208. [PubMed: 16568414]
- Allam MH, Said AF, El Samie Omran AA, El-Reheim DMA, Kasem AH. High sensitivity C-reactive protein: Its correlation with sputum cell counts in bronchial asthma. *Respiratory Medicine* 2009;103:1878–1884. [PubMed: 19836939]
- Anisman H, Zaharia MD, Meaney MJ, Merali Z. Do early-life events permanently alter behavioral and hormonal responses to stressors? *International Journal of Developmental Neuroscience* 1998;16(3-4):149–164. [PubMed: 9785112]
- Anthracopoulos MB, Karatza AA, Davlouros PA, Chiladakis JA, Manolis AS. Effects of two nebulization regimens on heart rate variability during acute asthma exacerbations in children. *Journal of Asthma* 2005;42:273–279. [PubMed: 16032936]
- Arck PC, Knackstedt MK, Blois SM. Current insights and future perspectives on neuro-endocrine-immune circuitry challenging pregnancy maintenance and fetal health. *Journal Reproduktionsmed Endokrinol* 2006;3(2):98–102.
- Baccarelli A, Wright RO, Bollati V, Tarantini L, Litonjua AA, Suh HH, Zanobetti A, Sparrow D, Vokonas PS, Schwartz J. Rapid DNA methylation changes after exposure to traffic particles. *American Journal of Respiratory and Critical Care Medicine* 2009;179:572–578. [PubMed: 19136372]
- Barnes PJ. Airway inflammation and autonomic control. *European Journal of Respiratory Diseases* 1986;69:80–87.
- Barnes PJ. Neural mechanisms in asthma. *British Medical Bulletin* 1992;48(1):149–168. [PubMed: 1352167]
- Barnes PJ. Corticosteroid resistance in airway disease. *Proceedings of the American Thoracic Society* 2004;1:264–268. [PubMed: 16113444]
- Bavis RW, Mitchell GS. Long-term effects of the perinatal environment on respiratory control. *Journal of Applied Physiology* 2008;104:1220–1229. [PubMed: 18187608]
- Belsky J. The determinants of parenting: A process model. *Child Development* 1984;55:83–96. [PubMed: 6705636]
- Beydon N, Davis SD, Lombardi E, Allen JL, Arets HGM, Aurora P, Bisgaard H, Davis M, Ducharme FM, Eigen H, Gappa M, Gaultier C, Gustafsson PM, Hall GL, Hantos Z, Healy MJR, Jones MH, Klug B, Carlsen KCL, McKenzie SA, Marchal F, Mayer OH, Merkus PJFM, Morris MG,

- Oostveen E, Pillow JJ, Seddon PC, Silverman M, Sly PD, Stocks J, Tepper RS, Vilozni D, Wilson NM. An official American Thoracic Society/European Respiratory Society Statement: Pulmonary function testing in preschool children. *American Journal of Respiratory and Critical Care Medicine* 2007;175:1304–1345. [PubMed: 17545458]
- Bosquet-Enlow M, Kullowatz A, Staudenmayer J, Spasojevic J, Ritz T, Wright RJ. Associations of maternal lifetime trauma and perinatal traumatic stress symptoms with infant cardiorespiratory reactivity to psychological challenge. *Psychosomatic Medicine* 2009;71:607–14. [PubMed: 19553287]
- Boyce WT, Ellis BJ. Biological sensitivity to context: I. An evolutionary-developmental theory of the origins and functions of stress reactivity. *Developmental Psychopathology* 2005;17:271–301.
- Brussee JE, Smit HA, Koopman LP, Wijga AH, Kerkhof M, Corver K, Vos AP, Gerritsen J, Grobbee DE, Brunekreef B, Merkus PJ, de Jongste JC. Interrupter resistance and wheezing phenotypes at 4 years of age. *American Journal of Respiratory and Critical Care Medicine* 2004;169:209–213. [PubMed: 14597483]
- Buijs RM, van der Vliet J, Garidou ML, Huitinga I, Escobar C. Spleen vagal denervation inhibits the production of antibodies to circulating antigens. *PLoS ONE* 2008;3(9):e3152. [PubMed: 18773078]
- Bush A. How early do airway inflammation and remodeling occur? *Allerology International* 2008;57:11–19.
- Butts CL, Sternberg EM. Neuroendocrine factors alter host defense by modulating immune function. *Cellular Immunology* 2008;252(1-2):7–15. [PubMed: 18329009]
- Caldji C, Diorio J, Meaney MJ. Variations in maternal care in infancy regulate the development of stress reactivity. *Biological Psychiatry* 2000;48:1164–1174. [PubMed: 11137058]
- Card JP, Levitt P, Gluhovsky M, Rinaman L. Early experience modifies the postnatal assembly of autonomic emotional motor circuits in rats. *Journal of Neuroscience* 2005;25(40):9102–9111. [PubMed: 16207869]
- Castro M, Ramirez MI, Gern JE, Cutting G, Redding G, Hagood JS, Whitsett J, Abman S, Raj JU, Barst R, Kato GJ, Gozal D, Haddad GG, Prabhakar NR, Gauda E, Martinez FD, Tepper R, Wood RE, Accurso F, Teague WG, Venegas J, Cole FS, Wright RJ. Strategic plan for pediatric respiratory diseases research: an NHLBI working group report. *Proceedings of the American Thoracic Society* 2009;15:1–10. [PubMed: 19131525]
- Cicchetti D, Rogosch FA. Diverse patterns of neuroendocrine activity in maltreated children. *Developmental Psychopathology* 2001;13:677–693.
- Coe CL, Lubach GR, Karaszewski JW. Prenatal stress and immune recognition of self and nonself in the primate neonate. *Biology of the Neonate* 1999;76:301–310. [PubMed: 10516397]
- Cohen G, Vella S, Jeffery H, Lagercrantz H, Katz-Salamon M. Cardiovascular stress hyperreactivity in babies of smokers and babies born preterm. *Circulation* 2008;118:1848–1853. [PubMed: 18852367]
- Coplan J, Andrews MW, Rosenblum LA, Owens MJ, Friedman S, Gorman JM, Nemeroff CB. Persistent elevations of cerebrospinal fluid concentrations of corticotropin-releasing factor in adult nonhuman primates exposed to early-life stressors: Implications for the pathophysiology of mood and anxiety disorders. *Proceedings of the National Academy of Sciences USA* 1996;93:1619–1623.
