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## Click synthesis of estradiol–cyclodextrin conjugates as cell compartment selective estrogens

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### Abstract

Cyclodextrin (CD) is a well known drug carrier and excipient for enhancing aqueous solubility. CDs themselves are anticipated to have low membrane permeability because of relatively high hydrophilicity and molecular weight. CD derivatization with 17-beta estradiol (E<sub>2</sub>) was explored extensively using a number of different click chemistries and the cell membrane permeability of synthetic CD–E<sub>2</sub> conjugate was explored by cell reporter assays and confocal fluorescence microscopy. *In simile* with reported dendrimer –E<sub>2</sub> conjugates, CD–E<sub>2</sub> was found to be a stable, extranuclear receptor selective estrogen that penetrated into the cytoplasm.

### Keywords

Cyclodextrin; Estrogen; Bioconjugate; Membrane; Nuclear receptor

## 1. Introduction

β-Cyclodextrins (CDs) are torus-shaped saccharides consisting of a shallow interior cavity, a primary face of 7 hydroxyls and a secondary face of 14 hydroxyl groups. CDs and modified CDs have many interesting biological properties due to their unique structure, which can be altered most readily by chemical modification of the primary face.<sup>1,2</sup> The hydrophobicity of the CD cavity has dominated thinking in terms of CD host–guest chemistry via formation of non-covalent inclusion complexes and native and commercially available CD derivatives, the latter generally heterogeneous compounds, have been used as efficient drug carriers.<sup>3</sup> Alternatively, the CD torus can be used as a scaffold to attach drug moieties in order to enhance their effectiveness by facilitating drug delivery or specificity; for example, CD–epoxysuccinyl peptide conjugates were reported as inhibitors of membrane-associated cathepsin B, known to be secreted in several tumor cell lines.<sup>4</sup> The synthesized CD–peptide conjugates were full inhibitors of extranuclear cysteine proteases and were reported not to be membrane-permeant. Other examples of CD conjugates as specific ligands of cell-surface receptors include opioid peptides,<sup>5</sup> gastrin peptides,<sup>6</sup> and mono- and oligosaccharides.<sup>7</sup> These approaches rely in part on the assumed membrane impermeability of CD conjugates, based upon the natural hydrophilicity of CD conjugates and a molecular weight (>1500) above the perceived limits of facile, passive membrane transport.

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The development of chemically and biologically stable estrogen–macromolecule conjugates has been a challenge of increasing importance given the interest in differentiating non-classical ligand-activated estrogen receptor (ER) actions versus the classical ER function as a ligand-activated nuclear receptor. Estrogens are able to elicit rapid responses that are thought to be mediated by extranuclear receptors that may be the classic ER, splice variants, or G-protein coupled receptors.<sup>8–10</sup> The non-classical, extranuclear action of E<sub>2</sub> elicit activity via cytosolic kinase cascades, which may have genomic effects via epigenetic actions or transcription factors other than ER.<sup>11,12</sup> The classical, genomic actions of ER as a nuclear receptor have been differentiated from membrane-associated ER activity by use of commercially available protein conjugates such as E<sub>2</sub>–BSA<sup>12–14</sup> and E<sub>2</sub>–peroxidase.<sup>15</sup> These conjugates have been widely used, but as remarked by Katzenellenbogen, they have critical limitations such as leakage of free E<sub>2</sub> either non-covalently bound or from biological degradation of the conjugate in cell cultures.<sup>8,13,16</sup> In order to overcome these problems and develop efficient E<sub>2</sub>–macromolecule conjugates, Katzenellenbogen and coworkers utilized a polyamidoamine (PAMAM) dendrimer as core architecture instead of a protein, leading to an E<sub>2</sub>–conjugate selective for extranuclear ER.

Click chemistry is commonly used today as a synonym for the Huisgen cycloaddition, although originally defined by Sharpless as encompassing simple synthetic steps giving quantitative conversions.<sup>17</sup> Quantitative procedures for formation of CD–conjugates avoid the complex chromatography required to remove by-products of incomplete derivatization. Herein, novel estradiol–CD conjugates were prepared using click chemistry and the estrogenic activity was assayed in order to test the ability of CD derivatives to provide selective probes of estrogenic activity.

## 2. Results and discussion

### 2.1. Synthesis of ACD conjugates

A CD–conjugate persubstituted at the C6 position of the primary face was the initial target. Aminocyclodextrins (ACDs) are a specific subset of persubstituted CDs, which may be readily prepared by direct amination of per-6-bromo-per-6-deoxy-CD or per-6-iodo-per-6-deoxy-CD. ACDs possess an annulus of amino groups in place of the ether oxygens of native CD and a corona of pendant side chains; at neutral pH the presence of the positively charged annulus diminishes the ability of the ACD cavity to include hydrophobic small molecules.<sup>18</sup> The ACD scaffold provides ACD derivatives with demonstrated, inherent biological activity.<sup>19</sup> Synthesis from the more reactive per-6-iodo-per-6-deoxy-CD was revealed by careful chromatography to yield an intramolecular cross-linked impurity,<sup>20</sup> whereas synthesis via the less reactive per-6-bromo-per-6-deoxy-CD is accomplished in quantitative steps requiring no chromatography (Scheme 1).<sup>21,22</sup> However, this ‘click’ strategy is limited to reactions with amines in the liquid phase or with high solubility in DMF,<sup>21</sup> an alternative click methodology is imine formation from reaction of aldehyde with per-6-amino-per-6-deoxy-β-CD (**3**), easily obtained from the per-azido derivative **2**, and subsequent reduction, ultimately leading to the desired CD–E<sub>2</sub> conjugate, 7EACD (Scheme 1).

EE<sub>2</sub> is a biologically active ER ligand of synthetic utility because of the reactivity of the terminal acetylene group.<sup>23</sup> Elaboration of EE<sub>2</sub> is readily achieved using palladium catalyzed Sonogashira cross coupling with halide reagents,<sup>24</sup> allowing attachment of a benzaldehyde moiety for ACD conjugation. Subsequent reduction of benzaldehyde **4** gave the benzyl alcohol **5** that was used as a control compound. The previously reported heterogeneous dendrimer E<sub>2</sub>–PAMAM conjugate was obtained by PAMAM condensation with **4** in MeOH at room temperature.<sup>8,25</sup> Similarly, condensation of ACD (**3**) with **4** was attempted, followed by in situ NaBH<sub>4</sub> reduction (Scheme 1; Table 1 entry 1). However, reaction did not go to completion, giving a heterogeneous mixture that, from mass spec and

NMR examination, contained (EE<sub>2</sub>)<sub>7</sub>ACD as major product in addition to smaller amounts of (EE<sub>2</sub>)<sub>6</sub>ACD and (EE<sub>2</sub>)<sub>5</sub>ACD (Fig. 1). Optimization of reaction conditions failed to yield a homogeneous product, however, semi-preparative RP-HPLC did yield conjugate 7EACD (**6**) that consists of (EE<sub>2</sub>)<sub>7</sub>ACD (>90%) with small amounts of (EE<sub>2</sub>)<sub>6</sub>ACD and (EE<sub>2</sub>)<sub>5</sub>ACD. Chromatographic and spectroscopic examination revealed no evidence for an inclusion complex between 7EACD and either EE<sub>2</sub> or the reaction by-product, **5**, compatible with the extensive washing and chromatography employed in synthesis, and the known propensity of ACD to former weak inclusion complexes.<sup>18</sup> 7EACD is comparable to the heterogeneous E<sub>2</sub>-dendrimer conjugate described by Katzenellenbogen.

The lack of a homogeneous product from the imine synthetic route (Scheme 1) was surprising given that the condensation of 2-pyridinecarboxaldehyde with **3** had been previously demonstrated in our lab to proceed to completion, yielding the persubstituted ACD (**7**) after reduction in situ (Table 1, entry 2).<sup>26</sup> To examine the limitations of this synthetic methodology for persubstituted ACD derivatives, other approaches were attempted. Condensation of **3** with 4-benzyloxybenzaldehyde yielded the desired product **8** (Table 1, entry 3). The possibility that complete reaction of **3** with **4** was hindered by the steric bulk of the ethynyl estradiol group was explored by using a longer linker arm between estradiol and aldehyde groups. However, neither condensation of **10** (Scheme 2) nor the synthon **9** with ACD (**3**) gave the desired imine products (Table 1, entries 5 and 6), indicating the preference in imine condensation for the more reactive arylaldehydes. Other alternatives were explored, including reaction via the known β-CD derivative, **15** (Scheme 2; Table 1, entry 7).<sup>27</sup> However, under all reactions conditions tested, the persubstituted product was the major, but not the sole product observed. The imine condensation reaction of **19** with **4** produced the monosubstituted E<sub>2</sub>-conjugate 1EACD (**20**; Table 1, entry 8), and a similar product (**21**) was obtained by the simple nucleophilic displacement of the tosyl group in mono-6-tosyl-β-CD using EE<sub>2</sub> derivative **12** (Table 1, entry 9). Both E<sub>2</sub>-conjugates **20** and **21** contained small amounts of unmodified β-CD carried through the synthesis as shown by NMR spectra and MALDI-TOF mass spectroscopic analysis.

## 2.2. Synthesis of 'qCD' conjugates

Cycloaddition click chemistry is currently used extensively for bioconjugation because of the efficient conversion and good yields in aqueous media.<sup>28–30</sup> Given the ease with which mono- and persubstituted azido-CD are prepared, this synthetic methodology was explored for E<sub>2</sub>-CD conjugates. The presence of seven triazole groups in the annulus of such cyclodextrin conjugates is likely to confer inherently different physicochemical properties to CD and ACD conjugates, hence we coin the click-derived conjugates as qCD derivatives.

A variety of click reactions were explored based upon literature precedent (Table 2),<sup>31,32</sup> monitoring conjugation products by MALDI-TOF spectroscopy. In situ [1,3]-cycloaddition ligation was unsuccessful from **1** (Table 2, entry 10), but was successful from mono-6-tosyl-β-CD giving the desired product **22** in 50% conversion along with unreacted β-CD contaminant **17** (Table 2, entry 11).<sup>33–35</sup> The reaction mixture of **2** with seven equivalent of EE<sub>2</sub> in the presence of CuI and DIEA changed color from green to brown coincident with the loss of the TLC spot for unreacted EE<sub>2</sub>. Reaction time was extended for one day beyond this indicator point, yielding quantitative conversion to 7EqCD (**23**) as a powder in 58% isolated yield after washing (Table 2, entry 12; Fig. 1). The monoazido-β-CD under the same reaction conditions gave 1EqCD (**24**) in 68% isolated yield (Table 2, entry 13). In all procedures, extensive washing was carried out to remove residual Cu ions. As a control compound for bioassay, a triazole modified estradiol was synthesized (qE<sub>2</sub>, **25**): methyl azide was prepared for in situ click ligation to EE<sub>2</sub>, resulting in the desired product **25** obtained in 60% yield after chromatography (Table 2, entry 14).

In order to estimate the membrane permeability of 7EqCD, rhodamine was coupled to the secondary face of 7EqCD (**26**; Table 2, entry 15). Non-covalent inclusion of rhodamine would be difficult to rule out in the rhodamine–7EqCD conjugates, therefore the triazole-CD derivative (**26**) was prepared and the rhodamine–qCD conjugate synthesized for comparison (Table 2, entries 16 and 17). Both compounds **23** and **26** were coupled with 1 mol equiv of fluorophore, either as the free carboxylic acid (rhodamine-19-perchlorate, yielding conjugates **23R'** and **26R'**, respectively) or as the succinimidyl ester (NHS–rhodamine, yielding products **23R** and **26R**, respectively). In each case, the presence of the monosubstituted rhodamine conjugate was demonstrated by MALDI-TOF spectroscopy.

