

Modulatory Effect of Rice Bran and Phytic Acid on Glucose Metabolism in High Fat-Fed C57BL/6N Mice

Soo Mi Kim¹, Catherine W. Rico¹, Sang Chul Lee² and Mi Young Kang^{1,*}

¹Department of Food Science and Nutrition, Kyungpook National University, Daegu 702-701, Republic of Korea

²Department of Agronomy, Kyungpook National University, Daegu 702-701, Republic of Korea

Received 21 December, 2009; Accepted 21 January, 2010; Published online 10 April, 2010

Summary The effect of dietary feeding of rice bran and phytic acid on the glucose metabolism in high fat-fed C57BL/6N mice fed was investigated. The mice were given with either a high fat diet only (HF group) or a high fat diet supplemented with rice bran (HF-RB group) or phytic acid (HF-PA group) for 7 weeks. The control mice (NC group) received a normal diet. At the end of the experimental period, the HF group exhibited substantially higher blood glucose level than the NC group. However, the HF-RB and HF-PA groups showed a marked decrease in the blood glucose level relative to HF mice. Furthermore, significantly higher glucokinase (GK) activity and lower phosphoenolpyruvate carboxykinase (PEPCK) activity were observed in HF-RB and HF-PA mice compared with that of the NC and HF ones. It was also found that the glucose-6-phosphatase (G6pase) activity and hepatic glycogen concentration were considerably higher in HF-RB and HF-PA groups, respectively, than that of the HF mice. These findings demonstrate that both rice bran and phytic acid could reduce the risk of high fat diet-induced hyperglycemia via regulation of hepatic glucose-regulating enzyme activities.

Key Words: rice bran, phytic acid, glucose metabolism, diabetes, high fat-fed mice

Introduction

Diabetes mellitus, a metabolic disease characterized by hyperglycemia and often associated with obesity, is one of the leading causes of death in most developed countries [1]. Its incidence has rapidly increased in epidemic proportions due to poor eating habit and sedentary lifestyle. Scientific studies have shown that chronic consumption of a high fat diet results in increased body weight and poor glucose regulation [2–4]. Individuals with higher intake of fat are more prone to develop glucose metabolism disorder, type 2 diabetes, or impaired glucose tolerance than those with lower fat intake [5]. A wide range of oral medicines are currently being used for treating diabetes, however, various

side effects and high rates of secondary failures have been associated with the available anti-diabetic medicine [6]. Thus, finding natural drugs with hypoglycemic activity has become stronger and more urgent.

Rice bran, a by-product of rice milling industry and commonly used as animal feed, has been the subject of many researches for the past years due to its health-promoting phytochemicals that have strong antioxidant activities [7–9] and hypocholesterolemic effects [10, 11]. It also contains high amount of phytic acid (9.5–14%) [12], a dietary fiber component found in most grains and legumes which has been shown to have antioxidant and anticancer properties [13–15]. Recent researches also revealed that phytic acid has hypoglycemic and antihyperlipidemic effects in diabetic mice [16, 17].

The high-fat diet-fed C57BL/6 mouse model is widely used by researchers in investigating the pathophysiology of impaired glucose tolerance and type 2 diabetes for the development of new treatments [18–20]. While the *in vitro*

*To whom correspondence should be addressed.

Tel/Fax: +82-53-950-6235

E-mail: mykang@knu.ac.kr

antioxidant potential of rice bran and phytic acid has been well documented, reports on their physiological functions in relation to glucose metabolism in animal models have been limited. Since oxidative stress is considered to be a key factor in the development of diabetes and its associated health disorders, the strong antioxidant activity of rice bran and phytic acid may be useful in preventing the development of diabetic hyperglycemia under a high fat diet condition. Hence, this study was conducted to investigate the effects of dietary feeding of rice bran and phytic acid on the glucose metabolism in high fat-fed C57BL/6N mice.