- Cunningham JN, Kliever W, Garner PW. Emotion socialization, child emotion understanding and regulation, and adjustment in urban African American families: differential associations across child gender. *Developmental Psychopathology* 2009;21:261–83.
- Dahl M, Tybjaerg-Hansen A, Vestbo J, Lange P, Nordestgaard BG. Elevated plasma fibrinogen associated with reduced pulmonary function and increased risk of chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care Medicine* 2001;164:1008–1011. [PubMed: 11587987]
- Danson EJ, Li D, Wang I, Dawson TA, Paterson DJ. Targeting cardiac sympatho-vagal imbalance using gene transfer of nitric oxide synthase. *Journal of Molecular and Cellular Cardiology* 2009;46:482–489. [PubMed: 19166856]

- Datti F, Datti M, Antunes E, Teixeira NA. Influence of chronic unpredictable stress on the allergic responses in rats. *Physiology of Behavior* 77(1):79–83. 2000.
- De Kloet ER. Hormones and the stressed brain. *Annals of the New York Academy of Sciences* 2004;1018:1–15. [PubMed: 15240347]
- de Weerth C, Buitelaar JK. Physiological stress reactivity in human pregnancy – a review. *Neuroscience and Biobehavioral Reviews* 2005;29:295–312. [PubMed: 15811500]
- DeBellis MD, Baum AS, Birmaher B, Keshavan MS, Eccard CH, Boring AM, et al. Developmental traumatology, Part 1: Biological stress systems. *Biological Psychiatry* 1999;9:1259–1270.
- Devereux G, Barker RN, Seaton A. Antenatal determinants of neonatal immune responses to allergens. *Clinical and Experimental Allergy* 2002;32(1):43–50. [PubMed: 12002736]
- Devereux G, Seaton A, Barker RN. In utero priming of allergen-specific helper T cells. *Clinical and Experimental Allergy* 2001;31:1686–1695. [PubMed: 11696044]
- El-Sheikh M, Erath SA, Buckhalt JA, Granger DA, Mize J. Cortisol and children's adjustment: the moderating role of sympathetic nervous system activity. *Journal of Abnormal Child Psychology* 2008;36:601–611. [PubMed: 18197472]
- Elenkov IJ, Chrousos GP. Stress Hormones, Th1/Th2 patterns, Pro/Anti-inflammatory Cytokines and Susceptibility to Disease. *Trends in Endocrinology and Metabolism* 1999;10(9):359–368. [PubMed: 10511695]
- Elenkov IJ, Iezzoni DG, Daly A, Harris AG, Chrousos GP. Cytokine dysregulation, inflammation and well-being. *Neuroimmunomodulation* 2005;12:255–269. [PubMed: 16166805]
- Elenkov IJ, Kvetnansky R, Hashiramoto A, Bakalov VK, Link AA, Zachman K, Crane M, Jezova D, Rovensky J, Dimitrov MA, Gold PW, Bonini S, Fleisher T, Chrousos GP, Wilder RL. Low- versus high-baseline epinephrine output shapes opposite innate cytokine profiles: presence of Lewis- and Fischer-like neurohormonal immune phenotypes in humans? *Journal of Immunology* 2008;181:1737–1745.
- Esposito E, Cuzzocrea S. The role of nitric oxide synthases in lung inflammation. *Current Opinion in Investigative Drugs* 2007;8:899–909.
- Essex MJ, Klein MH, Cho E, Kalin NH. Maternal stress beginning in infancy may sensitize children to later stress exposure: effects on cortisol and behavior. *Biological Psychiatry* 2002;52(8):776–784. [PubMed: 12372649]
- Feinberg AP. Phenotypic plasticity and the epigenetics of human disease. *Nature* 2007;447:433–440. [PubMed: 17522677]
- Field, T. The effects of mother's physical and emotional unavailability on emotions regulation. Vol. 208-227. University of Chicago Press; Chicago: 1994.
- Forsythe P, Ebeling C, Gordon JR, Befus AD, Vliagoftis H. Opposing effects of short- and long-term stress on airway inflammation. *American Journal of Respiratory and Critical Care Medicine* 2004;169:220–226. [PubMed: 14604839]
- Francis DD, Caldji C, Champagne F, Plotsky PM, Meaney MJ. The role of corticotropin-releasing factor-norepinephrine systems in mediating the effects of early experience on the development of behavioral and endocrine responses to stress. *Biological Psychiatry* 1999;46:1153–1166. [PubMed: 10560022]
- Franco Suglia S, Ryan L, Laden F, Dockery D, Wright RJ. Violence exposure, a chronic psychosocial stressor, and childhood lung function. *Psychosomatic Medicine* 2007;70:160–169. [PubMed: 18158365]
- Fryer AD, Jacoby DB. Muscarinic receptors and control of airway smooth muscle. *American Journal of Respiratory and Critical Care Medicine* 1998;158(5 Pt 3):S154–160. [PubMed: 9817739]
- Ganalingham MG, Mostyn A, Gardner DS, Stephenson T, Symonds TE. Developmental regulation of the lung in preparation for life after birth: hormonal and nutritional manipulation of local glucocorticoid action and uncoupling protein-2. *Journal of Endocrinology* 2006;188:375–386. [PubMed: 16522718]
- Garn H, Renz H. Epidemiological and immunological evidence for the hygiene hypothesis. *Immunobiology* 2007;212:441–452. [PubMed: 17544829]
- Gern JE. Viral and bacterial infections in the development and progression of asthma. *Journal of Allergy and Clinical Immunology* 2000;105:S497–502. [PubMed: 10669531]

- Gern JE, Rosenthal LA, Sorkness RL, Lemanske RF Jr. Effects of viral respiratory infections on lung development and childhood asthma. *Journal of Allergy and Clinical Immunology* 2005;15(4):668–674. [PubMed: 15805982]
- Gluckman PD, Hanson MA, Cooper C, Thornburg KL. Effect of in utero and early-life conditions on adult health and disease. *New England Journal of Medicine* 2008;359:61–73. [PubMed: 18596274]
- Goland RS, Jozak S, Warren WB, Conwell IM, Stark RI, Tropper PJ. Elevated levels of umbilical cord plasma corticotropin-releasing hormone in growth-related fetuses. *Journal of Clinical Endocrinology and Metabolism* 1993;77:1174–1179. [PubMed: 8077309]
- Gold DR, Wright R. Population disparities in asthma. *Annual Review of Public Health* 2005;26:89–113.
