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Plasticity in Singing Effort and its Relationship with Monoamine Metabolism in the Songbird Telencephalon

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Abstract

Factors intrinsic or extrinsic to individuals, such as their quality or the quality of competition in their social environment, can influence their communication signalling effort. We hypothesized that telencephalic monoamine secretion mediates the effects of a male's own quality and quality of his social environment on his sexual signalling effort. The duration of a male European starling's (Sturnus vulgaris) principal sexual signal, his song, positively correlates with several aspects of his quality, including his reproductive success, immunocompetence, and ability to attract mates. Therefore, the length of songs to which he is exposed reflects, in part, the quality of competition in his social environment. We manipulated the quality of the competitive environment by exposing male starlings to long or short songs for one week. We measured the length of songs produced by experimental males to gauge their quality, counted the number of songs they produced to gauge singing effort, and quantified telencephalic monoamine metabolism using HPLC. Singing effort increased with the length of the males' own songs and with the length of songs to which we exposed them. Norepinephrine metabolism in Area X of the song control system was negatively correlated with the subjects' mean song length and singing effort. Serotonin metabolism in the caudomedial mesopallium of the auditory telencephalon increased with the length of songs to which we exposed the subjects and with their singing effort. This raises the hypothesis that serotonin and norepinephrine secretion in the telencephalon help mediate the effects of extrinsic and intrinsic factors on signalling effort.

Keywords

bird song; monoamines; neuro	opiasticity; norepinephrine; seroton	ın

INTRODUCTION

The costs and benefits of producing communication signals can vary (Gil and Gahr, 2002), and therefore individuals should modulate their signalling effort (e.g. Sockman et al., 2005b) according to extrinsic factors, such as characteristics of the physical environment or the quality of competition among conspecifics, and to intrinsic factors, such as the signaller's own physiological state or competitive quality. Such context-dependent, behavioral

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plasticity requires the neural processing of these factors, followed by production of the appropriate motor programs involved in modulating signal production.

In most songbird species, adult males attract mates and repel competing males by producing learned vocalizations known as song (Catchpole and Slater, 1995). Song learning and production are controlled by a network of interconnected forebrain nuclei, including HVC (initialism used as a proper name), the Robust Nucleus of the Arcopallium (RA), the Magnocellular Nucleus of the Anterior Nidopallium (MAN), and Area X (Nottebohm et al., 1976). HVC and RA form part of a motor pathway involved in song production, whereas MAN and Area X form part of an anterior forebrain pathway involved in song learning and plasticity (Brainard, 2004). Auditory input from an individual's own songs or songs from another individual ascends via midbrain, thalamic, and telencephalic areas to the reciprocally connected caudomedial mesopallium (CMM) and caudomedial nidopallium (NCM) of the auditory telencephalon (Vates et al., 1996). The former provides input to the song control system via direct projections to HVC (Bauer et al., 2008).

Across vertebrate taxa, central monoamines mediate the effects of intrinsic and extrinsic factors on behavioral plasticity (Jacobs and Azmitia, 1992; Dave et al., 1998; Appeltants et al., 2002; Berridge and Waterhouse, 2003; Hurley et al., 2004; Bell et al., 2007), including plasticity in song output (Barclay and Harding, 1988; 1990; Barclay et al., 1991; 1996; Ball et al., 2003; Sasaki et al., 2006). For example, castration of zebra finches (*Taeniopygia guttata*) decreases norepinephrine turnover in the RA and male courtship behaviors, whereas hormone replacement increases norepinephrine turnover in the RA and restores courtship behavior to normal levels (Harding et al., 1983; Barclay and Harding, 1988; 1990). Furthermore, pharmacological depletion of norepinephrine levels in the RA results in a decrease in both the number of song bouts and the frequency of courtship displays (Barclay et al., 1991), suggesting that the stimulatory effects of elevated sex steroid hormones on these male sexual behaviors is mediated, at least in part, by norepinephrine metabolism in the RA.

In European starlings (Sturnus vulgaris), a male's mean song length reflects some aspects of his quality, in that it positively correlates with his reproductive success, immunocompetence, age, and ability to attract mates and repel competing males (Feare, 1984; Eens et al., 1991; Mountjoy and Lemon, 1991; 1996; Eens, 1997; Gentner and Hulse, 2000; Duffy and Ball, 2002). Therefore, to manipulate an extrinsic factor — the quality of competitors in the song environment — we chronically exposed male starlings to either long or short songs for 1 week. To assess an intrinsic factor — the male's own quality — we measured the length of songs produced during and immediately following this playback period. Although dichotomizing factors into the extrinsic and intrinsic is of dubious value, given that the two types of factors can interact to such an extent as to make them virtually inseparable, we use these terms as a simple way to distinguish between the quality of a signaller's perceived competitors and the signaller's own quality, respectively. To investigate the hypothesis that telencephalic monoamine secretion mediates the effects of a male's own quality and quality of his social environment on his signalling effort, we examined how singing effort was affected by the male's own song quality and song environment and then quantified the concentrations of serotonin, dopamine, and norepinephrine metabolites in the song control nuclei and auditory telencephalon.

MATERIALS AND METHODS

Animals and Housing

European starlings were captured on 12 December 2005 in Pennsylvania, USA (41.75°N 80.35°W) and transferred to large, outdoor flight cages at the University of North Carolina

at Chapel Hill (USA), where we conducted the university-approved study (IACUC protocol 04-241.2). For the entire study, we provided the birds with food (Daily Maintenance, Roudybush; Woodland, CA, USA) and water *ad libitum*. We identified males based on the presence of a blue proximal region of the bill (Kessel, 1951) and later confirmed their sex (see Experiment Procedure). On 10 May 2006, we paired 20 male subjects in 10 indoor cages on a 16 h light and 8 h dark (16 L : 8 D) photoperiod to drive the birds first through a reproductive-like state and then into a non-reproductive state (Nicholls et al., 1988), thus synchronizing the birds in their reproductive cycle. On 13 July, we changed the photoperiod to 8 L : 16 D to begin the process of re-instating sensitivity to reproductive stimuli (Nicholls et al., 1988).

Experiment Procedure

On 18 October, we transferred four subjects (two of the pairs mentioned above) individually into four sound-attenuation chambers located together in one room on an 11 L: 13 D photoperiod (Session 1). We had equipped each foam-lined chamber with a cage having two perches, an air intake and fan-driven exhaust, a fluorescent light that maintained the 11 L: 13 D photoperiod within the chamber, and a speaker (Pioneer Corp. TS-G1040R). We powered speakers by a daisy chain of four mono-block amplifiers interfaced with a computer.

The next day, we began the simultaneous broadcast of one of two acoustic treatments, a set of long songs or a set of short songs, (hereafter termed song-environment treatment; see Song Recordings Used for Playback), through each chamber's speaker (Table 1). We played the song at approximately 70 dB at 5 cm from the speaker to approximate the amplitude of songs that a free-living male would experience from a nearby conspecific male competitor. We balanced treatment levels between members of the original pairs described above and spatially interspersed among the chambers each replicate of the long song environment with each replicate of the short song environment. Using procedures described previously (Sockman and Salvante, 2008), we exposed the subjects to the song sets for 5.5 h per day for 7 days at partially randomized 30-min intervals during the photophase only and randomized the order of songs played within each 30-min period. Broadcasts began at the onset of the photophase each day. No more than two 30-min broadcasts occurred in a row (i.e., without at least one intervening, 30-min silent period), and no broadcast occurred during the last 30 min of the photophase each day. This approach, along with the use of an 11 L: 13 D photoperiod, was intended to mimic the natural sexual signalling environment of free-living starlings early in the breeding season. We broadcast white-noise in the room to help mask sound between chambers. On the 8th day of the session (hereafter termed day 8), the subjects received no song playback (Table 1).

