

CHROMOSOME COUNTS IN A POLYPLOID SERIES OF SACCHAROMYCES¹

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Direct determination of chromosome number in *Saccharomyces* is advantageous in the further development of yeast genetics. The enumeration of centromeres by deductions from tetrad analysis sets a minimum chromosome number (Lindgren, 1949; Lindgren and Lindgren, 1951*b*), but interchromosomal linkage (Shult and Lindgren, 1955) introduces complexities into the deduction of chromosomal numbers from genetic data. Experimental information bearing on the degree of ploidy arises from (1) morphological studies (Winge, 1935; Lindgren, 1949) and the characteristic growth patterns of the different members of the ploidy series (Townsend and Lindgren, 1954), (2) genetical analysis (Lindgren and Lindgren, 1951*a*; Roman *et al.*, 1951), (3) biochemical tests (Ogur *et al.* 1952; and Ogur, 1954*a, b*), and (4) irradiation data (Lucke and Sarachek, 1953), and although they are in general agreement, they do not reveal the chromosome number.

The present work describes the direct investigation of the chromosome number of diploid, triploid, and tetraploid *Saccharomyces*. The problem was made feasible by the convergence of several lines of experience in this laboratory:

(1) The perchloric acid-Giemsa technique of chromatin staining has yielded interpretable nuclear structures in yeast.

(2) Study of sporulation in yeast has indicated that the perchloric-Giemsa stained structures behave in a manner characteristic of chromosomes in higher organisms during meiosis I yielding discrete, rodlike, countable, chromatinic bodies.

(3) The availability of sporulating diploid, triploid, and tetraploid cultures has made it possible to count chromosomes at several levels of ploidy and to estimate the haploid number as

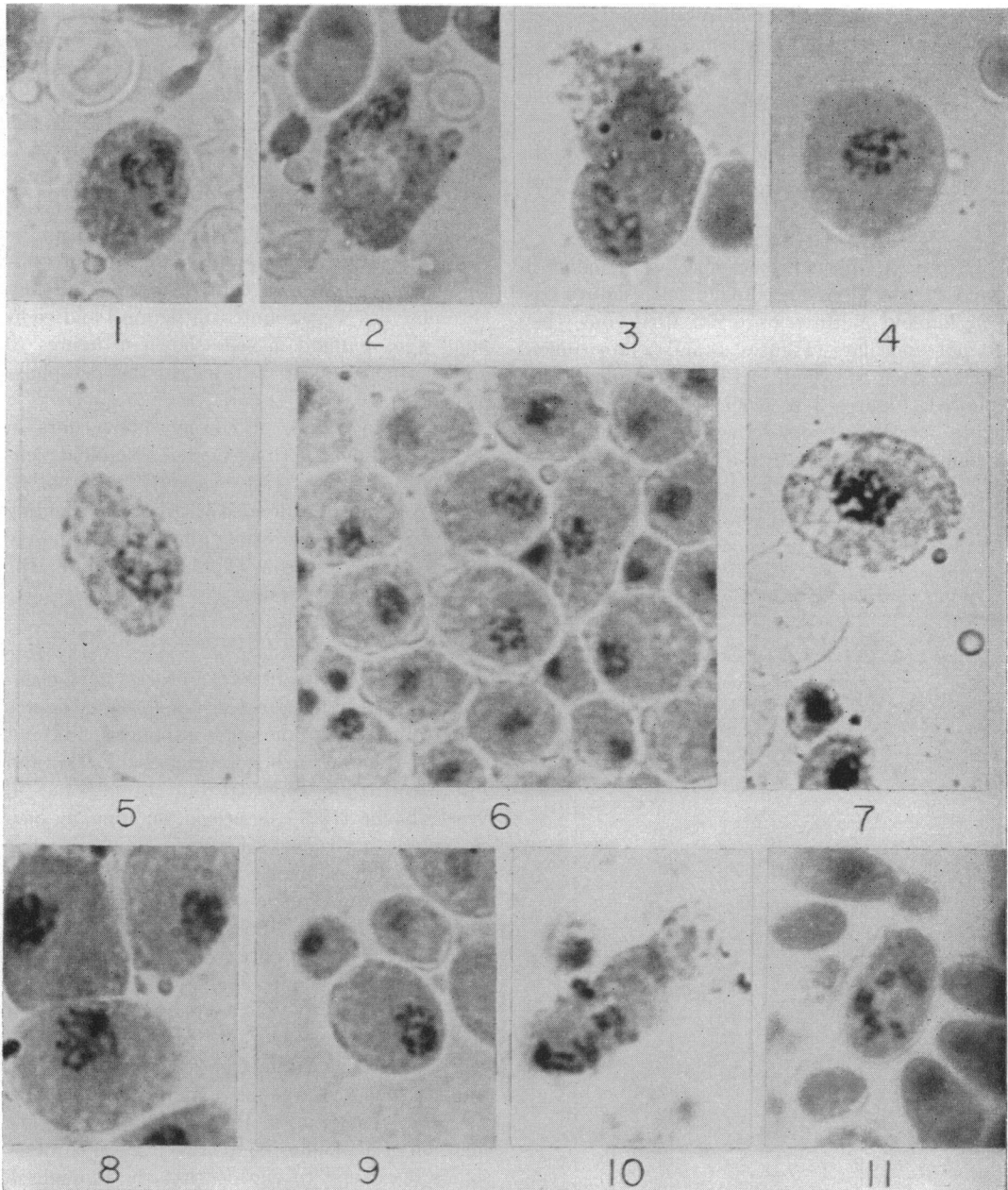
the mean chromosome count divided by the degree of ploidy.