- Gordis EB, Granger DA, Susman EJ, Trickett PK. Asymmetry between salivary cortisol and alpha-amylase reactivity to stress: relation to aggressive behavior in adolescents. *Psychoneuroendocrinology* 2006;31:976–987. [PubMed: 16879926]
- Grier DG, Halliday HL. Effects of glucocorticoids on fetal and neonatal lung development. *Treatments in Respiratory Medicine* 2004;3:295–306. [PubMed: 15606220]
- Gunnar MR, Bruce J, Hickman SE. Salivary cortisol response to stress in children. *Advances in Psychosomatic Medicine* 2001;22:52–60. [PubMed: 11477939]
- Gunnar MR, Donzella B. Social regulation of the cortisol levels in early human development. *Psychoneuroendocrinology* 2002;27:199–220. [PubMed: 11750779]
- Hanrahan JP, Tager IB, Segal MR, Tosteson TD, Castile RG, Van Vunakis H, Weiss ST, Speizer FE. The effect of maternal smoking during pregnancy on early infant lung function. *American Review of Respiratory Disease* 1992;145:1129–1135. [PubMed: 1586058]
- Hansen G, Berry G, DeKruyff RH, Umetsu DT. Allergen-specific Th1 cells fail to counterbalance Th2 cell-induced airway hyperreactivity but cause severe airway inflammation. *Journal of Clinical Investigation* 1999;103:175–183. [PubMed: 9916129]
- Harding R, Cock ML, Louey S, Joyce BJ, Davey MG, Albuquerque CA, Hooper SB, Maritz GS. The compromised intra-uterine environment: implications for future lung health. *Clinical and Experimental Pharmacology and Physiology* 2000;27:965–974. [PubMed: 11117232]
- Harrison OJ, Foley J, Bolognese BJ, Long E. r. Podolin PL, Walsh PT. Airway infiltration of CD4+CCR6+ Th17 type cells associated with chronic cigarette smoke induced airspace enlargement. *Immunology Letters*. 2008 Epub ahead of print.
- Hatzinger M, Brand S, Perren S, von Wyl A, von Klitzing K, Holsboer-Trachsler E. Hypothalamic-pituitary-adrenocortical (HPA) activity in kindergarten children: importance of gender and associations with behavioral/emotional difficulties. *Journal of Psychiatric Research* 2007;41:861–870. [PubMed: 16979188]
- Heaton T, Rowe J, Turner S, Aalberse RC, de Klerk N, Suriyaarachchi D, Serralha M, Holt BJ, Hollams E, Yerkovich S, Holt K, Sly PD, Goldblatt J, Le Souef P, Holt PG. An immunoepidemiological approach to asthma: identification of in-vitro T-cell response patterns associated with different wheezing phenotypes in children. *Lancet* 2005;365:142–149.
- Heim C, Newport DJ, Heit S, Graham YP, Wilcox M, Bonsall R, Miller AH, Nemeroff CB. Pituitary-adrenal and autonomic responses to stress in women after sexual and physical abuse in childhood. *Journal of the American Medical Association* 2000;284:592–597. [PubMed: 10918705]
- Herlenius E, Lagercrantz H. Development of neurotransmitter systems during critical periods. *Experimental Neurology* 2004;190(Suppl. 1):S8–21. [PubMed: 15498537]
- Hessl, D.; Dawson, G.; Frey, K.; Panagiotides, H.; Self, H.; Yamada, E., et al. A longitudinal study of children of depressed mothers: Psychobiological findings related to stress. In: Hann, DM.; Huffman, LC.; Lederhendler, KK.; Minecke, D., editors. *Advancing Research on Developmental Plasticity: Integrating the Behavioral Sciences and the Neurosciences of Mental Health*. National Institutes of Mental Health; Bethesda, MD: 1998. p. 256
- Hoffmeyer F, Raulf-Heimsoth M, Bruning T. Exhaled breath condensate and airway inflammation. *Current Opinion in Allergy and Clinical Immunology* 2009;9:16–22. [PubMed: 19532089]
- Holgate, ST. Asthma: a dynamic disease of inflammation and repair. In: Chadwick, DJ.; Cardew, G., editors. *The Rising Trends in Asthma*. John Wiley & Sons; West Sussex, England: 1997. p. 5-34.



- Holt PG, Upham JW, Sly PD. Contemporaneous maturation of immunologic and respiratory functions during early childhood: implications for development of asthma prevention strategies. *Journal of Allergy and Clinical Immunology* 2005;116:16–24. [PubMed: 15990766]
- Igosheva N, Klimova O, Anishchenko T, Glover V. Prenatal stress alters cardiovascular responses in adult rats. *Journal of Physiology* 2004;557(Pt 1):273–285. [PubMed: 15034122]
- Isenberg SA, Lehrer PM, Hochron S. The effects of suggestion and emotional arousal on pulmonary function in asthma: a review and hypothesis regarding vagal mediation. *Psychosomatic Medicine* 1992;54:192–216. [PubMed: 1565756]
- Ito K, Herbert C, Diegle JS, Vuppusetty C, Hansbro N, Thomas PS, Foster PS, Barnes PJ, Kumar RK. Steroid-resistant neutrophilic inflammation in a mouse model of an acute exacerbation of asthma. *American Journal of Respiratory Cellular and Molecular Biology* 2008;39:543–550.