We equipped each chamber with an omni-directional microphone (Sennheiser ME 62) that was interfaced with a computer to record and store the songs produced by the subjects. During the photophase of each of the first 8 days, we collected audio recordings of the songs produced by the subjects. We programmed the computer to store the recordings only if 27 consecutive peaks in the recording's oscillogram exceeded an amplitude threshold for at least 3 s (Sound Analysis Pro Software, version 1.02). Stored recordings began 20 ms prior to the peak train and ended when sound dropped below the amplitude threshold for 10 s. We selected the amplitude, peak-count, and time thresholds in order to minimize the recording of playback song and other sounds and to maximize the recording of the subjects' songs. Nonetheless, we generated a large number of audio files, many of which contained cage noises produced when the subjects moved in addition to the subjects' songs.

As part of another study, we then exposed one of the four subjects to a set of novel long songs and another subject to a set of novel short songs for 30 min (hereafter termed song-

stimulus treatment) beginning 4 h after the onset of the photophase on the 9th day of the session (see Song Recordings Used for Playback) (Table 1). We exposed the remaining two subjects to the song-stimulus treatment 30 min later. By balancing the song-stimulus treatment levels between members of the same song-environment treatment group, we generated four treatment groups (but see Song Recordings Used for Playback): 1) long song environment, long song stimulus; 2) long song environment, short song stimulus; 3) short song environment, long song stimulus; 4) short song environment, short song stimulus.

At 90 min following onset of the song-stimulus treatment, we weighed the subjects, rapidly decapitated them, and removed their brains. Using previously described protocols (Sockman and Salvante, 2008), we froze one hemisphere (alternating left and right with each treatment group), fixed the other hemisphere and stored both at -80° C. We removed testes from each individual and dried them at 60° C until they reached a constant dry mass. We repeated these procedures for a total of five sessions of four subjects each, resulting in a total of 20 subjects divided among five replicates for each of the four treatment groups, counterbalanced among the chambers and sessions.

Song Recordings Used for Playbacks

Details of the song recordings used for playback have been described previously (Gentner and Hulse, 2000). Briefly, for the song-environment treatment, a library of complete song bouts was recorded from a single, laboratory-housed male directing song at a female. From these, 12 songs were selected, which, based on length, were divided into two sets of six: a long-song set and a short-song set with mean song lengths of 55.2 and 26.0 s, respectively. The song sets used for the song-stimulus treatment were constructed in the same manner, except that the songs were recorded from a different male and the mean song lengths of the long-song and short-song sets were 55.6 and 25.4 s, respectively. Importantly, neither total song nor total silence duration differed between the long-song and short-song sets for either treatment, and therefore we exposed all males to the exact same amount of song, regardless of group. By necessity, the two levels of the treatment also differed in the repetition rate of individual songs. However, a previous study reported that song length (or a correlate thereof), but not repetition rate, affected expression of immediate, early genes in the auditory forebrain of female European starlings (Gentner et al., 2001). We assume that this sensitivity specifically to a correlate of song length over song repetition rate also applies to our dependent measures on males, but we do not know this with certainty.

We included the song-stimulus treatment as part of another study examining the effects of the song-environment treatment on neural sensitivity to the length of a novel song stimulus. Because our measurements of singing effort and mean song length preceded the song-stimulus treatment (see explanation below), they were not affected by it. Additionally, the two levels of the song-stimulus treatment were balanced across levels of the song-environment treatment. Consequently, we do not discuss the effects of the song-stimulus treatment further. However, because the brains were collected following exposure to the song-stimulus treatment, we measured monoamine metabolite levels after the subjects were exposed to the song-stimulus treatment. Consequently, we include song-stimulus treatment as a variable in all analyses of monoamine metabolite concentrations to control for its potential to affect monoamine metabolism.

Because both levels of our song-environment treatment were constructed from the recordings of a single male, we cannot extend our conclusions to the effects of song length in general (Kroodsma et al., 2001; Wiley, 2003). However, a previous study showed that females in a mate-choice context prefer this specific long-song set to this specific short-song set (Gentner and Hulse, 2000), and consequently our results are based on responses to

stimuli that are known to differ in their attractiveness to females and, presumably, to differ in the content of information about the competitive quality of the signaller.

Quantification of Male Singing Behavior

We used a male's song count as an indicator of his singing effort. For this study we used songs from day 8 only, because song count was most variable on this day. We defined a song as a series of at least two different motifs containing a combined total of at least 5 notes, wherein we defined a note as a continuous trace on a spectogram and a motif as a group of notes arranged in a fixed order (Adret-Hausberger and Jenkins, 1988; Eens et al., 1989). Although eleven birds sang on day 9, there are important reasons for not using day 9 in the analyses. First, it is difficult to interpret day 9 song because birds were sacrificed at different times during the day, and therefore, some males had longer than others to sing. Second, and most importantly, in this study we are interested in the effects of the songenvironment treatment, and not the song-stimulus treatment, on singing effort. Therefore, the best way to assess the effect of the song-environment treatment on singing effort without any confounding effects of the song-stimulus treatment was to focus on singing effort on day 8. Given these constraints, day 8 was also the time point nearest to that of our tissue collection and therefore the time point most relevant to our measurements of monoamine metabolites.

There were too many song files to analyze them all, so we employed the following sampling technique. Using a random number generator, we selected approximately 45% of the song files recorded on day 8. We distinguished song files from songs because individual files sometimes contained multiple songs, and individual songs sometimes spanned multiple files. The proportion of files selected from each individual reflected the proportion of all files that the individual produced. For example, if all 20 subjects produced a combined total of 1000 song files on day 8, we analyzed 450 of these files (45% of 1000 files = 450 files). If 25% of the total number of song files on day 8 came from individual X, then we randomly selected and analyzed 113 of individual X's day-8 song files (113 files = 25% of 450 song files analyzed from day 8). Within this random subset of song files, we then counted the number of songs and measured the lengths of their computer-drawn spectograms (using the software Raven, version 1.2.1, Cornell Lab of Ornithology).

Quantification of Monoamine Metabolites and Protein

When monoamines are secreted into the synapse, enzymes metabolize them. Thus, as indices of monamine secretion, we quantified levels of their principal metabolites (Moore, 1986). Using previously described protocols (Sockman and Salvante, 2008), we measured tissue concentrations of the dopamine metabolite 3,4-dihydroxyphenylacetic acid (DOPAC; hereafter termed dopamine metabolite), the norepinephrine metabolite 3-methoxy-4hydroxy-phenylglycol (MHPG; hereafter termed norepinephrine metabolite), and the serotonin metabolite 5-hydroxyindolacetic acid (5-HIAA; hereafter termed serotonin metabolite) using reversed-phase high-pressure liquid chromatography (HPLC) with electrochemical detection (Kilts et al., 1981). Briefly, we separated compounds using a Monochrom C18 3 μm column (100 × 4.6 mm, MetaChem) with a mobile phase (1 L, pH 3.9) consisting of sodium phosphate (7.1 g), citric acid (5.76 g), disodium EDTA (50 mg), sodium octyl sulfonate (350 mg) and methanol (13%). We maintained electrode potential at 650 mV with respect to an Ag/AgCl reference electrode. We prepared standard solutions containing a fixed amount (30 ng) of the internal standard (isoproterenol, Sigma) and variable amounts of each monoamine metabolite. We included a five-point standard curve in each assay ($5 \times 50 \,\mu\text{L}$ injections), and used linear regression to fit a line through the standard curve points ($R^2 > 0.99$ for each of the components in each assay).

