



Published in final edited form as:

Brain Res. 2010 August 6; 1347: 65–70. doi:10.1016/j.brainres.2010.06.001.

Clonidine, an α_2 receptor agonist, diminishes GABAergic neurotransmission to cardiac vagal neurons in the nucleus ambiguus

Kerry E. Philbin, Ryan J. Bateman, and David Mendelowitz

Department of Pharmacology and Physiology, The George Washington University, 2300 Eye Street NW, Washington, DC 20037

Abstract

In hypertension there is an autonomic imbalance in which sympathetic activity dominates over parasympathetic control. Parasympathetic activity to the heart originates from cardiac vagal neurons located in the nucleus ambiguus. Pre-sympathetic neurons that project to sympathetic neurons in the spinal cord are located in the ventral brainstem in close proximity to cardiac vagal neurons, and many of these pre-sympathetic neurons are catecholaminergic. In addition to their projection to the spinal cord, many of these pre-sympathetic neurons have axon collaterals that arborize into neighboring cardio-respiratory locations and likely release norepinephrine onto nearby neurons. Activation of α_2 adrenergic receptors in the central nervous system evokes a diverse range of physiological effects, including reducing blood pressure. This study tests whether clonidine, an α_2 adrenergic receptor agonist, alters excitatory glutamatergic, and/or inhibitory GABAergic or glycinergic synaptic neurotransmission to cardiac vagal neurons in the nucleus ambiguus. Cardiac vagal neurons were identified in a in-vitro brainstem slice preparation and synaptic events were recording using whole cell voltage clamp methodologies. Clonidine significantly inhibited GABAergic neurotransmission, but had no effect on glycinergic or glutamatergic pathways to cardiac vagal neurons. This diminished inhibitory GABAergic neurotransmission to cardiac vagal neurons would increase parasympathetic activity to the heart, decreasing heart rate and blood pressure. The results presented here provide a cellular substrate for the clinical use of clonidine as a treatment for hypertension as well as a role in alleviating post-traumatic stress disorder by evoking an increase in parasympathetic cardiac vagal activity, and a decrease in heart rate and blood pressure.

Keywords

ambiguus; clonidine; adrenergic; cardiac; vagal; parasympathetic

1. Introduction

Heart rate is determined by the activity of premotor cardioinhibitory vagal neurons (CVNs) located in the nucleus ambiguus. CVNs are the origin of parasympathetic innervation to the heart and dominate the control of heart rate (Mendelowitz, 1999; Mendelowitz and Kunze,

Correspondence to: David Mendelowitz, Department of Pharmacology and Physiology, George Washington University, 2300, Eye St NW, Washington, DC 20037, dmendel@gwu.edu, tel: 202-994-3466, FAX: 202-994-2870.

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1991). CVNs are intrinsically silent, and their activity is determined by synaptic activation from GABAergic, glycinergic and glutamatergic neurons, among others (Mendelowitz, 1996; Wang et al., 2001). In many cardiovascular diseases, including hypertension and heart failure, cardiac vagal activity is reduced and unresponsive (Vanoli et al., 1991).

In hypertension there is an autonomic imbalance in which sympathetic activity dominates over parasympathetic control. Pre-sympathetic neurons that project to sympathetic neurons in the spinal cord are located in the ventral brainstem in close proximity to CVNs, and many of these pre-sympathetic neurons are catecholaminergic (Loewy and Spyer, 1990). In addition to their projection to the spinal cord, many of these pre-sympathetic neurons have axon collaterals that arborize into neighboring cardio-respiratory locations. These axon collaterals likely release norepinephrine, as well as glutamate onto nearby neurons (Lipski et al., 1996).

Clonidine, an α_2 adrenergic agonist, is used clinically to treat hypertension. Activation of α_2 adrenergic receptors in the central nervous system evokes a diverse range of physiological effects, including reducing blood pressure and increasing sinus arrhythmia (Toader et al., 2009). Activation of α_2 adrenergic receptors with agonists, such as clonidine, inhibits adenylyl cyclase activity, resulting in a decrease in presynaptic Ca^{2+} concentrations and inhibition of norepinephrine release. Activation of α_2 adrenergic receptors in the ventro-lateral medulla elicits a decrease in brainstem sympathetic activity (Philipp et al., 2002).