- Jackson B, Kubzansky LD, Cohen S, Jacobs DR Jr, Wright RJ. Does harboring hostility hurt? Associations between hostility and pulmonary function in the Coronary Artery Risk Development in (Young) Adults (CARDIA) study. *Health Psychology* 2007;26:333–340. [PubMed: 17500620]
- Jackson B, Kubzansky LD, Cohen S, Weiss S, Wright RJ. A matter of life and breath: childhood socioeconomic status is related to young adult pulmonary function in the CARDIA study. *International Journal of Epidemiology* 2004a;33:271–278. [PubMed: 15082626]
- Jackson B, Wright RJ, Kubzansky LD, Weiss ST. Examining the influence of early life socioeconomic position on pulmonary function across the life span: where do we go from here? *Thorax* 2004b;59:186–188. [PubMed: 14985547]
- Jaenisch R, Bird A. Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nature Genetics* 2003;35:245–254. [PubMed: 12610534]
- Jansson T, Lambert GW. Effect of intrauterine growth restriction on blood pressure, glucose tolerance and sympathetic nervous system activity in the rat at 3–4 months of age. *Journal of Hypertension* 1999;17:1239–1248. [PubMed: 10489100]
- Joachim RA, Cifuentes LB, Sagach V, Quarcoo D, Hagen E, Arck PC, Fischer A, Klapp BF, Dinh QT. Stress induces substance P in vagal sensory neurons innervating the mouse airways. *Clinical and Experimental Allergy* 2006;36:1001–1010. [PubMed: 16911356]
- Joachim RA, Handjiski B, Blois SM, Hagen E, Paus R, Arck PC. Stress-induced neurogenic inflammation in murine skin skews dendritic cells towards maturation and migration. Key role of intercellular adhesion molecule-1/leukocyte function-associated antigen interactions. *American Journal of Pathology*. 2008 Epub ahead of print.
- Joachim RA, Noga O, Sagach V, Hanf G, Fliege H, Kocalevent RD, Peters EM, Klapp BF. Correlation between immune and neuronal parameters and stress perception in allergic asthmatics. *Clinical and Experimental Allergy* 2008;38:283–290. [PubMed: 18070153]
- Joachim RA, Quarcoo D, Arck PC, Herz U, Renz H, Klapp BF. Stress enhances airway reactivity and airway inflammation in an animal model of allergic bronchial asthma. *Psychosomatic Medicine* 2003;65:811–815. [PubMed: 14508025]
- Joachim RA, Sagach V, Quarcoo D, Dinh QT, Arck PC, Klapp BF. Neurokinin-1 receptor mediates stress-exacerbated allergic airway inflammation and airway hyperresponsiveness in mice. *Psychosomatic Medicine* 2004;66:564–571. [PubMed: 15272104]
- Kajekar R. Environmental factors and developmental outcomes in the lung. *Pharmacological Therapy* 2007;114:129–145.
- Kajekar R, Pieczarka EM, Smiley-Jewell SM, Schelegle ES, Fanucchi MV, Plopper CG. Early postnatal exposure to allergen and ozone leads to hyperinnervation of the pulmonary epithelium. *Respiratory Physiology and Neurobiology* 2007;155:55–63. [PubMed: 16616710]
- Kaufman J, Birmaher B, Perel J, Dahl RE, Moreci P, Nelson B, et al. The corticotropin-releasing hormone challenge in depressed abused, depressed nonabused, and normal control children. *Biological Psychiatry* 1997;42:669–679. [PubMed: 9325560]
- Kazuma N, Otsuka K, Miyakawa M, Shirase E, Matsuoka I, Muarta M. Seasonal variation in heart rate variability in asthmatic children. *Chronobiology International* 2000;17:503–511. [PubMed: 10908126]

- Kim EY, Battaile JT, Patel AC, You Y, Agapov E, Grayson MH, et al. Persistent activation of an innate immune response translates respiratory viral infection into chronic lung disease. *Nature Medicine* 2008;14:633–640.
- Kivlighan KT, DiPietro JA, Costigan KA, Laudenslager ML. Diurnal rhythm of cortisol during late pregnancy: associated with maternal psychological well-being and fetal growth. *Psychoneuroendocrinology* 2008;33:1225–1235. [PubMed: 18692319]
- Krozowski ZS, Provencher PH, Smith RE, Obeyesekere VR, Mercer WR, Albiston AL. Isozymes of 11 beta-hydroxysteroid dehydrogenase: which enzyme endows mineralocorticoid specificity? *Steroids* 1994;59:116–120. [PubMed: 8191539]
- Kubzansky LD, Sparrow D, Jackson B, Cohen S, Weiss ST, Wright RJ. Angry breathing: a prospective study of hostility and lung function in the Normative Aging Study. *Thorax* 2006;61:863–868. [PubMed: 16950835]
- Kubzansky LD, Wright RJ, Cohen S, Weiss ST, Rosner B, Sparrow D. Breathing Easy: A prospective study of optimism and pulmonary function in the Normative Aging Study. *Annals of Behavioral Medicine* 2002;24:345–353. [PubMed: 12434946]
- LeMay LG, Vander AJ, Kluger MJ. The effects of pentoxifylline on lipopolysaccharide (LPS) fever, plasma interleukin 6 (IL 6), and tumor necrosis factor (TNF) in the rat. *Cytokine* 1990;2:300–306. [PubMed: 2104230]
- Levenson JM, Sweatt JD. Epigenetic mechanisms: a common theme in vertebrate and invertebrate memory formation. *Cellular and Molecular Life Science* 2006;63:1009–1016.
- Li H, Burkhardt C, Heinrich UR, Brausch I, Xia N, Forstermann U. Histamine upregulates gene expression of endothelial nitric oxide synthase in human vascular endothelial cells. *Circulation* 2003;107:2348–2354. [PubMed: 12707234]
- Liao SY, Liao TN, Chiang BL, Huang MS, Chen CC, Chou CC, Hsieh KH. Decreased production of IFN gamma and increased production of IL-6 by cord blood mononuclear cells of newborns with a high risk of allergy. *Clinical and Experimental Allergy* 1996;26:397–405. [PubMed: 8732236]
- Liu D, Diorio J, Tannenbaum B, Caldji C, Francis DJ, Freedman A, et al. Maternal care, hippocampal glucocorticoid receptors, and hypothalamic-pituitary-adrenal response to stress. *Science* 1997;277:1659–1662. [PubMed: 9287218]
- Lowe LA, Simpson A, Woodcock A, Morris JC, Murray CS, Custovic A. Wheeze phenotypes and lung function in preschool children. *American Journal of Respiratory and Critical Care Medicine* 2009;171:231–237. [PubMed: 15502115]
- Lutgendorf SK, Moore MB, Bradley S, Shelton BJ, Lutz CT. Distress and expression of natural killer receptors on lymphocytes. *Brain Behavior and Immunity* 2005;19:185–194.