We cut 300-um sagittal sections from the frozen hemispheres on a cryostat maintained at -10°C, thaw mounted them onto glass microscope slides, and rapidly refroze them on pulverized dry ice. From each of three consecutive sections starting at the midline and using a chilled 1-mm (i.d.) custom-made, thin-walled stainless steel spring-loaded punch, we micropunched a region of the CMM, a caudo-dorsal region of the NCM (NCMd), and a ventral region of the NCM (NCMv), each for which the anatomical boundaries have been described (Sockman et al., 2002) (Fig. 1a). We also sampled from the four principal brain nuclei within the song control system (see introduction). We chose the areas from which to sample based on comparison with the canary and zebra finch brain atlases, and on anatomical boundaries defined by NISSL-staining of the song control nuclei in sections from the alternate, fixed hemisphere from the same birds (Sockman et al., 2009). We micropunched MAN from one section that was approximately 2100 µm from the midline and a region of Area X from three consecutive sections starting approximately 1500 µm from the midline (Fig. 1b). We also took micropunches from a region of HVC from four consecutive sections starting approximately 2400 µm from the midline, and from a region of the RA from two consecutive sections starting approximately 2400 µm from the midline (Fig. 1c).

We expelled the tissue punches into 1.9-ml polypropylene microcentrifuge tubes, froze them on dry ice and stored them at $-80\,^{\circ}\text{C}$ until assay. Immediately before assay we added mobile phase (225 µl) containing 30 ng isoproterenol to each tube. We sonicated the samples and then centrifuged them at 16,000 g for 15 minutes at 4°C. We aspirated the supernatant and injected 50 µl from each sample into the HPLC system. We calculated the metabolite contents by first correcting their peak heights for percent recovery of the internal standard (i.e., the height of the isoproterenol peak) for each sample, and then comparing the values to those obtained for the corresponding metabolites in the standard curve.

We measured the protein content of each sample by dissolving the remaining sample pellet in $100 \mu l \ 0.2 \ N$ NaOH and performing the Bradford protein-dye binding assay (Quick Start Bradford Protein Assay, Bio-Rad) with bovine serum albumin (Bio-Rad) as a standard on a μ Quant microplate spectrophotometer (BioTek) (Bradford, 1976).

Analyses

Our data consisted of a combination of fixed (e.g., song environment, mean song length) and hierarchically-structured random (e.g., individual nested within session) effects, each of which may differ from the others in its correlation structure. In addition, a few samples were lost during HPLC, rendering our dataset unbalanced. Therefore, we analyzed these data in a mixed, multilevel modeling framework using the software Stata IC 10.0 for the Macintosh (Stata Corporation, College Station, TX, USA), which readily accommodates unbalanced, hierarchically-structured combinations of fixed and random effects (Burton et al., 1998; Goldstein et al., 2002; Rabe-Hesketh and Skrondal, 2005). We primarily used Stata's command for multilevel mixed-effects linear regression but, for the song count data, we instead used the command for multilevel mixed-effects Poisson regression. These models estimated parameters with restricted maximum likelihood and used *z*-tests to test the null hypothesis that a coefficient equalled 0. For more information on the rationale for and approach to mixed, multilevel modeling frameworks, see Sockman et al. (2008).

For the majority of analyses, we nested observation (the individual bird's measurement) within session as a random intercept. However, for some metabolite responses in some brain areas, these initial models failed to converge on a solution. In these instances, we reanalyzed the data without the random intercept. With three monoamines of interest measured in each of seven brain areas, there is a large potential for spurious results. Therefore, to maximize the rigor of our analytical strategy, we established a three-branch decision tree to investigate

how the effects of song environment and mean song length on singing effort may be mediated by the secretion of monoamines in the auditory telencephalon and the song control nuclei (Fig. 2). Relationships resulting in p values greater than or equal to 0.05 were deemed unreliable and thus excluded from further consideration. For the first branch of this decision tree, we determined whether song environment, mean song length, or song stimulus predicted (p < 0.05) each monoamine metabolite within each brain area of interest. Also, if an area-specific monoamine metabolite was related to both song environment and mean song length, it had to have had a consistent relationship with both parameters (i.e., both positive or both negative) to be considered a potential candidate for mediating the effects of these parameters on singing effort. This is because singing effort increased with an increase in each parameter (see Results). This first branch of our decision tree resulted in a subset of area-specific metabolite concentrations that were associated with either song environment or mean song length (see Results).

The association of these area-specific metabolite concentrations with song environment or mean song length does not necessarily indicate that they are related in any way to the mediation of singing effort. At a minimum, the metabolite concentration should also be correlated with song count. Hence, in the second branch of our decision tree, we determined whether or not any of the above subset of area-specific monoamine metabolite concentrations predicted song count (Fig. 2). From the initial set of area-specific metabolite concentrations related to song environment or mean song length, this second approach generated a sub-subset of area-specific metabolite concentrations that were also related to song count.

Although a single area-specific metabolite concentration may correlate with both song count and with song environment or mean song length, the monoamine for that metabolite does not necessarily mediate the effects of these factors on singing effort. Additionally, the correlations should be in the same direction (both positive or both negative) because both song environment and mean song length were positively correlated with song count (see Results section below). Therefore, in the third branch of our decision tree, we identified only those area-specific metabolite concentrations that were correlated in the same direction with both singing effort and with at least one of either song environment or mean song length (Fig. 2).

We then assessed which of song environment, mean song length and the viable monoamine metabolite candidates from the third branch of our decision tree best predicted song count on day 8. For this final model, we also applied an information theoretic approach by calculating Akaike's Information Criterion (AIC, Burnham and Anderson, 2003) for a null model containing only an intercept and for each model in which we added the individual factors above in all possible combinations. A model's AIC value reflects its goodness of fit relative to its number of parameters, using the log likelihood of the model while penalizing for each parameter. The model with the most efficient combination of parameters is that with the lowest AIC value. Figures depict means \pm SEM and p-values from models described above.

RESULTS

Singing Behavior

Males exhibited considerable variation in song count (singing effort) on day 8 (range: 0 to 431 songs). Moreover, day-8 song count increased with the quality (length) of both the song environment (p < 0.001; Fig. 3a) and the male's own song (p < 0.001; Fig. 3b) (Table 2). Males that sang on day 8 (n = 18) exhibited over 2.5-fold inter-individual variation in mean song length (range: 16.5 to 43.1 s). However, we found no effect of song environment on

mean song length (z = -0.04, p > 0.2), indicating that our extrinsic factor did not significantly affect our intrinsic factor.

It is possible that the effect of song environment on song count may have been due to an effect on reproductive development. However, males in the long song environment (14.37 \pm 3.57 mg) had similar dry testes masses to those of males exposed to the short song environment (11.06 \pm 2.08 mg) (p > 0.2) and therefore, were not likely advanced reproductively, though, it should be noted that testis mass is at best a very gross measure of reproductive development.

What Factors Best Explain Variation in Singing Effort?