### 2.3. Summary of synthetic strategies

Elaboration of the ACD scaffold via imine formation with aryl aldehydes is efficient when the aldehyde can be used as a co-solvent or in high concentration, however other aldehydes do not give quantitative reaction. Monitoring of reactions by MALDI-TOF is useful to identify qualitatively the extent of multivalent substitution of CD conjugates, however, the intensity of signals in MALDI-TOF is not directly related to relative quantities of each multivalent ACD conjugate. The problematic ability of mass spec to provide quantitation has been previously described for CD derivatives.<sup>36</sup> Furthermore, only perfacially substituted CD derivatives give simple NMR spectra; incompletely substituted  $\beta$ -CD derivatives have 42 chemically and spectroscopically non-equivalent glucose carbons. From HPLC–UV chromatography, 7EACD was estimated predominantly to contain the desired persubstituted product,  $(EE_2)_7$ ACD, however the MALDI-TOF spectrum indicated the presence of both  $(EE_2)_6$ ACD and  $(EE_2)_5$ ACD isomers (Fig. 1). Cycloaddition click chemistry provided a more efficient route to perfacial modification, yielding qCD conjugates perfacially derivatized with  $EE_2$  on the primary face (Figs. 1 and 2). Four biocompatible  $E_2$ -CD conjugates were obtained for bioassay: 7EACD, 1EACD, 7EqCD, and 1EqCD.

### 2.4. Estrogenic activity of CD conjugates

The ER $\alpha$ -positive human endometrial Ishikawa cell line is routinely used to quantify estrogenic and antiestrogenic activity of test compounds.<sup>37</sup> In this cell line, the induction of alkaline phosphatase is an ER-mediated estrogenic response, allowing measurement of EC<sub>50</sub> values for estrogenic and IC<sub>50</sub> for antiestrogenic potency by enzyme assay. The measured EC<sub>50</sub> values for 1EACD, 1EqCD, 7EACD, and 7EqCD were 4.0  $\mu$ M, 4.0  $\mu$ M, 300 nM, and 200 nM, respectively; both polyvalent conjugates showed higher potency than monovalent derivatives (Fig. 3). There was no significant antiestrogenic activity or cytotoxicity observed from the test compounds at the concentrations studied (data not shown).  $EE_2$  derivatives **5** and **12** and ACD (**3**) represent control compounds for EACD: ACD was inactive, whereas **5** and **12** were 200-fold more potent than 7EACD (Fig. 3). Similarly, qE<sub>2</sub> and CD derivative **2** represent controls for EqCD; **2** was inactive, whereas qE<sub>2</sub> was a full estrogen (Fig. 3). All non-conjugated estrogen control compounds were substantially less potent than  $EE_2$  itself, for which EC<sub>50</sub> was measured as 28 pM in Ishikawa cells. Katzenellenbogen has referred to potency for  $E_2$ -dendrimer conjugates where observed potency is divided by the average number of  $E_2$  moieties in the conjugate.<sup>8</sup> Applying this measure, the potency for: **5**, 1EACD, and 7EACD were 27 nM, 4  $\mu$ M, and 2  $\mu$ M, respectively; and for qE<sub>2</sub>, 1EqCD, and 7EqCD were 27 nM, 4  $\mu$ M, and 1.4  $\mu$ M, respectively.

In the standard radioligand displacement assay for measuring relative binding affinity to recombinant human ER $\alpha$ , qE<sub>2</sub> was observed to be a weak ER $\alpha$  ligand (IC<sub>50</sub> = 0.4  $\pm$  0.1  $\mu$ M). In this assay, ER binding of EACD and EqCD derivatives was not observed within solubility limits, however, other compounds with submicromolar estrogenic potency in cell culture have shown low or negligible affinity in the radioligand displacement assay.<sup>38</sup>

Rapid signaling via non-classical estrogen-activated pathways has been recognized for some time in the CNS and cardiovascular systems.<sup>39,40</sup> However, the quantification of non-classical estrogenic activity is not trivial, in part because different molecular identities and cellular localization have been proposed for these receptors.<sup>9</sup> The debate on ligand-activated G-protein-coupled ER (GPR-ER, or GPR-30) has focused on the plasma membrane and endoplasmic reticulum,<sup>41</sup> whereas ER itself has been proposed to be active in the plasma membrane, cytoplasm, or mitochondria.<sup>42</sup> Rapid assays of MAPK pathway activation and intracellular Ca levels have been used for extranuclear ER activation, however, several pathways may elicit such responses. The classical activity of ER as a nuclear receptor can be assayed by E<sub>2</sub>-responsive gene reporter assays such as the alkaline phosphatase assay presented above, however, it has been shown that of E<sub>2</sub>-responsive genes, a full 25% are induced by E<sub>2</sub>-dendrimer conjugates.<sup>43</sup> Recently Tonetti and co-workers reported that T47D/PKC $\alpha$  cells (human breast cancer cells stably transfected with protein kinase C) were stimulated by E<sub>2</sub>-BSA, whereas the parent cell line (T47D/neo) was unresponsive.<sup>44</sup> Therefore, the activity of 7EqCD was assayed in both T47D cell lines (Fig. 4). The classical ER-dependent potency of 7EqCD in T47D/neo cells was low: EC<sub>50</sub> for 7EqCD was >100 nM in T47D/neo cells. However, the response of T47D/PKC $\alpha$  cells to 7EqCD was similar to that of E<sub>2</sub>-BSA; 7EqCD (10 nM) was a full estrogen, with the EC<sub>50</sub> estimated as 2 nM. Taken together, the data shown in Figure 4 suggest that 7EqCD at 2–10 nM may be useful as an extranuclear ER selective ligand and warrants further study.

## 2.5. Membrane permeability and mechanism of action

Rhodamine was conjugated to 7EqCD (**23R**, **23R'**) using two different coupling agents, and a control qCD (**26R**, **26R'**) was conjugated in the same way to provide comparison. Both fluorescent E<sub>2</sub>-conjugates retained estrogenic activity at higher concentrations as shown by measured EC<sub>50</sub> values in Ishikawa cells of  $1.5 \pm 0.47 \mu\text{M}$  and  $2.6 \pm 0.63 \mu\text{M}$  for **23R** and **23R'**, respectively. ER positive MCF-7 mammary cancer cells were incubated with either **23R**, **23R'**, or the control compound (**26R**) for 24 h at which time nuclei and conjugate were localized by confocal fluorescence microscopy (Fig. 5). Both 7EqCD conjugates were observed in the cytoplasm, but not the nucleus of MCF-7 cells, whereas no evidence for cell membrane permeation was obtained for the control compound.

The estradiol conjugates best described to date are E<sub>2</sub>-BSA, E<sub>2</sub>-peroxidase, and E<sub>2</sub>-dendrimers; at higher concentrations ( $\geq 10$  nM for protein conjugates and  $\geq 100$  nM for dendrimer conjugates) all three demonstrate classical estrogenic actions reasoned to result from leaching of non-covalently bound estrogen (E<sub>2</sub> or EE<sub>2</sub>) or degradation to release estrogens. There is no reason to expect that inclusion of EE<sub>2</sub> would be an important factor for either 7EACD or 7EqCD, because EE<sub>2</sub> has low reported affinity for  $\beta$ -CD ( $K_d = 8$  mM),<sup>45,46</sup> and a more polar CD annulus is known to reduce the affinity for hydrophobic aromatic guests further.<sup>18,21</sup> The presence of EE<sub>2</sub> in the 7EqCD product was tested in the same Ishikawa cell cultures used for assessment of classical estrogenic activity. For comparison, the presence of E<sub>2</sub> was measured in incubations of E<sub>2</sub>-BSA in Ishikawa cell culture. After four days, the cell supernatant was collected and assayed for EE<sub>2</sub> and E<sub>2</sub> using LC-MS/MS quantitation, against standard curves, of estrogens derivatized with dansyl chloride (Fig. 6). In the 7EqCD incubation, EE<sub>2</sub> was not observed above detection limits, but E<sub>2</sub> was observed in incubations of E<sub>2</sub>-BSA in cell cultures as anticipated from literature reports.

7EACD bears comparison to the heterogeneous E<sub>2</sub>-dendrimer conjugate prepared by Katzenellenbogen and co-workers, since both bioconjugates are prepared from a similar EE<sub>2</sub> synthon via an imine intermediate, with the same spacer unit linking macromolecular amine with EE<sub>2</sub>, and both are heterogeneous.<sup>8</sup> In MCF-7 cells, the E<sub>2</sub>-dendrimer induced E<sub>2</sub>-responsive genes, but E<sub>2</sub> itself was 3–4 orders of magnitude more potent than the conjugate (E<sub>2</sub>-dendrimer; EC<sub>50</sub>  $\sim 10^{-9}$ – $10^{-8}$  M).<sup>8</sup> These and other data were compatible with



permeation of the E<sub>2</sub>-dendrimer conjugate into the cytoplasm, but not into the nucleus. Recently, the combination of E<sub>2</sub>-BSA (membrane selective estrogen) and E<sub>2</sub>-dendrimer (extranuclear selective estrogen) conjugates has been used to distinguish membrane and extranuclear inputs into E<sub>2</sub>-responsive gene induction.<sup>43</sup>

### 3. Conclusions

The classical mechanism of estrogen action is defined by: (i) binding of an estrogenic ligand to ER in the cytoplasm; (ii) ER dimerization and translocation to the nucleus; (iii) complexation with the ERE domain of DNA; (iv) recruitment of coregulator proteins; (v) transcription and expression of E<sub>2</sub>-responsive gene products. Non-classical mechanisms utilize extranuclear receptors that are activated by estrogenic ligands, often via rapid signaling cascades, however these may also induce ER-dependent, genomic activity.<sup>43</sup> It is recognized that selective ligands are required to untangle the role of nuclear and extranuclear receptors and the cross-talk between classical and non-classical pathways.  $\beta$ -CD derivatives play an important role in drug delivery, and less frequently have been used as molecular scaffolds for biomimetics;<sup>47-49</sup> the expectation that these large, hydrophilic molecules seldom penetrate cellular membranes supports the exploration of E<sub>2</sub>-CD conjugates as cell compartment selective estrogens.

Several click chemistry approaches were attempted towards a  $\beta$ -CD persubstituted at the primary face with estradiol via a stable linker. Reactions using the amino-CD (ACD) scaffold proved insufficiently generalized; selected examples giving quantitative persubstitution, but others such as the ACD-estradiol conjugate, 7EACD, not providing an homogeneous persubstituted product. Conjugation via [1,3]-cycloaddition proved a suitable alternative method for preparation of a homogeneous, persubstituted E<sub>2</sub>-CD, namely 7EqCD. The four E<sub>2</sub>-CD conjugates studied in cell culture were not cytotoxic below solubility limits and all manifested estrogenic activity in cell culture. 7EqCD as an homogeneous E<sub>2</sub>-CD conjugate was preferred for further study. Labeling with rhodamine allowed localization of 7EqCD in the cytoplasm, but not the cell nucleus, suggesting that this E<sub>2</sub>-CD conjugate would provide a selective extranuclear estrogen comparable to E<sub>2</sub>-dendrimer conjugates. Furthermore, at concentrations giving negligible classical ER activity (1-10 nM), 7EqCD was shown to deliver activity in cell culture equivalent to E<sub>2</sub>, via a mechanism attributed in the literature to extranuclear ER. Thus, 7EqCD bears comparison to E<sub>2</sub>-dendrimer conjugates as a biological probe to explore extranuclear ER mechanisms; 7EqCD is the first homogeneous probe of this type.

## 4. Experimental

### 4.1. Synthesis of compounds

**4.1.1. Materials and general methods**—All chemicals and solvents were obtained from Sigma-Aldrich and Fisher unless otherwise stated and purified prior to use according to known procedures when necessary: 17 $\beta$ -ethynylestradiol (Steraloids, Newport, RI), NHS-rhodamine (PIERCE, Rockford, IL), and all other reagents were used as received. DMF and Et<sub>3</sub>N were distilled from CaH<sub>2</sub>.  $\beta$ -Cyclodextrin and all CD derivatives were dried under vacuum in a drying pistol over P<sub>2</sub>O<sub>5</sub> with refluxing acetone prior to use. Moisture sensitive reactions were performed under nitrogen using oven dried glassware. Analytical thin layer chromatography (TLC) was carried out on 60 Å silica gel-flexible baked plates (Whatman), and compounds were visualized by UV or by staining with I<sub>2</sub>, KMnO<sub>4</sub>, or phosphomolybdic acid solutions. Flash column chromatography was performed using silica gel (32-63  $\mu$ m, Selecto Scientific).