Materials and Methods

Animals and diet

Thirty-two male C57BL/6N mice of 4 weeks of age, weighing 12 g, were obtained from Orient Inc. (Seoul, Korea). They were individually housed in stainless steel cages in a room maintained at 25°C with 50% relative humidity and 12/12 h light/dark cycle and fed with a pelletized chow diet for 2 weeks after arrival. The mice were then randomly divided into 4 dietary groups ($n = 8$). The first and second groups were fed with a normal and high fat (17%, w/w) diets, respectively, while the other two groups were fed with high fat diet supplemented with either rice bran (RPC, Gimcheon, Korea) or phytic acid (Tsuno, Osaka, Japan). The composition of the experimental diet (Table 1) was based on the AIN-76 semisynthetic diet. The mice were fed for 7 weeks and allowed free access to food and water during the experimental period. The daily food intake of mice was constant (3 g/day) throughout the study.

Table 1. Composition of the experimental diets (%)

Component	Dietary group ¹			
	NC	HF	HF-RB	HF-PA
Casein	20	20	16.7	20
DL-Methionine	0.3	0.3	0.3	0.3
Sucrose	50	50	36.46	49.5
Corn starch	15			
Cellulose	5	5		5
Corn oil	5	3		3
Cholinbitartrate	0.2	0.2	0.2	0.2
Mineral mixture ²	3.5	3.5	1.34	3.5
Vitamin mixture ³	1	1	1	1
Lard		17	14	17
Rice bran			30	
Phytic acid				0.5
Total (%)	100	100	100	100

¹ NC, normal diet; HF, high fat diet; HF-RB, high fat diet + rice bran; HF-PA, high fat diet + phytic acid. ² AIN-76 mineral mixture. ³ AIN-76 vitamin mixture.

At the end of the experimental period, the mice were anaesthetized and sacrificed. Blood samples were collected and centrifuged at $1,000 \times g$ for 15 min at 4°C to obtain the plasma. The livers were removed, rinsed with physiological saline and stored at -70°C until analysis.

Measurement of blood glucose level

The blood glucose level in mice was measured using Accu-Chek Active Blood Glucose Test Strips (Roche Diagnostics GmbH, Mannheim, Germany). Blood samples were drawn from the tail vein of the mice before and after 3 and 7 weeks of feeding the animals with experimental diets.

Determination of insulin and glycogen levels

The insulin content was measured using radioimmunoassay kits (TMB Mouse Insulin ELISA kit, Sibayagi, Gunma, Japan). The glycogen concentration in liver was determined using the method described by Seifter *et al.* [21]. Fresh liver (100 mg) was mixed with 30% KOH and heated at 100°C for 30 min. The mixture was then added with 1.5 mL ethanol (95%, v/v) and kept overnight at 4°C. The pellet was mixed with 4 ml distilled water. A 500 μ L of the mixture was added with 0.2% anthrone (in 95% H₂SO₄) and the absorbance of the sample solution was measured at 620 nm. The results were calculated on the basis of a standard calibration curve of glucose.

Measurement of hepatic glucose-regulating enzyme activities

The hepatic enzyme source was prepared according to the method developed by Hulcher and Oleson [22]. Briefly, the liver tissue (0.3 g) was homogenized with 6 mL Tris buffer and the homogenates were then centrifuged at $600 \times g$ for 10 min at 4°C. The supernatant was ultracentrifuged at $100,000 \times g$ for 1 h at 4°C to isolate the cytosolic fraction for glucokinase (GK) and phosphoenolpyruvate carboxykinase (PEPCK) assay. The residue was mixed with 1 mL Tris buffer and re-ultracentrifuged at $100,000 \times g$ for 1 h at 4°C. The supernatant was discarded and the residue was collected for the G6pase assay. The GK activity was determined based from the method of Davidson and Arion [23] with slight modification. A 0.98 mL of the reaction mixture containing 50 mM HEPES-NaOH (pH 7.4), 100 mM KCl, 7.5 mM MgCl₂, 2.5 mM dithioerythritol, 10 mg/mL albumin, 10 mM glucose, 4 units of glucose-6-phosphate dehydrogenase, 50 mM NAD⁺ and 10 μ L cytosol was pre-incubated at 37°C for 10 min. The reaction was initiated with the addition of 10 μ L of 5 mM ATP and the mixture was incubated at 37°C for 10 min. The change in absorbance at 340 nm was recorded. The glucose-6-phosphatase (G6pase) activity was measured using the method described by Alegre *et al.* [24]. The reaction mixture contained 765 μ L of 131.58 mM HEPES-NaOH (pH 6.5), 100 μ L of 18 mM EDTA (pH 6.5), 100 μ L of 265 mM glucose-6-phosphate, 10 μ L of 0.2 M