Using our three-branch decision tree, we found four viable monoamine candidates whose area-specific metabolism was correlated with both singing effort and either song environment or mean song length and in a consistent direction (both positive or both negative) (Fig. 4). Concentrations of serotonin metabolite in the CMM and the NCMv and of dopamine metabolite in the NCMv were consistently positively related to both song environment (p = 0.011, Fig. 4a; p = 0.005, Fig. 4e; and p = 0.013, Fig. 4c, respectively) and song count on day 8 (p < 0.001, Fig. 4b; p < 0.001, Fig. 4f; and p < 0.001, Fig. 4d, respectively) (Table 3, 5). Similarly, concentrations of norepinephrine metabolite in Area X were negatively correlated with both mean song length (p = 0.014; Fig. 4g) and song count on day 8 (p < 0.001; Fig. 4h) (Table 4, 5). The relationship between dopamine metabolite in the NCMv and song count appears as though it might be driven by a single point with a high value for the concentration of dopamine metabolite (Fig. 4d). However, when that value is removed from the analysis, the relationship remains reliable (coefficient = 0.442 ± 0.104 , z = 4.23, p < 0.001). In addition to being related to song environment and song count, the concentration of serotonin metabolite in the NCMv was also negatively related to the songstimulus treatment (p = 0.033; Table 3).

If song environment affects singing effort via metabolism of serotonin in the CMM and the NCMv and dopamine in the NCMv, and if mean song length influences singing effort directly through norepinephrine metabolism in Area X, then we predict that when all of these factors are included in one model variation in singing effort should best be explained by monoamine metabolite levels as opposed to being explained by mean song length and song environment. In the full model, we found no reliable relationship between song count and dopamine metabolite levels in the NCMv or between song count and serotonin metabolite levels in the NCMv (both p > 0.2). Moreover, AIC values (not shown) indicated that the most efficient model did not include the concentration of dopamine metabolite in the NCMv or the concentration of serotonin metabolite in the NCMv as factors. However, AIC did support the inclusion of song environment (p = 0.002), mean song length (p < 0.001), the concentration of serotonin metabolite in the CMM (p = 0.019), and the concentration of norepinephrine metabolite in Area X (p < 0.001) in the model (Table 6). Therefore, serotonin metabolism in the CMM of male starlings may be involved in mediating the effects on singing effort of variation in the quality of competition in a male's social environment. Similarly, the association between a male's quality and his singing effort may be mediated, in part, by modulation of norepinephrine metabolism in Area X.

Additional Monoamine Metabolite Relationships

Although only two viable candidate monoamines emerged from our analytical approach described above, we discovered additional relationships between song environment, mean song length, song count, and monoamine metabolism in the auditory telencephalon and song control nuclei that did not fulfill all of the criteria in our decision tree, but may nonetheless provide insight into the relationships between region-specific monoamine metabolism and

plasticity in singing effort and other behaviors. We found that although males exposed to the long song environment had higher concentrations of norepinephrine metabolite in the CMM (p = 0.046; Table 3) and lower concentrations of dopamine metabolite in the MAN (p =0.027; Table 4) and HVC (p = 0.025; Table 4) than males exposed to the short song environment, song count on day 8 was not related to any of these monoamine metabolites (p > 0.2 in all cases; Table 5). Furthermore, the positive correlations between the concentrations of both serotonin metabolite in the NCMd and dopamine metabolite in Area X and song count on day 8 (p < 0.001 and p = 0.032; Table 5) were not consistent with their negative relationships with mean song length (p = 0.016; Table 3 and p = 0.013; Table 4). Consequently, we did not consider the metabolism of norepinephrine in the CMM, dopamine in the MAN, HVC and Area X, and serotonin in the NCMd as being directly involved in the neural pathways mediating the effects of song environment and mean song length on song count. However, the secretion of these monoamines in these brain areas may be influenced by the context-dependent changes in singing effort or be involved in the neural pathways mediating the effects of song environment and mean song length on other behaviors.

DISCUSSION

Context-dependent modulation of communication behavior is influenced by both extrinsic factors that can be attributed to aspects of the signalling environment and factors that are intrinsic to the signaller, such as his condition or quality. Birdsong is an excellent example of how the extrinsic physical and social environment and intrinsic quality, motivation, and reproductive state of the signaller influence communication behavior. However, how the songbird brain integrates these intrinsic and extrinsic factors and modulates signalling effort accordingly is poorly understood. The present study is consistent with the hypothesis that norepinephrine and serotonin metabolism in the telencephalon may, in part, mediate the effects of some intrinsic and extrinsic factors on male singing effort.

Male Song Quality, Norepinephrine in the Song Control System, and Singing Effort

Our results are consistent with the hypothesis that the association between a male's intrinsic song quality and his singing effort may be mediated by modulation of norepinephrine secretion in Area X. In male zebra finches electrophysiological and immediate early gene activity in Area X are lower when song is "directed" towards a conspecific, which often occurs in the context of courtship, than when the singer is not oriented towards a conspecific, and is thus singing "undirected" song (Jarvis et al., 1998; Hessler and Doupe, 1999). A proposed purpose of undirected singing is song practice (Jarvis et al., 1998). The increase in Area X activity during practice bouts may be due to Area X's perceptual role in processing information about a male's own song (Mello and Ribeiro, 1998; Doupe and Konishi, 1991), making for the intriguing possibility that this area is involved in mediating the influence of a male's own song quality on his singing effort.

Sex steroids, which are elevated during courtship, are likely involved in the mechanism linking intrinsic male song quality, norepinephrine secretion in Area X, and singing effort. Castration and hormone replacement studies on zebra finches suggest that the combination of low sex steroid levels and elevated norepinephrine secretion in Area X has a suppressive effect on singing effort (Barclay and Harding, 1988; 1990). Furthermore, pharmacological treatment that reduces noradrenergic input to the forebrain results in an increase in immediate early gene expression in Area X during directed singing (Castelino and Ball, 2005), further supporting the idea that norepinephrine suppresses neural activity in Area X.

Competitive Environment, Serotonin in the Auditory Telencephalon, and Singing Effort

Our findings are also consistent with the hypothesis that serotonin secretion in the CMM of male starlings may be involved in mediating the effects of extrinsic social factors on singing effort. The responsiveness of the songbird CMM to conspecific male song has been well documented (Mello et al., 1992; MacDougall-Shackleton et al., 1998; Ball et al., 2006; Sockman, 2007), and the sensitivity of the female's auditory telencephalon to variation in conspecific male song length is known to be modulated by the quality of songs in the acoustic environment (Sockman et al., 2002; 2005a). The CMM's direct projections to HVC (Bauer et al., 2008) may allow variation in serotonergically-modulated CMM activity to translate into plasticity in singing effort.

In the inferior colliculus, the principal midbrain nucleus of the mammalian auditory system, serotonin alters the magnitude of neuronal responses to relevant conspecific acoustic signals by modulating the range of frequencies to which the neurons are sensitive, via gain control and frequency tuning of specific neurons (Hurley and Pollak, 1999; 2001; 2005; Hurley et al., 2002; 2004). It has been hypothesized that serotonin's selective influence on auditory neurons could lead to differential pattern encoding for different auditory signals depending on the arousal state of the listener and the incoming auditory stimuli, which could ultimately lead to reconfiguration of the neural circuitry and responsiveness of the auditory system (Hurley et al., 2002; 2004). Thus, one hypothesis is that socially modulated serotonin metabolism in the CMM reshapes the HVC-projecting neurons in the CMM, changing the responsiveness of the auditory telencephalon and thus the song control system to conspecific song, resulting in an increase in singing effort.

Modulation of a Complex Communication Signal

We expected that if the effects of song environment and mean song length on singing effort were solely mediated by secretion of serotonin in the CMM and norepinephrine in Area X, respectively, then serotonin and norepinephrine secretion in these brain areas should explain variation in singing effort that would otherwise be attributed to the extrinsic and intrinsic factors. Contrary to our prediction, the best-fit model included all four parameters, and each explained a portion of the variation in singing effort. Indeed, the context-dependent modulation of singing effort is, not surprisingly, more complex than our simple dual-pathway model (forward solid arrows, Fig. 5). Additional mediating neural mechanisms must also be involved in the neural pathways through which male quality and social environment influence singing effort (dashed arrows, Fig. 5) because our intrinsic and extrinsic factors explained a portion of the variation in singing effort that was not attributable to secretion of norepinephrine in Area X or serotonin in the CMM. For example, our intrinsic and extrinsic factors may influence monoamine secretion in brain regions from which we did not sample.