Recent studies have shown that activation of α_2 receptors with clonidine significantly inhibits GABAergic inhibitory post-synaptic currents (IPSCs) in spinally projecting paraventricular nucleus neurons that can be prevented by the α_2 receptor antagonist yohimbine (Li et al., 2005). In contrast, clonidine has been shown to have little effect on glutamatergic excitatory post-synaptic currents (EPSCs) or miniature EPSCs (mEPSCs) in these spinally projecting paraventricular nucleus neurons (Li et al., 2005).

Despite its clinical significance, the effect of α_2 adrenergic receptor activation on synaptic inputs to parasympathetic cardiac vagal neurons has not yet been studied. The aim of the present study was to test whether activation of α_2 adrenergic receptors modulates excitatory glutamatergic, and/or inhibitory GABAergic or glycinergic synaptic neurotransmission to cardiac vagal neurons in the nucleus ambiguus.

2. RESULTS

Application of the α_2 adrenergic receptor agonist clonidine (10 μM) did not change holding current in CVNs, but evoked a significant decrease in the frequency of GABAergic neurotransmission to CVNs, see figure 1 (from 5.6 ± 0.9 Hz to 4.7 ± 0.9 Hz, $n=9$, $P<0.05$). In addition, clonidine (10 μM) significantly diminished the amplitude of GABAergic events in CVNs (from 61.5 ± 8.1 pA to 49.8 ± 4.0 pA, Fig. 1, $n=9$, $P<0.05$). To test if this inhibition of GABAergic neurotransmission evoked by clonidine was due to activating α_2 adrenergic receptors, the selective α_2 adrenergic receptor antagonist yohimbine (5 μM) was applied prior to clonidine application. Yohimbine by itself evoked a significant decrease in GABAergic IPSC frequency from 5.1 ± 1.0 Hz to 2.4 ± 0.8 Hz, see figure 2, top ($n=9$, $P<0.05$). Subsequent bath application of clonidine (10 μM) in the presence of the yohimbine bath did not significantly change the frequency or amplitude of GABAergic events, see figure 2, top ($n=9$, $P>0.05$). While yohimbine prevented the responses to clonidine, as expected, it was surprising that yohimbine by itself inhibited GABAergic IPSC frequency. Although the mechanism responsible for the inhibition of GABAergic neurotransmission by yohimbine is unknown, it is likely the result of yohimbine mediated 5-HT_{1A} receptor activation as yohimbine has also been shown to activate 5-HT_{1A} receptors. (Newman-Tancredi et al.,

1998). An additional series of experiments were conducted with the more selective α_2 adrenergic receptor antagonist atipamezole (1 μM), see figure 2, bottom. Atipamezole, as did yohimbine, prevented the inhibition of GABAergic neurotransmission to CVNs elicited by the alpha-2 receptor agonist clonidine. Atipamezole also elicited a much smaller, but statistically significant, inhibition of GABA ISPC frequency. This is consistent with atipamezole having a higher selectivity for alpha-2 receptors over 5-HT_{1A} receptors than yohimbine, but both antagonists have some potency for activation of 5HT_{1A} receptors (Newman-Tancredi et al., 1998).

To further study the mechanism by which clonidine and activation of alpha-2 receptors modulates GABAergic neurotransmission to CVNs, the voltage-gated sodium channel blocker tetrodotoxin (TTX, 1 μM) was included in the perfusate. Inclusion of TTX abolished action potential dependent synaptic neurotransmission and limited the remaining events to spontaneous miniature inhibitory post-synaptic currents (mIPSCs). There was no significant change in either GABAergic mIPSC frequency or amplitude upon application of clonidine in the presence of TTX (control: 2.6 ± 0.6 Hz, clonidine: 2.4 ± 0.5 Hz; See Fig. 3, n=14, $P > 0.05$).

To further characterize the role of clonidine on inhibitory neurotransmission to CVNs, clonidine (10 μM) was applied while glycinergic IPSCs were isolated. Clonidine did not elicit any significant change in glycinergic IPSC frequency or amplitude (Control: 3.4 ± 0.3 Hz, Clonidine: 3.5 ± 0.2 Hz; See Fig. 3, n=12, $P > 0.05$).

The effect of clonidine (10 μM) was also tested on excitatory neurotransmission to CVNs. Application of clonidine, while isolating for glutamatergic EPSCs, did not produce any significant change in the frequency or amplitude of EPSC events (Control: 2.8 ± 0.4 Hz, Clonidine: 2.5 ± 0.3 Hz; See Fig. 3, n=17, $P > 0.05$).