NADP⁺, 0.6 IU/mL mutarotase and 0.6 IU/mL glucose dehydrogenase. After pre-incubation at 37°C for 3 min, the mixture was added with 5 µL microsome and incubated at 37°C for 4 min. The change in absorbance at 340 nm was measured. The PEPCK activity was determined based from the method developed by Bentle and Lardy [25]. The reaction mixture consisted of 72.92 mM sodium Hepes (pH 7.0), 10 mM dithiothreitol, 500 mM NaHCO₃, 10 mM MnCl₂, 25 mM NADH, 100 mM IDP, 200 mM PEP, 7.2 unit of malic dehydrogenase and 10 µL cytosol. The enzyme activity was determined based from the decrease in the absorbance of the mixture at 350 nm at 25°C.

Statistical analysis

All data are presented as the mean ± SE. The data was evaluated by one-way ANOVA using a Statistical Package for Social Sciences software program (SPSS Inc., Chicago, IL) and the differences between the means were assessed using Duncan's multiple range test. Statistical significance was considered at $p < 0.05$.

Results

Body weight gain

Prior to feeding the mice with the experimental diets, all the animals exhibited similar body weights. However, at the end of the experimental period, a substantial increase in the body weight of HF mice was observed relative to that of the control group (data not shown). The HF-RB and HF-PA groups, on the other hand, showed considerably lower body weights than the HF group, suggesting that both rice bran and phytic acid could control the increase in body weight of mice under high fat diet condition.

Blood glucose levels

The initial blood glucose levels in mice prior to feeding with experimental diets did not significantly differ among the groups (Fig. 1). However, high-fat feeding resulted in a marked increase in the glucose level of mice after 3 and 7 weeks. On the final week, the HF-RB and HF-PA mice exhibited substantially lower glucose level compared with the HF group.

Insulin and glycogen levels

There was no significant difference in the insulin level among the mice groups (Table 2). Supplementation with rice bran in the diet did not significantly change the glycogen content in mice. On the other hand, the hepatic glycogen content was considerably higher in HF-PA group than that of the control and HF mice.

Hepatic glucose-regulating enzyme activities

The hepatic GK enzyme activity was significantly higher

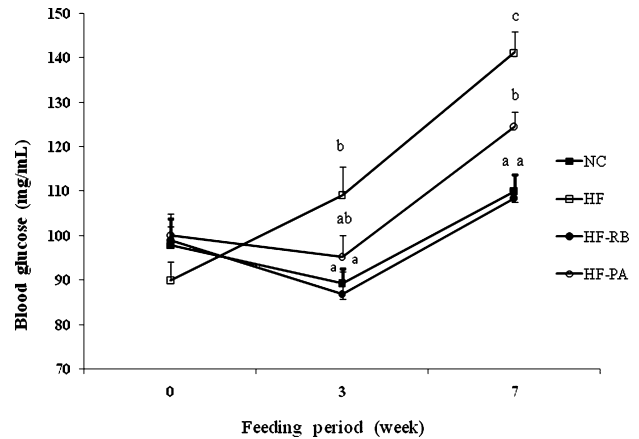


Fig. 1. Effect of rice bran and phytic acid supplementation on the blood glucose level in high fat fed-mice. Means not sharing a common superscript are significantly different at $p < 0.05$ ($n = 8$). NC, normal diet; HF, high fat diet; HF-RB, high fat diet + rice bran; HF-PA, high fat diet + phytic acid.