MATERIALS AND METHODS

Three Carbondale strains of *Saccharomyces* (1) diploid strain 14268 x 8256, (2) triploid strain 11296 x 13894 and (3) tetraploid strain 11294 x 11296 were grown for 24 hr at 30 C on agar slants of the following composition: KH_2PO_4 , 2.0 g; $(\text{NH}_4)_2\text{SO}_4$, 2.0 g; MgSO_4 , 0.5 g; glucose, 20.0 g; basamin (Anheuser-Busch, Inc.), 5.0 g; agar, 20.0 g; and distilled water, 1,000 ml. These cultures were used to inoculate slants of a sporulation medium similar to the acetate medium of Stantial (1928, 1935) and Elder (1933, 1937) which was described by Adams (1949): sodium acetate, 10.0 g; liquid yeast extract (Anheuser-Busch, Inc.), 2.5 ml; glucose, 1.0 g; agar, 30.0 g; and distilled water, 1,000 ml. These cultures were incubated at 24 C for periods of time varying from 16 to 24 hr. From these cultures, cells in almost every phase of sporulation were obtained. Experience has shown that the chromosomes are most clearly distinguishable during the presporulation phase.

The cells were washed from the slants with approximately 5 ml of distilled water, centrifuged, and the supernatant decanted and stained by the perchloric-Giemsa method (Lindgren *et al.*, 1956). Wet mounts were squashed according to a suggestion by Dr. Jack Shultz (personal communication) as follows: The slide and coverslip were coated lightly with silicone grease by polishing with a tissue after having placed a small amount of silicone stopcock grease on each. One or two loopfuls of a cell suspension were applied to the coated side of the slide, the coverslip applied, and blotted with paper. During the blotting operation, considerable pressure was applied with the thumb in a rotary direction. The coverslip was then hammered vigorously with a small rubber mallet. To prevent drying,

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Figures 1-11. Figures 1 and 2. Diploid Saccharomyces in late prophase of meiosis I showing 8 chromosomes.

Figures 3 to 5. Triploid Saccharomyces in late prophase of meiosis I showing 12, 13, and 11 chromosomes, respectively.

Figure 6. Cells in synchrony.

Figures 7 to 11. Tetraploid Saccharomyces cells in prophase and metaphase. Sixteen chromosomes were counted in the cell shown in figure 9, and 8 pairs were counted in figures 10 and 11, respectively.

with each containing two chromosomes. His figures indicate that he observed an optical section of the chromatin coated spindle (Lindgren, *et al.*, 1956).

Sinoto and Yuasa (1941) and DeLamater (1950) reported four chromosomes in the budding phase of the strains of *Saccharomyces* which they studied. Levan, however, in 1947 stated that there were at least ten chromosomes. Leitz (1951) found three chromosomes in the haplophase and six in the diplophase of cells of *Saccharomyces priorianus*.

Subramaniam (1948, 1951) states that the haploid number of chromosomes in yeast is one and that variations are due to "endopolyploidy." Mundkur (1954) failed to find discrete chromosomal bodies in vegetative cells fixed by freeze-drying and stained with Feulgen and concluded that there are no microscopically demonstrable chromosomes in yeast and that all previous demonstrations were due to harsh chemical fixatives.

Previous work in this laboratory on the vegetative tetraploid cells (Lindgren *et al.*, 1956) failed to reveal countable chromosomes in contrast to the present success with cells at late prophase of meiosis I.

SUMMARY

Observation of sporulating diploid, triploid, and tetraploid strains of *Saccharomyces*, fixed with modified Carnoy solution, hydrolyzed in perchloric acid, and stained with Giemsa, revealed nuclei with definite rodlike chromosomes. Counts of the chromosomes when correlated to the corresponding degrees of ploidy showed that the haploid number is 4.

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