- Lutz W, Sulkowski WJ. Vagus nerve participates in regulation of the airways: inflammatory response and hyperreactivity induced by occupational asthmogens. *International Journal of Occupational Medicine and Environmental Health* 2004;17:417–431. [PubMed: 15852756]
- Lyons-Ruth K, Block DE. The disturbed caregiving system: Relations among childhood trauma, maternal caregiving, and infant affect and attachment. *Infant Mental Health Journal* 1996;17:257–275.
- Mannino DM, Braman S. The epidemiology and economics of chronic obstructive pulmonary disease. *Proceedings of the American Thoracic Society* 2007;4:502–506. [PubMed: 17878461]
- Martin JA, Kung HC, Matthews TJ, Hoyert DL, Strobino DM, Guyer B, et al. Annual summary of vital statistics. *Pediatrics* 2008;121:788–801. [PubMed: 18381544]
- Marwick JA, Wallis G, Meja K, Kuster B, Bouwmeester T, Chakravarty P, et al. Oxidative stress modulates theophylline effects on steroid responsiveness. *Biochemistry Biophysics Research Communications* 2008;377:797–802.
- Martinez FD, Wright AL, Taussig LM, Holberg CJ, Halonen M, Morgan WJ. Asthma and wheezing in the first six years of life. *New England Journal of Medicine* 1995;332:133–138. [PubMed: 7800004]
- McEwen BS. Protective and damaging effects of stress mediators: the good and bad sides of the response to stress. *Metabolism: Clinical and Experimental* 2002;51(6 Suppl 1):2–4. [PubMed: 12040533]

- McGowan PO, Sasaki A, D'Alessio AC, Dymov S, Labonte B, Szyf M, et al. Epigenetic regulation of the glucocorticoid receptor in human brain associated with childhood abuse. *Nature Neuroscience* 2009;12:342–348.
- McKinley L, Alcorn JF, Peterson A, DuPont RB, Kapadia S, Logar A, et al. Th17 cells mediate steroid-resistant airway inflammation and airway hyperresponsiveness in mice. *Journal of Immunology* 2008;181:4089–4097.
- McLeod TM, Lopez-Figueroa AL, Lopez-Figueroa MO. Nitric oxide, stress, and depression. *Psychopharmacology Bulletin* 2001;35:24–41. [PubMed: 12397868]
- Meaney MJ, Szyf M. Maternal care as a model for experience-dependent chromatin plasticity? *Trends in Neuroscience* 2005;28:456–463.
- Miles EA, Warner JA, Jones AC, Colwell BM, Bryant TN, Warner JO. Peripheral blood mononuclear cell proliferative responses in the first year of life in babies born to allergic parents. *Clinical and Experimental Allergy* 1996;26:780–788. [PubMed: 8842551]
- Miller BD, Wood BL. Influence of specific emotional states on autonomic reactivity and pulmonary function in asthmatic children. *Journal American Academy Child Adolescent Psychiatry* 1997;36:669–677.
- Miller BD, Wood BL. Emotions and family factors in childhood asthma: psychobiologic mechanisms and pathways of effect. *Advances in Psychosomatic Medicine* 2003;24:131–160. [PubMed: 14584352]
- Miller GE, Chen E, Zhou ES. If it goes up, must it come down? Chronic stress and the hypothalamic-pituitary-adrenal axis in humans. *Psychological Bulletin* 2007;133:25–45. [PubMed: 17201569]
- Miller GE, Cohen S, Ritchey AK. Chronic Psychological Stress and the Regulation of Pro-Inflammatory Cytokines: A Glucocorticoid-Resistance Model. *Health Psychology* 2002;21:531–541. [PubMed: 12433005]
- Miller GE, Gaudin A, Zysk E, Chen E. Parental support and cytokine activity in childhood asthma: the role of glucocorticoid sensitivity. *Journal of Allergy and Clinical Immunology* 2009;123:824–830. [PubMed: 19181373]
- Miller RL, Ho SM. Environmental epigenetics and asthma: current concepts and call for studies. *American Journal of Respiratory and Critical Care Medicine* 2008;177:567–573. [PubMed: 18187692]
- Mitchell GS, Johnson SM. Neuroplasticity in respiratory motor control. *Journal of Applied Physiology* 2003;94:358–374. [PubMed: 12486024]
- Miossec P, Korn T, Kuchroo VK. Interleukin-17 and Type 17 Helper T Cells. *New England Journal of Medicine* 2009;361:888–898. [PubMed: 19710487]
- Montuschi P. Analysis of exhaled breath condensate in respiratory medicine: methodological aspects and potential clinical applications. *Therapeutic Advances in Respiratory Disease* 2007;1:5–23. [PubMed: 19124344]
- Morgan WJ, Stern DA, Sherrill DL, Guerra S, Holberg CJ, Guilbert TW, et al. Outcome of asthma and wheezing in the first 6 years of life: follow-up through adolescence. *American Journal of Respiratory and Critical Care Medicine* 2005;172:1253–1258. [PubMed: 16109980]
- Morris AS, Silk JS, Steinberg L, Myers SS, Robinson LR. The role of family context in the development of emotion regulation. *Social Development* 2007;16:361–388. [PubMed: 19756175]
- Mossman TR, Coffman RL. Th1 and Th2 cells: different patterns of lymphokine secretion lead to different functional properties. *Annual Reviews in Immunology* 1989;7:145–173.
- Mossman TR, Sad S. The expanding universe of T-cell subsets: Th1, Th2 and more. *Immunology Today* 1996;17:138–146. [PubMed: 8820272]
- Nakano M, Onozuka K, Yamasu H, Zhong WF, Nakano Y. Protective effects of cytokines in murine Salmonella. *Advances in Experimental Medicine Biology* 1992;319:89–95.