Table 2. Insulin and glycogen concentrations¹ in mice fed with high fat diet supplemented with rice bran and phytic acid

Dietary group ²	Insulin (ng/mL)	Glycogen (mg/g liver)
NC	1.58 ± 0.01 ^a	2.55 ± 0.16 ^a
HF	1.45 ± 0.01 ^a	3.14 ± 0.21 ^a
HF-RB	1.87 ± 0.02 ^a	3.91 ± 1.33 ^{ab}
HF-PA	2.41 ± 0.04 ^a	5.77 ± 0.92 ^b

¹ Values are means ± SE ($n = 8$). Means in the same column not sharing a common superscript are significantly different at $p < 0.05$, which pertains to the "a and b", in the table. ² NC, normal diet; HF, high fat diet; HF-RB, high fat diet + rice bran; HF-PA, high fat diet + phytic acid.

in mice fed with rice bran and phytic acid than that of the control and HF-fed ones (Fig. 2). While high fat feeding did not significantly affect the activities of G6pase and PEPCK enzymes (Figs. 3 and 4), dietary feeding of rice bran suppressed the elevation of G6pase activity (Fig. 3). Moreover, both HF-RB and HF-PA groups exhibited significantly lower PEPCK activity than that of the NC and HF ones (Fig. 4).

Discussion

In vitro studies on the antioxidant capacity of rice bran and phytic acid revealed that these plant components have strong antioxidant activity [13, 26]. Since diabetes is a free radical-mediated disease, the effect of rice bran and phytic acid on the glucose metabolism in high fat-fed mice was investigated. In the present study, a high fat-diet resulted

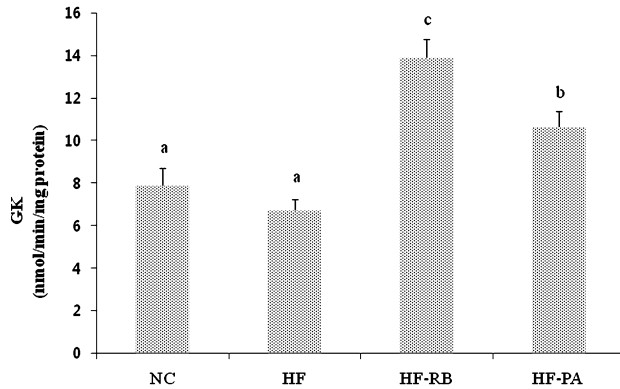


Fig. 2. Effect of rice bran and phytic acid supplementation on the hepatic GK enzyme activity in high fat fed-mice. Bars not sharing a common superscript are significantly different at $p < 0.05$ ($n = 8$). NC, normal diet; HF, high fat diet; HF-RB, high fat diet + rice bran; HF-PA, high fat diet + phytic acid.

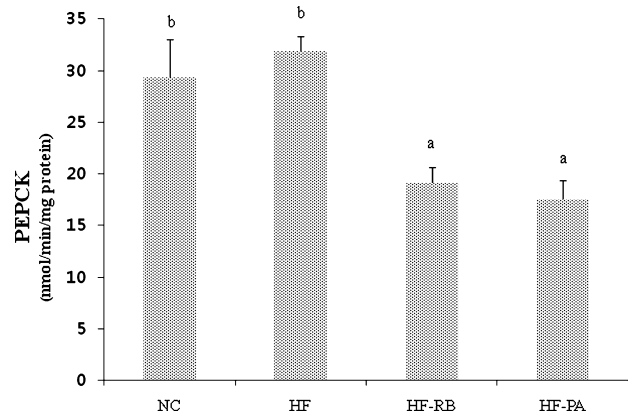


Fig. 4. Effect of rice bran and phytic acid supplementation on the hepatic PEPCK enzyme activity in high fat fed-mice. Bars not sharing a common superscript are significantly different at $p < 0.05$ ($n = 8$). NC, normal diet; HF, high fat diet; HF-RB, high fat diet + rice bran; HF-PA, high fat diet + phytic acid.

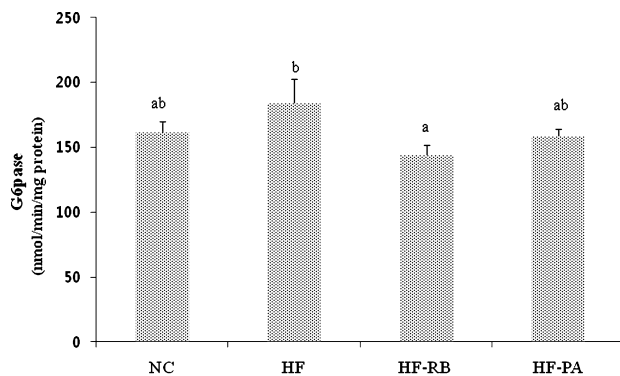


Fig. 3. Effect of rice bran and phytic acid supplementation on the hepatic G6pase enzyme activity in high fat fed-mice. Bars not sharing a common superscript are significantly different at $p < 0.05$ ($n = 8$). NC, normal diet; HF, high fat diet; HF-RB, high fat diet + rice bran; HF-PA, high fat diet + phytic acid.