3. DISCUSSION

Despite the clinical significance of α_2 adrenergic receptors in brainstem cardiovascular control, little is known about the role of α_2 adrenergic receptors in modulating synaptic neurotransmission to parasympathetic CVNs that dominate the control of heart rate. Our results demonstrate that activation of α_2 adrenergic receptors with clonidine decreases GABAergic neurotransmission to CVNs, but clonidine has no significant effect on CVN holding current, or glycinergic or glutamatergic neurotransmission to CVNs. These results focused on CVNs are similar to the effects of clonidine in other neurons. In Purkinje cells located in the cerebellum as well parvocellular neurons located in hypothalamic paraventricular nuclei, the primary action of clonidine was decreased GABAergic neurotransmission (Han et al., 2002; Hirono and Obata, 2006).

Since clonidine changed GABAergic IPSCs but not GABAergic mIPSCs in CVNs, it is unlikely clonidine directly activates pre-synaptic receptors at GABAergic nerve terminals surrounding CVNs. Rather, the decrease in GABAergic IPSCs is most likely the result of activation of α_2 adrenergic receptors located precedent to the GABAergic synapse upon CVNs, perhaps at the GABAergic cell body or prior synapses in the GABAergic brainstem pathway. As there are only a few selective sites of origin of GABAergic neurons that project to CVNs, it is likely α_2 adrenergic receptors alters the activity of GABAergic neurons originating in either the immediate vicinity of the nucleus ambiguus and/or in the area of the nucleus tractus solitarius that contain GABAergic neurons that project to CVNs (Frank et al., 2009).

In order to further test if the decrease in GABAergic IPSCs resulted from activation of α_2 adrenergic receptors, the selective α_2 adrenergic receptor antagonist yohimbine was applied

prior to clonidine application. As expected, yohimbine prevented the clonidine mediated inhibition. However surprisingly there was a significant decrease in GABAergic IPSCs seen upon application of yohimbine by itself. Although the mechanism responsible for the inhibition by yohimbine is unknown, it is likely the result of yohimbine mediated 5-HT_{1A} receptor activation. Although generally considered an α_2 adrenergic receptor antagonist, yohimbine has also been shown to have significant 5-HT_{1A} receptor affinity (Newman-Tancredi et al., 1998). Prior work has shown activation of 5-HT_{1A} receptors cause significant decreases in GABAergic transmission to CVNs (Wang et al., 2007). Therefore, the significant decreases in GABAergic events upon application of yohimbine can most likely be attributed to 5-HT_{1A} receptor activation. The more selective α_2 adrenergic receptor antagonist atipamezole also prevented the inhibition of GABAergic neurotransmission to CVNs elicited by the alpha-2 receptor agonist clonidine. Atipamezole also elicited a much smaller, but statistically significant, inhibition of GABA ISPC frequency. This is consistent with atipamezole having a higher selectivity for alpha-2 receptors over 5-HT_{1A} receptors than yohimbine, but both antagonists have some potency for activation of 5HT_{1A} receptors (Newman-Tancredi et al., 1998).

The origin and location of adrenergic neurons and synaptic terminals that activate the α_2 adrenergic receptors and alter GABAergic neurotransmission to CVNs described in this study are unknown. However one intriguing potential source of adrenergic input is the pre-sympathetic neurons that are located in the ventral brainstem in close proximity to CVNs. Many of these pre-sympathetic neurons are catecholaminergic (Loewy and Spyer, 1990) and possess axon collaterals that arborize into other cardiorespiratory medullary regions, including within the nucleus ambiguus, in addition to their projection to sympathetic neurons in the spinal cord (Standish et al., 1995; Ter Horst et al., 1996; Ter Horst et al., 1993). Adrenergic (including α_2 and β_1 subtypes) receptors have been found within the nucleus ambiguus and shown to play a role in parasympathetic control of cardiovascular function (Gurtu et al., 1983; Paschalis et al., 2009).

Clonidine is clinically useful as an anti-hypertension treatment, and also has been shown to be beneficial in the treatment of post-traumatic stress disorder (Boehnlein and Kinzie, 2007). Our results show the likely site of action of clonidine in the brainstem is via diminishing GABAergic neurotransmission to CVNs. This diminished inhibitory neurotransmission to CVNs would increase parasympathetic activity to the heart, decreasing heart rate and blood pressure. Although the mechanism by which clonidine affects sympathetic control has been previously examined, this is the first evidence suggesting a mechanism by which activation of α_2 adrenergic receptors in the brainstem increase parasympathetic activity. The results presented here provide a cellular substrate for the clinical use of clonidine as a treatment for hypertension as well as a role in alleviating post-traumatic stress disorder by evoking an increase in parasympathetic cardiac vagal activity, and a decrease in heart rate and blood pressure.