- Navratil M, Plavec D, Dodig S, Jelcic Z, Nogalo B, Erceg D, Turkaj M. Markers of systemic and lung inflammation in childhood asthma. *Journal of Asthma* 2009;46:822–888. [PubMed: 19863287]
- Nockher WA, Renz H. Neurotrophins and asthma: novel insight into neuroimmune interaction. *Journal of Allergy and Clinical Immunology* 2006;117:67–71. [PubMed: 16387586]

- Nogueira PJ, Ferreira HHA, Antunes E, Teixeira NA. Chronic mild prenatal stress exacerbates the allergen-induced airway inflammation in rats. *Mediators of Inflammation* 1999;8:119–122. [PubMed: 10704150]
- O'Connor GT, Weiss ST, Tager IB, Speizer FE. The effect of passive smoking on pulmonary function and nonspecific bronchial responsiveness in a population based sample of children and young adults. *American Review of Respiratory Disease* 1987;135:800–804. [PubMed: 3565928]
- Oberlander TF, Weinberg J, Papsdorf M, Grunau R, Misri S, Devlin AM. Prenatal exposure to maternal depression, neonatal methylation of human glucocorticoid receptor gene (NR3C1) and infant cortisol stress responses. *Epigenetics* 2008;3:97–106. [PubMed: 18536531]
- Okuyama K, Ohwada K, Sakurada S, Sato N, Sora I, Tamura G, et al. The distinctive effects of acute and chronic psychological stress on airway inflammation in a murine model of allergic asthma. *Allergology International* 2007;56:29–35. [PubMed: 17259807]
- Ownby DR, Johnson CC, Peterson EL. Exposure to dogs and cats in the first year of life and risk of allergic sensitization at 6 to 7 years of age. *Journal of the American Medical Association* 2002;288:963–972. [PubMed: 12190366]
- Oya H, Kawamura T, Shimizu T, Bannai M, Kawamura H, Minagawa M, et al. The differential effect of stress on natural killer T (NKT) and NK cell function. *Clinical and Experimental Immunology* 2000;121:384–390. [PubMed: 10931157]
- Peters JM, Franco Suglia S, Platts-Mills TAE, Hosen J, Wright RJ. Psychological stress modifies the influence of prenatal allergen exposure on cord blood IgE: The Boston ACCESS project. *American Journal of Respiratory and Critical Care Medicine* 2008;177(Abstracts Issue):A231.
- Pichavant M, Goya S, Meyer EH, Johnston RA, Kim HY, Matangkasombut P, et al. Ozone exposure in a mouse model induces airway hyperreactivity that requires the presence of natural killer T cells and IL-17. *Journal of Experimental Medicine* 2008;205:385–393. [PubMed: 18250191]
- Pincus-Knackstedt MK, Joachim RA, Blois SM, Douglas AJ, Orsal AS, Klapp BF, et al. Prenatal stress enhances susceptibility of murine adult offspring toward airway inflammation. *Journal of Immunology* 2006;177:8484–8492.
- Pinkerton KE, Joad JP. Influence of air pollution on respiratory health during perinatal development. *Clinical and Experimental Pharmacology and Physiology* 2006;33:269–272. [PubMed: 16487273]
- Pohunek P, Warner JO, Turzikova J, Kudrman J, Roche WR. Markers of eosinophilic inflammation and tissue re-modeling in children before clinically diagnosed bronchial asthma. *Pediatric Allergy and Immunology* 2005;16:43–51. [PubMed: 15693911]
- Powell ND, Bailey MT, Mays JW, Stiner-Jones LM, Hanke ML, Padgett DA, et al. Repeated social defeat activates dendritic cells and enhances Toll-like receptor dependent cytokine secretion. *Brain Behavior and Immunity* 2009;23:225–231.
- Prescott SL. The development of respiratory inflammation in children. *Paediatric Respiratory Reviews* 2006;7:89–96. [PubMed: 16765293]
- Prescott SL, Macaubas C, Holt BJ, Smallacombe TB, Loh R, Sly PD, et al. Transplacental priming of the human immune system to environmental allergens: universal skewing of initial T cell responses toward the Th2 cytokine profile. *Journal of Immunology* 1998;160:4730–4737.
- Propper C, Moore GA, Mills-Koonce WR, Halpern CT, Hill-Soderlund AL, Calkins SD, et al. Gene-environment contributions to the development of infant vagal reactivity: the interaction of dopamine and maternal sensitivity. *Child Development* 2008;79:1377–1394. [PubMed: 18826531]
- Pryce CR, Ruedi-Bettschen D, Dettling AC, Feldon J. Early life stress: long-term physiological impact in rodents and primates. *News Physiological Science* 2002;17:150–155.
- Quarcoo D, Pavlovic S, Joachim RA. Stress and airway reactivity in a murine model of allergic airway inflammation. *Neuroimmunomodulation* 2009;16:318–324. [PubMed: 19571592]
- Rangasamy T, Guo J, Mitzner WA, Roman J, Singh A, Fryer AD, et al. Disruption of Nrf2 enhances susceptibility to severe airway inflammation and asthma in mice. *Journal of Experimental Medicine* 2005;202:47–59. [PubMed: 15998787]

- Rasmussen R, Mikkelsen D, Hancox RJ, Lambrechtsen J, Nybo M, Hansen HS, Stersted HC. High-sensitivity C-reactive protein is associated with reduced lung function in young adults. *European Respiratory Journal* 2009;33:382–388. [PubMed: 19010993]
- Rasmussen F, Taylor DR, Flannery EM, Cowan JO, Greene JM, Herbison GP, Sears MR. Risk factors for airway remodeling in asthma manifested by a low postbronchodilator FEV1/vital capacity ratio: a longitudinal population study from childhood to adulthood. *American Journal of Respiratory and Critical Care Medicine* 2002;165:1480–1488. [PubMed: 12045120]
- Reinisch JM, Simon NG, Karwo WG, Gandelman R. Prenatal exposure to prednisone in humans and animals retards intra-uterine growth. *Science* 1978;202:436–438. [PubMed: 705336]
- Ridker PM, Cushman M, Stampfer MJ, Tracy RP, Hennekens CH. Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. *New England Journal of Medicine* 1997;336:973–979. [PubMed: 9077376]
- Ritz T. Probing the psychophysiology of the airways. Physical activity, experienced emotion, and facially expressed emotion. *Psychophysiology* 2004;41:809–821. [PubMed: 15563334]
- Ritz T, Kullowatz A. Effects of stress and emotion on lung function in health and asthma. *Current Respiratory Medicine Review* 2005;1:208–219.