**4.1.2. Instrumentation**—NMR spectra were recorded on Bruker Ultrashield (400 MHz) or Avance (300 MHz) spectrometers using CDCl<sub>3</sub>, DMSO-*d*<sub>6</sub>, or CD<sub>3</sub>CN at 300 K. Variable temperature <sup>1</sup>H NMR at 323 K was performed on 7EqCD in order to improve chemical splitting resolution. Chemical shifts of <sup>1</sup>H NMR spectra were reported using TMS (0 ppm) as a reference, and <sup>13</sup>C NMR spectra were referenced to internal deuterated NMR solvents. All <sup>13</sup>C NMR spectra of CD derivatives were obtained from overnight runs at optimized condition (D1; 3–5 s, TD; 32 K, SI; 265 K). Analytical HPLC (C8 column, 5 μm, 4.6 × 150 mm, Agilent Technologies) was carried out with Perkin–Elmer (Fremont, CA) equipment, and semi-preparative reversed-phase C8 column (5 μm, 10 × 250 mm, VYDAC) was used for purification on the same equipment. Matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) mass spectra were obtained on a Voyager (Applied Biosystems Co. Ltd) using DHB as a matrix. Mass spectra for monomer precursors were recorded on an Agilent 1100 series LC/MSD ion trap instrument, using APCI as ionization method.

**4.1.3. Per-6-azido-6-deoxy-β-cyclodextrin (2)<sup>50</sup>**—Per-6-iodo-β-cyclodextrin (**1**; 4.5 g, 2.36 mmol) obtained by literature procedures was dissolved in DMF (51 mL), and sodium azide (1.32 g, 20.3 mmol) was added. The suspension was stirred under N<sub>2</sub> atmosphere at 70 °C for 20 h. The resulting yellow solution was concentrated under reduced pressure to half volume, and was poured into 90% EtOH (280 mL) to precipitate the final product, a fine white powder that was filtered and was washed with H<sub>2</sub>O and dried in a P<sub>2</sub>O<sub>5</sub> drying pistol for one day (2.8 g, 90% yield). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ<sub>ppm</sub> 5.91 (br d, *J* = 6.7 Hz, 7H), 5.76 (s, 7H), 4.92 (s, 7H), 3.80–3.70 (m, 14H), 3.61–3.56 (m, 14H), 3.40–3.33 (m, 14H). <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ<sub>ppm</sub> 102.05, 83.20, 72.60, 72.00, 70.33, 51.33. MALDI-TOF: calcd C<sub>42</sub>H<sub>63</sub>O<sub>28</sub>N<sub>21</sub> [M]<sup>+</sup> 1310.08, found [M+Na]<sup>+</sup> 1333.91.

**4.1.4. Per-6-amino-6-deoxy-β-cyclodextrin (3)**—Per-6-azido-β-cyclodextrin **2** (1.53 g, 1.17 mmol) was dissolved in DMF (30.8 mL), and Ph<sub>3</sub>P (4.85 g, 18.5 mmol) was added. The evolution of N<sub>2</sub> was observed by the formation of bubbles in the reaction vessel. After 1 h, the evolution of N<sub>2</sub> ceased, 35% aqueous NH<sub>3</sub> solution (6.0 mL) was added dropwise into the solution. After the addition of aqueous NH<sub>3</sub> was completed, the reaction mixture turned to a white suspension that was stirred at room temperature for 18 h. The resulting mixture was condensed under vacuum at 60 °C to one fourth of volume, and then EtOH (77 mL) was added to precipitate the product. The white powder was washed with EtOH (77 mL, three times) and dried for one day (1 g, 82% yield). For NMR spectroscopy, the free amino cyclodextrin was converted to amine salt by addition of diluted HCl until the pH had reached 6. <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ<sub>ppm</sub> 5.21 (d, *J* = 2.4 Hz, 7H), 4.25 (ddd, 7H), 4.03 (t, *J* = 2.7 Hz, 7H), 3.72 (dd, *J* = 2.4, 7.5 Hz, 7H), 3.63 (t, *J* = 6.9 Hz, 7H), 3.49 (dd, *J* = 2.1, 9.9 Hz, 7H), 3.33–3.28 (dd, *J* = 5.7 Hz, 7H), <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ<sub>ppm</sub> 100.94, 81.69, 71.67, 71.15, 67.29, 39.72. MALDI-TOF: calcd C<sub>42</sub>H<sub>77</sub>N<sub>7</sub>O<sub>28</sub> [M]<sup>+</sup> 1128.10, found [M+Na]<sup>+</sup> 1151.96.

**4.1.5. 17α-[(4-Formylphenyl)ethynyl-estra-1,3,5-(10)-triene-3,17β-diol, EE<sub>2</sub>-Ph-CHO (4)**—Compound **4** was synthesized by literature procedures.<sup>25</sup> After silica gel column chromatography (EtOAc/Hx, 1:2) and vacuum drying, a fine beige powder was obtained (945 mg, 70% yield). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ<sub>ppm</sub> 10.00 (s, 1H), 8.99 (s, 1H), 7.89 (d, *J* = 8.4 Hz, 2H), 7.62 (d, *J* = 8.1, 7.2 Hz, 2H), 6.51 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.43 (d, *J* = 2.4 Hz, 1H), 5.59 (s, 1H), 2.70–1.23 (m, 14H), 0.82 (s, 3H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ<sub>ppm</sub> 192.5, 154.9, 137.2, 135.2, 131.9, 130.2, 129.7, 128.9, 126.1, 114.9, 112.8, 99.2, 83.6, 78.8, 49.5, 47.4, 43.4, 33.0, 29.2, 27.0, 26.2, 22.6, 12.9. APCI-MS: calcd for C<sub>27</sub>H<sub>28</sub>O<sub>3</sub> 400.52, found [M-H]<sup>-</sup> 399.70.

**4.1.6. 17 $\alpha$ -[(4-Hydroxymethylphenyl)]ethynyl-estra-1,3,5-(10)-triene-3,17 $\beta$ -diol (5)**—EE<sub>2</sub>-Ph-CHO **4** (470 mg, 1.17 mmol) was dissolved in MeOH (4 mL) and DMF (1.3 mL). Under N<sub>2</sub> atmosphere NaBH<sub>4</sub> (110 mg, 2.91 mmol) was quickly added. The dark brown reaction mixture was stirred at room temp for 3 h. After removal of solvent, the residue was purified by silica gel column chromatography (EtOAc/Hx, 2:1) giving a white solid (423 mg, 90% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  9.00 (s, 1H), 7.37 (d, *J* = 8 Hz, 2H), 7.30 (s, *J* = 7.6 Hz 2H), 7.07 (d, *J* = 8.4 Hz, 1H), 6.51 (d, *J* = 8.4 Hz, 1H), 6.44 (s, 1H), 5.44 (s, 1H), 5.25 (s, 1H), 4.50 (s, 2H), 2.71 (s, 2H), 2.32 (d, *J* = 11.6 Hz, 1H), 2.24–1.69 (m, 9H), 1.37–1.19 (m, 5H), 0.86 (m, 1H), 0.81 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  155.0, 142.7, 137.2, 131.0, 130.3, 126.5, 126.1, 121.1, 115.0, 112.7, 94.5, 84.1, 78.6, 62.5, 49.4, 47.2, 43.4, 39.0, 33.0, 29.3, 27.1, 26.3, 22.7, 13.0. APCI-MS: calcd for C<sub>27</sub>H<sub>30</sub>O<sub>3</sub> 402.53, found [M-H]<sup>-</sup> 401.50.

**4.1.7. Per-6-(estradiol-17-ethynylbenzylamino)-6-deoxy- $\beta$ -cyclodextrin (6)**—Per-6-amino- $\beta$ -cyclodextrin (50 mg, 0.044 mmol) and EE<sub>2</sub>-Ph-CHO **4** (176 mg, 3.08 mmol) were dissolved in DMF (1 mL), and the reaction mixture was refluxed at room temperature or 65 °C for 4–10 d under N<sub>2</sub> atmosphere. The resulting brown imine suspension was reduced in situ by freshly prepared 0.1 M NaBH<sub>4</sub> solution (2.1 mL) at 0 °C and allowed to stir for 2 h. The warmed reaction mixture was poured into acetone (25 mL) to precipitate the desired product. The beige solid was repeatedly washed and filtered until no corresponding alcohol **5** from EE<sub>2</sub>-Ph-CHO reduction was detected on TLC. The resulted precipitates were collected and dried yielding a beige powder (51 mg, 30% yield). MALDI-TOF: *m/z* calcd for C<sub>231</sub>H<sub>273</sub>N<sub>7</sub>O<sub>42</sub> 3818.9, C<sub>204</sub>H<sub>245</sub>N<sub>7</sub>O<sub>40</sub> 3434.7, C<sub>177</sub>H<sub>217</sub>N<sub>7</sub>O<sub>38</sub> 3050.5; found [M+Na]<sup>+</sup> 3843.46, 3458.81, 3073.22.

**4.1.8. Per-6-(pyridine-2-ylmethylamino)-6-deoxy- $\beta$ -cyclodextrin (7)**—2-Pyridinecarboxaldehyde (33.5  $\mu$ L, 0.35 mmol) was syringed into a suspension of **3** (50 mg, 0.044 mmol) in DMF (100  $\mu$ L). The yellow reaction solution was stirred at room temperature for 5 hr. The resulting imine was reduced in situ by freshly prepared 0.1 M NaBH<sub>4</sub> solution (2.58 mL) at 0 °C and allowed to stir for 2 h. The warmed reaction mixture was poured into Et<sub>2</sub>O (50 mL) to precipitate product. After washing and filtration, the white precipitate was collected (60 mg, 77% yield). For NMR assay, the HBr salt was prepared after acidification with 5% HBr to bring pH to 4.0. After water was evaporated, the beige powder was taken for NMR spectroscopy. <sup>13</sup>C NMR (75 MHz, HBr salt in D<sub>2</sub>O)  $\delta_{\text{ppm}}$  148.1, 138.5, 124.3, 123.6, 100.9, 81.8, 71.7, 71.2, 67.3, 50.8, 48.4. MALDI-TOF: calcd for C<sub>84</sub>H<sub>112</sub>N<sub>14</sub>O<sub>28</sub> 1765.89, found [M+Na]<sup>+</sup> mass 1788.31.

**4.1.9. Per-6-(4-(benzyloxy)benzylamino)-6-deoxy- $\beta$ -cyclodextrin (8)**—4-Benzyloxybenzaldehyde (312 mg, 1.47 mmol) was added to a suspension of **3** (40 mg, 0.035 mmol) in DMF (580  $\mu$ L) and MeOH (1.17 mL). The reaction solution was stirred at room temperature for seven days. The resulting imine was reduced in situ with freshly prepared 0.1 M NaBH<sub>4</sub> solution (2.1 mL) stirring at 0 °C for 2 h. The warmed reaction mixture was poured into acetone (25 mL) to precipitate the product. After repetitive washing followed by filtration, a white precipitate was collected and dried (71 mg, 81% yield). MALDI-TOF: calcd for C<sub>140</sub>H<sub>161</sub>N<sub>7</sub>O<sub>35</sub> 2501.84, found [M+Na]<sup>+</sup> 2523.94.

**4.1.10. 4-(4-Iodophenyl) butanal (9)**—Compound **9** was prepared by adaptation of literature procedures.<sup>51,52</sup> To a mixture of 1,4-diiodobenzene (5 g, 0.015 mol), palladium(II) acetate (68 mg, 0.304 mmol), sodium hydrogen carbonate (3.2 g, 0.038 mol), and tetra-*N*-butyl ammonium chloride (4.2 g, 0.015 mol) purged with nitrogen gas was added DMF until dissolution. 3-Butene-1-ol (2 mL, 0.023 mol) was added via syringe, and the mixture stirred for 2 d at 40 °C. During the reaction, the mixture turned from orange to



dark brown in color. After cooling acetone (20 mL) was added, and the resulting precipitates were removed by filtration. This procedure was repeated until no more precipitate was produced, and the filtrates then concentrated. Purification by silica gel chromatography (EtOAc/Hx, 1:7) afforded a light-yellow oil (1.08 g, yield 26%; 80:20 to branched aldehyde).  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta_{\text{ppm}}$  9.72 (s, 1H), 7.57 (d,  $J = 8.4$  Hz, 2H), 6.90 (d,  $J = 8.1$  Hz, 2H), 2.56 (t,  $J = 7.8, 7.2$  Hz, 2H), 2.41 (td,  $J = 7.2, 1.2$  Hz, 2H), 1.89 (p,  $J = 7.2, 7.5$  Hz, 2H).  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta_{\text{ppm}}$  202.0, 141.0, 137.5, 130.6, 101.78, 81.24, 72.43, 69.74, 68.93, 59.61, 21.23.