Norepinephrine secretion in Area X and serotonin secretion in the CMM are also likely influenced by intrinsic and extrinsic factors other than the male's own quality and the quality of perceived competitors (dotted arrows, Fig. 5). Noradrenergic fibers innervating the songbird auditory telencephalon and song control system project from cell bodies located within the locus coeruleus and subcoeruleus regions (Mello et al., 1998;Appeltants et al., 2000;Ball et al., 2004). Telencephalic serotonergic fibers likely originate from the raphe nuclei, as they do in mammals (Bobillier et al., 1976;Halaris et al., 1976;Moore et al., 1978;Azmitia and Segal, 1978;Klepper and Herbert, 1991;Jacobs and Azmitia, 1992). These monoaminergic cell bodies and fibers receive input from a variety of brain regions, including the hypothalamus and regions of the limbic system (Jacobs and Azmitia, 1992;Ball et al., 2004). Consequently, the extent of monoaminergic secretion in Area X and

the CMM may be indirectly influenced by a multitude of intrinsic and extrinsic factors that modulate the signalling properties of these afferent connections.

Factors in addition to the quality of the male and his competitive environment also must influence his singing effort, and these other factors' effects may be mediated by neural mechanisms other than, or in addition to, monoaminergic activity in the auditory telencephalon and song control system (dotted-dashed arrow, Fig. 5). Central and peripheral levels of sex steroids are elevated during the period of courtship and mate acquisition, often reflect individual condition and quality, and respond rapidly to social interactions (Balthazart, 1983; Wingfield et al., 1993; Ball and Balthazart, 2004; Remage-Healey et al., 2008). Variation in levels of sex steroids and their biologically active metabolites could lead to differential up-regulation and activation of androgen and estrogen receptors within the song control system and consequently modulate monoaminergic activity in these areas and singing activity in response to the male's own condition or social interactions (Schlinger, 1997; Ball et al., 2003; Gahr, 2004; Harding, 2004; Meitzen et al., 2007; Remage-Healey et al., 2008). The signaller's sexual motivation and arousal may also play a role in regulating signalling behavior in the context of reproduction. Numerous studies have reported relationships between the expression of sexually motivated behaviors, including male birdsong, and variation in gene expression, morphology, and neurotransmitter and neuroendocrine activity in brain regions that are associated with motivation and arousal, such as the nucleus taeniae, area ventralis of Tsai, medial bed nucleus of the stria terminalis, ventromedial nucleus of the hypothalamus, and the medial preoptic area, which includes the medial preoptic nucleus (Hull et al., 1999; Riters and Ball, 1999; Riters et al., 2000,2004; Maney and Ball, 2003; Sewards and Sewards, 2003; Heimovics and Riters, 2005,2006,2007; Balthazart and Ball, 2007; Heimovics et al., 2009). Future studies examining the neural mechanisms through which other extrinsic and intrinsic factors influence singing effort are crucial for broadening our understanding of how variability in the signaller's environment or in the signaller itself can shape adaptive behavioral plasticity.

Finally, variation in singing effort may itself elicit the observed differences in norepinephrine secretion in Area X, serotonin secretion in the CMM, and other area-specific monoamine metabolite levels that were individually related to singing effort, but either did not explain variation in singing effort in the final model or exhibited contradictory relationships with song count and mean song length or song environment, and therefore were not subjected to further analyses (see Materials and Methods and Results) (reverse solid arrows, Fig. 5). Numerous studies have shown that context-dependent variation in singing (e.g., singing directed or undirected song) or hearing this variation in one's own songs can cause changes in the song-control and auditory-processing brain regions, including differences in neural activity, immediate early gene expression, and monoamine metabolism (Jarvis and Nottebohm, 1997; Jarvis et al., 1998; Hessler and Doupe, 1999; reviewed in Doupe et al., 2005; Sasaki et al., 2006). This may explain the significant relationships between singing effort and monoamine metabolism in brain regions that have previously been associated with either context-dependent singing, such as dopamine secretion in Area X (Hessler and Doupe, 1999; Sasaki et al., 2006), or processing contextdependent variation in song, such as serotonin secretion in the NCMv and NCMd and dopamine secretion in the NCMd (Jarvis and Nottebohm, 1997; Sockman and Salvante, 2008).

This study implicates norepinephrine and serotonin secretion within the male starling song control system and auditory telencephalon, respectively, as potential mediators of the effects of intrinsic and extrinsic factors on the modulation of birdsong production. Although some of the relationships we have shown are correlative, identifying these relationships are a critical first step in the discovery of neural mechanisms underlying context-dependent

plasticity in singing effort. Future studies involving experimental manipulation of monoaminergic activity in specific brain regions should provide insight into the potential of these neurotransmitters as direct regulators of plasticity in communication signalling.

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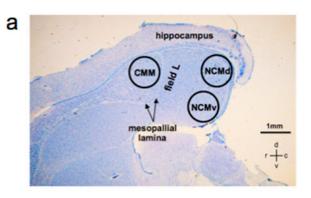
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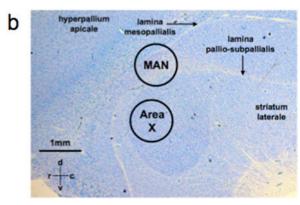
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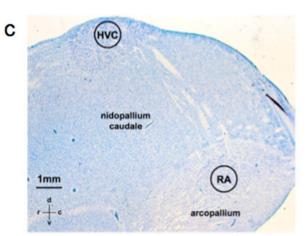


Figure 1. Photomicrographs of regions within the forebrain of European starlings sampled for HPLC analysis of central monoamine metabolism. Sample punches (1 mm i.d.) taken from a) the caudomedial mesopallium (CMM), dorsal caudomedial nidopallium (NCMd), and ventral caudomedial nidopallium (NCMv) of the auditory telencephalon, and b) the magnocellular nucleus of the anterior nidopallium (MAN), Area X, c) the robust nucleus of the arcopallium (RA) and HVC (initialism used as a proper name) of the song control system.

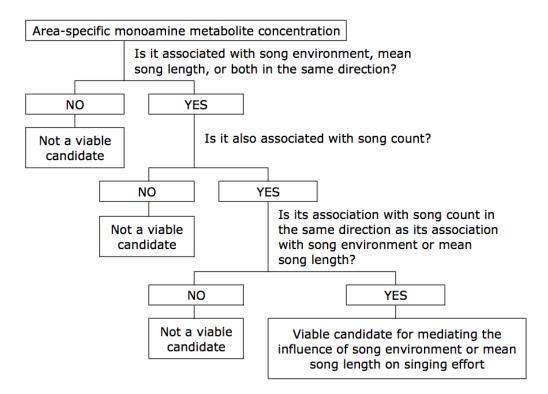


Figure 2.Three-branch decision tree for determining viable area-specific monoamine metabolite candidates for mediating the influence of song environment and mean song length on singing effort.

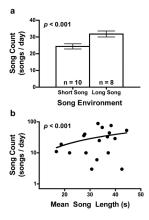


Figure 3. Change in song count with respect to a) song environment (mean \pm SE) and b) the length of evoked songs (Y axis has a log-10 scale because count data tend to follow a Poisson distribution)..