4. Experimental Procedure

Individual CVNs located in the nucleus ambiguus were identified by the presence of the fluorescent tracer, and differential interference contrast optics along with infrared illumination and infrared-sensitive video detection cameras were used to gain better spatial resolution. The pipettes were filled with a solution consisting of KCl (150 mM), MgCl₂ (4 mM), EGTA (10 mM), Na-ATP (2 mM), HEPES (10mM) at a pH of 7.3 for inhibitory GABAergic and glycinergic events, and CsCl (130 mM), HEPES (10 mM), EGTA (10 mM), CaCl₂ (1 mM), and MgCl₂ (1 mM) at a pH of 7.3 for recording glutamatergic events. CVNs were studied by means of the whole-cell patch-clamp technique and were voltage clamped at a holding potential of -80 mV.

GABAergic IPSCs were isolated by adding strychnine (1 μ M), a glycinergic receptor antagonist, 6-cyano-7-nitroquinoxaline-2,3-dione ([CNQX] 50 μ M), a non-NMDA antagonist, and D-2-Amino-5 phosphonovalerate ([AP5] 50 μ M), a NMDA receptor antagonist, to the perfusate. Gabazine (25 μ M), a GABA_A receptor antagonist, CNQX (50 μ M) and AP5 (50 μ M) were included in the perfusate to isolate glycinergic IPSCs. Glutamatergic EPSCs were isolated by adding gabazine (25 μ M) and strychnine (1 μ M). In addition to gabazine and strychnine, (TTX) (1 μ M) was added to the perfusate in order to block action potential dependent events and isolate mEPSCs or mIPSCs. Drugs were applied after a 5–10 min control period and only one experiment was conducted in each slice. Clonidine (10 μ M), an α_2 agonist, was applied for a period of 5 min by inclusion in the perfusate. In two different sets of experiments yohimbine (5 μ M), and atipamezole (1 μ M), both α_2 adrenergic receptor antagonists, were added to the perfusate 5 min before the addition of clonidine (10 μ M). Concentrations of agonists and antagonists were chosen based on their pharmacological profiles and common dosage in the literature. Preliminary results were conducted with different concentrations of clonidine (ranging from 0.1 to 100 μ M) and 10 μ M clonidine provided the most consistent and non-saturating responses in our preliminary experiments. MiniAnalysis (Synaptosoft version 4.3.1) was used to analyze all experiments. The threshold for the GABAergic, glycinergic and glutamatergic events was a threshold of 5 times the root mean square of noise. Results are presented as mean \pm S.E. and statistically compared with a paired Student's t-test (for significance of difference *P<0.05), except for the experiments with yohimbine in which an ANOVA with repeated measures was performed.

In an initial surgery, 2–5 day old Sprague-Dawley rats were anesthetized with hypothermia to slow the heart and aid in recovery. A right thoracotomy was performed to expose the heart and the retrograde tracer, rhodamine (XRITC, Molecular Probes, 2% solution, 20–50 μ L), was then injected into the pericardial sac to retrogradely label CVNs.

On the day of the experiment, one to three days after the injection of the fluorescent tracer, the animal was anesthetized with isofluorane and sacrificed by cervical dislocation. The brain was rapidly removed and immersed in a cold HEPES buffer (4°C) with the following composition: NaCl (140 mM), KCl (5 mM), CaCl₂ (2 mM), glucose (5 mM), HEPES (10 mM). The buffer was continuously oxygenated with 100% O₂. Using a dissection microscope, the hindbrain was isolated. The brain was glued to a stage and placed in the slicing chamber of a vibratome filled with the same buffer solution. Coronal slices 500–600 μ m in thickness were cut. The slices were then mounted in a perfusion chamber and submerged in a perfusate with the following composition: NaCl (125 mM), KCl (3 mM), CaCl₂ (2 mM), NaHCO₃ (26 mM), glucose (5 mM), HEPES (5 mM) oxygenated with 95% O₂/5% CO₂ gas mixture. The osmolarity of all solutions was 285–290 mosM, and the pH was maintained between 7.35 and 7.4. All animal procedures were performed in compliance with the institutional guidelines at The George Washington University and were in accordance with the recommendations of the Panel on Euthanasia of the American Veterinary Medical Association and the NIH publication (85–23, revised 1996) 'Guide for the Care and Use of Laboratory Animals'. The minimal number of animals was used and attention was given to minimize any possible discomfort.