#### 4.1.11. $17\alpha$ -[(4-Butanalphenyl)ethynyl]-estra-1,3,5 (10)-triene-3,17 $\beta$ -diol (10)—

Compound **10** was prepared by literature procedures<sup>25</sup> as a light-yellow powder (900 mg, 60% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta_{\text{ppm}}$  9.75 (s, 1H), 7.45 (d,  $J = 8$  Hz, 2H), 7.25 (d,  $J = 8$  Hz, 2H), 7.20 (d,  $J = 8$  Hz, 2H), 6.91 (s, 1H), 6.69 (d,  $J = 8$  Hz, 1H), 6.62 (s, 1H), 3.64 (s, 1H), 2.85 (m, 2H), 2.69 (t,  $J = 8$  Hz, 2H), 2.48–2.46 (m, 4H), 1.96–1.86 (m, 6H), 1.49–1.34 (m, 4H), 0.98 (s, 3H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{DMSO}-d_6$ )  $\delta_{\text{ppm}}$  203.2, 154.9, 141.8, 137.1, 131.2, 130.3, 128.6, 126.1, 120.5, 114.9, 112.7, 94.5, 84.0, 78.6, 49.4, 47.3, 43.4, 42.3, 40.4, 39.0, 34.2, 32.9, 29.2, 27.0, 26.3, 23.2, 12.9. APCI-MS: calcd  $\text{C}_{30}\text{H}_{32}\text{O}_3$  442.25, found  $[\text{M}-\text{H}]^-$  442.10.

#### 4.1.12. Per-6-(4-iodobenzylamino)-6-deoxy- $\beta$ -cyclodextrin (11)—ACD 3 (50 mg,

0.044 mmol) and 4-iodobenzaldehyde (85.1 mg, 0.352 mmol) were dissolved in DMF (0.1 mL), and the reaction mixture was stirred at 60–70 °C for 18 h under  $\text{N}_2$  atmosphere. The resulted brown imine solution was reduced in situ by freshly prepared 0.1 M  $\text{NaBH}_4$  solution (2.1 mL) at 0 °C and allowed to stir for 2 h. The warmed reaction mixture was poured into acetone (25 mL) to precipitate the product as beige powder that was repeatedly washed and filtered until no corresponding alcohol from 4-iodobenzaldehyde was detected by TLC. The precipitates were collected, and overnight drying gave a light-yellow powder that by MALDI-TOF analysis gave signals corresponding to a mixture of 1–7-fold substitution.

#### 4.1.13. $17\alpha$ -(4-Aminophenyl)ethynyl-estra-1,3,5 (10)-triene-3,17 $\beta$ -diol (12)<sup>53–55</sup>

—A solution of  $\text{Pd}(\text{OAc})_2$  (80.7 mg, 0.169 mmol) and  $\text{P}(\text{Ph})_3$  (90.4 g, 0.338 mmol) in TEA (20 mL) was stirred under  $\text{N}_2$  for 10 min.  $\text{CuI}$  (65.7 mg, 0.338 mmol) and 4-iodoaniline (748 mg, 3.376 mmol) were added, and after 5 min, 17 $\beta$ -ethynylestradiol (1 g, 3.376 mmol) was transferred. The dark brown mixture was stirred at room temperature for 5 h. TEA was evaporated, and the residue was purified by silica gel column chromatography ( $\text{CH}_2\text{Cl}_2/\text{MeOH}$ , 20:1). After drying under vacuum, yellow powder was obtained (1.10 g, 84% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta_{\text{ppm}}$  8.98 (s, 1H), 7.06 (s, 2H), 6.50 (d,  $J = 7.2$  Hz, 2H), 6.43 (s, 1H), 5.39 (s, 2H), 5.25 (s, 1H), 3.33 (s, 1H), 2.70 (s, 2H), 2.31 (d,  $J = 9.6$  Hz, 1H), 2.15–1.68 (m, 9H), 1.31 (m, 4H), 0.79 (s, 3H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{CN}$ )  $\delta_{\text{ppm}}$  154.9, 148.8, 137.1, 132.3, 130.3, 126.1, 114.9, 113.6, 112.7, 109.1, 91.4, 85.2, 78.6, 49.2, 47.1, 43.4, 32.9, 29.2, 27.0, 26.3, 22.6, 12.9. APCI-MS: calcd For  $\text{C}_{26}\text{H}_{29}\text{NO}_2$  387.2, found  $[\text{M}-\text{H}]^-$  386.6.

#### 4.1.14. (2-Phthalimidoethyl) isothiuroniumhydrobromide (13)—Compound **13**

was synthesized according to literature procedures.<sup>54,55</sup> as a white solid (5.1, 79% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta_{\text{ppm}}$  9.14 (s, 4H), 7.91 (m, 4H), 3.39 (t,  $J = 5.2$  Hz, 6.4 Hz, 2H), 3.55 (t,  $J = 5.2$  Hz, 6.4 Hz, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{DMSO}-d_6$ )  $\delta_{\text{ppm}}$  169.2, 167.6, 134.7, 131.4, 123.2, 36.0, 29.3. APCI-MS: calcd for  $\text{C}_{11}\text{H}_{12}\text{BrN}_3\text{O}_2\text{S}$  328.98, found  $[\text{M}-\text{HBr}]^+$  249.80.

**4.1.15. Per-6-(phthalimidoethylthio)-6-deoxy- $\beta$ -cyclodextrin (14)**—Compound **14** was synthesized according to literature procedures<sup>27,56</sup> as a white solid (112 mg, 17% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  7.71 (s, 4H), 5.84 (d, *J* = 23 Hz, 2 Hz, 2H), 4.86 (s, 1H), 3.94 (br s, 1H), 3.81 (m, 2H), 3.59 (m, 1H), 3.42–3.33 (m, overlaps with DOH), 3.04 (d, *J* = 11.6 Hz, 1H), 2.91–2.78 (m, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  167.4, 134.2, 131.3, 122.9, 102.1, 84.2, 72.5, 72.2, 71.2, 36.9, 33.0, 30.8. MALDI-TOF: calcd for C<sub>112</sub>H<sub>119</sub>N<sub>7</sub>O<sub>42</sub>S<sub>7</sub> 2459.66, found [M+Na]<sup>+</sup> 2482.62.

**4.1.16. Per-6-(2-aminoethylthio)-6-deoxy- $\beta$ -cyclodextrin (15)**—Compound **15** was prepared by literature procedures<sup>27</sup> as a white powder (50 mg, 81% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  5.45 (d, *J* = 24, 4 Hz, 2H), 4.51 (br s, 1H), 3.40 (br s, 1H), 3.19 (t, *J* = 8.4 Hz, 8.8 Hz, 1H), 3.02–2.48 (m, overlaps with DOH), 2.09 (s, 1H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  102.1, 84.5, 72.6, 72.2, 71.4, 38.6, 32.8, 29.7. MALDI-TOF: calcd for C<sub>56</sub>H<sub>105</sub>N<sub>7</sub>O<sub>28</sub>S<sub>7</sub> 1548.94, found [M]<sup>+</sup> 1548.90.

**4.1.17. Per-6-(estradiol-17-(4-((2-(propylthio)ethylamino)-methyl)phenyl)ethynyl)-6-deoxy- $\beta$ -cyclodextrin (16)**—Compounds **15** (40 mg, 0.0258 mmol) and **4** (310 mg, 0.774 mmol) were dissolved in DMF, and stirred at 60–70 °C for 10 days. 2 mL of aqueous NaOH (1 N in MeOH) and stirred for a further 2.5 h. After cooling and concentration, the residue was precipitated in acetone (25 mL) giving a beige powder that was repeatedly washed and filtered until no trace of **4** was detected by TLC. The precipitates were dried to give a light-yellow solid product containing **16** as the major product in a heterogeneous mixture. MALDI-TOF: calcd for C<sub>245</sub>H<sub>301</sub>N<sub>7</sub>O<sub>42</sub>S<sub>7</sub> 4240.49, found [M+H<sub>2</sub>O+H]<sup>+</sup> 4259.84.

**4.1.18. Mono-6-(*p*-tolylsulfonyl)-6-deoxy- $\beta$ -cyclodextrin (17)**—Compound **17** was prepared by adaptation of literature procedures<sup>57</sup> as a white powder was obtained (1 g, 10% yield). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  7.76 (d, *J* = 7.6 Hz, 2H), 7.44 (d, *J* = 7.2 Hz, 2H), 5.71 (br s, 14H), 4.84 (s, 4H), 4.77 (s, 3H), 4.33 (d, *J* = 10.4 Hz, 3H), 4.20 (m, 3H), 3.66–3.51 (m, overlaps with HOD), 3.38–3.22 (m, overlaps with HOD), 2.09 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  144.9, 132.7, 129.9, 127.6, 102.0 (m), 81.5 (m), 73.1–72.8, 60.0, 21.3. MALDI-TOF: calcd C<sub>49</sub>H<sub>76</sub>O<sub>37</sub>S [M]<sup>+</sup> 1289.19, found [M+Na]<sup>+</sup> 1313.43.

**4.1.19. Mono-6-azido-6-deoxy- $\beta$ -cyclodextrin (18)**<sup>57</sup>—Compound **17** (350 mg, 0.271 mmol) and NaN<sub>3</sub> (189 mg, 2.90 mmol) were suspended in H<sub>2</sub>O (3.53 mL), which was heated and to a clear solution with stirring at 80 °C for 15 h. The reaction mixture was cooled, poured into acetone (60 mL) to precipitate the product as a white powder that was dried in a drying pistol at 56 °C under vacuum. A pure white solid was obtained (492.8 mg, 90% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  5.73 (br s, 14 H), 4.84 (m, 7 H), 3.75–3.55 (m, 28 H), 3.41–3.28 (m, overlaps with HOD), <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  102.0 (m), 83.0, 81.6 (m), 73.0 (m), 72.4, 72.2, 72.1, 70.2, 60.2 (m), 51.1. MALDI-TOF: calcd C<sub>42</sub>H<sub>69</sub>N<sub>3</sub>O<sub>34</sub> [M]<sup>+</sup> 1160.00, found [M+Na]<sup>+</sup> 1183.74.

**4.1.20. Mono-6-amino-6-deoxy- $\beta$ -cyclodextrin hydrochloride (19)**<sup>57</sup>—Compound **18** (300 mg, 0.259 mmol) and PPh<sub>3</sub> (116 mg, 0.442 mmol) were dissolved in DMF (5.6 mL) to which 30% NH<sub>4</sub>OH (2.7 mL) was added by syringe. The reaction mixture was stirred at room temperature for 18 h, and poured into acetone (50 mL) yielding after successive washes and drying a white powder (200.0 mg, 68% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  5.15 (br s, 7 H), 4.06–4.00 (m, 21 H), 3.73–3.63 (m, 10 H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  101.8, 101.5, 82.8, 81.0, 80.8, 73.1, 73.0, 72.1, 71.8, 71.7, 60.2, 41.2. MALDI-TOF: calcd C<sub>42</sub>H<sub>71</sub>NO<sub>34</sub> [M]<sup>+</sup> 1134.01, found [M+Na]<sup>+</sup> 1157.57.

**4.1.21. Mono-6-(17-estradiolphenylmethylamino)-6-deoxy- $\beta$ -cyclodextrin (20)**

—Compounds **19** (50 mg, 0.044 mmol) and **4** (370.6 mg, 0.926 mmol) were dissolved in DMF (1 mL), and the reaction mixture was refluxed at 65 °C for 3 d under N<sub>2</sub>. The resulting brown imine suspension was reduced in situ by the addition of a freshly prepared 0.1 M NaBH<sub>4</sub> solution (3 mL) at 0 °C with stirring for 2 h. The warmed reaction mixture was precipitated into acetone (25 mL) and the precipitate repeatedly washed and filtered until no presence of **5** was detected by TLC. After drying a white powder was obtained (30 mg, 45% yield). MALDI-TOF: calcd for C<sub>69</sub>H<sub>99</sub>NO<sub>36</sub> 1518.53, found [M+Na]<sup>+</sup> 1541.97.

**4.1.22. Mono-6-(estradiol-17-ethynylbenzylamino)-6-deoxy- $\beta$ -cyclodextrin (21)**

—Compounds **17** (100 mg, 0.078 mmol) and **12** (630 mg, 1.63 mmol) were dissolved in DMF (0.5 mL), and the reaction mixture was refluxed at 60–70 °C for 7 d under N<sub>2</sub>. The solvent was evaporated under vacuum, and the dark brown residue was precipitated in acetone (50 mL). The beige powder was repeatedly washed and filtered until no trace of compound **12** was detected by TLC. The precipitates were collected, and overnight drying gave a light-yellow solid (51 mg, 43%). MALDI-TOF: calcd for C<sub>69</sub>H<sub>99</sub>NO<sub>36</sub> 1518.53, found [M+Na]<sup>+</sup> 1541.60.