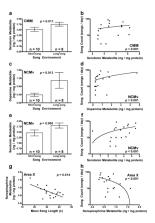


Figure 4.

Region-specific monoamine candidates for mediating the effects of song environment and mean song length on singing effort. Serotonin metabolism in the CMM (a, b), dopamine metabolism in the NCMv (c, d) and serotonin metabolism in the NCMv (e, f) were positively associated with both song environment and song count. Norepinephrine metabolism in Area X was negatively associated with both g) mean song length and h) song count. Y axes showing song counts have a log-10 scale because count data tend to follow a Poisson distribution.

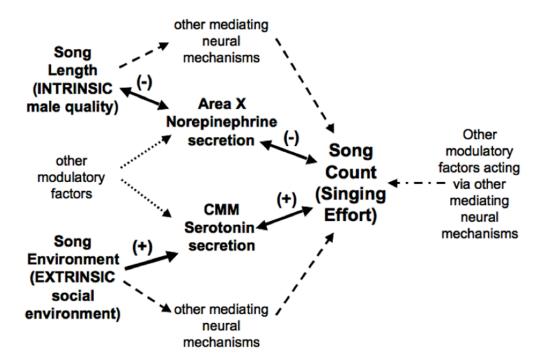


Figure 5. Hypothetical model by which an individual's mean song length, its song environment, and other potential intrinsic and extrinsic factors may influence singing effort (see Fig. 1 for abbreviations). Norepinephrine secretion in Area X and serotonin secretion in the CMM may, in part, mediate the influence of male quality and the quality of the perceived social environment, respectively, on singing effort (forward solid arrows). Alternatively, the variation in singing effort may elicit the observed changes in norepinephrine secretion in Area X and serotonin secretion in the CMM (reverse solid arrows). Male quality and the quality of the perceived social environment also likely influence singing effort through alternate neural pathways (dashed arrows). Other intrinsic and extrinsic factors may also affect secretion of norepinephrine in Area X and serotonin in the CMM (dotted arrows), through which they may ultimately influence singing effort. The unexplained variation in singing effort can be attributed to alternative intrinsic and extrinsic modulatory factors acting on additional neural pathways to regulate plasticity in singing effort (dotted-dashed arrow).

Table 1

Song Treatments and Experimental Design

Treatment	Duration	Male Recorded		Experimen	tal Design		
Song Environment	7 days	a	long song (n=1)	long song (n=1)	short song (n=1)	short song (n=1)	x 5 sessions
Day 8 (no song	1 day		silence (n=1)	silence (n=1)	silence (n=1)	silence	x 5 sessions
playback)			\$	\$	\$	4	
Song Stimulus	30 minutes	b	long song (n=1)	short song (n=1)	long song (n=1)	short song (n=1)	x 5 sessions

Table 2 Statistical effects of song environment * and mean song length (s) on song count (n = 18 males).

Independent variables	Estimate	Standard Error	z Score	<i>p</i> -value
Song Environment	0.4657	0.0874	5.33	< 0.001
Mean Song Length	0.1029	0.0121	8.48	< 0.001
Intercept	-0.2731	0.4784	-0.57	> 0.2

^{*}Coded 0 for short song, 1 for long song

Table 3

Statistical effects of song environment*, song stimulus*, and mean song length (s) on monoamine metabolite levels (ng/mg protein) in the caudomedial mesopallium (CMM), dorsal caudomedial nidopallium (NCMd), and ventral caudomedial nidopallium (NCMv) of the auditory telencephalon (n = 18 males).