- Ritz T, Steptoe A. Emotion and pulmonary function in asthma: reactivity in the field and relationship with laboratory induction of emotion. *Psychosomatic Medicine* 2000;62:808–815. [PubMed: 11139001]
- Robinson DS, Hamid Q, Ying S, et al. Predominant Th2 like bronchoalveolar T-lymphocyte population in atopic asthma. *New England Journal of Medicine* 1992;326:298–304. [PubMed: 1530827]
- Sagiyama K, Tsuchida M, Kawamura H, Wang S, Li C, Bai X, et al. Age-related bias in function of natural killer T cells and granulocytes after stress: reciprocal association of steroid hormones and sympathetic nerves. *Clinical and Experimental Immunology* 2004;135:56–63. [PubMed: 14678265]
- Schulte-Herbruggen O, Nassenstein C, Lommatzsch M, Quarcoo D, Renz H, Braun A. Tumor necrosis factor-[alpha] and interleukin-6 regulate secretion of brain-derived neurotrophic factor in human monocytes. *Journal Neuroimmunology* 2005;160:204–209.
- Seckl J. Glucocorticoids, feto-plaental 11-beta-hydroxysteroid dehydrogenase type 2, and the early life origins of adult disease. *Steroids* 1997;62:89–94. [PubMed: 9029721]
- Seckl JR. Glucocorticoid programming of the fetus: adult phenotypes and molecular mechanisms. *Molecular and Cellular Endocrinology* 2001;185:61–71. [PubMed: 11738795]
- Shaheen S. The beginnings of chronic airflow obstruction. *British Medical Bulletin* 1997;53:58–70. [PubMed: 9158284]
- Sin DD, Man SF. Fueling the fire - systemic inflammation and development of lung disease in the general community. *International Journal of Epidemiology* 2006;35:1008–1010. [PubMed: 16641128]
- Stern DA, Morgan WJ, Wright AL, Guerra S, Martinez FD. Poor airway function in early infancy and lung function by age 22 years: A non-selective longitudinal cohort study. *Lancet* 2007;3770:758–764. [PubMed: 17765525]
- Sternberg EM. Neural-immune interactions in health and disease. *Journal of Clinical Investigation* 1997;100:2641–2647. [PubMed: 9389725]
- Sternberg EM. Neuroendocrine regulation of autoimmune/inflammatory disease. *Journal of Endocrinology* 2001;169:429–435. [PubMed: 11375112]
- Sternberg EM. Neural regulation of innate immunity: a coordinated nonspecific host response to pathogens. *Nature Reviews in Immunology* 2006;6:318–328.
- Sternthal MJ, Bosquet Enlow M, Cohen S, Jacobson Canner M, Staudenmayer J, Tsang K, Wright RJ. Maternal interpersonal trauma and cord blood IgE in an inner-city cohort: a life course perspective. *Journal of Allergy and Clinical Immunology* 2009;124:954–60. [PubMed: 19748657]
- Stock P, Akbari O, Berry G, Freeman GJ, Dekruyff RH, Umetsu DT. Induction of T helper type 1-like regulatory cells that express Foxp3 and protect against airway hyper-reactivity. *Nature Immunology* 2004;5:1149–1156. [PubMed: 15448689]

- Suarez CJ, Parker NJ, Finn PW. Innate immune mechanism in allergic asthma. *Current Allergy and Asthma Reports* 2008;8:451–459. [PubMed: 18682113]
- Sunyer J, Pistelli R, Plana E, Andreani M, Baldari F, Kotz M, Koenig W, Peddannen J, Peters A, Forastier F. Systemic inflammation, genetic susceptibility and lung function. *European Respiratory Journal* 2008;32:92–97. [PubMed: 18385179]
- Sutherland TJ, Taylor DR, Sears MR, Cowan JO, McLachlan CR, Filsell S, Williamson A, Greene JM, Poulton R, Hancox RJ. Association between exhaled nitric oxide and systemic inflammatory markers. *Annals of Allergy Asthma and Immunology* 2007;99:534–539.
- Szyf M, McGowan P, Meaney MJ. The social environment and the epigenome. *Environmental and Molecular Mutagenesis* 2008;49:46–60. [PubMed: 18095330]
- Takamura M, Matsumoto H, Nimi A, Ueda T, Matsuoka H, Yamaguchi M, Jinnai M, Muro S, Hirai T, Ito Y, Nakamura T, Mio T, Chin K, Mishima M. High sensitivity C-reactive protein in asthma. *European Respiratory Journal* 2006;27:908–912. [PubMed: 16707391]
- Talge NM, Donzella B, Gunnar MR. Fearful temperament and stress reactivity among preschool-aged children. *Infant and Child Development* 2008;17:427–445. [PubMed: 19122850]
- Tang MLK, Kemp AS, Thorburn J, Hill DJ. Reduced interferon-gamma secretion in neonates and subsequent atopy. *Lancet* 1994;344:983–985. [PubMed: 7934430]
- Tarantini L, Bonzini M, Apostoli P, Pegoraro V, Bollati V, Marinelli B, et al. Effects of particulate matter on genomic DNA methylation content and iNOS promoter methylation. *Environmental Health Perspectives* 2009;117(2):217–222. [PubMed: 19270791]
- Taussig LM, Wright AL, Holberg CJ, Halonen M, Morgan WJ, Martinez FD. Tucson Children's Respiratory Study: 1980 to present. *Journal of Allergy and Clinical Immunology* 2003;111:661–675. [PubMed: 12704342]
- Tracey KJ. The inflammatory reflex. *Nature* 2002;420(19 December):853–859. [PubMed: 12490958]
- Turato G, Barbato A, Baraldo S, Zanin ME, Bazzan E, Lokar-Oliani K, et al. Nonatopic children with multitrigger wheezing have airway pathology comparable to atopic asthma. *American Journal of Respiratory and Critical Care Medicine* 2008;178:476–482. [PubMed: 18511700]
- Turner JD, Pelascini LP, Macedo JA, Muller CP. Highly individual methylation patterns of alternative glucocorticoid receptor promoters suggest individualized epigenetic regulatory mechanisms. *Nucleic Acids Research* 2008;36:7207–7218. [PubMed: 19004867]
- Umetsu DT, McIntire JJ, Akbari O, Macaubas C, DeKruyff RH. Asthma: an epidemic of dysregulated immunity. *Nature Immunology* 2002;3(8):715–720. [PubMed: 12145657]
- Udem, BJ.; Weinreich, D. Neuroimmune interactions in the lung. In: Bienenstock, J.; Goetzle, EJ.; Blennerhassett, MG., editors. *Autonomic neuroimmunology*. Taylor & Francis; New York: 2003. p. 279-294.