Acknowledgments

Supported by NIH grants HL49965, 59895 and 72006 to D.M.

Abbreviations

CVNs	cardioinhibitory vagal neurons
IPSCs	inhibitory post-synaptic currents
EPSCs	excitatory post-synaptic currents
mEPSCs	miniature EPSCs
mIPSCs	miniature IPSCs
TTX	tetrodotoxin

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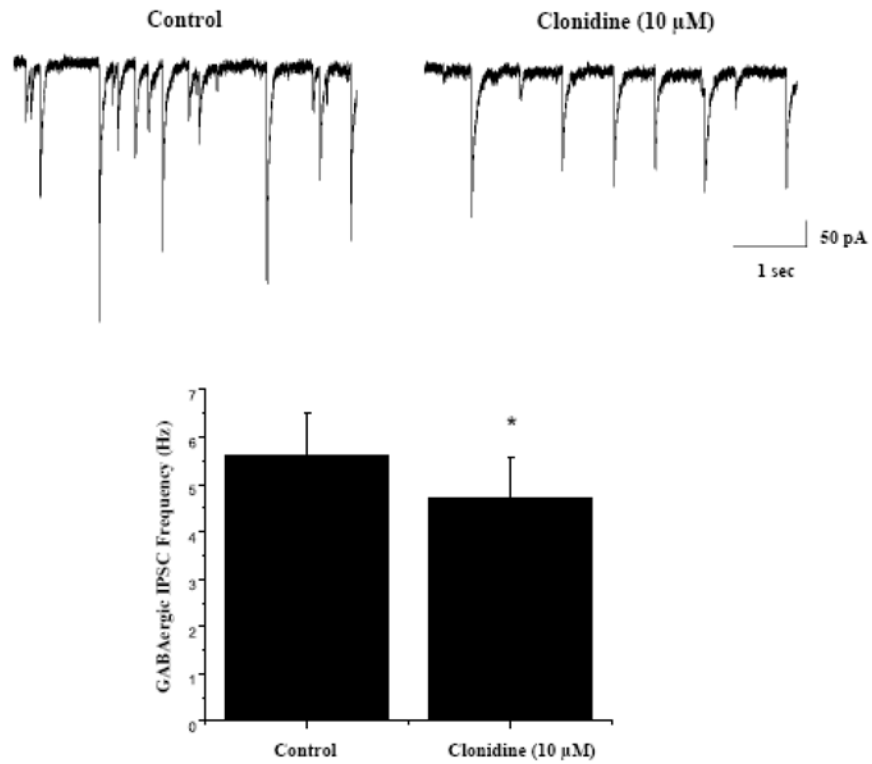


Figure 1.

As shown in representative traces, top, and the average results from 9 CVNs, bottom, inclusion of the α_2 adrenergic receptor agonist clonidine (10 μ M) in the perfusate evoked a significant decrease in the frequency of GABAergic neurotransmission to CVNs (from 5.6 ± 0.9 Hz to 4.7 ± 0.9 Hz, $p < 0.05$).