Independent variables Estimate Standard Error z Score p-value Dopamine metabolite in CWT Song Environment 0.4193 0.3936 1.07 > 0.2 Song-Stimulus Treatment −0.6609 0.4050 −1.63 0.103 Mean Song Length 0.0024 0.0348 0.07 > 0.2 Intercept 0.7962 1.2062 0.66 > 0.2 Norepinephrine metabolite in CMM Tuge 0.1242 1.99 0.046 Song-Stimulus Treatment 0.1035 0.1275 0.81 > 0.2 Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001 Serotonin metabolite in CMM Song-Stimulus Treatment −0.5574 0.4137 −1.35 0.178 Mean Song Length −0.0218 0.0309 −0.71 > 0.2 Intercept 4.0433 1.0817 > 0.2 Song-Stimulus Treatment −0.0996 0.1273 −0.78 > 0	Area-specific monoamine metabolite								
Song Environment 0.4193 0.3936 1.07 > 0.2 Song-Stimulus Treatment −0.6609 0.4050 −1.63 0.103 Mean Song Length 0.0024 0.0348 0.07 > 0.2 Intercept 0.7962 1.2062 0.66 > 0.2 Norepinephrine metabolite in CMM Song Environment 0.2476 0.1242 1.99 0.046 Song-Stimulus Treatment 0.1035 0.1275 0.81 > 0.2 Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001 Serotonin metabolite in CMM Song-Stimulus Treatment −0.0574 0.4137 −1.35 0.178 Mean Song Length −0.0218 0.0309 −0.71 > 0.2 Intercept 4.0433 1.0817 3.74 < 0.001 Dopamine metabolite in NCMd Song-Stimulus Treatment −0.0196 0.1273 −0.78 > 0.2	Independent variables	Estimate	Standard Error	z Score	<i>p</i> -value				
Song-Stimulus Treatment	Dopamine metabolite in CMM								
Mean Song Length 0.0024 0.0348 0.07 > 0.2 Intercept 0.7962 1.2062 0.66 > 0.2 Norepinephrine metabolite in CMM Song Environment 0.2476 0.1242 1.99 0.046 Song-Stimulus Treatment 0.1035 0.1275 0.81 > 0.2 Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001	Song Environment	0.4193	0.3936	1.07	> 0.2				
Intercept 0.7962 1.2062 0.66 > 0.2 Norepinephrine metabolite in CMM CMM Song Environment 0.2476 0.1242 1.99 0.046 Song-Stimulus Treatment 0.1035 0.1275 0.81 > 0.2 Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001 Serotonin metabolite in CMW Song Environment 1.0236 0.4031 2.54 0.011 Song-Stimulus Treatment −0.5574 0.4137 −1.35 0.178 Mean Song Length −0.0218 0.0309 −0.71 > 0.2 Intercept 4.0433 1.0817 3.74 < 0.001 Dopamine metabolite in NCMd Song Environment 0.1325 0.1238 1.07 > 0.2 Mean Song Length −0.0160 0.0105 −1.52 0.128 Intercept 0.8565 0.3651 2.35 0.019 Norepinephrine metabolite in NCMd <td>Song-Stimulus Treatment</td> <td>-0.6609</td> <td>0.4050</td> <td>-1.63</td> <td>0.103</td>	Song-Stimulus Treatment	-0.6609	0.4050	-1.63	0.103				
Norepinephrine metabolite in CMM Song Environment 0.2476 0.1242 1.99 0.046 Song-Stimulus Treatment 0.1035 0.1275 0.81 > 0.2 Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001	Mean Song Length	0.0024	0.0348	0.07	> 0.2				
Song Environment 0.2476 0.1242 1.99 0.046 Song-Stimulus Treatment 0.1035 0.1275 0.81 > 0.2 Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001	Intercept	0.7962	1.2062	0.66	> 0.2				
Song-Stimulus Treatment 0.1035 0.1275 0.81 > 0.2 Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001	Norepinephrine metabolite in CMM								
Mean Song Length −0.0135 0.0095 −1.42 0.155 Intercept 1.1084 0.3334 3.32 < 0.001	Song Environment	0.2476	0.1242	1.99	0.046				
Intercept 1.1084 0.3334 3.32 < 0.001 Serotonin metabolite in CMM Song Environment 1.0236 0.4031 2.54 0.011 Song-Stimulus Treatment −0.5574 0.4137 −1.35 0.178 Mean Song Length −0.0218 0.0309 −0.71 >0.2 Intercept 4.0433 1.0817 3.74 <0.001	Song-Stimulus Treatment	0.1035	0.1275	0.81	> 0.2				
Serotonin metabolite in CMM Song Environment 1.0236 0.4031 2.54 0.011 Song-Stimulus Treatment −0.5574 0.4137 −1.35 0.178 Mean Song Length −0.0218 0.0309 −0.71 >0.2 Intercept 4.0433 1.0817 3.74 <0.001	Mean Song Length	-0.0135	0.0095	-1.42	0.155				
Song Environment 1.0236 0.4031 2.54 0.011 Song-Stimulus Treatment −0.5574 0.4137 −1.35 0.178 Mean Song Length −0.0218 0.0309 −0.71 >0.2 Intercept 4.0433 1.0817 3.74 <0.001	Intercept	1.1084	0.3334	3.32	< 0.001				
Song-Stimulus Treatment -0.5574 0.4137 -1.35 0.178 Mean Song Length -0.0218 0.0309 -0.71 >0.2 Intercept 4.0433 1.0817 3.74 < 0.001	Serotonin metabolite in CM	M							
Mean Song Length −0.0218 0.0309 −0.71 >0.2 Intercept 4.0433 1.0817 3.74 <0.001	Song Environment	1.0236	0.4031	2.54	0.011				
Intercept 4.0433 1.0817 3.74 < 0.001 Dopamine metabolite in NCMd Song Environment 0.1325 0.1238 1.07 > 0.2 Song-Stimulus Treatment −0.0996 0.1273 −0.78 > 0.2 Mean Song Length −0.0160 0.0105 −1.52 0.128 Intercept 0.8565 0.3651 2.35 0.019 Norepinephrine metabolite in NCMd Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length −0.0127 0.0153 −0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept	Song-Stimulus Treatment	-0.5574	0.4137	-1.35	0.178				
Dopamine metabolite in NCMd Song Environment 0.1325 0.1238 1.07 > 0.2 Song-Stimulus Treatment -0.0996 0.1273 -0.78 > 0.2 Mean Song Length -0.0160 0.0105 -1.52 0.128 Intercept 0.8565 0.3651 2.35 0.019 Norepinephrine metabolite in NCMd Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length -0.0127 0.0153 -0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment -1.3394 0.7053 -1.90 0.058 Mean Song Length -0.1266 0.0526 -2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Mean Song Length	-0.0218	0.0309	-0.71	> 0.2				
Song Environment 0.1325 0.1238 1.07 > 0.2 Song-Stimulus Treatment −0.0996 0.1273 −0.78 > 0.2 Mean Song Length −0.0160 0.0105 −1.52 0.128 Intercept 0.8565 0.3651 2.35 0.019 Norepinephrine metabolite in NCMd Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length −0.0127 0.0153 −0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Intercept	4.0433	1.0817	3.74	< 0.001				
Song-Stimulus Treatment -0.0996 0.1273 -0.78 > 0.2 Mean Song Length -0.0160 0.0105 -1.52 0.128 Intercept 0.8565 0.3651 2.35 0.019 Norepinephrine metabolite in NCMd Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length -0.0127 0.0153 -0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment -1.3394 0.7053 -1.90 0.058 Mean Song Length -0.1266 0.0526 -2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Dopamine metabolite in NC	Md							
Mean Song Length −0.0160 0.0105 −1.52 0.128 Intercept 0.8565 0.3651 2.35 0.019 Norepinephrine metabolite in NCMd Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length −0.0127 0.0153 −0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Song Environment	0.1325	0.1238	1.07	> 0.2				
Intercept 0.8565 0.3651 2.35 0.019 Norepinephrine metabolite in NCMd Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length −0.0127 0.0153 −0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Song-Stimulus Treatment	-0.0996	0.1273	-0.78	> 0.2				
Norepinephrine metabolite in NCMd Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length −0.0127 0.0153 −0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Mean Song Length	-0.0160	0.0105	-1.52	0.128				
Song Environment 0.0989 0.1834 0.54 > 0.2 Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length −0.0127 0.0153 −0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Intercept	0.8565	0.3651	2.35	0.019				
Song-Stimulus Treatment 0.0170 0.1885 0.09 > 0.2 Mean Song Length −0.0127 0.0153 −0.83 > 0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Norepinephrine metabolite i	•							
Mean Song Length −0.0127 0.0153 −0.83 >0.2 Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment −1.3394 0.7053 −1.90 0.058 Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Song Environment	0.0989	0.1834	0.54	> 0.2				
Intercept 1.1905 0.5319 2.24 0.025 Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment -1.3394 0.7053 -1.90 0.058 Mean Song Length -0.1266 0.0526 -2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Song-Stimulus Treatment	0.0170	0.1885	0.09	> 0.2				
Serotonin metabolite in NCMd Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment -1.3394 0.7053 -1.90 0.058 Mean Song Length -0.1266 0.0526 -2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Mean Song Length	-0.0127	0.0153	-0.83	> 0.2				
Song Environment 0.6173 0.6872 0.90 > 0.2 Song-Stimulus Treatment -1.3394 0.7053 -1.90 0.058 Mean Song Length -0.1266 0.0526 -2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Intercept	1.1905	0.5319	2.24	0.025				
Song-Stimulus Treatment -1.3394 0.7053 -1.90 0.058 Mean Song Length -0.1266 0.0526 -2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Serotonin metabolite in NC	Md							
Mean Song Length −0.1266 0.0526 −2.40 0.016 Intercept 8.0397 1.8441 4.36 < 0.001	Song Environment	0.6173	0.6872	0.90	> 0.2				
Intercept 8.0397 1.8441 4.36 < 0.001	Song-Stimulus Treatment	-1.3394	0.7053	-1.90	0.058				
•	Mean Song Length	-0.1266	0.0526	-2.40	0.016				
Dopamine metabolite in NCMv	Intercept	8.0397	1.8441	4.36	< 0.001				
-	Dopamine metabolite in NC	Dopamine metabolite in NCMv							
Song Environment 1.1129 0.4504 2.47 0.013	Song Environment	1.1129	0.4504	2.47	0.013				
Song-Stimulus Treatment -0.8647 0.4623 -1.87 0.061	Song-Stimulus Treatment	-0.8647	0.4623	-1.87	0.061				
Mean Song Length -0.0099 0.0349 -0.28 > 0.2	Mean Song Length	-0.0099	0.0349	-0.28	> 0.2				
Intercept 1.1888 1.2230 0.97 > 0.2	Intercept	1.1888	1.2230	0.97	> 0.2				
Norepinephrine metabolite in NCMv	Norepinephrine metabolite i	in NCMv							
Song Environment 0.4327 0.1642 2.63 0.008	Song Environment	0.4327	0.1642	2.63	0.008				

Area-specific monoamine metabolite							
Independent variables	Estimate	Standard Error	z Score	<i>p</i> -value			
Song-Stimulus Treatment	-0.2832	0.1688	-1.68	0.093			
Mean Song Length	-0.0326	0.0138	-2.37	0.018			
Intercept	1.7773	0.4786	3.71	< 0.001			
Serotonin metabolite in NCMv							
Song Environment	1.4067	0.5038	2.79	0.005			
Song-Stimulus Treatment	-1.1028	0.5181	-2.13	0.033			
Mean Song Length	-0.0460	0.0437	-1.05	> 0.2			
Intercept	4.9051	1.5161	3.24	< 0.001			

^{*} Coded 0 for short song, 1 for long song

Table 4

Statistical effects of song environment*, song-stimulus*, and mean song length (s) on monoamine metabolite levels (ng/mg protein) in the magnocellular nucleus of the anterior nidopallium (MAN; n = 18 males), Area X (n = 18 males), HVC (initialism used as a proper name; n = 16 males), and the robust nucleus of the arcopallium (RA; n = 18 males) of the song control system.