in a marked increase in the blood glucose level in mice. However, both rice bran and phytic acid were able to suppress the increase in glucose concentration. Previous investigations also showed that rice bran powder and its water soluble fraction could decrease serum glucose level in human subjects with diabetes [27, 28]. It was also found that the fiber components of rice bran isolated from watery extract exhibited hypoglycemic effect in mice with normal serum glucose [29]. Likewise, Lee *et al.* [17] reported that dietary feeding of phytic acid resulted in a significant reduction in the blood glucose levels in diabetic mice.

The antihyperglycemic action of rice bran and phytic acid is probably associated with the marked enhancement of GK activity and inhibition of G6pase and PEPCK in the liver.

Hepatic GK enzyme plays a major role in the regulation of glucose homeostasis [30]. An increase in the expression of hepatic GK could cause an increase in the utilization of blood glucose for energy production or glycogen storage in the liver [31], thereby resulting in reduced blood glucose level. G6pase and PEPCK enzymes, on the other hand, regulate gluconeogenesis and glucose output in the liver [30, 32]. Hence, a decrease in G6pase and PEPCK activities signifies a decrease in hepatic glucose production. Although no significant difference was observed in the insulin levels among the groups, the results suggest that mice fed with rice bran and phytic acid had a tendency towards higher insulin levels than animals fed with high fat diet alone. An enhanced rate of glycogenesis was observed in HF-PA mice, as manifested by a significant increase in the hepatic glycogen concentration. The HF-RB group also showed a relatively higher glycogen content than HF and NC mice, although the difference is not statistically significant. The activities of hepatic glucose-regulating enzymes are reported to be partly regulated by insulin. High levels of insulin were shown to inhibit hepatic glucose production via stimulation of GK gene transcription and glycogen synthesis and inhibition of gluconeogenesis [31, 32].

A high fat diet negatively affects glucose metabolism and the regulation of blood glucose is essential in preventing the development of diabetes. Hyperglycemia promotes the formation of reactive oxygen species (ROS), which cause cellular damage [33]. The liver undergoes free radical-mediated injury in diabetes, and an increased ROS is related to the damage of hepatic glucose-regulating enzymes [34]. Antioxidants have long been recognized as a means of treating diabetes as they were shown to reduce blood glucose concentrations by protecting the cells against the

toxic effects of ROS under hyperglycemic condition [35, 36]. Although the antioxidant potential of rice bran and phytic acid has been well documented, further studies are still needed to elucidate the underlying mechanism of the antihyperglycemic action of rice bran and phytic acid in order to have a better understanding of their therapeutic potential.

Conclusion

Results of this study demonstrate that rice bran and phytic acid could improve the blood glucose metabolism in high fat-fed C57BL/6N mice. The enhanced glucose metabolism may be partly due to the activation of GK, and inhibition of G6pase and PEPCK enzymes in the liver. The hypoglycemic effect of rice bran and phytic acid illustrates that these plant components may be beneficial for the treatment of diabetic hyperglycemia.