**4.1.23. Mono-6-(estradiol-17-(1*H*-1,2,3-triazol-4-yl))-6-deoxy- $\beta$ -cyclodextrin (22)**

—Compound **17** (100 mg, 0.078 mmol), 17 $\beta$ -ethynylestradiol (23.1 mg, 0.078 mmol), and sodium azide (6.6 mg, 0.10 mmol) were dissolved in DMF and H<sub>2</sub>O (3:1, 0.9 mL). A solution of CuSO<sub>4</sub>·5H<sub>2</sub>O (400 mg, 1.60 mmol) and sodium ascorbate (360 mg, 1.82 mmol) was prepared separately in the same DMF/H<sub>2</sub>O solvent system (2 mL) and added via syringe. The brown suspension was stirred at 60–70 °C for four days. The solvent was evaporated, and the residue was precipitated in H<sub>2</sub>O (50 mL). This washing process was repeated three times for complete removal of residual copper ions. After centrifugation at 4 °C (4000 rpm, 20 min), a fine beige powder was obtained (60 mg, 53%). MALDI-TOF: calcd for C<sub>62</sub>H<sub>93</sub>N<sub>3</sub>O<sub>36</sub> 1456.42, found [M+Na]<sup>+</sup> 1479.44.

**4.1.24. Per-6-(estradiol-17-(1*H*-1,2,3-triazol-4-yl))-6-deoxy- $\beta$ -cyclodextrin (23)**

—Compound **2** (100 mg, 0.076 mmol) and EE<sub>2</sub> (157.6 mg, 0.532 mmol) were dissolved in 50% of MeOH and DMF (1.9 mL). CuI (72.4 mg, 0.38 mmol) was added, and DIEA (66.2  $\mu$ L) added by syringe to the reaction mixture. The green suspension was stirred at room temperature for 1 d until there was no EE<sub>2</sub> detected by TLC. The solvent was evaporated under vacuum, and the residue was dissolved with 6 mL of aqueous NH<sub>4</sub>OH (30%). The deep blue solution was precipitated into cold acetone (50 mL). This washing process was repeated three times for removal of residual copper. After addition of H<sub>2</sub>O and centrifugation at 4 °C three times (4000 rpm, 20 min), a fine beige powder was obtained (150 mg, 58%). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>, 323 K)  $\delta_{\text{ppm}}$  8.61 (br s, 1H), 7.78 (s, 1H), 6.88 (d, *J* = 8.1 Hz, 1H), 6.46 (s, 1H), 5.61 (s, 3H), 5.12 (s, 1H), 4.66 (s, 2H), 4.60 (s, 2H), 4.14 (m, 1 H), 3.74 (t, *J* = 8.7 Hz, 2H), 3.27–3.15 (m, overlaps with DOH), 2.653 (s, 3H), 2.20–1.26 (18H), 0.903 (s, 3H), 0.69 (m, 1H). <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>, 300 K)  $\delta_{\text{ppm}}$  154.8, 153.6, 137.1, 130.4, 125.9, 124.4, 114.9, 112.6, 101.5, 82.0, 81.0, 72.4, 71.8, 69.5, 49.5, 47.6, 46.8, 43.1, 37.1, 32.6, 30.7, 29.2, 27.2, 26.1, 23.4, 14.3. MALDI-TOF: *m/z* calcd for C<sub>182</sub>H<sub>231</sub>N<sub>21</sub>O<sub>42</sub> 3383.7, found [M+H<sub>2</sub>O]<sup>+</sup> 3402.90. HRMS: *m/z* calcd for C<sub>182</sub>H<sub>233</sub>N<sub>21</sub>O<sub>42</sub> [MH<sub>2</sub>]<sup>2+</sup> 1692.3366; found 1692.3302.

**4.1.25. Mono-6-(17-estradiol-17-(1*H*-1,2,3-triazol-4-yl))-6-deoxy- $\beta$ -cyclodextrin (24)**

—Mono-6-azido- $\beta$ -cyclodextrin **2** (100 mg, 0.086 mmol) and 17 $\beta$ -ethynylestradiol (25.5 mg, 0.086 mmol) were dissolved in 50% of MeOH and DMF (2.15 mL). CuI (82.0 mg, 0.43 mmol) was added, and the reaction mixture formed brown suspension. DIEA (75.0  $\mu$ L) was added via syringe into the reaction mixture. The green suspension was stirred at

room temperature for one day until there was no EE<sub>2</sub> detected on TLC. The solvent was evaporated under vacuum, and the residue was dissolved with 6 mL of aqueous NH<sub>4</sub>OH (30%). The deep blue colored solution was precipitated into cold Ac<sub>2</sub>O (50 mL). This washing process was repeated three times for the complete removal of copper ions. After centrifugation at 4 °C (4000 rpm, 20 min), fine beige powder was obtained, 1EqCD (80 mg, 64% yield). MALDI-TOF: calcd for C<sub>62</sub>H<sub>93</sub>N<sub>3</sub>O<sub>36</sub> 1456.42, found [M+Na]<sup>+</sup> 1479.20.

**4.1.26. 17 $\alpha$ -(17-1,2,3-Triazolmethyl)ethynyl-estra-1,3,5 (10)-triene-3,17 $\beta$ -diol (25)**—This procedure was modified from the literature.<sup>31</sup> Dimethylsulfate (90 mL, 0.676 mmol) was added to the solution of 17 $\beta$ -ethynylestradiol (100 mg, 0.338 mmol) and sodium azide (46 mg, 0.676 mmol) in *t*-BuOH/H<sub>2</sub>O (0.61/0.2 mL) at 0 °C. CuSO<sub>4</sub> (4.22 mg, 5 mol %) in 200  $\mu$ L of water and sodium ascorbate (6.7 mg, 10 mol%) in 200  $\mu$ L of water were added stepwisely. This reaction suspension was gradually warmed up to room temperature and stirred for 1 d. It was worked up by addition of water and extracted with EtOAc. The combined organic layers were washed with brine, and the solvent was removed. The resulting mixture was evaporated under pressure, and the residue was purified by silica gel column chromatography (EtOAc/Hx, 1:5). After drying under vacuum, white powder was obtained (71.7 mg, 60% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  8.94 (s, 1H), 7.79 (s, 1H), 6.95 (d, *J* = 8 Hz, 1H), 6.46 (d, *J* = 8.4 Hz, 1H), 6.40 (s, 1H), 5.04 (s, 1H), 4.01 (s, 3H), 2.68 (m, 2H), 2.50 (m, 1H), 2.10 (d, *J* = 12 Hz, 1H), 1.88–1.70 (m, 5H), 1.42–1.26 (m, 6H), 0.95 (s, 3H), 0.65 (t, *J* = 12 Hz, 11.6 Hz, 1H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  154.8, 154.2, 137.1, 130.4, 126.0, 123.5, 114.8, 112.6, 81.1, 47.5, 46.6, 43.1, 37.1, 36.0, 32.6, 29.2, 27.2, 26.1, 23.5, 14.3.

**4.1.27. Per-6-(4-hydroxymethyl-1,2,3-triazolmethyl)-6-deoxy- $\beta$ -cyclodextrin (26)**—Azide **2** (50 mg, 0.038 mmol) and CuI (36.0 mg, 0.19 mmol) were dissolved in 50% of MeOH and DMF (1.0 mL) to which propagyl alcohol (15.8  $\mu$ L, 0.27 mmol) and DIEA (33.0  $\mu$ L, 0.19 mmol) were added by syringe, and the mixture then stirred at room temperature for 1 d. The resulting green suspension was clarified using 2.0 mL of aqueous 30%NH<sub>4</sub>OH, and Chelex 100 sodium form was added to the deep blue solution to complex copper ions. The solution was allowed to stir vigorously overnight, and the yellow supernatant obtained was concentrated to give a beige powder (11 mg, 15% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  7.80 (s, 1H), 6.0 (br s, 4H), 5.1 (m, 6H), 4.6–4.3 (m, 11H), 4.0 (m, 3H), 3.7 (t, *J* = 8.8, 8.4 Hz, 4H), 2.9 (br s, 11H), 2.1 (s, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta_{\text{ppm}}$  147.6, 124.1, 101.5, 82.5, 72.5, 71.8, 69.7, 54.6, 49.3, MALDI-TOF: *m/z* calcd for C<sub>63</sub>H<sub>91</sub>N<sub>21</sub>O<sub>35</sub> 1701.6, found [M+Mg]<sup>+</sup> 1726.0.

**4.1.28. General rhodamine conjugation procedure**—Standard synthetic procedures were used. The appropriate qCD derivative (50 mg), the appropriate rhodamine coupling reagent (1 equiv of NHS–rhodamine for **23R**, **26R** and rhodamine-19-perchlorate for **23R'**, **26R'**) and DMAP (1 equiv) were dissolved in DMF (1 mL) under N<sub>2</sub>. The reaction mixture was cooled to 0 °C and EDC–HCl salt (1 equiv) was quickly added and the solution allowed to warm, before stirring overnight at room temperature in the dark. The resulting dark red suspension was precipitated in acetone (15 mL) giving a pink powder after filtration, which was washed twice with acetone followed by centrifugation at 4 °C for 1 h (10,000 rpm). The collected solids were lyophilized overnight to give a pink solid and kept at –20 °C in the dark.

**4.1.29. Rhodamine coupled 7EqCD (23R)**—Yield: 35 mg, 61%. MALDI-TOF: calcd for C<sub>210</sub>H<sub>259</sub>N<sub>23</sub>O<sub>46</sub> 3841.43, found [M+Na]<sup>+</sup> 3864.41.

**4.1.30. Rhodamine coupled 7EqCD (23R')**—Yield: 36 mg, 63%. MALDI-TOF: calcd  $C_{209}H_{259}N_{23}O_{43}$  3781.42, found  $[M+Na]^+$  3804.41.

**4.1.31. Rhodamine coupled per-6-(4-hydroxymethyl-1,2,3-triazolmethyl)-6-deoxy- $\beta$ -cyclodextrin (26R)**—Yield: 30 mg, 47%. MALDI-TOF: calcd for  $C_{91}H_{119}N_{23}O_{38}$  2143.05, found  $[M+K]^+$  2181.75.

**4.1.32. Rhodamine coupled per-6-(4-hydroxymethyl-1,2,3-triazolmethyl)-6-deoxy- $\beta$ -cyclodextrin (26R')**—Yield: 30 mg, 49%. MALDI-TOF: *m/z* calcd for  $C_{90}H_{119}N_{23}O_{36}$  2099.04, found  $[M+K]^+$  2138.15.

## 4.2. Bioassay procedures

**4.2.1. Cell culture**—The Ishikawa cell line was provided by Dr. R. B. Hochberg (Yale University, New Haven, CT) and the cells were grown in Dulbecco's Modified Eagle medium (DMEM/F12) containing 1% sodium pyruvate, 1% non-essential amino acids (NEAA), 1% glutamax-1, 0.05% insulin, and 10% heat-inactivated FBS. All cultures were maintained at 37 °C with 5% CO<sub>2</sub>:95% air. For phenol red-free complete medium, estrogen-stripped FBS was used. Human breast cancer cell line T47D:A18 is a hormone-responsive clone: the stable transfectant cell lines T47D:A18/neo and T47D:A18/PKC $\alpha$ <sup>58</sup> were maintained in RPMI 1640 (phenol red) supplemented with 10% fetal bovine serum-containing G418 (500  $\mu$ g/mL). Before transient transfection, T47D:A18/neo and T47D:A18/PKC $\alpha$  cells were maintained for three days in phenol red-free, E<sub>2</sub>-depleted RPMI 1640 supplemented with G418. After stripping, cells were transiently transfected by electroporation as described previously.<sup>58</sup>

**4.2.2. Measurement of ERE activation**—T47D cells were transiently transfected by electroporation as described previously.<sup>58</sup> Briefly,  $8 \times 10^6$  cells were harvested and resuspended in 0.5 mL serum-free, phenol red-free RPMI 1640. ERE-tk-Luc plasmid containing the luciferase reporter gene controlled by the ERE (5  $\mu$ g) and  $\beta$ -galactosidase ( $\beta$ -gal; 1  $\mu$ g) expressing plasmid pCMV $\beta$  were added to the cell suspension and incubated and processed. On the following day, medium containing E<sub>2</sub> ( $10^{-9}$  mol/L), 7EqCD or qE<sub>2</sub> ( $10^{-9}$ – $10^{-7}$  mol/L), or vehicle (ethanol) was added. Luciferase activities were measured using Luciferase Reporter Gene Assay System from Applied Biosystems.  $\beta$ -gal signals were measured with Galacto-Light Plus assay systems (Applied Biosystems). After 18 h incubation, cells were washed with ice-cold PBS and lysed in the lysis buffer provided. The cell lysates were cleared by centrifugation and luciferase activity and  $\beta$ -gal signals were determined after adding corresponding substrates and read by a Monolight 2010 luminometer (Analytical Luminescence Laboratory). The results were normalized relative to pRL-TK activity, to account for transfection efficiency, by dividing the sum of the luciferase activity by the sum of the  $\beta$ -gal signals.