Area-specific monoamine metabolite								
Independent variables	Estimate	Standard Error	z Score	<i>p</i> -value				
Dopamine metabolite in MAN								
Song Environment	-1.0367	0.4692	-2.21	0.027				
Song-Stimulus Treatment	-0.2369	0.4847	-0.49	> 0.2				
Mean Song Length	0.0500	0.0496	1.01	> 0.2				
Intercept	0.4566	1.7160	0.27	> 0.2				
Norepinephrine metabolite	in MAN							
Song Environment	-0.1294	0.3855	-0.34	> 0.2				
Song-Stimulus Treatment	0.2359	0.3956	0.60	> 0.2				
Mean Song Length	-0.0094	0.0295	-0.32	> 0.2				
Intercept	1.4070	1.0344	1.36	0.174				
Serotonin metabolite in MA	N							
Song Environment	-1.6444	1.3901	-1.18	> 0.2				
Song-Stimulus Treatment	1.7541	1.4298	1.23	> 0.2				
Mean Song Length	-0.0390	0.1214	-0.32	> 0.2				
Intercept	4.6247	4.2144	1.10	> 0.2				
Dopamine metabolite in Are	ea X							
Song Environment	-0.8380	2.2441	-0.37	> 0.2				
Song-Stimulus Treatment	-4.7413	2.3031	-2.06	0.040				
Mean Song Length	-0.4259	0.1719	-2.48	0.013				
Intercept	34.4193	6.0221	5.72	0.001				
Norepinephrine metabolite	Norepinephrine metabolite in Area X							
Song Environment	-0.0110	0.1277	-0.09	> 0.2				
Song-Stimulus Treatment	0.1298	0.1314	0.99	> 0.2				
Mean Song Length	-0.0278	0.0113	-2.45	0.014				
Intercept	1.5569	0.3929	3.96	< 0.001				
Serotonin metabolite in Are	a X							
Song Environment	0.0985	0.4842	0.20	> 0.2				
Song-Stimulus Treatment	-0.6392	0.4970	-1.29	0.198				
Mean Song Length	-0.0553	0.0371	-1.49	0.136				
Intercept	4.8048	1.2994	3.70	< 0.001				
Dopamine metabolite in HV	C							
Song Environment	-0.2669	0.1189	-2.24	0.025				
Song-Stimulus Treatment	0.0106	0.1207	0.09	> 0.2				
Mean Song Length	-0.0133	0.0086	-1.54	0.123				
Intercept	1.0362	0.2998	3.46	< 0.001				
Norepinephrine metabolite in HVC								

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Area-specific monoamine metabolite Independent variables Estimate Standard Error z Score p-value Song Environment -0.0207 0.1139 -0.18 > 0.2 Song-Stimulus Treatment -0.13550.1157 -1.17> 0.2 -0.55-0.00470.0085 > 0.2 Mean Song Length Intercept 0.7531 0.2951 2.55 0.011 Serotonin metabolite in HVC Song Environment 0.3211 0.5966 0.54 > 0.2 Song-Stimulus Treatment -0.63540.6057 -1.05> 0.2 Mean Song Length 0.0395 0.0431 0.92 > 0.2 Intercept 2.6759 1.5042 1.78 0.075 Dopamine metabolite in RA Song Environment 0.2704 0.1618 1.67 0.095 Song-Stimulus Treatment -0.17210.1660 -1.04> 0.2 Mean Song Length -0.02360.0124 -1.900.057 Intercept 1.3618 0.4342 3.14 0.002 Norepinephrine metabolite in RA Song Environment 0.1155 0.1055 1.09 > 0.2 -0.0653> 0.2 Song-Stimulus Treatment 0.1089 -0.60-0.00190.0107 -0.17> 0.2 Mean Song Length 0.3066 0.3709 0.83 > 0.2 Intercept Serotonin metabolite in RA Song Environment 0.8813 1.1273 0.78 > 0.2 > 0.2 Song-Stimulus Treatment -0.87091.1570 -0.75Mean Song Length -0.01100.0863 -0.13> 0.2

5.8994

3.0252

1.95

0.051

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Intercept

^{*}Coded 0 for short song, 1 for long song

Table 5

Statistical effects of monoamine metabolite levels (ng/mg protein) in select regions of the auditory telencephalon (see Table 3 for abbreviations) and song control system (see Table 4 for abbreviations) on song count (n = 16 males for dopamine metabolite in HVC; n = 18 males for all other monoamine metabolites). Comparisons were made between the directions of the significant relationships (i.e. positive or negative correlations) and the direction of the previously reported significant relationships between the monoamine metabolite of interest and either song environment or mean song length.

Independent Variables	Estimate	Standard Error	z Score	<i>p</i> -value
Norepinephrine metabolite in CMM	-0.1485	0.1752	-0.85	> 0.2
Intercept	3.2895	0.3057	10.76	< 0.001
Serotonin metabolite in CMM	0.4769	0.0552	8.64	< 0.001
Intercept	1.4405	0.3809	3.78	< 0.001
CONSISTENT with positive relations	hip with son	g environment		
Serotonin metabolite in NCMd	0.1784	0.0278	6.43	< 0.001
Intercept	2.5322	0.3046	8.31	< 0.001
INCONSISTENT with negative relati	onship with i	mean song length		
Dopamine metabolite in NCMv	0.2145	0.0348	6.16	< 0.001
Intercept	2.9645	0.2451	12.10	< 0.001
CONSISTENT with positive relations	hip with son	g environment		
Serotonin metabolite in NCMv	0.0200	0.0100	5.78	< 0.001
Intercept	2.3342	0.2451	7.99	< 0.001
CONSISTENT with positive relations	hip with son	g environment		
Dopamine metabolite in MAN	-0.0204	0.0366	-0.56	> 0.2
Intercept	3.1957	0.2833	11.28	< 0.001
Norepinephrine metabolite in Area X	-0.5311	0.1469	-3.61	< 0.001
Intercept	3.5351	0.2633	13.43	< 0.001
CONSISTENT with negative correlat	ion with mea	n song length		
Dopamine metabolite in Area X	0.0020	0.0010	2.15	0.032
Intercept	2.8397	0.3128	9.08	< 0.001
INCONSISTENT with negative corre	lation with m	nean song length		
Dopamine metabolite in HVC	-0.0217	0.1763	-0.12	> 0.2
Intercept	3.1811	0.2879	11.05	< 0.001

Table 6

The best-fit model based on our data of the modulatory factors and central monoaminergic mechanisms influencing song count (n = 18 males; See Table 3 for abbreviation).

Independent variables	Estimate	Standard Error	z Score	p-value
Serotonin metabolite in CMM (ng/mg protein)	0.1737	0.0744	2.34	0.019
Norepinephrine metabolite in Area X (ng/mg protein)	-0.7725	0.2098	-3.68	< 0.001
Song environment*	0.3807	0.1242	3.07	0.002
Mean song length (s)	0.0949	0.0131	7.26	< 0.001
Intercept	-0.0767	0.5355	-0.14	> 0.2

 $^{^*}$ Coded 0 for short song, 1 for long song