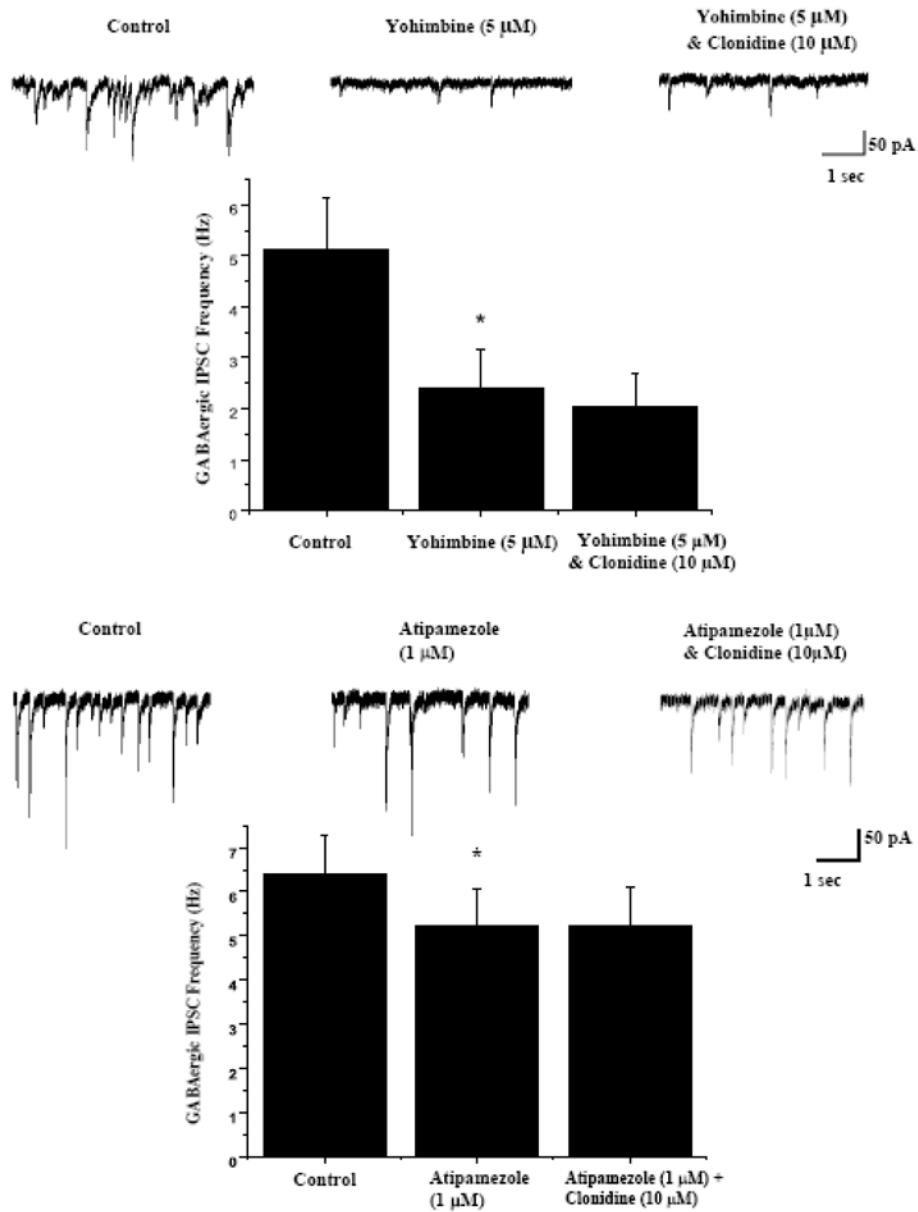


Figure 2.

The selective α_2 adrenergic receptor antagonist yohimbine (5 μ M) was applied prior to clonidine application to examine if there is any endogenous activation of α_2 adrenergic receptors and if the responses to clonidine could be blocked by the α_2 adrenergic receptor antagonist yohimbine. As shown in representative traces, top, and the average results from 9 CVNs, bottom, yohimbine prevented the inhibition by clonidine, but surprisingly, also evoked a significant decrease in GABAergic IPSC frequency (from 5.1 ± 1.0 Hz to 2.4 ± 0.8 Hz, $p < 0.05$) when applied by itself.

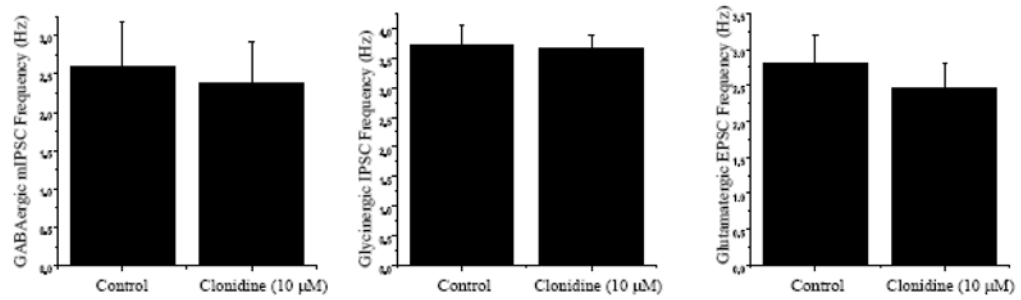


Figure 3.

To further study the mechanism by which clonidine modulates GABAergic neurotransmission to CVNs and isolate GABAergic mIPSCs TTX was included in the perfusate. As shown in the left panel, in the presence of TTX there was no significant change in GABAergic mIPSC frequency upon application of clonidine. As shown in the middle panel, clonidine had no significant effect on glycinergic IPSCs in CVNs, and as shown in the right panel, application of clonidine did not produce any significant change in the frequency of glutamatergic EPSC events.