- Vallee M, Mayo W, Dellu F, Le Moal M, Simon H, Maccari S. Prenatal stress induces high anxiety and postnatal handling induces low anxiety in adult offspring: correlation with stress-induced corticosterone secretion. *Journal of Neuroscience* 1997;17:2626–2636. [PubMed: 9065522]
- van den Toorn LM, Overbeek SE, de Jongste JC, Leman K, Hoogsteden HC, Prins JB. Airway inflammation is present during clinical remission of atopic asthma. *American Journal of Respiratory and Critical Care Medicine* 2001;164:2107–2113. [PubMed: 11739143]
- Veres TZ, Rochlitzer S, Shevchenko M, Fuchs B, Prenzler F, Nassenstein C, et al. Spatial interactions between dendritic cells and sensory nerves in allergic airway inflammation. *American Journal of Respiratory Cellular and Molecular Biology* 2007;37:553–561.
- von Hertzen LC. Maternal stress and T-cell differentiation of the developing immune system: possible implications for the development of asthma and atopy. *Journal of Allergy and Clinical Immunology* 2002;109(6):923–928. [PubMed: 12063519]
- von Mutius E. Childhood experiences take away your breath as a young adult. *American Journal of Respiratory and Critical Care Medicine* 2002;165:1467–1468. [PubMed: 12045117]
- Wang Y, McCusker C. Neonatal exposure with LPS and/or allergen prevents experimental allergic airways disease: development of tolerance using environmental antigens. *Journal of Allergy and Clinical Immunology* 2006;118:143–151. [PubMed: 16815150]
- Weaver IC, Cervoni N, Champagne FA, D'Alessio AC, Sharma S, Seckl JR, et al. Epigenetic programming by maternal behavior. *Nature Neuroscience* 2004;7:847–854.

- Weaver IC, Szyf M, Meaney MJ. From maternal care to gene expression: DNA methylation and the maternal programming of stress responses. *Endocrine Research* 2002;28:699. [PubMed: 12530685]
- Wenzel SE, Busse WW. Severe asthma: lessons from the Severe Asthma Research Program. *Journal of Allergy and Clinical Immunology* 2007;119:14–21. quiz 22-13. [PubMed: 17208583]
- Wenzel SE, Schwartz LB, Langmack EL, Haliday JL, Trudeau JB, Gibbs RL, et al. Evidence that severe asthma can be divided pathologically into two inflammatory subtypes with distinct physiologic and clinical characteristics. *American Journal of Respiratory and Critical Care Medicine* 1999;160:1001–1008. [PubMed: 10471631]
- Wignarajah D, Cock ML, Pinkerton KE, Harding R. Influence of intrauterine growth restriction on airway development in fetal and postnatal lung. *Pediatric Research* 2002;51:681–688. [PubMed: 12032261]
- Wills-Karp M. Immunologic basis of antigen-induced airway hyperresponsiveness. *Annual Review in Immunology* 1999;17:255–281.
- Willwerth BM, Schaub B, Tantisira KG, Gold DR, Palmer LJ, Litonjua AA, et al. Prenatal, perinatal, and heritable influences on cord blood immune responses 2006;96:445–453.
- Wonnacott KM, Bonneau RH. The effects of stress on memory cytotoxic T lymphocyte-mediated protection against herpes simplex virus infection at mucosal sites. *Brain Behavior and Immunity* 2002;16:104–117.
- Wright R, Bosquet EM. Maternal Stress and perinatal programming in the expression of atopy. *Expert Reviews in Clinical Immunology* 2008;4:535–538.
- Wright RJ. Stress and Atopic Disorders. *Journal of Allergy and Clinical Immunology* 2005;116:1301–1306. [PubMed: 16337463]
- Wright RJ. Prenatal maternal stress and early caregiving experiences: implications for childhood asthma risk. *Pediatric and Perinatal Epidemiology* 2007;21:8–14. [PubMed: 17935570]
- Wright RJ, Cohen RT, Cohen S. The impact of stress on the development and expression of atopy. *Current Opinions in Allergy and Clinical Immunology* 2005;5:23–29.
- Wright RJ, Finn PW, Contreras JP, Cohen S, Wright RO, Staudenmayer J, Weiss ST, Gold DR. Chronic caregiver stress and IgE expression, allergen-induced proliferation and cytokine profiles in a birth cohort predisposed to atopy. *Journal of Allergy and Clinical Immunology* 2004;113:1051–1057. [PubMed: 15208584]
- Wright RJ, Visness CM, Calatroni A, Grayson MH, Gold DR, Sandel MT, Lee-Parritz L, Wood RA, Kattan M, Bloomberg GR, Burger M, Togias A, Witter FR, Sperling RS, Sadovsky Y, Gern JE. Prenatal maternal stress and cord blood innate and adaptive cytokine responses in an inner-city cohort. *American Journal of Respiratory and Critical Care Medicine*. in press.
- Wrona D. Neural-immune interactions: an integrative view of the bidirectional relationship between the brain and immune systems. *Journal of Neuroimmunology* 2006;172(1-2):38–58. [PubMed: 16375977]
- Yamasu K, Shimada Y, Sakaizumi M, Soma G, Mizuno D. Activation of the systemic production of tumor necrosis factor after exposure to acute stress. *European Cytokine Network* 1992;3(4):391–398. [PubMed: 1421011]
- Yehuda R, Bierer LM. Transgenerational transmission of cortisol and PTSD risk. *Progress in Brain Research* 2008;167:121–134. [PubMed: 18037011]
- Zaehner T, Plueshke V, Frey BM, Frey FJ, Ferrari P. Structural analysis of the 11 beta-hydroxysteroid dehydrogenase type 2 gene in end-stage renal disease. *Kidney International* 2000;58:1413–1419. [PubMed: 11012876]
- Zhang Y, Zhang Y, Miao J, Hanley GC, Sun X, Chen T, et al. Chronic restraint stress promotes immune suppression through toll-like receptor 4-mediated phosphoinositide 3-kinase signaling. *Journal of Neuroimmunology* 2008;204:13–19. [PubMed: 18814920]
- Zietkowski Z, Tomasiak-Lozowska MM, Skiepkowski R, Mroczko B, Szmitkowski M, Bodzenta-Lukaszyk A. High-sensitivity C-reactive protein in the exhaled breath condensate and serum in stable and unstable asthma. *Respiratory Medicine* 2009;103:379–385. [PubMed: 19010654]