References

- [1] McGill, M. and Felton, A.M.: New global recommendations: a multidisciplinary approach to improving outcomes in diabetes. *Prim. Care Diabetes*, **1**, 49–55, 2007.
- [2] Alsaif, M.A. and Duwaih, M.M.S.: Influence of dietary fat quantity and composition on glucose tolerance and insulin sensitivity in rats. *Nutr. Res.*, **24**, 417–425, 2004.
- [3] Messier, C., Whately, K., Liang, J., Du, L., and Puissant, D.: The effects of high-fat, high-fructose, and combination diet on learning, weight, and glucose regulation in C57BL/6 mice. *Behav. Brain Res.*, **178**, 139–145, 2007.
- [4] Petro, A.E., Cotter, J., Cooper, D.A., Peters, J.C., Surwit, S.J., and Surwit, R.S.: Fat carbohydrate and calories in the development of diabetes and obesity in the C57BL/6J mouse. *Metabolism*, **53**, 454–457, 2004.
- [5] Lichtenstein, A.H. and Schwab, U.S.: Relationship of dietary fat to glucose metabolism. *Atherosclerosis*, **150**, 227–243, 2000.
- [6] Inzucchi, S.E.: Oral antihyperglycemic therapy for type 2 diabetes: scientific review. *J. Am. Med. Assoc.*, **287**, 360–372, 2002.
- [7] Chotimarkorn, C., Benjakul, S., and Silalai, N.: Antioxidant components and properties of five long-grained rice bran extracts from commercial available cultivars in Thailand. *Food Chem.*, **111**, 636–641, 2008.
- [8] Devi, R.R., Jayalekshmy, A., and Arumughan, C.: Antioxidant efficacy of phytochemical extracts from defatted rice bran in the bulk oil system. *Food Chem.*, **104**, 658–664, 2007.
- [9] Iqbal, S., Bhangar, M.I., and Anwar, F.: Antioxidant properties and components of some commercially available varieties of rice bran in Pakistan. *Food Chem.*, **93**, 265–272, 2005.
- [10] Kahlon, T.S., Chow, F.I., and Sayre, R.N.: Cholesterol lowering properties of rice bran. *Cereal Food World*, **39**, 99–103, 1994.
- [11] Revilla, E., Santa Maria, C., Miramontes, E., Bautista, J., Garcia-Martinez, A., Cremades, O., Cert, R., and Parrado, J.: Nutraceutical composition, antioxidant activity and hypocholesterolemic effect of water-soluble enzymatic extract from rice bran. *Food Res. Int.*, **42**, 387–393, 2009.
- [12] Martin-Tereso, J., Gonzalez, A., Laar, H.V., Burbano, C., Pedrosa, M.M., Mulder, K., den Hartog, L.A., and Verstegen, M.W.A.: *In situ* ruminal degradation of phytic acid in formaldehyde-treated rice bran. *Animal Feed Sci. Technol.*, **152**, 286–297, 2009.
- [13] Graf, E. and Eaton, J.W.: Antioxidant functions of phytic acid. *Free Radic. Biol. Med.*, **8**, 61–69, 1990.
- [14] Shamsuddin, A.M.: Anti-cancer function of phytic acid. *Int. J. Food Sci. Technol.*, **37**, 769–782, 2002.
- [15] Shamsuddin, A.M. and Vucenic, I.: IP₆ and inositol in cancer prevention and therapy. *Curr. Cancer Ther. Rev.*, **1**, 259–269, 2005.
- [16] Lee, S.H., Park, H.J., Cho, S.Y., Jung, H.J., Cho, S.M., Cho, Y.S., and Lillehoj, H.S.: Effects of dietary phytic acid on serum and hepatic lipid levels in diabetic KK mice. *Nutr. Res.*, **25**, 869–876, 2005.
- [17] Lee, S.H., Park, H.J., Chun, H.K., Cho, S.Y., Cho, S.M., and Lillehoj, H.S.: Effects of dietary phytic acid on the blood glucose level in diabetic KK mice. *Nutr. Res.*, **26**, 474–479, 2006.
- [18] Schreyer, S.A., Wilson, D.L., and LeBoeuf, R.C.: C57BL/6 mice fed high fat diets as models for diabetes-accelerated atherosclerosis. *Atherosclerosis*, **136**, 17–24, 1998.
- [19] Surwit, R.S., Kuhn, C.M., Cochrane, C., McCubbin, J.A., and Feinglos, M.N.: Diet induced type II diabetes in C57BL/6J mice. *Diabetes*, **37**, 1163–1167, 1988.
- [20] Winzell, M.S. and Ahren, B.: The high fat diet-fed mouse: a model for studying mechanisms and treatment of impaired glucose tolerance and type 2 diabetes. *Diabetes*, **53**, S215–S219, 2004.
- [21] Seifter, S., Dayton, S., Navic, B., and Muntwyler, E.: The estimation of glycogen with the anthrone reagent. *Arch. Biochem.*, **25**, 191–200, 1950.
- [22] Hulcher, F.H. and Oleson, W.H.: Simplified spectrophotometric assay for microsomal 3-hydroxy-3-methylglutaryl CoA reductase by measurement of coenzyme A. *J. Lipid Res.*, **14**, 625–631, 1973.
- [23] Davidson, A.L. and Arion, W.J.: Factors underlying significant underestimations of glucokinase activity in crude liver extracts: physiological implications of higher cellular activity. *Arch. Biochem. Biophys.*, **253**, 156–167, 1987.
- [24] Alegre, M., Ciudad, C.J., Fillat, C., and Guinovart, J.J.: Determination of glucose-6-phosphatase activity using the glucose dehydrogenase-coupled reaction. *Anal. Biochem.*, **173**, 185–189, 1988.
- [25] Bentle, L.A. and Lardy, H.A.: Interaction of anions and divalent metal ions with phosphoenolpyruvate carboxykinase. *J. Biol. Chem.*, **251**, 2916–2921, 1976.
- [26] Lai, P., Li, K.Y., Lu, S., and Chen, H.H.: Phytochemicals and antioxidant properties of solvent extracts from Japonica rice bran. *Food Chem.*, **117**, 538–544, 2009.