**4.2.3. Measurement of induction of alkaline phosphatase**—The procedure of Liu et al. was used as described previously.<sup>59</sup> Test samples (10  $\mu$ L at varying concentrations in DMSO) were added to determine EC<sub>50</sub> values, and the cells in a total volume of 200  $\mu$ L media/well were incubated at 37 °C for four days. For the determination of antiestrogenic activity,  $2 \times 10^{-8}$  M estradiol was added to the media. The enzyme activity was measured by reading the release of *p*-nitrophenyl phosphate at 405 nm every 15 s with a 10 s shake between readings for 16 readings using a Power Wave 200 microplate scanning spectrophotometer (Bio-Tek Instruments, Winooski, VT). For estrogenic determination, the percent induction as compared with the estradiol control was calculated. For antiestrogenic determination, the percent induction was calculated compared to the background induction. The sulforhodamine (SRB) assay was used as a measure of cytotoxicity in parallel with



alkaline phosphatase (ALP) induction assays using ~1000 cells per well in a 96-well plate ( $5 \times 10^6$  cells/mL). Cells were treated with the same samples used in the ALP induction assay. The plates were read using the endpoint mode at 515 nm. Calculation of the percent cytotoxicity is essential to rule out cell death as a causal factor in reduced induction and apparent antiestrogenic activity.

**4.2.4. ER $\alpha$  competitive binding assay**—Competitive displacement of [ $^3\text{H}$ ] estradiol ( $\text{E}_2$ ) radioligand (Amersham Biosciences, Piscataway, NJ) from full-length recombinant human (h) ER $\alpha$  (PanVera/Invitrogen, Carlsbad, CA) was assayed as described.<sup>59</sup> Briefly, the reaction mixture consisted of 5  $\mu\text{L}$  of compound in DMSO, 5  $\mu\text{L}$  of hER $\alpha$  (0.5 pmol) in ER binding buffer [consisting of 10 mM Tris-HCl (pH 7.5), 10% glycerol, 2 mM dithiothreitol, 1 mg/mL BSA], 5  $\mu\text{L}$  of 'Hot Mix' [400 nM, prepared fresh using 95 Ci/mmol [ $^3\text{H}$ ] estradiol, diluted in 1:1 ethanol/ER binding buffer; obtained from NEN Life Science Products (Boston, MA)] and 85  $\mu\text{L}$  of ER binding buffer. The incubation was carried out with test compounds at room temperature for 2 h. Radioactivity was counted using a Beckman LS 5801 liquid scintillation counter (Schamburg, IL). The  $\text{IC}_{50}$  was calculated from a concentration–response data obtained in triplicate experiments.

**4.2.5. Fluorescence confocal microscopy**—MCF-7 cells were grown ( $10^6$  cells/mL) on each of eight wells on a sterile Nunc<sup>TM</sup> chambered cover glass and incubated for 48 h at 37 °C with 5%  $\text{CO}_2$  in phenol-free medium supplemented with 10% stripped fetal bovine serum medium. Cells were incubated with rhodamine conjugates (100 nM) for 24 h at 37 °C, 5%  $\text{CO}_2$ . Cells were rinsed twice with PBS to remove the unincorporated dye and 0.2  $\mu\text{g}/\text{mL}$  Hoechst stain was added to the cells to detect nuclear staining. Imaging was performed with a Zeiss LSM 510 laser-scanning confocal microscope with the detector gain adjusted to eliminate the background autofluorescence. The fluorescence signal from rhodamine conjugate was monitored with a 544 nm argon/krypton laser and a 576 nm band pass filter. Hoechst nuclear staining signal was monitored with a 345 nm UV laser and 420 nm band pass filter. A  $\times 63$  (1.2 numerical aperture) water immersion objective was used for all experiments. Images were analyzed using the analysis tool provided in the Zeiss biophysical software package.

## Acknowledgments

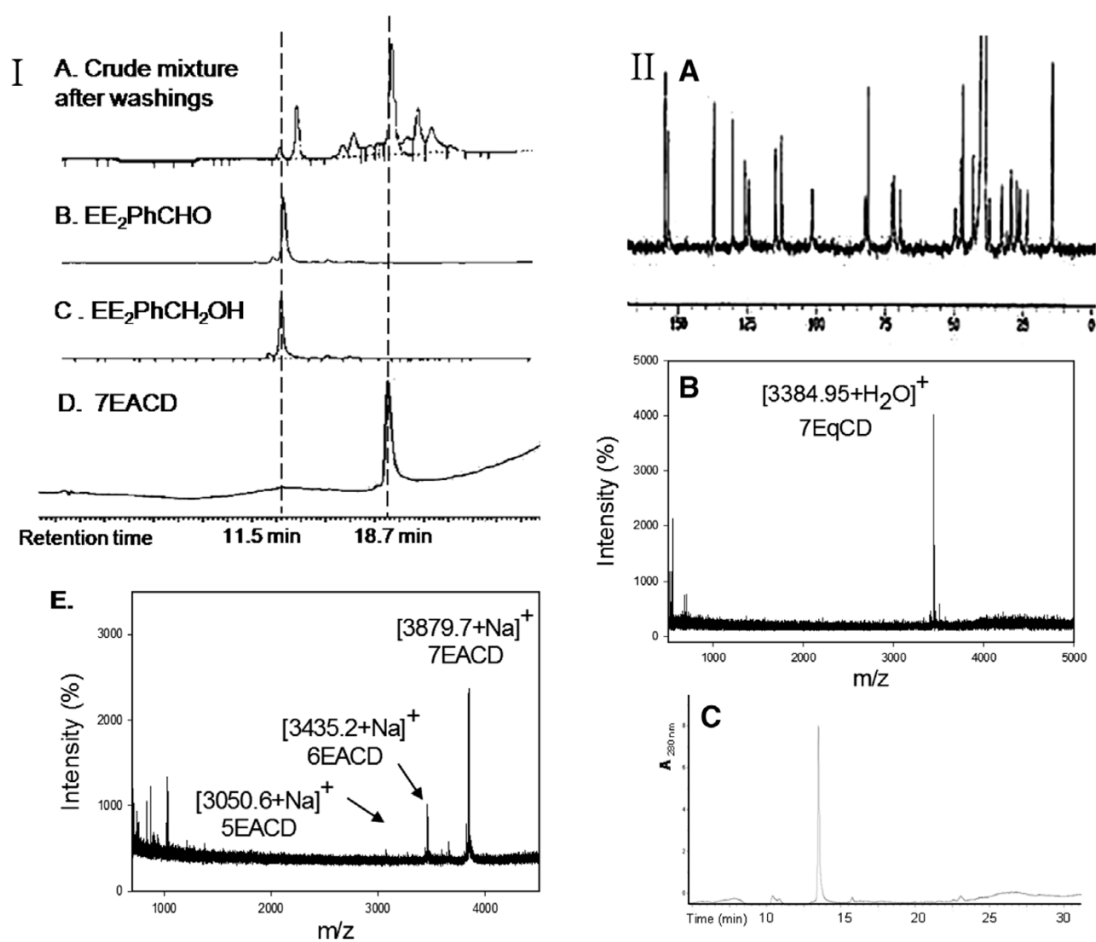
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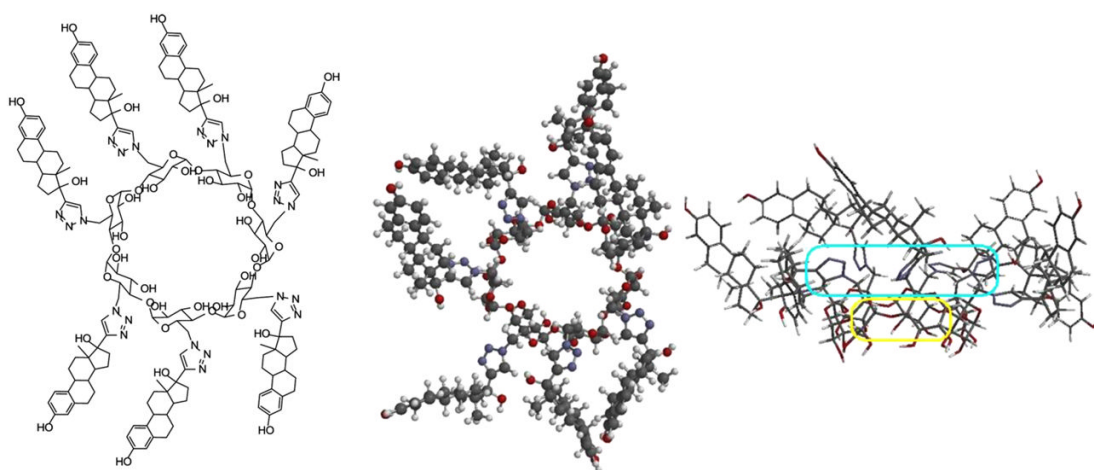
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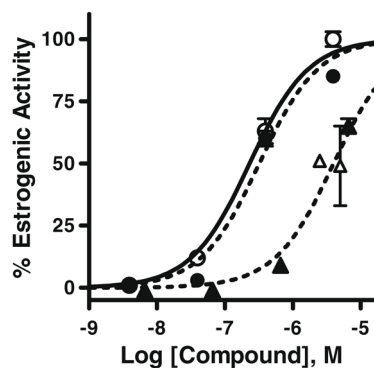
**Figure 1.**

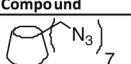
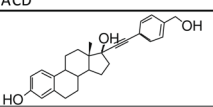
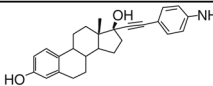
(I) (A–D) HPLC–UV chromatograms of crude and purified 7EACD product compared to synthetic monomer standards; (E) MALDI–TOF mass spectrum of purified 7EACD. (II) (A) <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) spectrum of 7EqCD ( $\delta$  ppm) 154.8, 153.6, 137.1, 130.4, 125.9, 124.4, 114.9, 112.6, 101.5, 82.0, 81.0, 72.4, 71.8, 69.5, 49.5, 47.6, 46.8, 43.1, 37.1, 32.6, 30.7, 29.2, 27.2, 26.1, 23.4, 14.3); (B) MALDI–TOF spectrum of 7EqCD; (C) HPLC–UV chromatogram of 7EqCD, detection at 280 nm,  $m/z = 1261.7$  MHz<sup>3+</sup>).



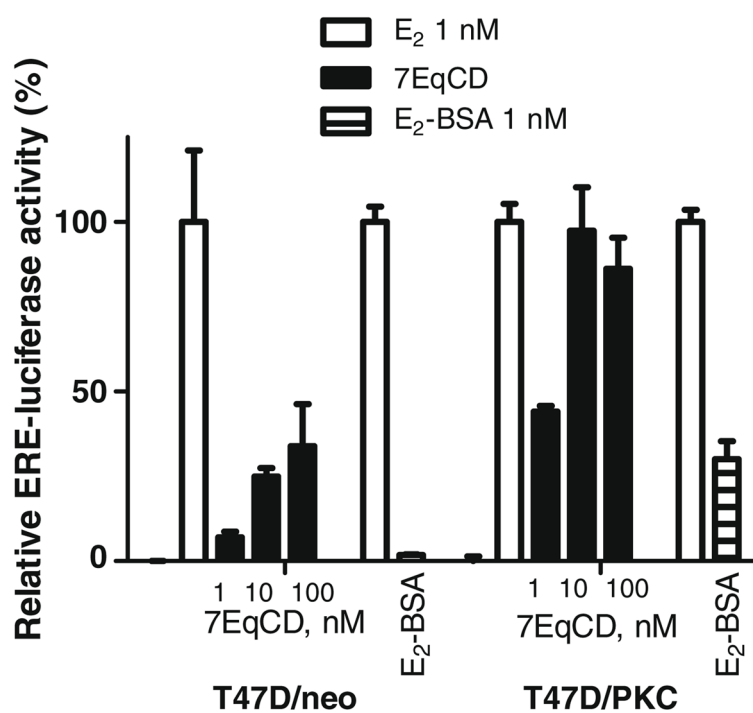
**Figure 2.** Structure of 7EqCD and one representative conformer optimized at the AM1 level in Spartan 04 (Wavefunction Inc., CA) showing top and side views with small cavity (yellow), annulus of triazole groups (cyan), and large flexible corona of pendant estradiol groups visible.



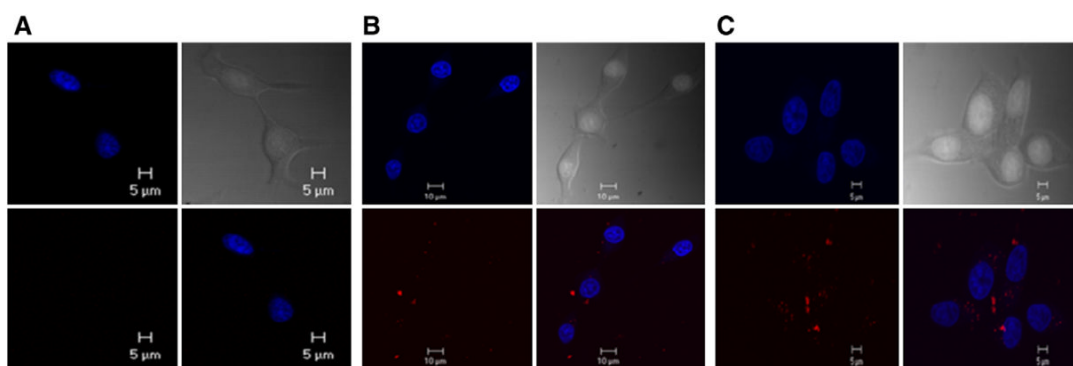


	Compound	EC <sub>50</sub>
2		na <sup>a</sup>
3	ACD	na
5		27 nM
6	7EACD	300 nM
12		10 nM
20	1EACD	4.0 μM
23	7EqCD	200 nM
24	1EqCD	4.0 μM
25	qE <sub>2</sub>	27 nM
a. Not active (na)		

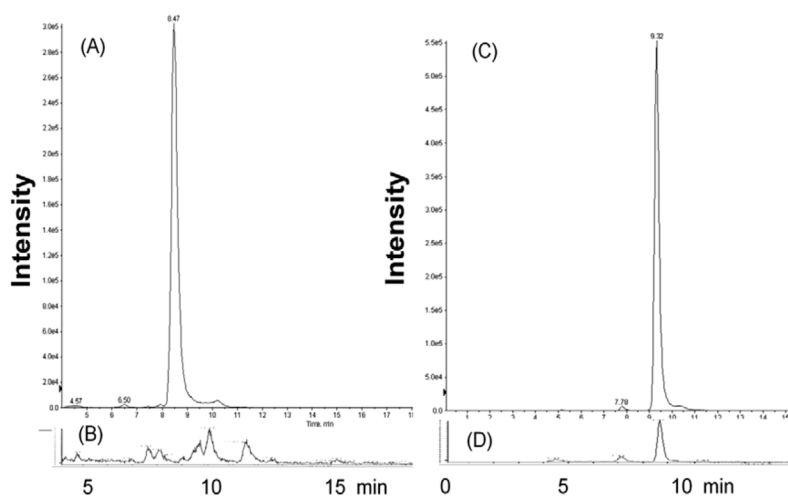
**Figure 3.** Estrogenic activity determined measuring alkaline phosphatase activity induced in ER(+) Ishikawa cells. Left: concentration–response for cells treated with 1EACD (closed triangles), 1EqCD (open triangles), 7EACD (closed circles), 7EqCD (open circles): 100% estrogenic activity corresponds to E<sub>2</sub> (1 nM). Data show mean and s.d. from triplicate experiments. Right: table of potency for all compounds for which concentration–response was measured in Ishikawa cells.



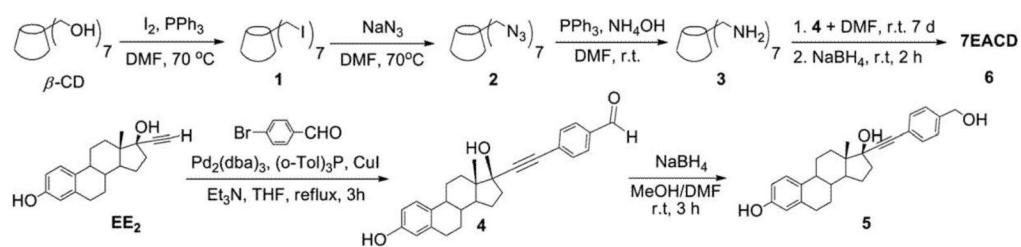
**Figure 4.** Activation of ERE and induction of luciferase activity in T47D/neo and T47D/PKC $\alpha$  cell lines by 7EqCD compared to E<sub>2</sub>-BSA. In all experiments, relative luciferase activity was corrected for transfection efficiency and normalized to vehicle (0%) and E<sub>2</sub> (1 nM; 100%). Data show mean and s.d. from triplicate measurements on two separate cell passages.



**Figure 5.** Localization of (A) **26R**, (B) **23R**, and (C) **23R'** in ER positive MCF-7 cells by confocal microscopy. Drugs (100 nM) were incubated with cells for 24 h. Clockwise from bottom-left: Ex/Em: 544/576 nm rhodamine-conjugates; Ex/Em: 345 nm/420 nm nuclear marker; differential interference contrast image; overlay of rhodamine and nuclear dyes showing cytoplasmic localization of rhodamine conjugates for **23R** and **23R'**. Data shown are representative of several images of different cells and planes.

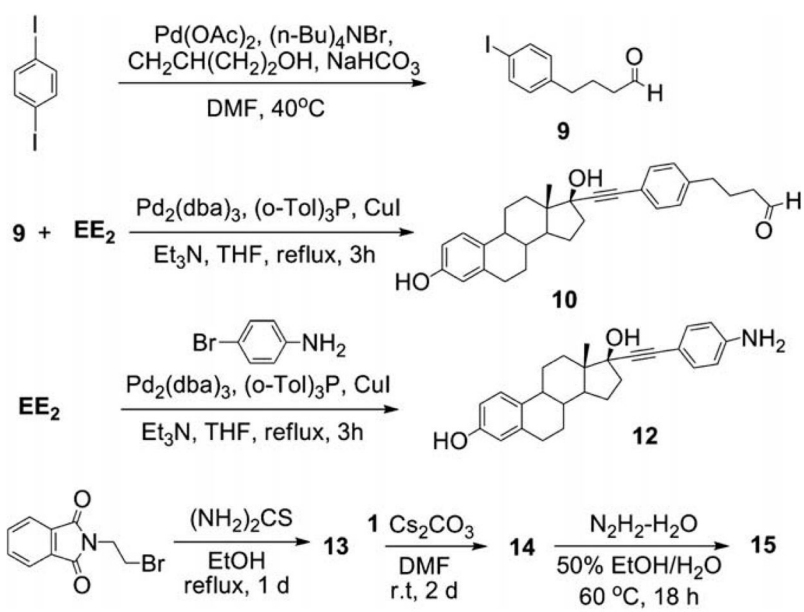


**Figure 6.** LC-MS/MS analysis of free estrogen in cell culture supernatant after four days incubation of E<sub>2</sub>-BSA or 7EqCD. Positive ion electrospray selected reaction monitoring (SRM) chromatograms of estrogens as dansyl derivatives: (A) EE<sub>2</sub> (70 nM) incubated in Ishikawa cell culture (peak intensity  $3.0 \times 10^5$ ); (B) 7EqCD (10 nM) incubated with Ishikawa cells (maximum y-axis intensity  $1.4 \times 10^3$ ) – EE<sub>2</sub> peak was not observed above detection limits (<50 pM); SRM conditions EE<sub>2</sub>-dansyl·[MH<sup>+</sup>]  $m/z$  530→171 collision energy 53 eV; (C) E<sub>2</sub> (70 nM) incubated in Ishikawa cell culture (peak intensity  $5.5 \times 10^5$ ); (D) E<sub>2</sub>-BSA (1 μg/mL) incubated with Ishikawa cells (peak intensity  $1.1 \times 10^4$ ) showing approximately 1 nM based upon standard curves; SRM conditions E<sub>2</sub>-dansyl [MH<sup>+</sup>]  $m/z$  506→171 collision energy 57 eV.



**Scheme 1.**  
Modifications of  $\beta$ -cyclodextrin.

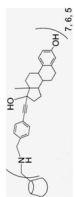




**Scheme 2.**  
Synthesis of **EE**<sub>2</sub> derivatives and synthons.

Reaction condition	Product
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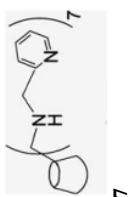
60–70 °C, 10 d

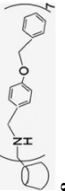
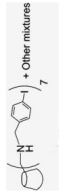
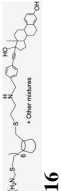
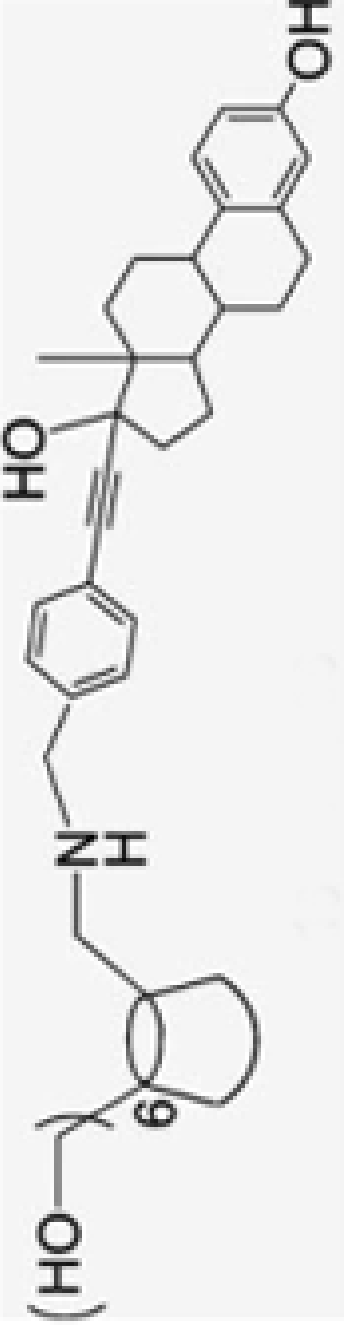


7EACD,6

Reaction condition	Product
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rt, 5 h

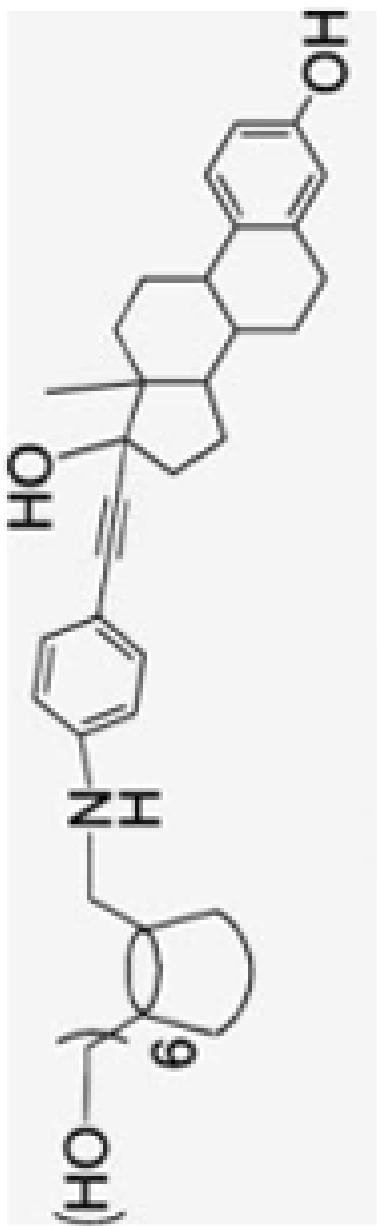


Reaction condition	Product
rt, 7 d	 <b>8</b>
60–70 °C, 18 h	 <b>11</b> + Other mixtures
60–70 °C, 18 h	S.M recovered
60–70 °C, 10 d	S.M recovered
60–70 °C, 10 d	 <b>16</b> + Other mixtures
60–70 °C, 3 d	 <b>6</b>