- [27] Qureshi, A.A., Sami, S.A., and Khan, F.A.: Effects of stabilized rice bran, its soluble and fiber fractions on blood glucose levels and serum lipid parameters in humans with diabetes mellitus Types I and II. *J. Nutr. Biochem.*, **13**, 175–187, 2002.
- [28] Tazakori, Z., Dehghan, M.H., Iranparvar, M., Zare, M., Foladi, N., and Mohammadi, R.: Effect of rice bran powder on blood glucose levels and serum lipid parameters in diabetes patient II. *Res. J. Biol. Sci.*, **2**, 252–255, 2007.
- [29] Hikino, H., Takahashi, M., Oshima, Y., and Konno, C.: Isolation and hypoglycemic activity of orizabrans A, B, C and D glycans of *Oryza sativa* bran. *Planta Medica*, **54**, 1–3, 1988.
- [30] Akiyama, S., Katsumata, S., Suzuki, K., Ishimi, Y., Wu, J., and Uehara, M.: Dietary hesperidin exerts hypoglycemic and hypolipidemic effects in streptozotocin-induced marginal type 1 diabetic rats. *J. Clin. Biochem. Nutr.*, **46**, 87–92, 2010.
- [31] Iynedjian, P.B., Gjinovci, A., and Renold, A.E.: Stimulation by insulin of glucokinase gene transcription in liver of diabetic rats. *J. Biol. Chem.*, **263**, 740–744, 1988.
- [32] Friedman, J.E., Sun, Y., Ishizuka, T., Farrell, C.J., McCormack, S.E., Herron, L.M., Hakimi, P., Lechner, P., and Yun, J.S.: Phosphoenolpyruvate carboxykinase (GTP) gene transcription and hyperglycemia are regulated by glucocorticoids in genetically obese *db/db* transgenic mice. *J. Biol. Chem.*, **272**, 31475–31481, 1997.
- [33] Hong, J.H., Cha, Y.S., and Rhee, S.J.: Effects of the cellcultured *Acanthopanax senticosus* extract on antioxidative defense system and membrane fluidity in the liver of type 2 diabetes mouse. *J. Clin. Biochem. Nutr.*, **45**, 101–109, 2009.
- [34] Lelli, S.M., San, L.C., Viale, M.D., and Mazzetti, M.B.: Response of glucose metabolism enzymes in an acute porphyria model role of reactive oxygen species. *Toxicology*, **216**, 49–58, 2005.
- [35] Kaneto, H., Kajimoto, Y., Miyagawa, J., Matsuoka, T., Fujitani, Y., Umayahara, Y., Hanafusa, T., Matsuzawa, Y., Yamasaki, Y., and Hori, M.: Beneficial effects of antioxidants in diabetes: possible protection of pancreatic beta-cells against glucose toxicity. *Diabetes*, **48**, 2398–2406, 1999.
- [36] Modak, M., Dixit, P., Londhe, J., Ghaskadbi, S., and Devasagayam, T.P.A.: Indian herbs and herbal drugs used for the treatment of diabetes. *J. Clin. Biochem. Nutr.*, **40**, 163–173, 2007.