1EACD,20

Reaction condition	Product
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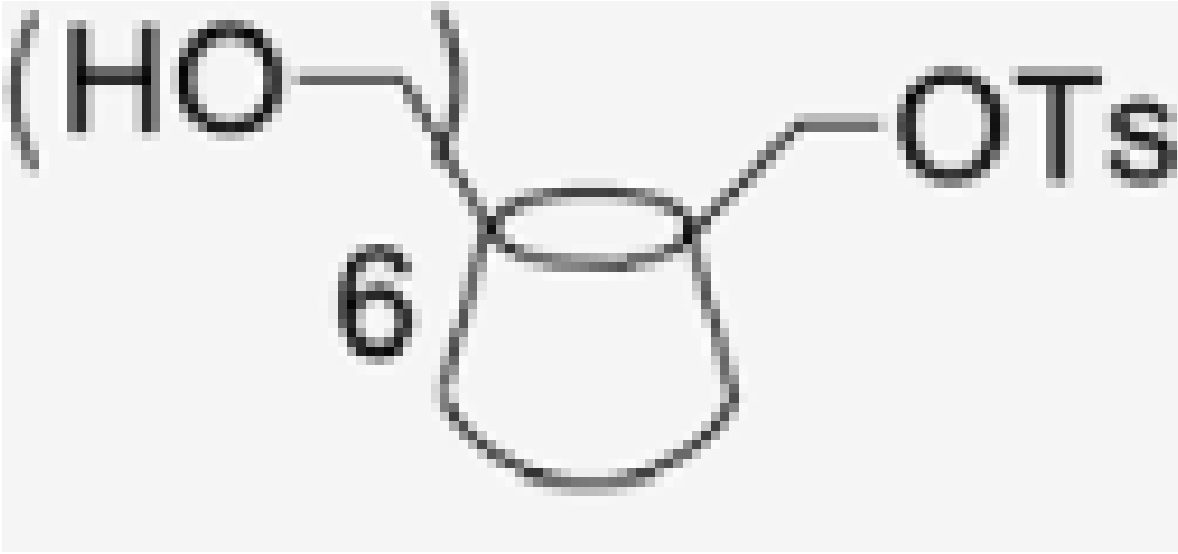
60–70 °C, 7 d



21

Table 2

Click chemistry routes to qCD derivatization

Entry	CD
10 <sup>g</sup>	
1	
11 <sup>h</sup>	
17	

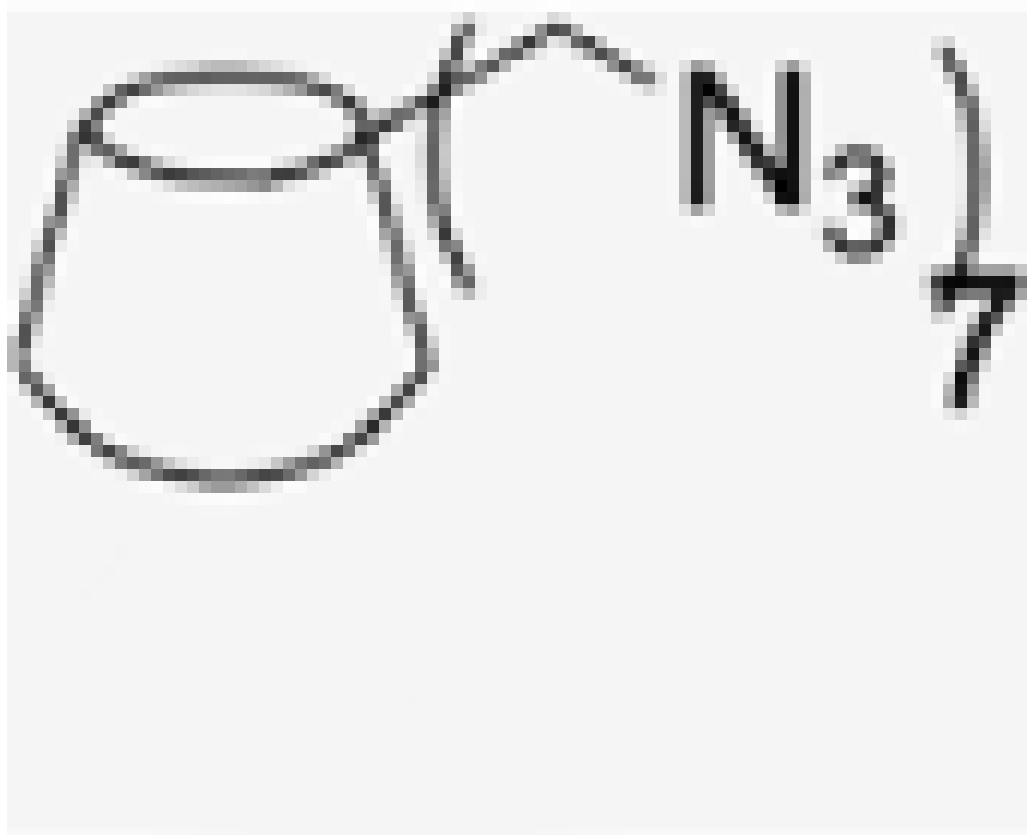


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Entry CD

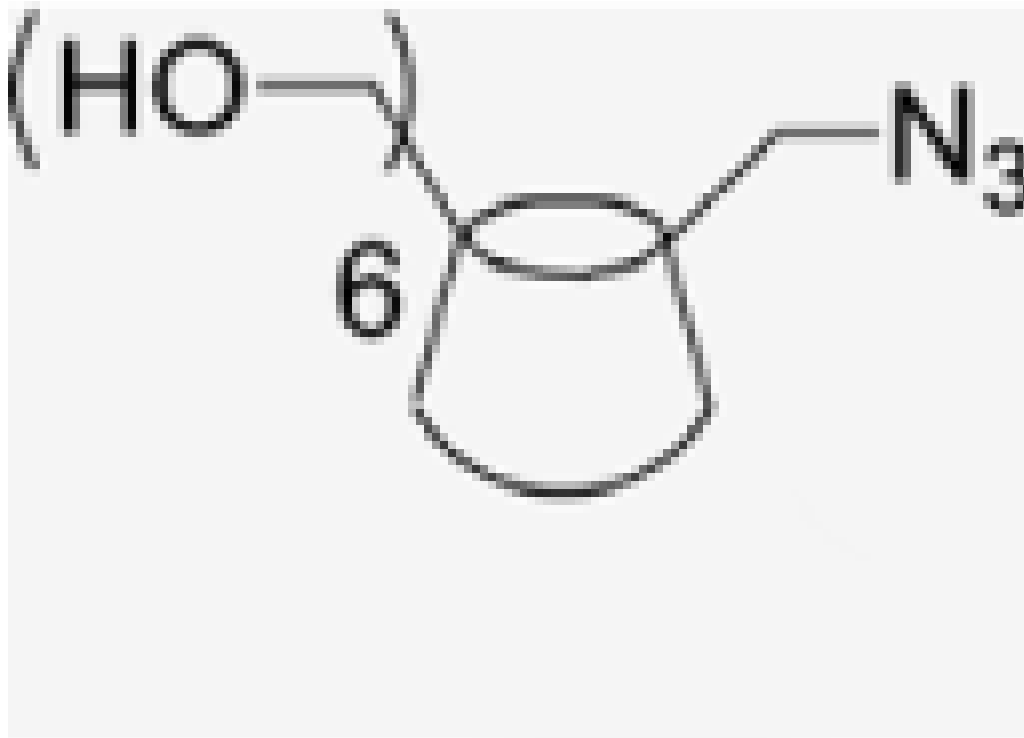
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12<sup>i</sup>



2

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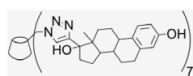
**Entry** CD13<sup>i</sup>

18

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Entry	CD
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14<sup>j</sup>15<sup>k</sup>**7EqCD,23**

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Entry	CD
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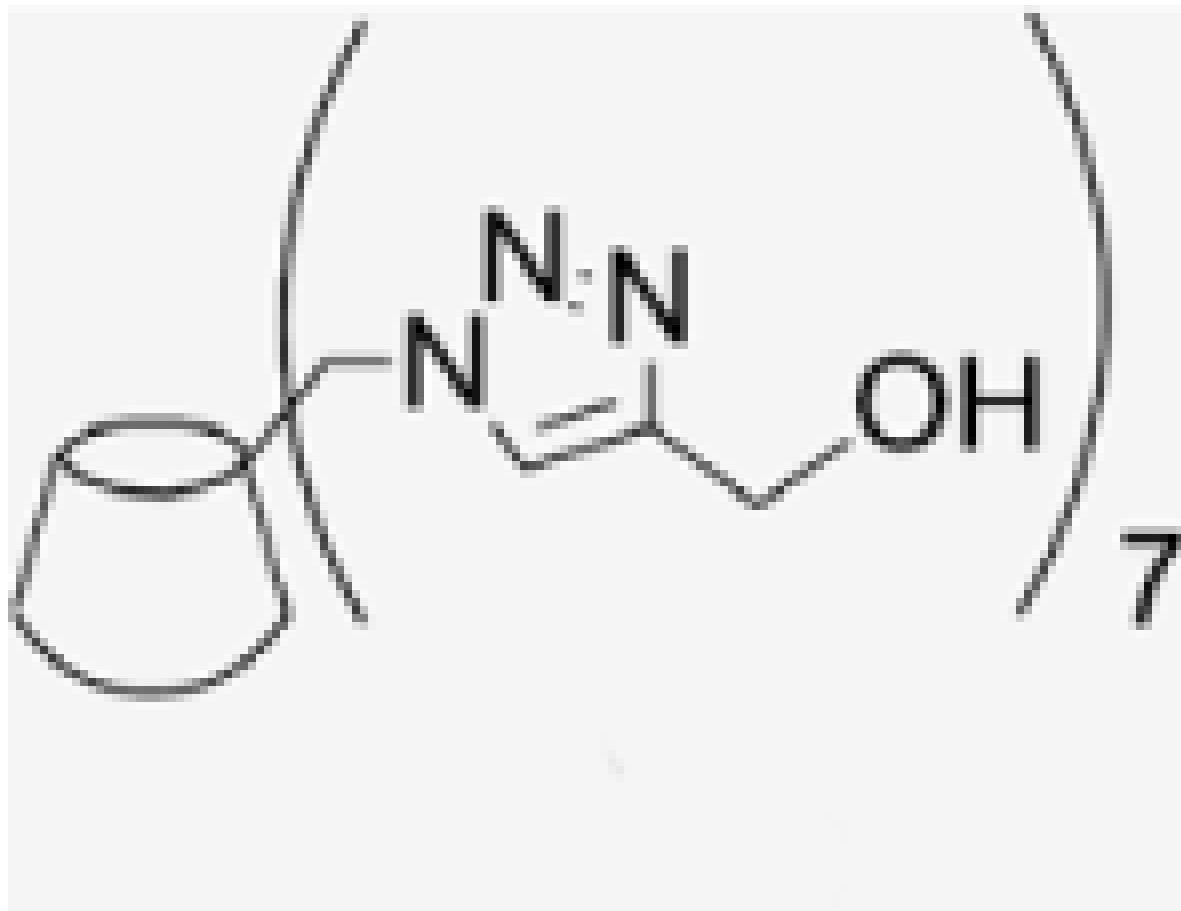
16<sup>i</sup>

2

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**Entry** CD

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17<sup>k</sup>

26

<sup>g</sup>DMF/H<sub>2</sub>O (3:1), 60 °C, 4 d.<sup>h</sup>DMF/H<sub>2</sub>O (3:1), 60 °C, 4 d.<sup>i</sup>DMF/MeOH, rt, 1 d.<sup>j</sup>*t*-BuOH/H<sub>2</sub>O (3:1), rt, 1 d.<sup>k</sup>DMAP/DMF, EDC-HCl/0 °C.